One Week's Treatment With the Long-Acting Glucagon-Like Peptide 1 Derivative Liraglutide (NN2211) Markedly Improves 24-h Glycemia and α- and β-Cell Function and Reduces Endogenous Glucose Release in Patients with Type 2 Diabetes

Kristine B. Degn,[1] Claus B. Juhl,[1] Jeppe Sturis,[2] Grethe Jakobsen,[2] Birgitte Brock,[1] Visvanathan Chandramouli,[3] Joergen Rungby,[1] Bernard R. Landau,[3] and Ole Schmitz[1]

Glucagon-like peptide 1 (GLP-1) is potentially a very attractive agent for treating type 2 diabetes. We explored the effect of short-term (1 week) treatment with a GLP-1 derivative, liraglutide (NN2211), on 24-h dynamics in glycemia and circulating free fatty acids, islet cell hormone profiles, and gastric emptying during meals using acetaminophen. Furthermore, fasting endogenous glucose release and gluconeogenesis (3-^{3}H-glucose infusion and ^2H_2O ingestion, respectively) were determined, and aspects of pancreatic islet cell function were elucidated on the subsequent day using homeostasis model assessment and first- and second-phase insulin response during a hyperglycemic clamp (plasma glucose ~16 mmol/l), and, finally, on top of hyperglycemia, an arginine stimulation test was performed. For accomplishing this, 13 patients with type 2 diabetes were examined in a double-blind, placebo-controlled crossover design. Liraglutide (6 μg/kg) was administered subcutaneously once daily. Liraglutide significantly reduced the 24-h area under the curve for glucose (P = 0.01) and glucagon (P = 0.04), whereas the area under the curve for circulating free fatty acids was unaltered. Twenty-four-hour insulin secretion rates as assessed by deconvolution of serum C-peptide concentrations were unchanged, indicating a relative increase. Gastric emptying was not influenced at the dose of liraglutide used. Fasting endogenous glucose release was decreased (P = 0.04) as a result of a reduced glycogenolysis (P = 0.01), whereas gluconeogenesis was unaltered. First-phase insulin response and the insulin response to an arginine stimulation test with the presence of hyperglycemia were markedly increased (P < 0.001), whereas the proinsulin/insulin ratio fell (P = 0.001). The disposition index (peak insulin concentration after intravenous bolus of glucose multiplied by insulin sensitivity as assessed by homeostasis model assessment) almost doubled during liraglutide treatment (P < 0.01). Both during hyperglycemia per se and after arginine exposure, the glucagon responses were reduced during liraglutide administration (P < 0.01 and P = 0.01). Thus, 1 week's treatment with a single daily dose of the GLP-1 derivative liraglutide, operating through several different mechanisms including an ameliorated pancreatic islet cell function in individuals with type 2 diabetes, improves glycemic control throughout 24 h of daily living, i.e., prandial and nocturnal periods. This study further emphasizes GLP-1 and its derivatives as a promising novel concept for treatment of type 2 diabetes. *Diabetes* 53:1187–1194, 2004

Type 2 diabetes is characterized by insulin resistance and progressive islet cell dysfunction leading to insulin deficiency (1). Increased hepatic glucose release is considered to play a key role in fasting, as well as postprandial hyperglycemia, and increased gluconeogenesis seems to be essential in this scenario (2,3). In addition, elevated glucagon levels (4,5) and a reduced response of glucagon-like peptide 1 (GLP-1) to meals (6) are common features.

The importance of aggressive glucose-lowering therapy to prevent late diabetes complications in type 2 diabetes has been convincingly established (7,8). However, the U.K. Prospective Diabetes Study also demonstrated that the antidiabetic treatment used failed to maintain acceptable glycemic control in the vast majority of the patients, emphasizing the need for more effective antidiabetic agents.

GLP-1 is an incretin hormone secreted from the intestinal mucosa in response to meal ingestion (9). Insulin secretion is stimulated, and glucagon secretion is inhibited; both actions are glucose dependent. Also, GLP-1 has trophic effects on pancreatic β-cells and inhibits their apoptosis (10). Furthermore, it delays gastric emptying (9,11) and may even be a satiety factor. These observations support GLP-1 as a novel candidate for treatment of type 2 diabetes, and the beneficial effects of GLP-1 on

From the [1]Department of Endocrinology (M & C), University Hospital of Aarhus, and Department of Clinical Pharmacology, University of Aarhus, Aarhus, Denmark; [2]Novo Nordisk A/S, Bagsværd, Denmark; and [3]Case Western Reserve University School of Medicine, Cleveland, Ohio.

Address correspondence and reprint requests to Ole Schmitz, MD, Department of Medicine M (Endocrinology & Diabetes), University Hospital of Aarhus, AKH, Nørrebrogade 42-44 DK-8000 Aarhus C, Denmark. E-mail: ole.schmitz@iekf.au.dk.

Received for publication 14 October 2003 and accepted in revised form 23 January 2004.

AUC, area under the curve; DPP-IV, dipeptidylpeptidase IV; EGR, endogenous glucose release; ELISA, enzyme-linked immunosorbent assay; FFA, free fatty acid; GLP-1, glucagon-like peptide 1; GLY, glycogenolysis; GNG, gluconeogenesis; HMT, hexamethylenetetramine; HOMA, homeostasis model assessment; ISR, insulin secretion rate; OHA, oral hypoglycemic agent; t_{max}, time of occurrence for maximum drug concentration.

© 2004 by the American Diabetes Association.

MPI EXHIBIT 1081 PAGE 1

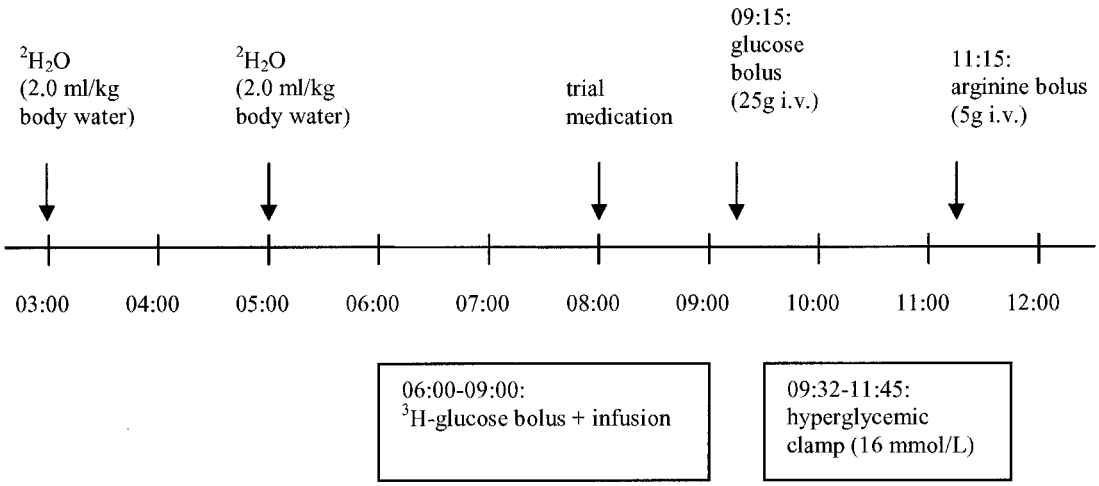


FIG. 1. Flow sheet illustrating the procedures on study day 9. See text for details.

glycemic control in individuals with type 2 diabetes have been demonstrated (12,13). However, native GLP-1 has a very short half-life because of its rapid degradation by the ubiquitous enzyme dipeptidylpeptidase IV (DPP-IV) (14), making GLP-1 per se unsuitable as a therapeutic drug. Derivatives that are resistant to DPP-IV are one way to overcome this failing. Liraglutide is an acylated GLP-1 derivative that binds to albumin. This limits its sensitivity to DPP-IV and delays absorption from its injection site. A prolonged pharmacokinetic profile in humans is attained, the half-life after subcutaneous administration being ~12 h (15,16). A single-dose study demonstrated significant glucose lowering in individuals with type 2 diabetes in both the fasting and postprandial states (16), and in a 12-week repeated-dose study, liraglutide lowered blood glucose to the same extent as a sulfonylurea compound (glimepiride) and the prevalence of hypoglycemia was very low (17).

The present study was undertaken to gain further insight into GLP-1 derivatives as novel antidiabetic drugs. We sought to do this by administering liraglutide to individuals with type 2 diabetes for 7 days and assessing the impact on 24-h glucose and hormone levels during daily life conditions, gastric emptying rate, endogenous glucose release (EGR), and various aspects of the pancreatic islet cell function.

RESEARCH DESIGN AND METHODS

The protocol was approved by the local ethics committee and performed in accordance with the Helsinki Declaration.

Patients. Thirteen patients (5 women and 8 men) with type 2 diabetes according to World Health Organization criteria were examined. Their age (mean ± SD) was 56.4 ± 9.2 years, BMI was 31.2 ± 3.6 kg/m^2, last measured HbA_{1c} before inclusion was 7.3 ± 0.4% (normal range <6.4%), and the duration of diabetes was 3.0 ± 2.6 years (range 5 months to 8 years). At study entry, six patients were treated with diet alone and seven were additionally treated with oral hypoglycemic agents (OHAs; sulfonylurea n = 3, metformin n = 3, sulfonylurea and metformin n = 1). Concomitant medications were angiotensin-converting enzyme inhibitors (n = 4), thiazide diuretics (n = 1), β-blockers (n = 1), calcium antagonists (n = 1), HMG-CoA reductase inhibitors (n = 2), and acetyl salicylic acid (n = 3). Three patients had retinopathia simplex, and three had nephropathy (microalbuminuria).

Experimental design. This crossover trial was double-blinded, placebo-controlled, and randomized. After inclusion, the patients discontinued their OHA for 2 weeks before beginning study medication. Liraglutide (6 μg/kg body wt) or placebo was injected subcutaneously into the abdomen once daily (at ~0745) for 9 days using a NovoPen (1.5 with Novofine 30-G, 0.3- to 8-mm needle) as the dispensing device. After 7 days of treatment, the patients arrived at the clinical research unit at 2200. In the next 2 days (days 8 and 9), the following experiments were carried out during continuous treatment.

On day 8 at 0730, a catheter was placed in an antecubital vein for blood sampling. Three standard meals were served at 0800, 1200, and 1800, to be finished within 20 min. Breakfast contained 2,660 kJ (protein 14 E%, carbohydrate 55 E%, and fat 31 E%), lunch contained 2,865 kJ (protein 16 E%, carbohydrate 50 E%, and fat 34 E%), and dinner contained 3,397 kJ (protein 28 E%, carbohydrate 53 E%, and fat 19 E%). With breakfast and dinner, 1 g of acetaminophen dissolved in 150 ml of water was given to assess gastric emptying rate (18). Serum acetaminophen was determined every 15–30 min during the following 4 h, and area under the curve (AUC) for acetaminophen ($AUC_{acetaminophen}$) and the time of occurrence for maximum drug concentration (t_{max}) were calculated. During day 8, blood was collected with changing intervals for determination of glucose, insulin, C-peptide, proinsulin (only fasting and after breakfast), glucagon, free fatty acids (FFAs), and liraglutide.

On day 9, the following procedures were performed (Fig. 1).

Gluconeogenesis. At 0300 and 0500, patients drank 2 ml of ^2H_2O/kg body water (99.9% H; Sigma Aldrich; body water was estimated to be 50% of total body weight in women and 60% in men). Water ingested ad libitum thereafter was enriched with 0.4% ^2H_2O to maintain isotopic steady state. At 0800, 0830, and 0900, blood was drawn for determination of gluconeogenesis (GNG).

EGR. EGR was estimated by use of an isotope dilution technique. At 0600, a bolus of 3-^{3}H-glucose (adjusted priming: 6 μCi × plasma glucose level in mmol/l) was given followed by a continuous infusion (0.3 μCi/min) until 0900. Blood samples were drawn before the bolus and every 15 min from 0800 to 0900 for determination of glucose ^{3}H specific activity. Steady state was achieved during this period in both situations, the average specific activity of glucose being 2,078, 2,180, and 2,160 cpm/mg at time points 0800, 0830, and 0900 during active treatment and 1,927, 1,822, and 1,906 cpm/mg at the same time points during placebo.

First-phase insulin secretion. First-phase insulin secretion was determined using an intravenous glucose bolus. At 0915, 25 g of glucose was administered intravenously over 2 min. Blood was then collected every 2–5 min until 0932 for determining glucose and insulin.

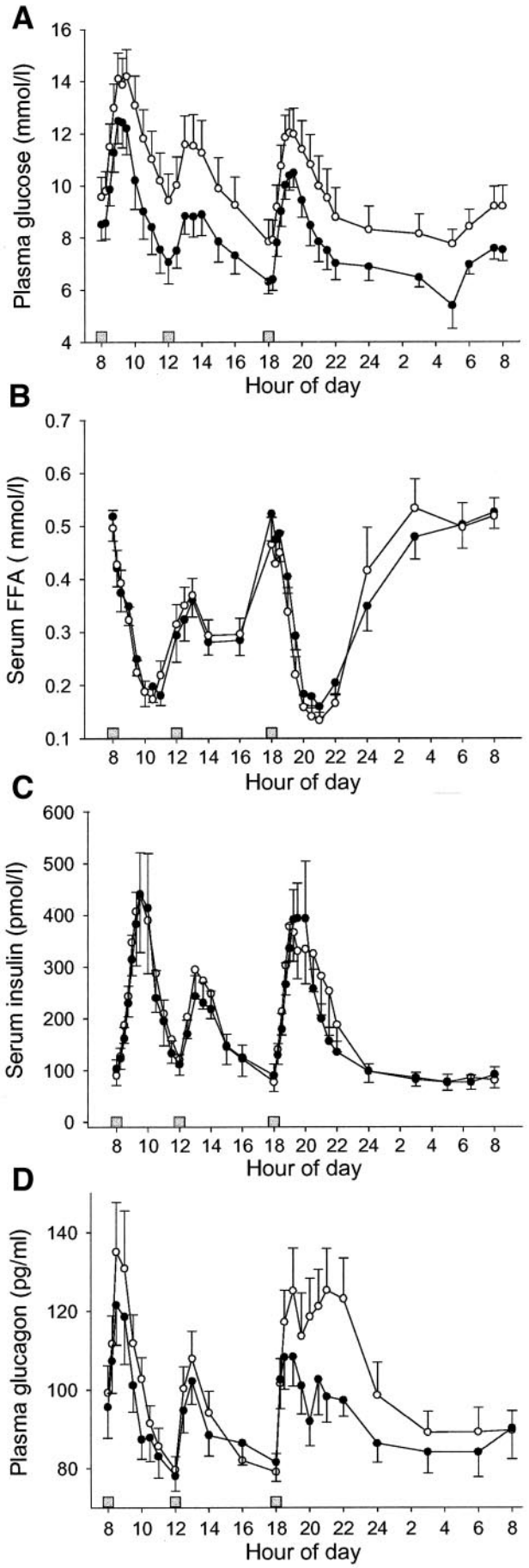
Hyperglycemic clamp. From 0932 to 1145, 20% glucose was infused to maintain plasma glucose at 16 mmol/l. After 75 min, when approximate steady state was attained, blood was drawn every 10 min (from 1045 to 1115) for determining plasma glucose and serum insulin, C-peptide, and proinsulin.

Arginine stimulation test. At 1115, a bolus of 5 g of arginine was given intravenously over 30 s followed by collection of blood every 5 min until 1145 for determining circulating glucose, insulin, C-peptide, glucagon, and proinsulin.

Safety. Adverse events, vital signs, hematology, and biochemistry were monitored throughout the study.

Assays. All biochemical analyses were performed in duplicate. Plasma glucose was measured immediately on a glucose analyzer (Beckman Instruments, Palo Alto, CA) using the glucose oxidase technique. All other serum and plasma samples were stored at −20°C (C-peptide at −80°C) until analyzed. Serum insulin was determined using a highly specific and sensitive two-site

MPI EXHIBIT 1081 PAGE 2



enzyme-linked immunosorbent assay (ELISA; DAKO Diagnostics, Cambridgeshire, U.K.), and serum C-peptide was measured by a two-site monoclonal-based ELISA with an intra-assay coefficient of variation of 5.1% (DAKO Diagnostics). Serum proinsulin was analyzed by specific immunoassay with no cross-reaction with insulin and C-peptide (DAKO Diagnostics). Serum acetaminophen was determined by high-performance liquid chromatography after extraction. Plasma glucagon was measured using a radioimmunoassay kit (Linco Research). Liraglutide concentrations were analyzed by ELISA using a monoclonal antibody against GLP-1/liraglutide as capture antibody and another monoclonal antibody specific for the NH_2-terminal part of GLP-1/liraglutide for detection. Before the latter analysis, samples were incubated at 37°C to remove endogenous GLP-1, liraglutide being stable toward this incubation. Determination of 3-^{3}H-glucose activity was as previously described (19). Measurement of deuterium enrichment at carbons 2 and 5 glucose was as described by Landau et al. (20). In brief, 15 ml of blood was diluted with 30 ml of demineralized water and deproteinized using 15 ml of 0.3 N $ZnSO_4$ and 15 ml of 0.3 N $Ba(OH)_2$. The samples were then centrifuged at 2,000 rpm for 15 min, and the pellet was diluted in 15 ml of demineralized water to wash out the remaining glucose. Glucose was isolated by high-performance liquid chromatography. For determination of deuterium enrichment on C5, glucose was first converted to xylose, and the carbon 5 with its hydrogens was cleaved by periodate oxidation to formaldehyde, which was condensed with ammonium hydroxide to form hexamethylenetetramine (HMT). The ^{2}H bound to C2 of glucose was isolated after conversion of glucose to ribitol-5-phosphate and arabitol-5-phosphate and treated to form HMT. Enrichments in the HMTs were measured using a Hewlett-Packard mass spectrometry system with standards of HMTs analyzed along with the unknown samples.

Calculations and statistics. Fasting data are presented as the mean values at 0730 and 0800 on days 8 and 9. Furthermore, fasting plasma glucose on day 9 is presented. AUCs of substrates and hormones were calculated by applying the trapezoidal rule and represent the total AUC. EGR was determined using Steele's equations for steady state, as modified by Finegood et al. (21). The fractional contribution of GNG to glucose production equals the ratio between deuterium bound to carbon 5 in glucose and that bound to carbon 2. The rate of GNG in the fasting state was obtained by multiplying the mean EGR from 0800 to 0900 by the mean fractional contribution of GNG during that same hour. The rate of glycogenolysis (GLY) was calculated by subtracting the rate of GNG from total EGR. Insulin secretion rates (ISRs) were estimated by mathematical analysis (deconvolution) of peripheral C-peptide concentrations using a two-compartment model, as described by Polonsky et al. (22) and the standard C-peptide kinetic parameters published by Van Cauter et al. (23). This model allows accurate estimation of ISR also under non–steady-state conditions. Another estimate of β-cell function and an estimate of insulin resistance were calculated by homeostasis model assessment (HOMA-B and R) (24), using glucose and insulin values after a 14-h long fast and 24 h after the last liraglutide injection. As a supplement to the latter, insulin sensitivity was calculated by dividing glucose infusion rate during steady state of the hyperglycemic clamp with mean serum insulin level. Finally, an analog to the disposition index was calculated by multiplying 1/HOMA-R by the peak insulin concentrations during the first-phase insulin secretion test (25).

Statistical analysis was performed using a mixed model with sequence of treatment, visit, and treatment as fixed factors and patient as a random factor. Differences were considered significant at $P < 0.05$. All results are given as liraglutide versus placebo.

RESULTS

Twenty-four-h profiles of substrates and hormones. During the 24-h profile, the average concentration of plasma glucose ($P = 0.01$) and postprandial plasma glucose concentrations after all three meals were decreased by ~20% ($P = 0.01$, $P = 0.02$, and $P = 0.02$, respectively) during liraglutide treatment. Fasting plasma glucose also declined (Fig. 2, Table 1).

Circulating concentrations of fasting insulin, 24-h AUC insulin, postprandial insulin concentrations, and ISR were comparable during treatment with liraglutide and placebo, despite the lower glycemia during active treatment suggesting a relative increase in insulin secretion. The ratio of

FIG. 2. Twenty-four-hour profiles of plasma glucose (*A*), serum FFAs (*B*), serum insulin (*C*), and plasma glucagon (*D*) (means ± SEM), day 8. ○, placebo; ●, liraglutide; ▢, meals. (Statistical details are listed in Table 1.)

MPI EXHIBIT 1081 PAGE 3

TABLE 1
Twenty-four-hour profiles and fasting values

	Liraglutide	Placebo	*P*
Fasting values			
Plasma glucose (mmol/l)	8.06 ± 0.52	9.39 ± 0.76	0.078
Plasma glucose day 9 (mmol/l)	7.56 ± 0.42	9.20 ± 0.78	0.025
FFA (mmol/l)	0.52 ± 0.03	0.51 ± 0.03	0.536
Insulin (pmol/l)	95.6 ± 15.1	87.2 ± 17.9	0.513
ISR (nmol/h)	17.7 ± 2.0	16.1 ± 2.1	0.206
Proinsulin/insulin ratio	0.17 ± 0.04	0.27 ± 0.05	0.009
Glucagon (pg/ml)	92.8 ± 7.2	94.5 ± 5.6	0.586
24-h total AUCs			
Plasma glucose (mmol · l^{-1} · h)	187.5 ± 14.0	232.3 ± 21.9	0.014
FFA (mmol · l^{-1} · h)	8.54 ± 0.51	8.65 ± 0.68	0.876
Insulin (pmol · l^{-1} · h)	3,854 ± 581	4,154 ± 881	0.375
ISR (nmol)	566.1 ± 55.1	561.6 ± 72.6	0.982
Glucagon (pg · ml^{-1} · h)	2,179 ± 118	2,371 ± 135	0.037
Postprandial total AUCs			
Plasma glucose 8–12 h (mmol · l^{-1} · h)	38.66 ± 3.52	47.51 ± 3.95	0.010
Plasma glucose 12–16 h (mmol · l^{-1} · h)	32.52 ± 2.92	41.89 ± 4.54	0.017
Plasma glucose 18–22 h (mmol · l^{-1} · h)	33.76 ± 2.50	41.10 ± 3.94	0.016
Insulin 8–12 h (pmol · l^{-1} · h)	999 ± 173	1,056 ± 253	0.512
Insulin 12–16 h (pmol · l^{-1} · h)	723 ± 107	808 ± 162	0.312
Insulin 18–22 h (pmol · l^{-1} · h)	1,017 ± 160	1,117 ± 221	0.392
Proinsulin/insulin ratio 8–12 h	0.12 ± 0.03	0.19 ± 0.04	0.008
Glucagon 8–12 h (pg · ml^{-1} · h)	383.0 ± 22.8	413.4 ± 25.6	0.080
Glucagon 12–16 h (pg · ml^{-1} · h)	362.6 ± 18.6	374.2 ± 19.5	0.384
Glucagon 18–22 h (pg · ml^{-1} · h)	397.3 ± 23.9	470.1 ± 35.2	0.009

Data are means ± SEM. Twenty-four-hour AUCs were calculated from samples obtained from 0800 to 0800. Fasting values were calculated from samples taken at 0730 and 0800 on days 8 and 9.

24-h $AUC_{insulin}/AUC_{glucose}$ was significantly higher during liraglutide administration ($P < 0.05$), whereas only a trend was observed in the fasting insulin/glucose ratio ($P < 0.15$). The fasting proinsulin/insulin ratio and the proinsulin/insulin ratio after the breakfast meal were markedly reduced during liraglutide administration ($P < 0.01$).

Liraglutide exposure significantly reduced the 24-h AUC of glucagon ($P = 0.04$), primarily as a result of a marked reduction in glucagon concentrations after the protein-rich evening meal ($P < 0.01$). Fasting plasma glucagon concentrations were unaltered. The 24-h AUC insulin/glucagon ratio did not differ in the two situations (1.78 ± 0.28 pmol/μg vs. 1.72 ± 0.32 pmol/μg; $P = 0.20$). Fasting value and 24-h AUC of FFAs were similar in the two regimens.

Gastric emptying rate. $AUC_{acetaminophen}$ during the breakfast meal (210 ± 16 vs. 210 ± 12 μmol · l^{-1} · h) and $AUC_{acetaminophen}$ during dinner (178 ± 10 vs. 179 ± 9 μmol · l^{-1} · h) were almost identical. Similarly, t_{max} did not show a significant difference after breakfast (40 ± 8 min during liraglutide treatment vs. 32 ± 4 min during placebo) or dinner (62 ± 10 vs. 50 ± 7 min).

EGR. The rate of EGR in the fasting state was significantly reduced by liraglutide (1.92 ± 0.06 vs. 2.13 ± 0.09 mg · kg^{-1} · min^{-1}; $P = 0.04$). This was due to diminished GLY (0.83 ± 0.04 vs. 1.02 ± 0.04 mg · kg^{-1} · min^{-1}; $P = 0.01$). In contrast, GNG did not change (Fig. 3).

Islet cell function. β-Cell function in the fasting state, as assessed by HOMA-B analysis, was increased by 30% during liraglutide administration ($P = 0.01$). First-phase insulin response after the intravenous glucose bolus was increased by ~60% ($P < 0.01$). During steady state of the hyperglycemic clamp, there was a 2- to 3.5-fold increase in mean insulin concentration, whereas mean circulating glucagon concentration was reduced by ~20% ($P < 0.01$). Analogously, the insulin response after arginine infusion was substantially increased during liraglutide treatment ($P < 0.01$), whereas the glucagon response was reduced ($P = 0.01$). The proinsulin/insulin ratio after liraglutide administration was reduced by 40–50% during the hyperglycemic clamp (Fig. 4, Table 2).

Insulin sensitivity and disposition index. Insulin sensitivity (as calculated by HOMA-R and glucose infusion rate/mean insulin level) was unaltered during the two treatment regimens. The disposition index increased substantially, almost doubling after liraglutide ($P < 0.01$; Table 2).

Pharmacokinetic properties. The half-life of liraglutide in steady state was 17.9 h, and t_{max} was 10.1 ± 3 h (Fig. 5).

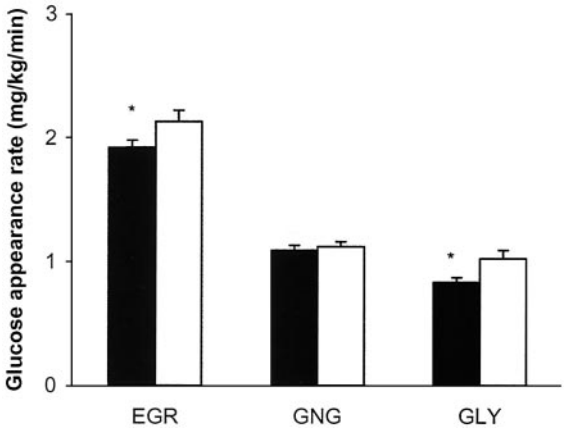


FIG. 3. Fasting EGR, GNG, and GLY, day 9. Data are means ± SEM. □, placebo; ■, liraglutide; *$P < 0.05$, liraglutide vs. placebo.

MPI EXHIBIT 1081 PAGE 4

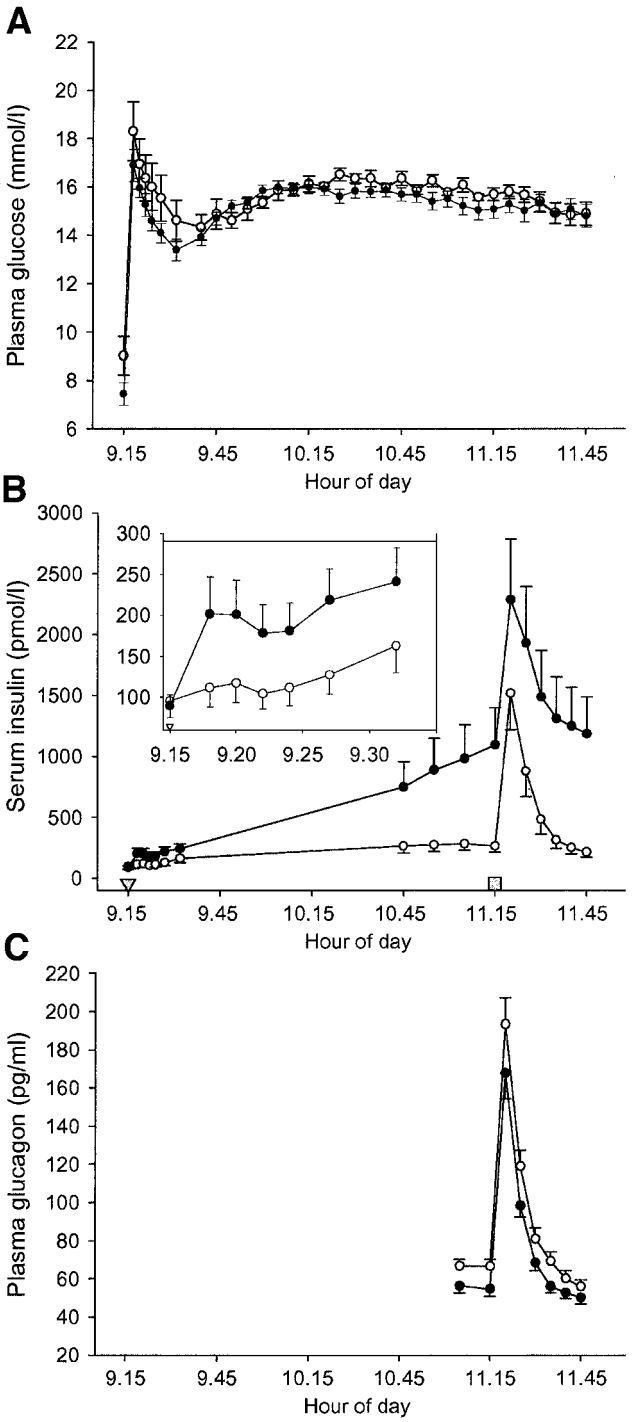


FIG. 4. Plasma glucose (*A*) and serum insulin (*B*) concentrations during an intravenous glucose bolus, hyperglycemic clamp, and arginine stimultion test, day 9. ○, placebo; ●, liraglutide, ▽, glucose bolus (25 g); ▨, arginine bolus (5 g). Plasma glucagon (*C*) is depicted from 1105. Steady state of hyperglycemic clamp was in the interval 1045–1115. The insert in the middle panel is a blow-up of the first-phase insulin response. Data are means ± SEM. (Statistical details are listed in Table 2.)

Safety. No hypoglycemic episodes occurred. Three patients experienced gastrointestinal adverse events (nausea and abdominal pain) during the treatment period. In two cases, the symptoms disappeared after the first days of treatment, and in the remaining case, the severity of the gastrointestinal discomfort declined throughout the treatment week. These three were among the patients in whom the highest serum concentrations of liraglutide were found. No other safety concerns were identified.

DISCUSSION

In the present study, we investigated the effects of 1 week of treatment with the GLP-1 derivative liraglutide (NN2211) on 24-h substrate and hormone profiles under conditions that simulate daily living, on EGR, and on pancreatic islet cell function. A major finding is a markedly reduced circadian plasma glucose level during liraglutide treatment exhibited by fasting, prandial, and nocturnal concentrations. When evaluating the average reduction in plasma glucose (~2 mmol/l), it is important to emphasize that 6 of the 13 patients, even after OHA withdrawal, had a fasting plasma glucose <8.3 mmol/l, i.e., the level above which action should be taken (26). Moreover, this 1-week study was probably too brief to harvest the additional advantageous effects of decreased glucose toxicity, i.e., no improvement in insulin sensitivity. Finally, the same dose (6 μg/kg body wt) of liraglutide was used in all patients. It is likely that some patients would have a further improvement in blood glucose lowering using a higher dose. The current study thus confirms that liraglutide possesses the widely known beneficial effects of native GLP-1 on glycemia in patients with type 2 diabetes (12,13,27) and demonstrates that once-daily dosing of liraglutide is sufficient to ensure 24-h effectiveness (vide infra, pharmacokinetics). It is also important to highlight that no episodes of hypoglycemia occurred despite some of the patients' exhibiting a remarkable glycemic control during liraglutide treatment.

Improved glycemic control during liraglutide treatment is probably orchestrated by changes in insulin and glucagon secretion. Basal and prandial ISRs were unchanged despite the substantial reduction in glycemia, clearly indicating improved β-cell function during daily life conditions. This is in accordance with previous studies exploring native GLP-1 actions (12,13,27) and demonstrates that GLP-1 and/or GLP-1 derivatives augment the ability of the β-cell to respond to prevailing prandial stimuli, e.g., glycemia and glucose dynamics.

A second important observation is that liraglutide significantly reduces 24-h circulating glucagon levels. In particular, the large postprandial glucagon excursion after the protein-rich evening meal was amply decreased. An impaired postprandial inhibition of glucagon is a common feature in type 2 diabetes (5,28), and this contributes notably to the postprandial hyperglycemia (4). Of particular note is that hyperglycemia per se inhibits glucagon secretion, and the decrement in plasma glucagon during liraglutide administration was seen despite a lower glycemic level.

The lowering effect of native GLP-1 on upper gastrointestinal motility is well established (29). In a single-dose study using liraglutide in patients with type 2 diabetes, reduced gastric emptying rate was also reported (16). In contrast, we did not in the current study observe delayed gastric emptying during liraglutide administration. The data achieved from the acetaminophen assessment technique has been proved to agree with data from using much more sophisticated methods for assessment of gastric

MPI EXHIBIT 1081 PAGE 5

TABLE 2
Islet cell function tests, insulin sensitivity, and disposition index

	Liraglutide	Placebo	*P*
Intravenous glucose tolerance test			
Insulin total AUC 9.15–9.32 h (pmol · l^{-1} · h)	55.45 ± 9.93	34.26 ± 6.4	0.008
Maximal insulin concentration 9.15–9.32 h (pmol/l)	262.6 ± 47.5	166.5 ± 32.4	0.007
Hyperglycemic clamp, steady state			
Insulin concentration (pmol/l)	929.6 ± 262.9	271.9 ± 53.3	0.015
Proinsulin/insulin ratio	0.09 ± 0.02	0.18 ± 0.03	0.001
Glucagon concentration (pg/ml)	55.5 ± 3.7	66.7 ± 3.5	0.005
GIR/average serum insulin (arbitrary units)	0.13 ± 0.03	0.15 ± 0.04	0.392
Arginine stimulation test			
Insulin total AUC 11.15–11.45 h (pmol · l^{-1} · h)	799.6 ± 190	307.2 ± 65.5	0.004
Maximal insulin concentration (pmol/l)	2,539 ± 523	1,518 ± 296	0.005
Glucagon total AUC 11.15–11.45 h (pg · l^{-1} · h)	40.7 ± 2.5	48.7 ± 3.0	0.012
Maximal glucagon concentration (pg/ml)	162.0 ± 13.7	193.4 ± 14	0.034
HOMA analysis			
β-Cell function (% of normal)	59.95 ± 8.84	46.07 ± 9.86	0.010
Insulin resistance (fold normal)	4.09 ± 0.84	4.74 ± 1.15	0.373
Disposition index (pmol/l*HOMA-R)	80.4 ± 14	41.9 ± 5	0.008

Data are means ± SEM. GIR, glucose infusion rate.

emptying (30). A possible explanation behind the apparent discrepancy between the present and earlier data might be that the effect on upper gastrointestinal motility undergoes tachyphylaxis. However, two studies of liraglutide/native GLP-1 in animals (31) and humans (12) report an effect on gastric emptying after several weeks of treatment. Thus, it is conceivable that the lack of effect of liraglutide on gastric emptying observed in our study may be due to the dose used. Probably the dose-response relationships between blood glucose–lowering effects and delaying gastric emptying differ, the latter being rightward shifted.

Another novel finding is the decline in fasting EGR after liraglutide administration. This could be due to an increased insulin/glucagon ratio. However, a significant increase was not observed in peripheral blood. It is essential to emphasize that circulating FFA levels did not differ in the two situations. In our diabetic individuals, fasting EGR was within what may be defined as the normal range. Presumably, a 10% reduction in fasting EGR (~10 g glucose overnight) shall have an effect on fasting glycemic levels. It is also reasonable to assume that a much greater decrease in EGR will be present during daytime, taking into consideration the importance of dynamic changes in circulating glucagon on hepatic glucose handling in the prandial state (4,32,33).

Our interest was also in exploring the influence of liraglutide treatment on the contribution of GNG and GLY to EGR. The method allowed us to examine only fasting glucose kinetics. Several authors reported that the elevated EGR of individuals with type 2 diabetes is mainly attributable to an augmented GNG (2,3,34,35). The current study showed that the restraining effect of the GLP-1 derivative liraglutide on fasting EGR was solely due to inhibition of GLY, fasting GNG being unchanged. The opposite result might have been suspected, namely a reduction of GNG. However, studies, e.g., by Cherrington et al. (36), have demonstrated that the inhibitory effect of physiological insulin concentrations (in contrast to supraphysiological levels) on hepatic glucose production is primarily mediated through decreased GLY, whereas GNG

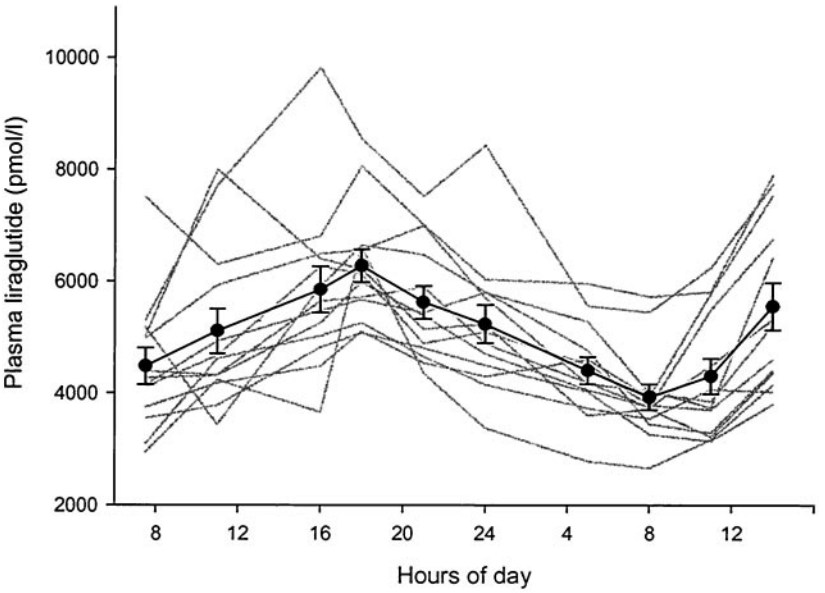


FIG. 5. Concentration profiles of liraglutide. Dosing time was 0730. Gray lines indicate individual concentration profiles; black line indicates mean ± SEM.

MPI EXHIBIT 1081 PAGE 6

is less sensitive to the direct action of insulin. Similarly, the stimulatory effect of glucagon on hepatic glucose release is primarily due to glycogen breakdown, whereas the hormone has less effect on GNG (36,37). Very high portal insulin levels are capable of suppressing hepatic GNG, but this may primarily be due to an indirect effect via decreased lipolysis and hence plasma FFA levels (36).

Again, although there was no significant change in the concentrations of fasting levels of insulin and glucagon, these were relatively augmented or decreased, respectively, considering the lowered level of glycemia. Furthermore, peripheral blood does not necessarily precisely reflect portal concentration. The effect of GLP-1/GLP-1 derivatives on GLY and GNG in patients with type 2 diabetes and more deranged glycemic control, i.e., with a substantial elevation of EGR, remains unknown.

Progressive impaired β-cell function leading to insulin deficiency is a key feature of type 2 diabetes (1,38). Both a reduction in β-cell mass and a functional abnormality of the islet cells seem to be responsible for the islet cell's deterioration (39). It seems to be an ongoing process hardly influenced by concomitant glucose lowering treatment (38). Loss of first-phase insulin response to glucose is an early β-cell defect, followed by a weakened basal and second-phase insulin output as the disease progresses (39,40). This, combined with an abnormal circadian glucagon secretory pattern (5,33) and diminished ability of hyperglycemia to suppress glucagon secretion, demonstrates the necessity for opposing both the α- and β-cell malfunction in the treatment of type 2 diabetes.

Improved β-cell function in patients with type 2 diabetes has been reported after 6–12 h of GLP-1 infusion as assessed by first- and second-phase insulin response (41). A 6-week study demonstrated similar effects on β-cell function, strongly indicating maintenance of this beneficial GLP-1 action (12). We found a dramatic improvement in β-cell function after 8 days of liraglutide treatment. All β-cell function tests were substantially ameliorated, indicating an effect on first- and second-phase insulin response to glucose as well as (near) maximal insulin secretory capacity. In addition, we found an almost 50% reduction of the proinsulin/insulin ratio in the circulation during both fasting and stimulation. The latter observation may indicate an improved processing of insulin in the β-cell. In fact, in the current study, liraglutide treatment almost led to a normalized fasting proinsulin/insulin ratio. An elevated proinsulin/insulin ratio is a cardinal feature in type 2 diabetes and the pre-diabetic state (42,43) and is a well-established indicator of β-cell dysfunction (44). Furthermore, liraglutide treatment led to an almost doubling of the disposition index, which reflects the ability of the β-cells to adapt to the contemporary insulin resistance (glucose allostasis) (45), again emphasizing the powerful effects of GLP-1/GLP-1 derivatives on the β-cell.

Another important impact of liraglutide on pancreatic glucoregulatory function was the significant inhibition of glucagon secretion during hyperglycemia and in particular during arginine infusion, demonstrating that liraglutide efficiently improves the secretory pattern of both α- and β-cells in the pancreatic islets of patients with type 2 diabetes, and it is anticipated that these beneficial actions will be maintained during long-term treatment. Very conceivable, the impact of GLP-1 derivatives on islet-cell function and consequently on glycemic control will further benefit from long-term treatment both directly via the GLP-1 trophic effects on the β-cells and indirectly via decreased glucose toxicity.

The pharmacokinetic properties of liraglutide were evaluated by means of a 30-h plasma profile during steady state. The plasma half-life was ~18 h, and t_{max} was ~10 h postdosing. Medication was in this study given in the morning, resulting in the peak plasma concentration at ~1800. Both the 24-h lasting beneficial effect on glycemia and the relatively high plasma concentrations maintained for 24 h clearly document that once-daily dosing is sufficient. Dosing at bedtime is another interesting option. Whether it is superior to morning dosing needs to be clarified.

In summary, we report that once-daily dosing with the GLP-1 derivative liraglutide for 1 week in individuals with type 2 diabetes results in *1*) substantially decreased glucose concentration lasting 24 h and is ascribable to a relative increase in insulin secretion and decrease in glucagon concentration; *2*) reduced EGR, as a result of diminished GLY; and *3*) substantially improved AIR and maximal insulin secretory capacity. During longer lasting treatment, these promising antidiabetic effects may be further supplemented by an increase in β-cell mass and the benefit achieved by weight loss.

ACKNOWLEDGMENTS

The Danish Diabetes Association, the Institute of Experimental Clinical Research; University of Aarhus; Novo Nordisk A/S, Copenhagen; and National Institutes of Health Grant 14507 supported this study financially.

Ms. Annette Mengel, Ms. Lene Ring, and Ms. Elsebeth Hornemann are thanked for outstanding technical assistance.

REFERENCES

1. Ferrannini E: Insulin resistance versus insulin deficiency in non-insulin-dependent diabetes mellitus: problems and prospects. *Endocr Rev* 19:477–490, 1998
2. Consoli A, Nurjhan N, Capani F, Gerich J: Predominant role of gluconeogenesis in increased hepatic glucose production in NIDDM. *Diabetes* 38:550–557, 1989
3. Magnusson I, Rothman DL, Katz LD, Shulman RG, Shulman GI: Increased rate of gluconeogenesis in type II diabetes mellitus. A 13C nuclear magnetic resonance study. *J Clin Invest* 90:1323–1327, 1992
4. Shah P, Vella A, Basu A, Basu R, Schwenk WF, Rizza RA: Lack of suppression of glucagon contributes to postprandial hyperglycemia in subjects with type 2 diabetes mellitus. *J Clin Endocrinol Metab* 85:4053–4059, 2000
5. Reaven GM, Chen YD, Golay A, Swislocki AL, Jaspan JB: Documentation of hyperglucagonemia throughout the day in nonobese and obese patients with noninsulin-dependent diabetes mellitus. *J Clin Endocrinol Metab* 64:106–110, 1987
6. Vilsboll T, Krarup T, Deacon CF, Madsbad S, Holst JJ: Reduced postprandial concentrations of intact biologically active glucagon-like peptide 1 in type 2 diabetic patients. *Diabetes* 50:609–613, 2001
7. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33): U.K. Prospective Diabetes Study (UKPDS) Group. *Lancet* 352:837–853, 1998
8. Gaede P, Vedel P, Larsen N, Jensen GV, Parving HH, Pedersen O: Multifactorial intervention and cardiovascular disease in patients with type 2 diabetes. *N Engl J Med* 348:383–393, 2003
9. Nauck MA: Is glucagon-like peptide 1 an incretin hormone? *Diabetologia* 42:373–379, 1999
10. Stoffers DA, Kieffer TJ, Hussain MA, Drucker DJ, Bonner-Weir S, Habener

MPI EXHIBIT 1081 PAGE 7

JF, Egan JM: Insulinotropic glucagon-like peptide 1 agonists stimulate expression of homeodomain protein IDX-1 and increase islet size in mouse pancreas. *Diabetes* 49:741–748, 2000
11. Flint A, Raben A, Astrup A, Holst JJ: Glucagon-like peptide 1 promotes satiety and suppresses energy intake in humans. *J Clin Invest* 101:515–520, 1998
12. Zander M, Madsbad S, Madsen JL, Holst JJ: Effect of 6-week course of glucagon-like peptide 1 on glycaemic control, insulin sensitivity, and beta-cell function in type 2 diabetes: a parallel-group study. *Lancet* 359: 824–830, 2002
13. Meneilly GS, Grieg N, Tildesley H, Habener JF, Egan JM, Eliha D: Effects of 3 months of continuous subcutaneous administration of glucagon-like peptide 1 in elderly patients with type 2 diabetes. *Diabetes Care* 26:2835–2841, 2003
14. Deacon CF, Nauck MA, Toft-Nielsen M, Pridal L, Willms B, Holst JJ: Both subcutaneously and intravenously administered glucagon-like peptide I are rapidly degraded from the NH_2-terminus in type II diabetic patients and in healthy subjects. *Diabetes* 44:1126–1131, 1995
15. Agerso H, Jensen LB, Elbrond B, Rolan P, Zdravkovic M: The pharmacokinetics, pharmacodynamics, safety and tolerability of NN2211, a new long-acting GLP-1 derivative, in healthy men. *Diabetologia* 45:195–202, 2002
16. Juhl CB, Hollingdal M, Sturis J, Jakobsen G, Agerso H, Veldhuis J, Porksen N, Schmitz O: Bedtime administration of NN2211, a long-acting GLP-1 derivative, substantially reduces fasting and postprandial glycemia in type 2 diabetes. *Diabetes* 51:424–429, 2002
17. Matthews D, Madsbad S, Schmitz O, Lagendorf KW, Jakobsen G: The long-acting GLP-1 derivative, NN2211, a new agent for the treatment of type 2 diabetes (Abstract). *Diabetes* 51 (Suppl. 2):A84, 2002
18. Clements JA, Heading RC, Nimmo WS, Prescott LF: Kinetics of acetaminophen absorption and gastric emptying in man. *Clin Pharmacol Ther* 24: 420–431, 1978
19. Hother-Nielsen O, Mengel A, Moller J, Rasmussen O, Schmitz O, Beck-Nielsen H: Assessment of glucose turnover rates in euglycaemic clamp studies using primed-constant [3-3H]-glucose infusion and labelled or unlabelled glucose infusates. *Diabet Med* 9:840–849, 1992
20. Landau BR, Wahren J, Chandramouli V, Schumann WC, Ekberg K, Kalhan SC: Contributions of gluconeogenesis to glucose production in the fasted state. *J Clin Invest* 98:378–385, 1996
21. Finegood DT, Bergman RN, Vranic M: Estimation of endogenous glucose production during hyperinsulinemic-euglycemic glucose clamps: comparison of unlabeled and labeled exogenous glucose infusates. *Diabetes* 36:914–924, 1987
22. Polonsky KS, Licinio-Paixao J, Given BD, Pugh W, Rue P, Galloway J, Karrison T, Frank B: Use of biosynthetic human C-peptide in the measurement of insulin secretion rates in normal volunteers and type I diabetic patients. *J Clin Invest* 77:98–105, 1986
23. Van Cauter E, Mestrez F, Sturis J, Polonsky KS: Estimation of insulin secretion rates from C-peptide levels: comparison of individual and standard kinetic parameters for C-peptide clearance. *Diabetes* 41:368–377, 1992
24. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC: Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 28:412–419, 1985
25. Kahn SE, Prigeon RL, McCulloch DK, Boyko EJ, Bergman RN, Schwartz MW, Neifing JL, Ward WK, Beard JC, Palmer JP, Porte D Jr: Quantification of the relationship between insulin sensitivity and beta-cell function in human subjects: evidence for a hyperbolic function. *Diabetes* 42:1663–1672, 1993
26. American Diabetes Association: Standards of medical care for patients with diabetes mellitus. *Diabetes Care* 25 (Suppl. 1):S33–S49, 2002
27. Larsen J, Hylleberg B, Ng K, Damsbo P: Glucagon-like peptide-1 infusion must be maintained for 24 h/day to obtain acceptable glycemia in type 2 diabetic patients who are poorly controlled on sulphonylurea treatment. *Diabetes Care* 24:1416–1421, 2001
28. Butler PC, Rizza RA: Contribution to postprandial hyperglycemia and effect on initial splanchnic glucose clearance of hepatic glucose cycling in glucose-intolerant or NIDDM patients. *Diabetes* 40:73–81, 1991
29. Nauck MA, Wollschlager D, Werner J, Holst JJ, Orskov C, Creutzfeldt W, Willms B: Effects of subcutaneous glucagon-like peptide 1 (GLP-1 [7-36 amide]) in patients with NIDDM. *Diabetologia* 39:1546–1553, 1996
30. Naslund E, Bogefors J, Gryback P, Jacobsson H, Hellstrom PM: Gastric emptying: comparison of scintigraphic, polyethylene glycol dilution, and paracetamol tracer assessment techniques. *Scand J Gastroenterol* 35:375–379, 2000
31. Ribel U, Larsen MO, Rolin B, Carr RD, Wilken M, Sturis J, Westergaard L, Deacon CF, Knudsen LB: NN2211: a long-acting glucagon-like peptide-1 derivative with anti-diabetic effects in glucose-intolerant pigs. *Eur J Pharmacol* 451:217–225, 2002
32. Singhal P, Caumo A, Carey PE, Cobelli C, Taylor R: Regulation of endogenous glucose production after a mixed meal in type 2 diabetes. *Am J Physiol Endocrinol Metab* 283:E275–E283, 2002
33. Dinneen S, Alzaid A, Turk D, Rizza R: Failure of glucagon suppression contributes to postprandial hyperglycaemia in IDDM. *Diabetologia* 38:337–343, 1995
34. Wajngot A, Chandramouli V, Schumann WC, Ekberg K, Jones PK, Efendic S, Landau BR: Quantitative contributions of gluconeogenesis to glucose production during fasting in type 2 diabetes mellitus. *Metabolism* 50:47–52, 2001
35. Gastaldelli A, Baldi S, Pettiti M, Toschi E, Camastra S, Natali A, Landau BR, Ferrannini E: Influence of obesity and type 2 diabetes on gluconeogenesis and glucose output in humans: a quantitative study. *Diabetes* 49:1367–1373, 2000
36. Cherrington AD: Banting Lecture 1997: control of glucose uptake and release by the liver in vivo. *Diabetes* 48:1198–1214, 1999
37. Magnusson I, Rothman DL, Gerard DP, Katz LD, Shulman GI: Contribution of hepatic glycogenolysis to glucose production in humans in response to a physiological increase in plasma glucagon concentration. *Diabetes* 44: 185–189, 1995
38. Turner R, Cull C, Holman R: United Kingdom Prospective Diabetes Study 17: a 9-year update of a randomized, controlled trial on the effect of improved metabolic control on complications in non-insulin-dependent diabetes mellitus. *Ann Intern Med* 124:136–145, 1996
39. Porte D Jr: Banting Lecture 1990: β-cells in type II diabetes mellitus. *Diabetes* 40:166–180, 1991
40. Mahler RJ, Adler ML: Clinical review 102: type 2 diabetes mellitus: update on diagnosis, pathophysiology, and treatment. *J Clin Endocrinol Metab* 84:1165–1171, 1999
41. Ahren B, Larsson H, Holst JJ: Effects of glucagon-like peptide-1 on islet function and insulin sensitivity in noninsulin-dependent diabetes mellitus. *J Clin Endocrinol Metab* 82:473–478, 1997
42. Nyholm B, Walker M, Gravholt CH, Shearing PA, Sturis J, Alberti KG, Holst JJ, Schmitz O: Twenty-four-hour insulin secretion rates, circulating concentrations of fuel substrates and gut incretin hormones in healthy offspring of type II (non-insulin-dependent) diabetic parents: evidence of several aberrations. *Diabetologia* 42:1314–1323, 1999
43. Kahn SE, Halban PA: Release of incompletely processed proinsulin is the cause of the disproportionate proinsulinemia of NIDDM. *Diabetes* 46:1725–1732, 1997
44. Roder ME, Porte D Jr, Schwartz RS, Kahn SE: Disproportionately elevated proinsulin levels reflect the degree of impaired B cell secretory capacity in patients with noninsulin-dependent diabetes mellitus. *J Clin Endocrinol Metab* 83:604–608, 1998
45. Stumvoll M, Tataranni PA, Stefan N, Vozarova B, Bogardus C: Glucose allostasis. *Diabetes* 52:903–909, 2003

MPI EXHIBIT 1081 PAGE 8