PROVISIONAL APPLICATION FOR PATENT COVER SHEET — Page 1 of 2

This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

SUBMITTED BY EFS-WEB ON 19 MAY 2006

INVENTOR(S)

Given Name (first and middle [if any])	Family Name or Surname	Residence (City and either State or Foreign Country)
MARK E.	DEEM	Mountain View, CA
HANSON	GIFFORD	Woodside, CA

Additional inventors are being named on the _____ *separately numbered sheets attached hereto*

TITLE OF THE INVENTION (500 characters max)

METHODS AND SYSTEMS FOR TOXIN DELIVERY TO THE NASAL CAVITY

Direct all correspondence to: **CORRESPONDENCE ADDRESS**

☒ The address corresponding to Customer Number: **20350**

OR

☐ Firm or Individual Name

Address

City	State	Zip
Country	Telephone	Email

ENCLOSED APPLICATION PARTS *(check all that apply)*

☒ Application Data Sheet. See 37 CFR 1.76

☒ Specification *Number of Pages* 30

☒ Drawing(s) *Number of Sheets* 20

☐ CD(s), Number of CDs _____

☒ Other (specify) Return Postcard

Fees Due: Filing fee of $200 ($100 for small entity). If the specification and drawings exceed 100 sheets of paper, an application size fee is also due, which is $250 ($125 for small entity) for each additional 50 sheets or fraction thereof. See 35 U.S.C. 41(a)(1)(G) and 37 CFR 1.16(s).

METHOD OF PAYMENT OF THE FILING FEE AND APPLICATION SIZE FEE FOR THIS PROVISIONAL APPLICATION FOR PATENT

☒ Applicant claims small entity status. See 37 CFR 1.27.

☐ A check or money order is enclosed to cover the filing fee and application size fee (if applicable).

☐ Payment by credit card. Form PTO-2038 is attached.

☒ The Director is hereby authorized to charge the filing fee and application size fee (if applicable) or credit any overpayment to Deposit Account Number: 20-1430. A duplicative copy of this form is enclosed for fee processing.

100

TOTAL FEE AMOUNT ($)

USE ONLY FOR FILING A PROVISIONAL APPLICATION FOR PATENT

60778218 v1

NEURENT 1011

PTO/SB/16 (10-05)

The invention was made by an agency of the United States Government or under a contract with an agency of the United States Government.

☒ No.

☐ Yes, the name of the U.S. Government agency and the Government contract number are: _____

WARNING:

Petitioner/applicant is cautioned to avoid submitting personal information in documents filed in a patent application that may contribute to identity theft. Personal information such as social security numbers, bank account numbers, or credit card numbers (other than a check or credit card authorization form PTO-2038 submitted for payment purposes) is never required by the USPTO to support a petition or an application. If this type of personal information is included in documents submitted to the USPTO, petitioners/applicants should consider redacting such personal information from the documents before submitting them to the USPTO. Petitioner/applicant is advised that the record of a patent application is available to the public after publication of the application (unless a non-publication request in compliance with 37 CFR 1.213(a) is made in the application) or issuance of a patent. Furthermore, the record from an abandoned application may also be available to the public if the application is referenced in a published application or an issued patent (see 37 CFR 1.14). Checks and credit card authorization forms PTO-2038 submitted for payment purposes are not retained in the application file and therefore are not publicly available.

SIGNATURE		Date	May 19, 2006
TYPED or PRINTED NAME	James M. Heslin	REGISTRATION NO. (*if appropriate*)	29,541
TELEPHONE	650-326-2400	Docket Number:	020979-003700US

60778218 v1

Application Data Sheet

Application Information

Application number::	
Filing Date::	05/19/2006
Application Type::	Provisional
Subject Matter::	Utility
Title::	METHODS AND SYSTEMS FOR TOXIN DELIVERY TO THE NASAL CAVITY
Attorney Docket Number::	020979-003700US
Request for Early Publication::	No
Request for Non-Publication::	No
Total Drawing Sheets::	20
Small Entity?::	Yes
Petition included?::	No
Secrecy Order in Parent Appl.::	No

Applicant Information

Applicant Authority Type::	Inventor
Primary Citizenship Country::	US
Status::	Full Capacity
Given Name::	MARK
Middle Name::	E.
Family Name::	DEEM
City of Residence::	Mountain View
State or Province of Residence::	CA
Country of Residence::	US
Street of Mailing Address::	685 Sierra Avenue
City of Mailing Address::	Mountain View
State or Province of mailing address::	CA
Country of mailing address::	US
Postal or Zip Code of mailing address::	94041

Applicant Authority Type::	Inventor
Primary Citizenship Country::	US
Status::	Full Capacity
Given Name::	HANSON
Family Name::	GIFFORD
City of Residence::	Woodside
State or Province of Residence::	CA
Country of Residence::	US
Street of Mailing Address::	3180 Woodside Road
City of Mailing Address::	Woodside
State or Province of mailing address::	CA
Country of mailing address::	US
Postal or Zip Code of mailing address::	94062

Correspondence Information

Correspondence Customer Number::	20350

Representative Information

Representative Customer Number::	20350

Assignee Information

Assignee Name::	The Foundry Inc.
Street of mailing address::	199 Jefferson Drive
City of mailing address::	Menlo Park
State or Province of mailing address::	CA
Country of mailing address::	US
Postal or Zip Code of mailing address::	94025

This sheet is not to be filed with the USPTO, but retained in the prosecution file as a record of the iManage number. Fields having no information may be deleted from the ADS. For example, if there is no foreign priority claim, the foreign priority text may be deleted from the ADS.

60778213 v1

Progenitor Toxin
Active Toxin Type A
LC
Proteolytic Cleavage
HC

FIG 1

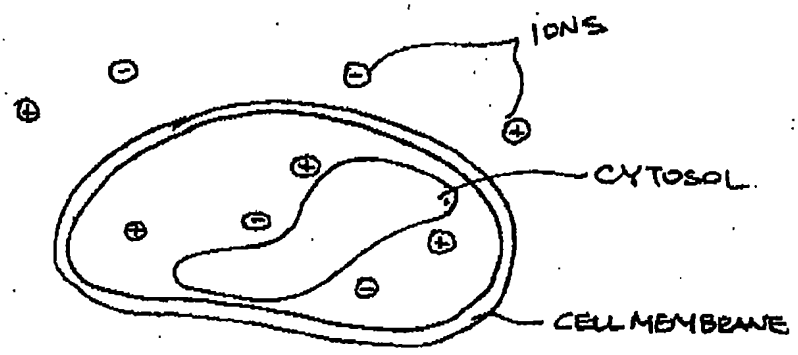
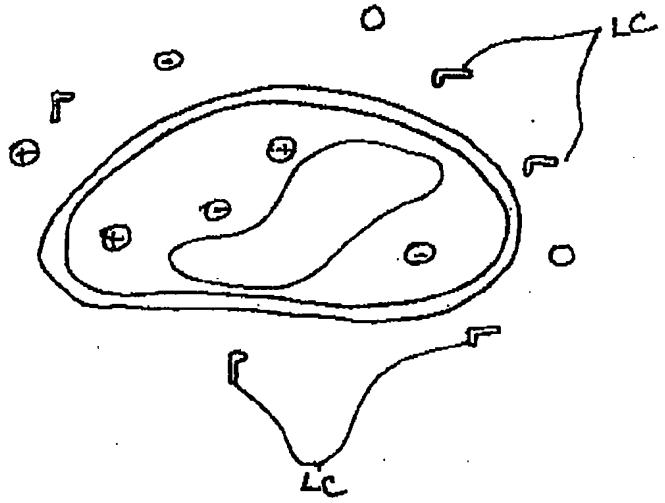


FIG 2A



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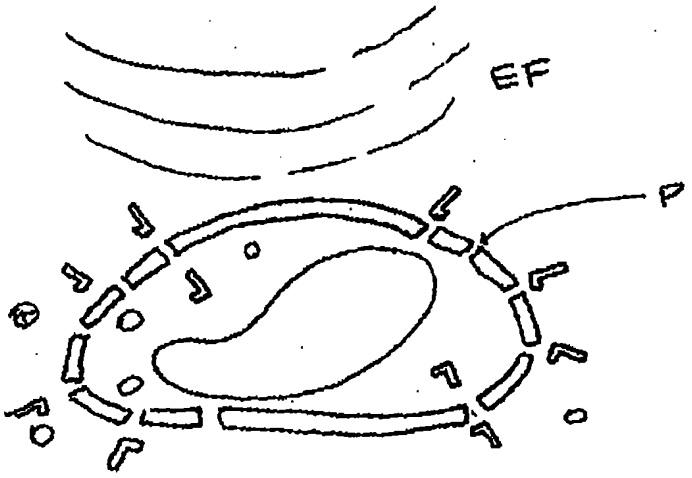


FIG 3A

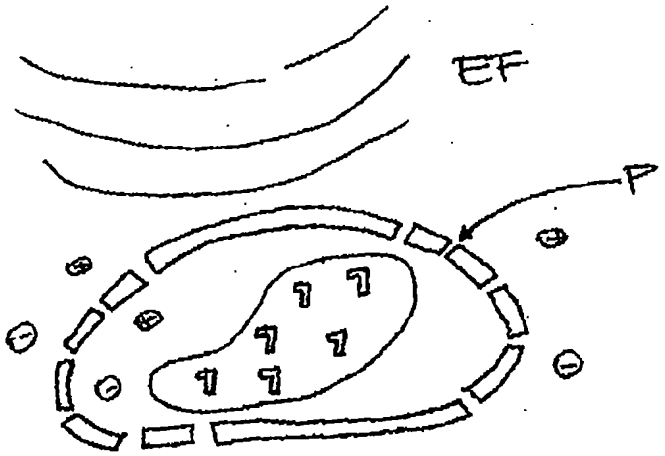


FIG 3B

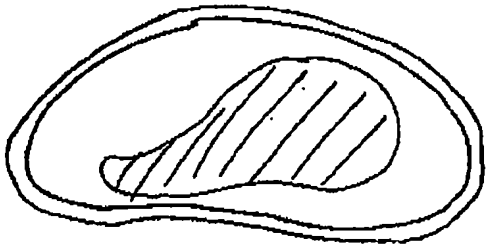


FIG 4

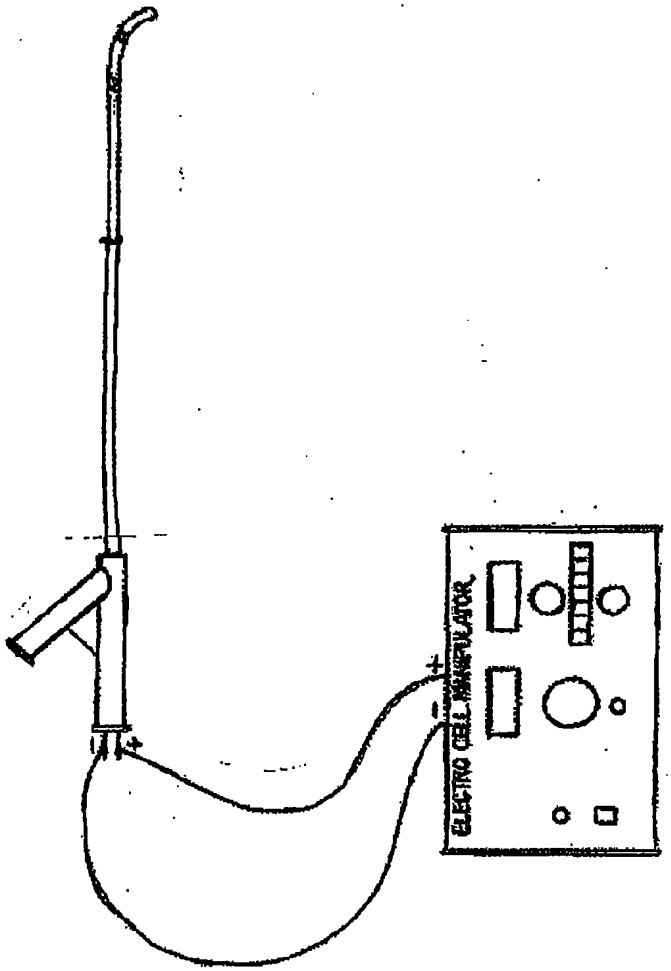
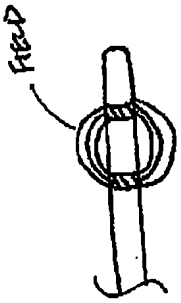


FIG 5



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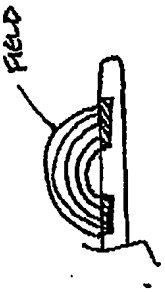

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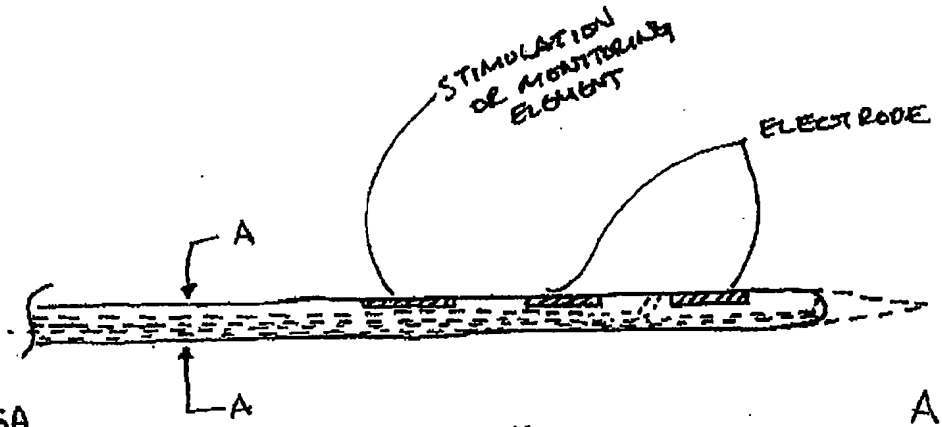

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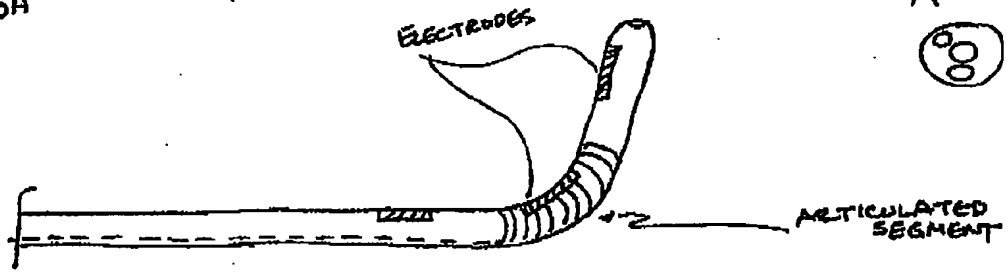

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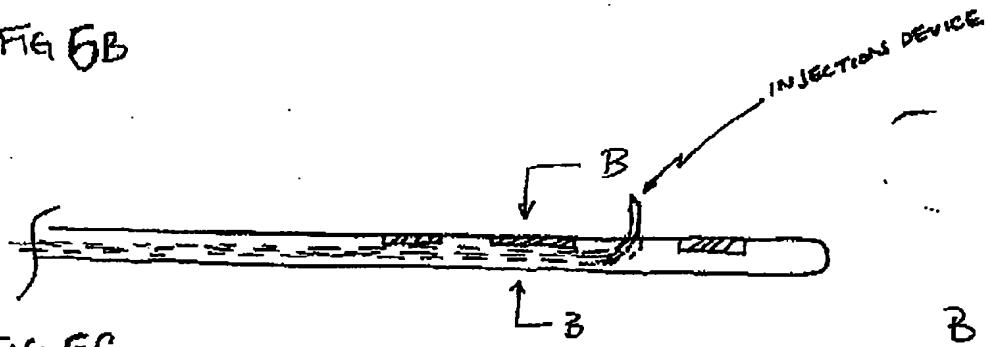


FIG 6C

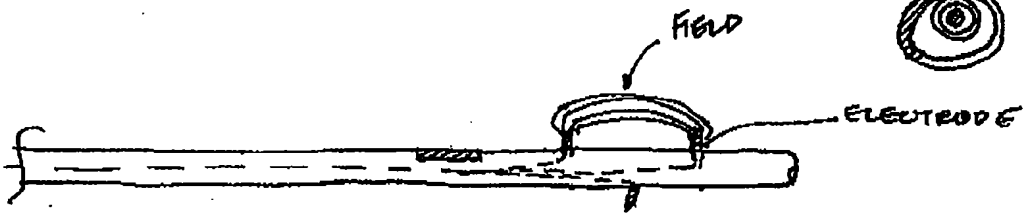


FIG 6D

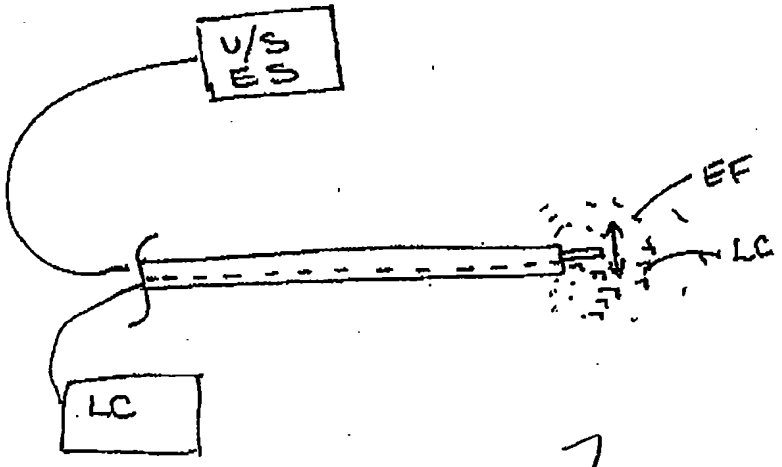


FIG ~~8~~ 7

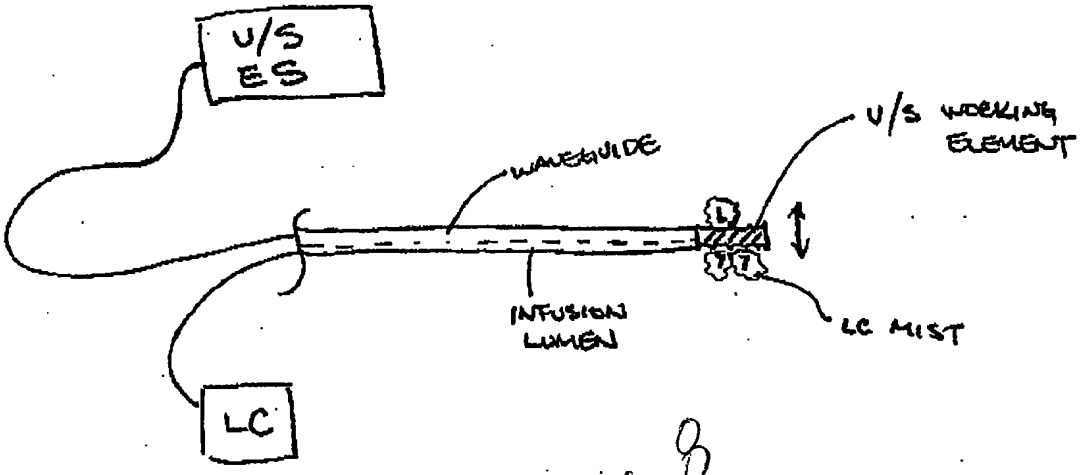


FIG ~~11~~ 8

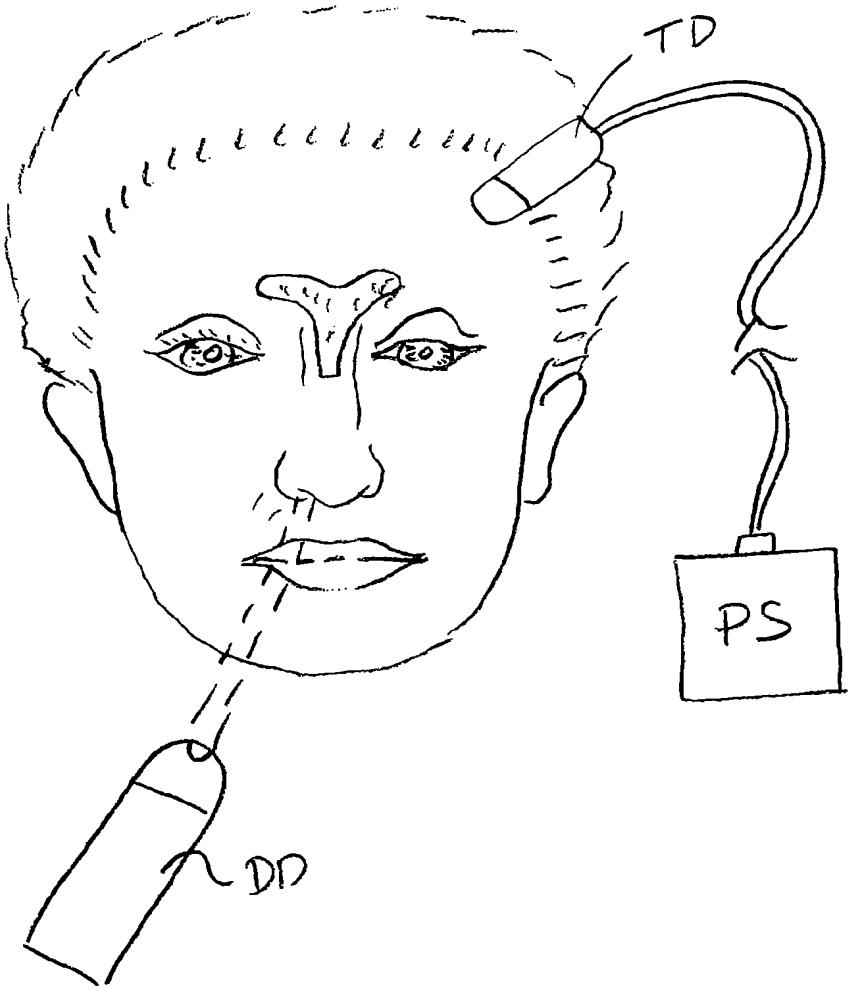


Fig-9

LC8

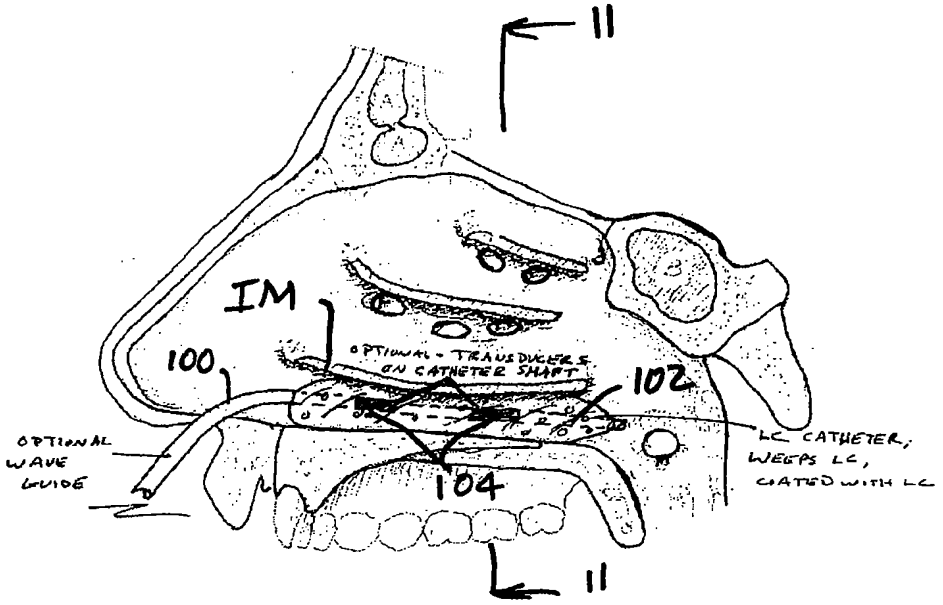
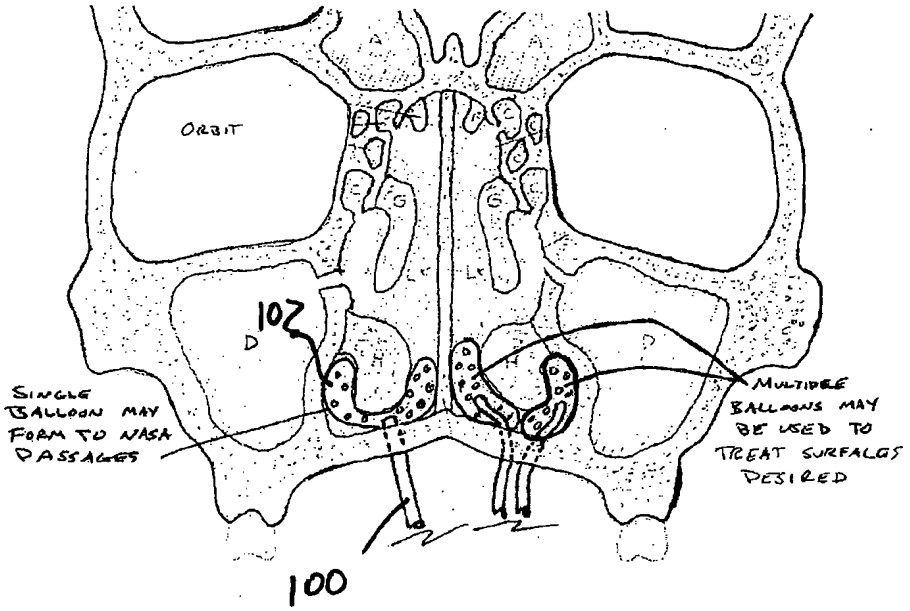


FIG. 10



LC9

FIG. 11

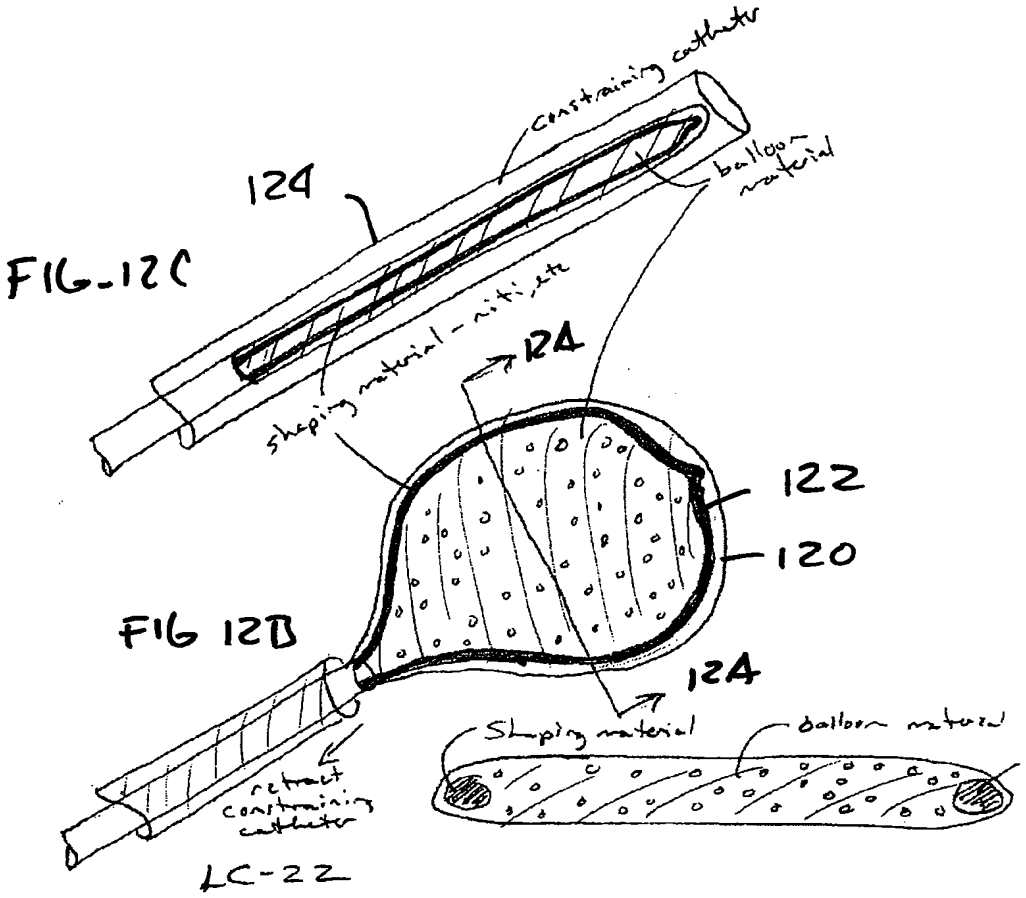

constraining catheter
124
balloon material
FIG-12C
shaping material - nitinole
124
122
120
FIG 12B
124
Shaping material
balloon material
retract constraining catheter
LC-22

FIG. 12A

LC 4

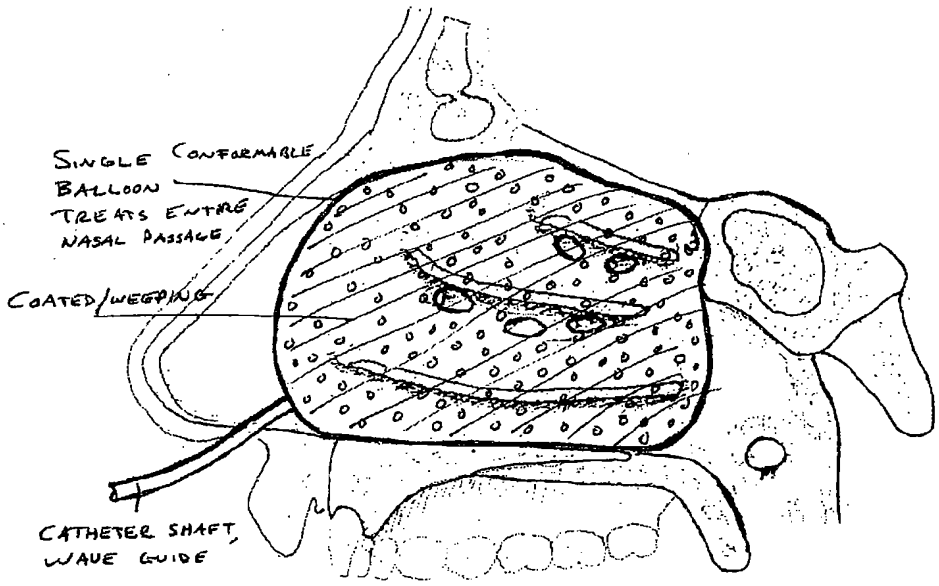
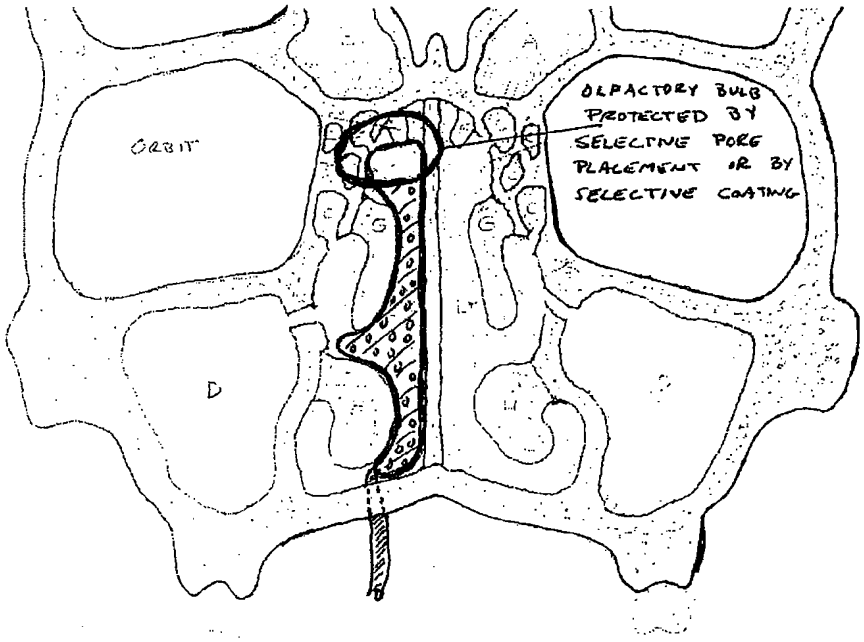


FIG. 13



LC 5

FIG. 14

LC 6

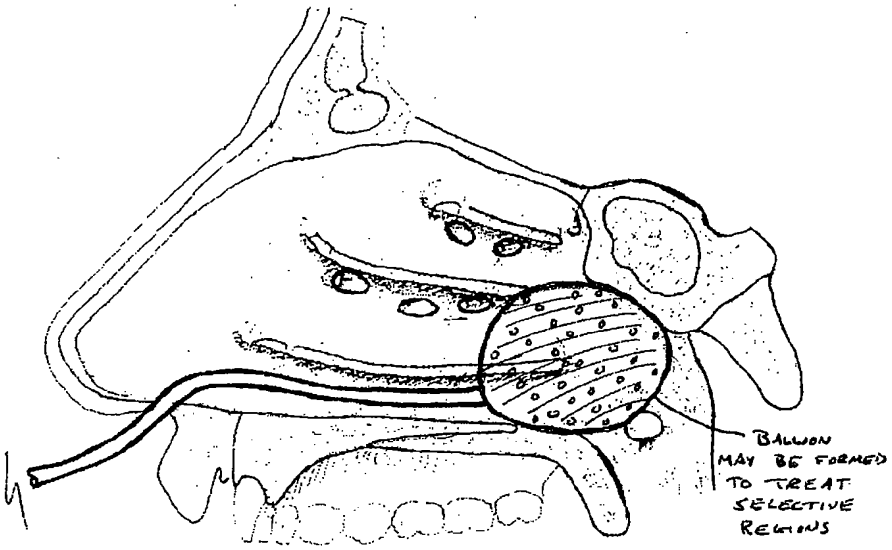
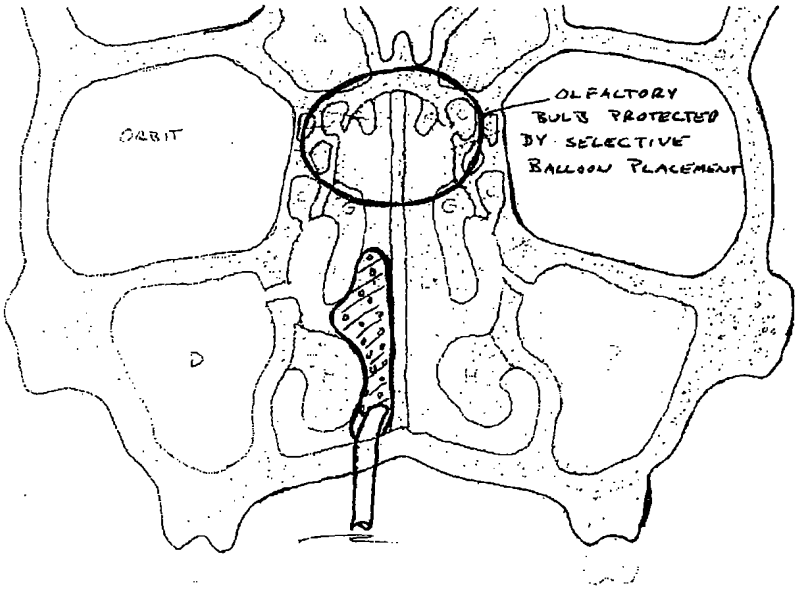
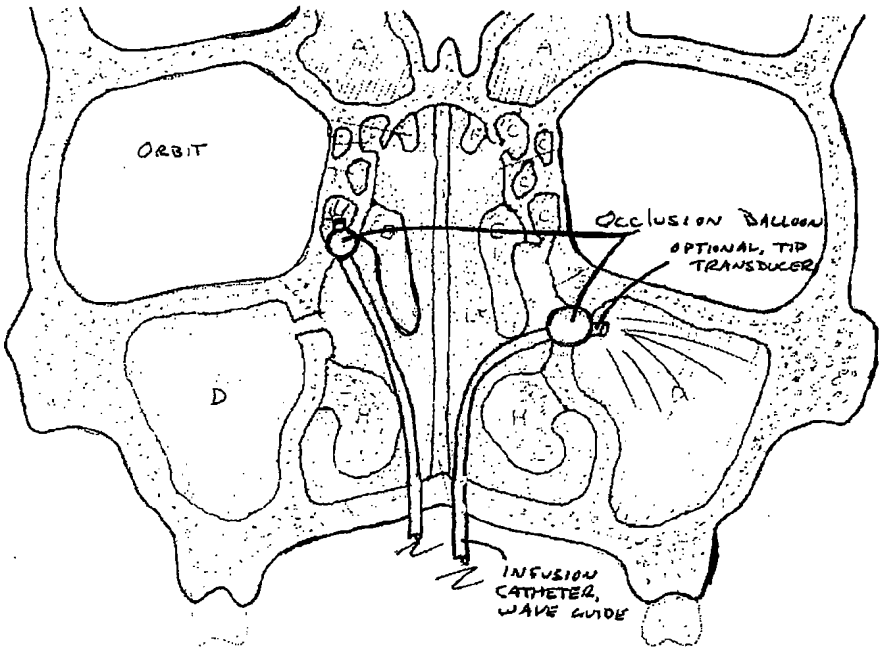


FIG. 15



LC 7

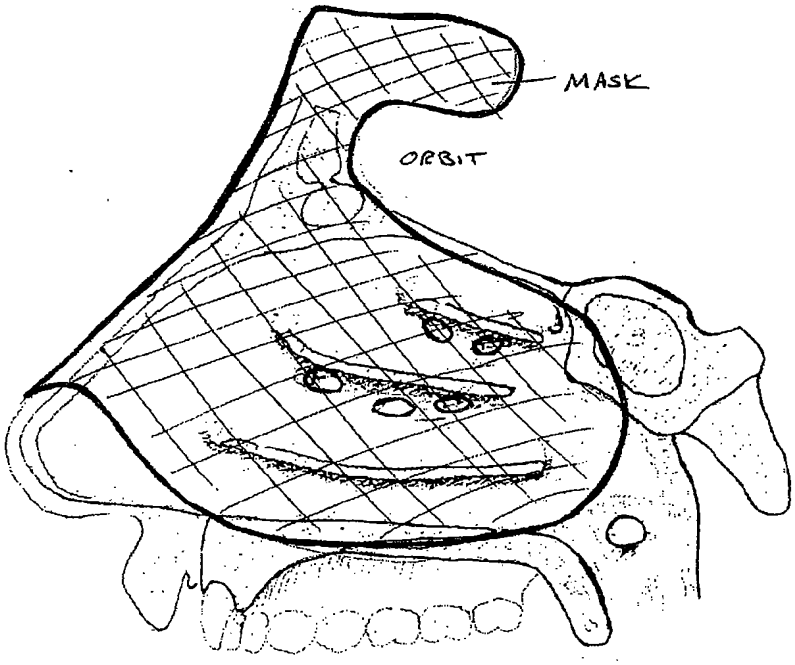
FIG. 16



LC3

FIG. 17

LC.20



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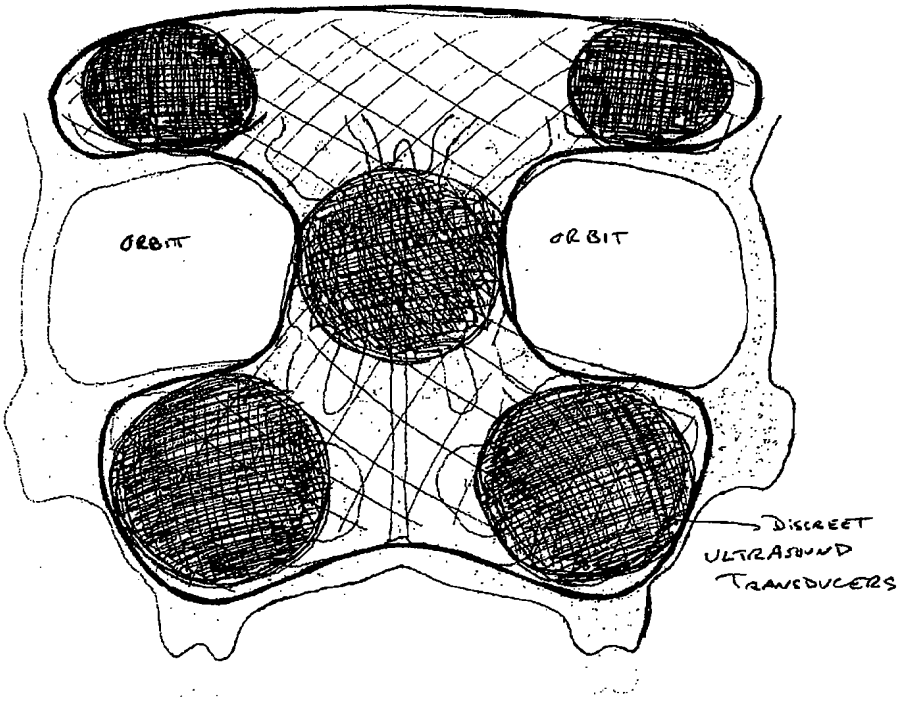


LC-21

FIG. 19

LC-10

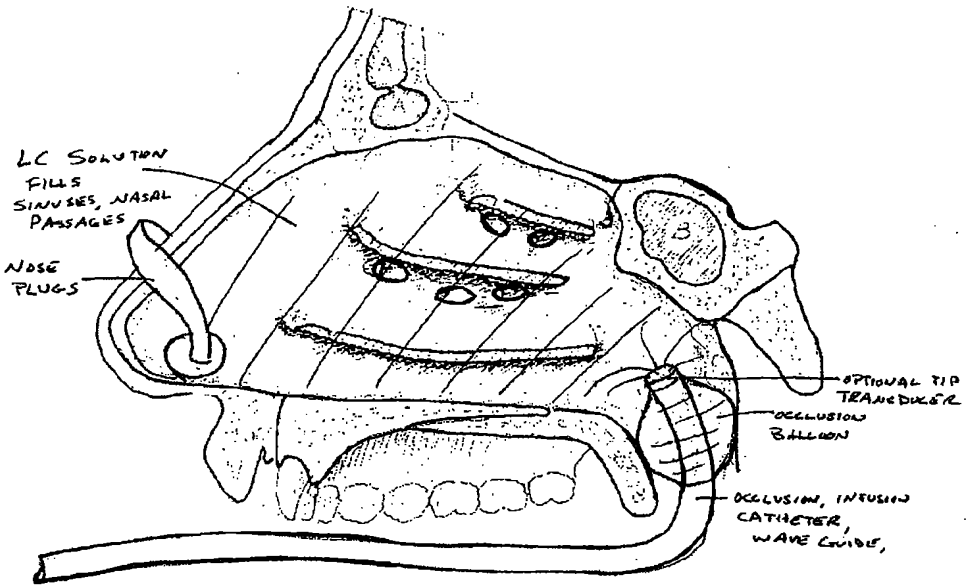
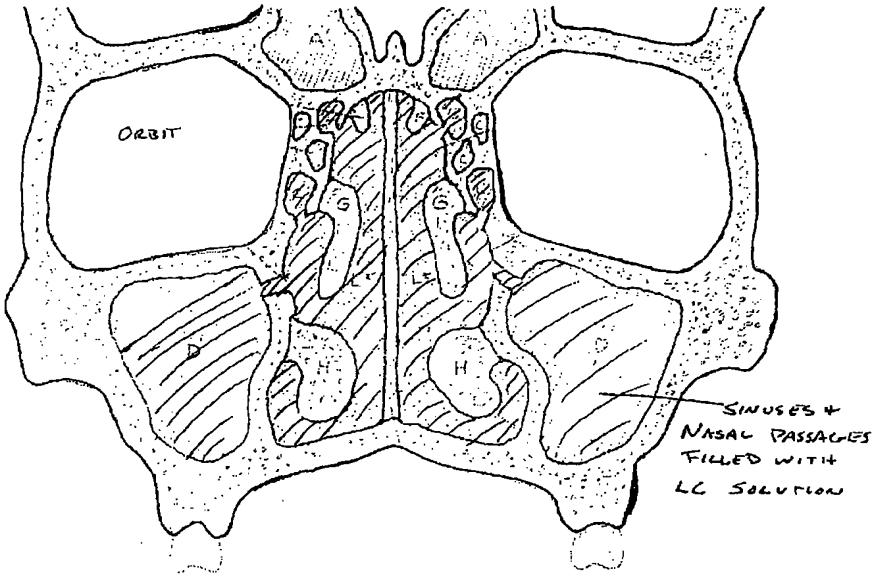


FIG. 20



LC·11

FIG. 21

LC·14

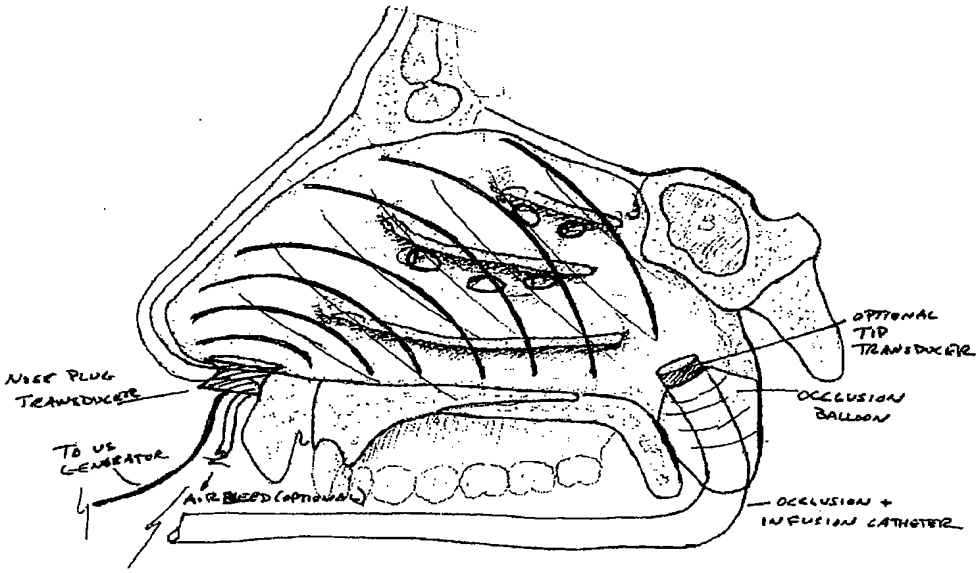
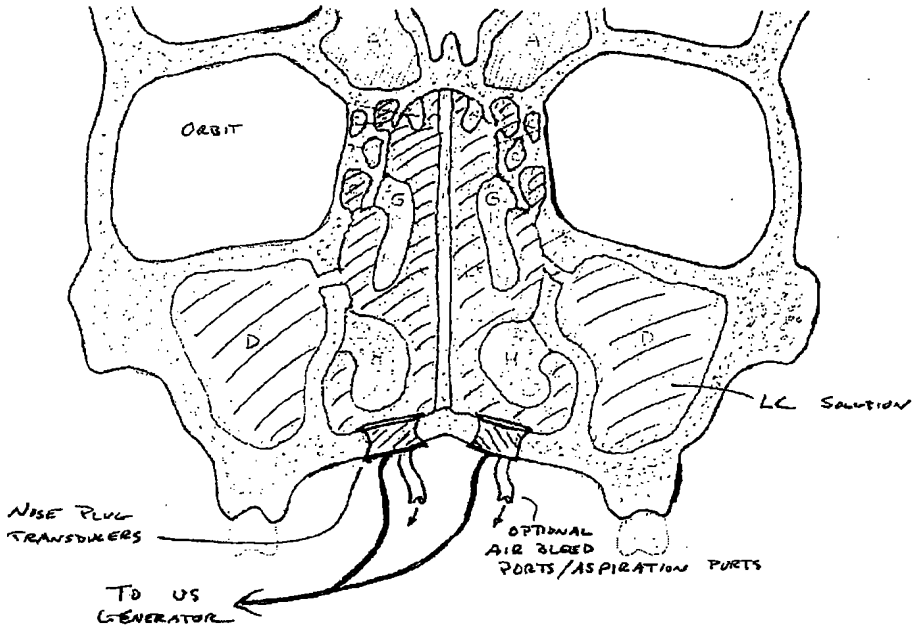


FIG. 22



LC.15

FIG. 23

LC-12

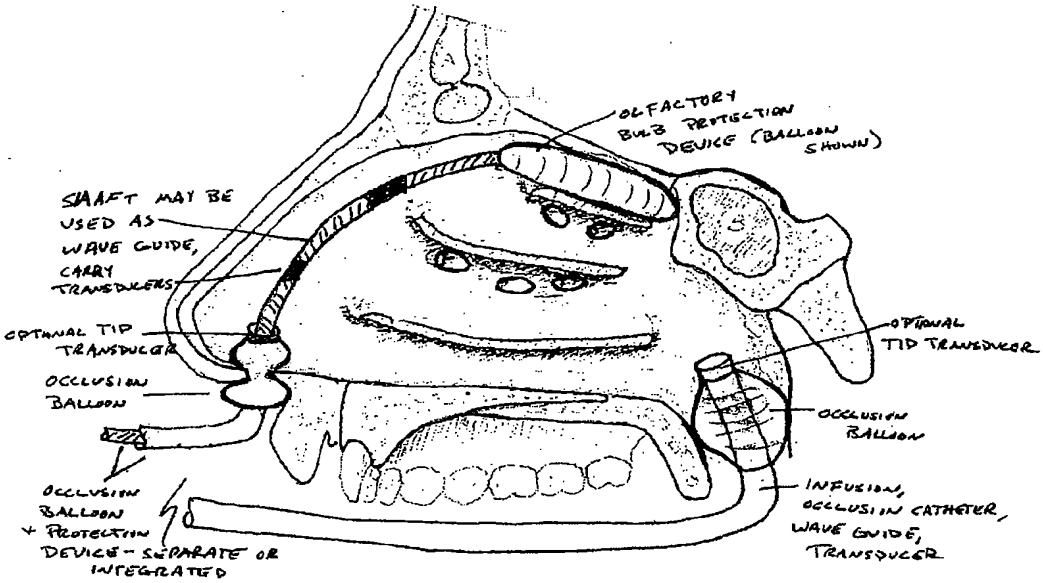
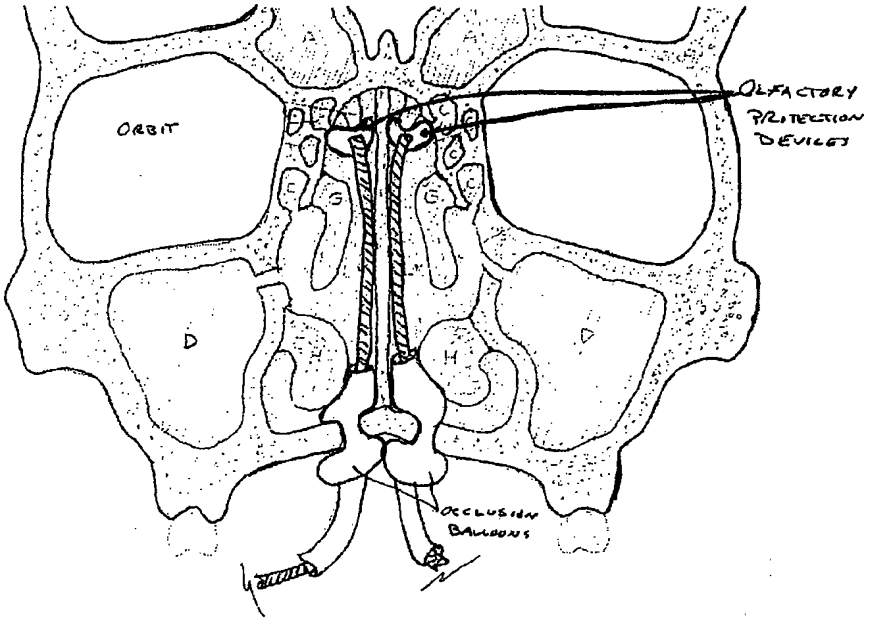


FIG. 24



LC-13

FIG. 25

LC 1

Protect,

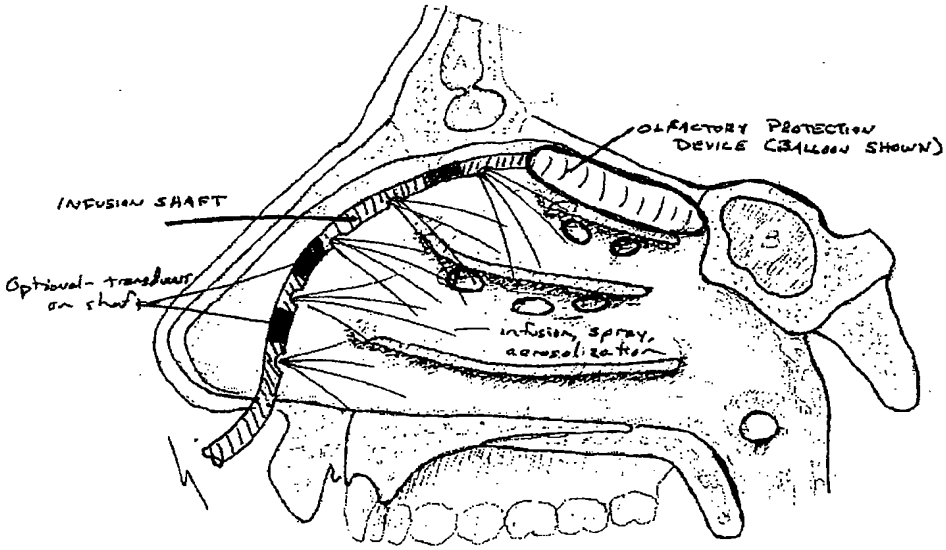
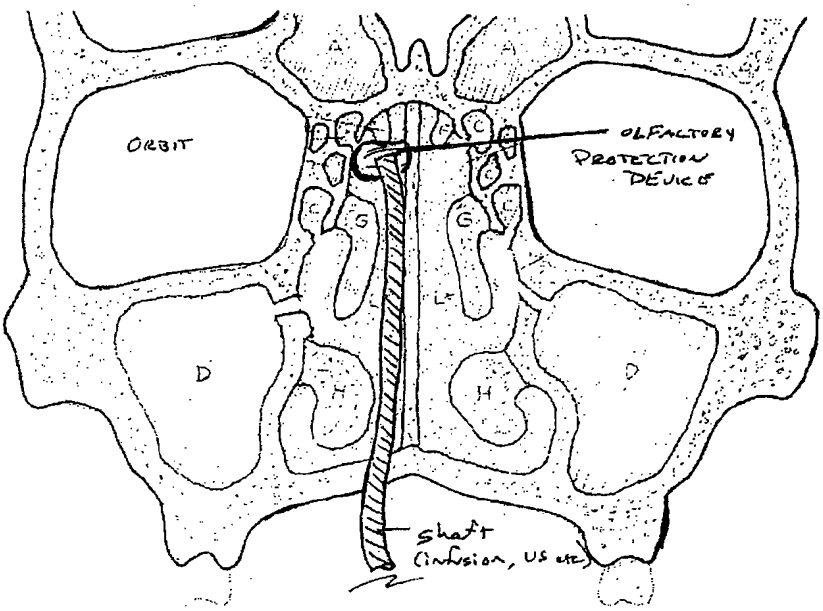
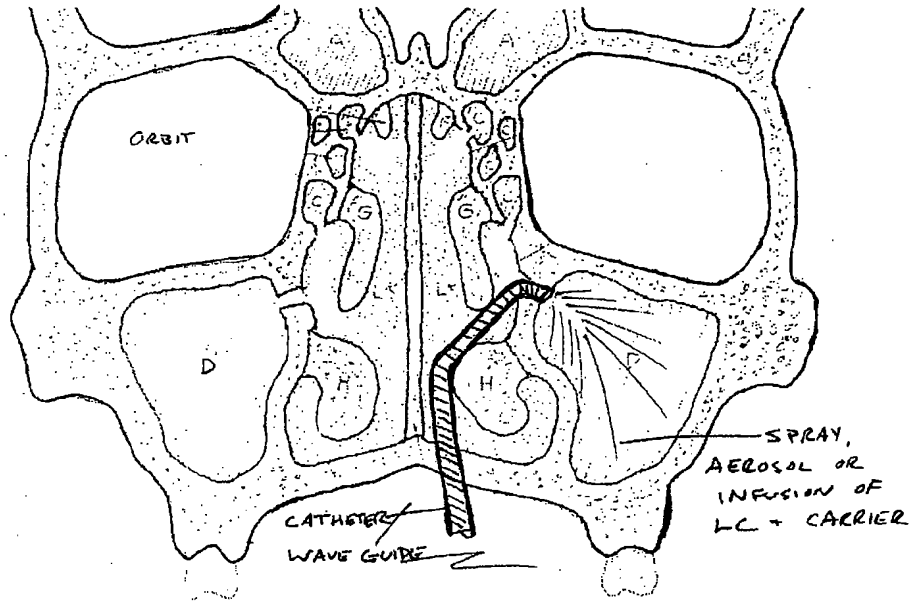


FIG. 26



LC2

FIG. 27



LC19

FIG. 28

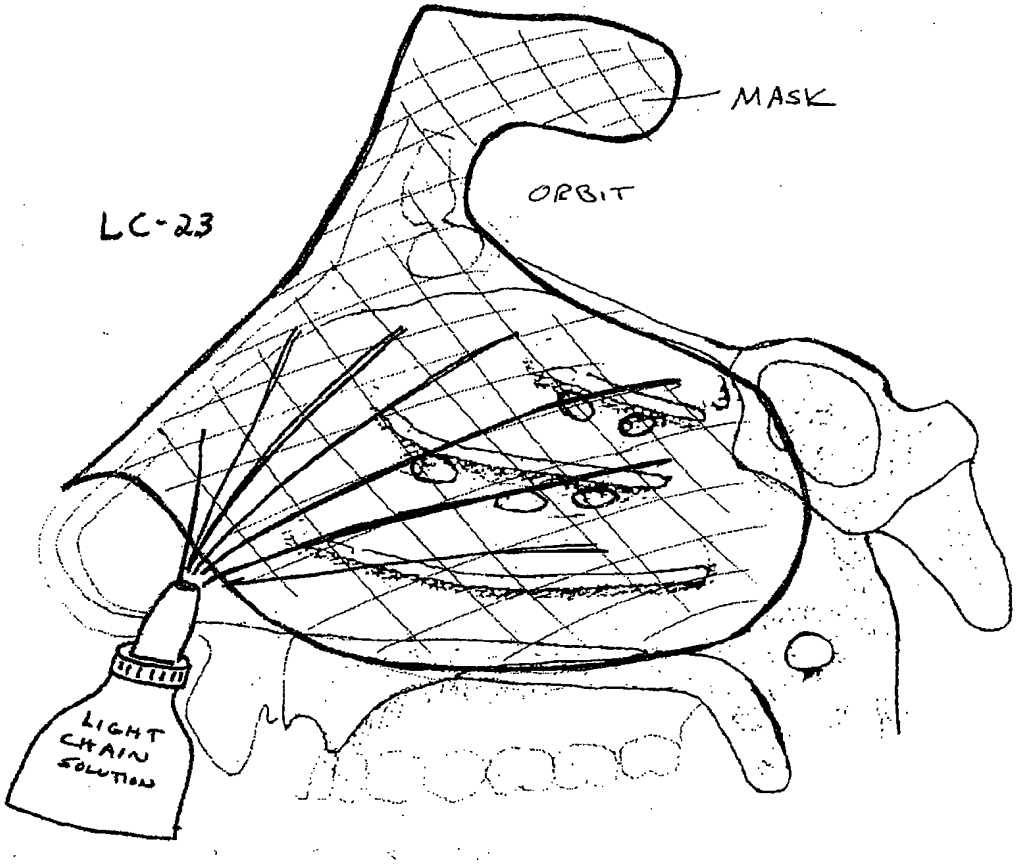
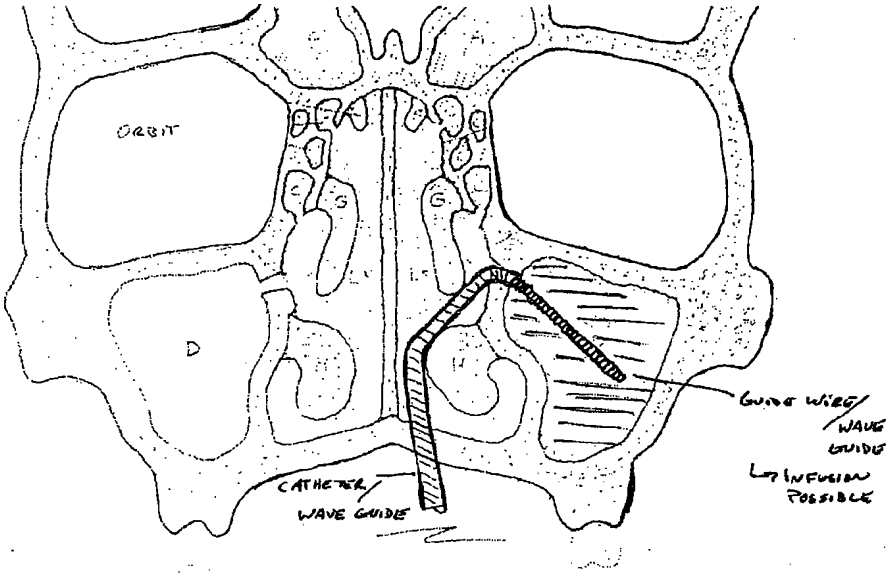
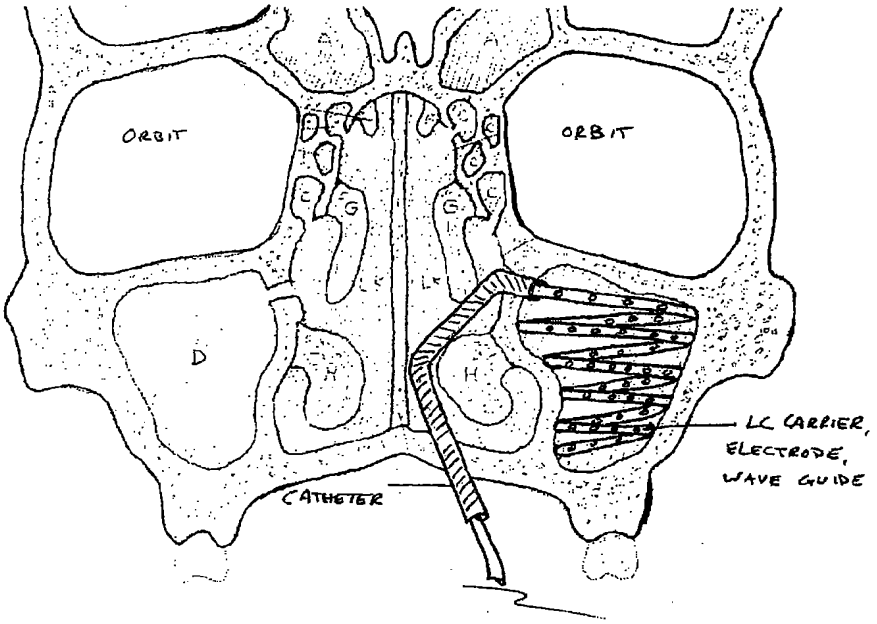


FIG. 29



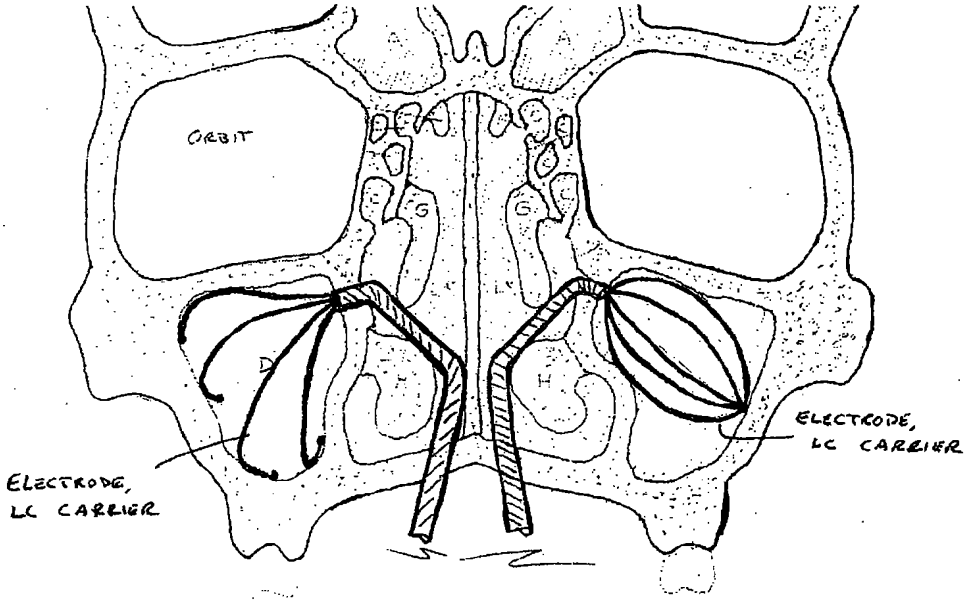
LC-18

FIG. 30



LC.17

FIG. 31



LC.16

FIG. 32

Attorney Docket No.: 020979-003700US

PROVISIONAL PATENT APPLICATION

METHODS AND SYSTEMS FOR TOXIN DELIVERY TO THE NASAL CAVITY

Inventors: MARK E. DEEM, a citizen of The United States
residing at 685 Sierra Avenue
Mountain View, California 94041; and

HANSON GIFFORD, a citizen of The United States
residing at 3180 Woodside Road
Woodside, California 94062.

Assignee: The Foundry Inc.
199 Jefferson Drive
Menlo Park, CA 94025

Status: Small Entity

60777918 v1

TOWNSEND and TOWNSEND and CREW LLP
Two Embarcadero Center, Eighth Floor
San Francisco, California 94111-3834
Tel: 650-326-2400

METHODS AND SYSTEMS FOR TOXIN DELIVERY TO THE NASAL CAVITY

CROSS-REFERENCES TO RELATED APPLICATIONS

[0001] The disclosure of this application is related to prior provisional Application No. 60/702,077, (Attorney Docket No. 020979-003400US), filed on July 22, 2005, the full disclosure of which is incorporated herein by reference.

BACKGROUND OF THE INVENTION

[0002] 1. Field of the Invention. The present invention relates generally to medical methods and systems. More particularly, the present invention relates to methods and systems for delivering toxins, such as botulinum toxin light chain fragments, to target cells in a nasal cavity.

[0003] Rhinitis, which includes the symptoms of rhinorrhea, is a condition resulting from inflammation and swelling of the patient's mucus membranes which line the nasal cavity. Rhinitis and/or rhinorrea can arise from a number of conditions, most often results from allergies to pollen, dust, seasonal allergens or other airborne substances, but can also be caused by anatomic pathologies such as blockages (as in the case of sinusitis). Symptoms may include sneezing, itching, nasal congestion, and a runny nose.

[0004] While numerous treatments for rhinitis have been proposed over the years, no single treatment is optimum for all patients or all conditions. Most commonly, hay fever and other forms of rhinitis are treated with antihistamines which block the inflammatory response. While effective, many antihistamines can cause drowsiness, are effective for only a limited amount of time, and present the patient with an on-going cost to continuously purchase the drugs.

[0005] Recently, a longer term therapy for rhinitis which relies on the use of botulinum toxin for blocking mucus production by mucus-producing cells in the nasal membrane have been proposed. Botulinum and other neurotoxins are capable of disabling adrenergic cells, including goblet cells which are responsible for the majority of mucus production in the nasal cavity membrane. Dr. Ira Sanders has demonstrated that introduction of intact botulinum toxin molecules into the nasal passages of canines can reduce mucus secretion by a significant amount.

[0006] While the experimental work of Dr. Sanders holds promise for long term rhinitis treatment, it faces a number of challenges before it is suitable for wide spread use in humans. In particular, botulinum toxin is a neurotoxin which could have significant negative effects on a patient if accidentally released outside of the targeted nasal passages. Inadvertent distribution of the toxin to muscles of the oropharynx, mouth, tongue, or elsewhere could result in serious complications to the patient. Additionally, the use of botulinum-soaked gauze pads for delivering the toxin to the nasal cavities, as demonstrated by Dr. Sanders, will have limited ability to uniformly and selectively deliver the botulinum to the regions having high concentrations of preferred target cells, such as goblet cells in the nasopharynx.

[0007] For these reasons, it would be desirable to provide improved methods and systems for delivering toxins, such as botulinum and active botulinum fragments, to the nasal membrane of a patient, particularly a patient suffering from rhinitis or other conditions associated with nasal inflammation and conditions, such as sinus headaches and migraine headaches. The methods and systems should be capable of providing for selective and repeatable delivery of the toxins to defined target areas within the nasal cavities, including particular paranasal sinuses, the nasopharynx, and in some cases substantially the entire nasal cavity. The systems and methods should provide for the safe and effective delivery of the toxins, and in particular should reduce or eliminate the risk of toxin being delivered to non-targeted tissues outside of the nasal cavity. At least some of these objectives will be met by the inventions described hereinbelow.

[0008] 2. Description of the Background Art. U.S. Patent No. 5,766,605, to Sanders et al. has been described above. Sharri et al. (1995) *Otolaryngol. Head Neck Surg.* 112: 566-571 also reports the work of Dr. Sanders described in the '605 patent. Ünal et al. (2002) *Acta Otolaryngol* 123: 1060-1063 describes the injection of botulinum toxin A into the turbinates of patients suffering from allergic rhinitis. See also, U.S. 6,974,578. The purification and possible therapeutic uses of botulinum light chain are described in US2004/0151741, US2005/0019346, and Chaddock et al. (2002) *Protein Expression and Purification* 25: 219-228. Energy-medicated transdermal delivery of intact botulinum toxin is suggested in US2005/007441 and 2004/0009180. The use of catheters and other devices for the energy-medicated delivery of botulinum light chain is described in commonly owned co-pending provisional application 60/702,077 (Attorney Docket No. 020979-003400US, filed July 22, 2005, the full disclosure of which has previously been incorporated herein by reference.

BRIEF SUMMARY OF THE INVENTION

[0009] The present invention provides treatments for any disease or condition for which rhinorrhea is a result or symptom.

[0010] Rhinorrhea is the term describing the eflluence of mucus from the lining of the nasal passages, nasopharynx, or paranasal sinuses. Rhinorrhea can be a symptom of a number of diseases such as the common cold, sinusitis or rhinitis, Rhinitis (inflammation of the airways) falls into two major categories – allergic and non-allergic (or vasomotor) rhinitis. Each can have several subcategories. Sinusistis is an infection or inflammation of the paranasal sinuses. Sinusitis may have a number of different causes, and can be the result of chronic inflammation of the nasal passages, for example as a result of chronic rhinitis.

[0011] Allergic rhinitis is an immunologic response modulated by IgE and characterized predominantly by sneezing, rhinorrhea, nasal congestion, and pruritus of the nose. It may be seasonal (a condition commonly referred to as hay fever) or perennial. The seasonal form is caused by allergens released during tree, grass, or weed pollination, whereas the perennial form is caused by allergies to animal dander, dust mites, or mold spores with or without associated pollinosis. Data also suggest that urban air pollutants from automobiles and other sources may have an adjunctive effect. (from attached articles by Nathan et al)

[0012] Nonallergic rhinitis is a diagnosis of rhinitis without any immunoglobulin E (IgE) mediation, as documented by allergen skin testing. Hence, the rhinorrhea, sneezing, pruritus, and congestion do not result from allergy or hypersensitivity and continue to persist, whether continuously or sporadically. Nonallergic rhinitis affects 5-10% of the population. Nonallergic rhinitis has 7 basic subclassifications, including infectious rhinitis, nonallergic rhinitis with eosinophilia syndrome (NARES), occupational rhinitis, hormonal rhinitis, drug-induced rhinitis, gustatory rhinitis, and vasomotor rhinitis. Patients may or may not present with the same symptoms seen in allergic rhinitis.

[0013] According to the present invention, botulinum toxin, ricin, exotoxin A, diphtheria toxin, cholera toxin, tetanus toxin, other neurotoxins, and active fragments thereof are delivered to a patient's nasal membrane while applying energy to target cells within the membrane under conditions which cause a reversible (or in some instances non-reversible) poration of the cell membranes to enhance delivery of the toxin into the cells. The region where the toxin is introduced may comprise any portion of the nasal cavity, such as a single paranasal sinus or portion thereof, a main nasal passage, two or more paranasal sinuses, or in

some cases may comprise substantially the entire nasal cavity of the patient. A particular target region for the toxin may comprise the nasopharynx which is at the back of the nasal passage. The nasopharynx comprises a cluster of goblet cells which are responsible for mucus secretion and which are susceptible to the disabling mechanism of the botulinum toxin and other neurotoxins.

[0014] The energy is preferably selectively applied to a targeted region containing a variety of cell types, including goblet cells, epithelial cells, ciliated and non-ciliated columnar cells, basal cells, and less or no energy applied to untargeted regions. It will be appreciated that the energy may be applied to regions of the nasal membrane which are the same or different from the regions to which the toxin has been introduced. By controlling the delivery area of both the toxin delivery and the energy delivery, the methods and apparatus of the present invention can more specifically target the goblet and other recipient cells of interest while minimizing the amount of toxin which enters non-targeted cells. That is, only those cells in the nasal membrane which are exposed to both the toxin and the applied energy will preferentially be permeablized or porated to receive the toxin within the cytoplasm of the cell.

[0015] The toxin to be delivered may comprise any neurotoxin capable of disabling mucus secretion in goblet cells and other mucus-producing nasal cells. Preferably, the toxin comprises botulinum toxin, although other toxins such as ricin, exotoxin A, diphtheria toxin, cholera toxin, tetanus toxin, other neurotoxins, and active fragments thereof may also find use. In preferred aspects of the present invention, only an active fragment of the toxin will be delivered to the nasal cavity. Botulinum toxin and the other toxins listed above commonly comprise both a heavy chain and a light chain. The heavy chain is responsible for binding to the target cells and mediating passage of the light chain into the cytoplasm of the target cells. By delivering only the light or active chain of these toxins (after removal of the heavy chain or recombinant production of only the light chain), the risk of accidental delivery of the toxin to non-target cells is greatly reduced. Delivery of the active or light chain fragments into the target cells, according to the present invention, is mediated and enhanced by the selective application of an energy which porates the cell membrane to allow entry of the light chain or active fragment. The presently preferred botulinum light chain fragment may be derived from any one of the seven presently known botulinum types A-G.

[0016] Any type of energy which is capable of reversibly permeablizing or porating the cell wall to allow passage of the toxin molecule, either whole toxin or preferably light chain

fragment, into the cell cytoplasm may be applied to the cell membrane. Thus, energy may comprise various forms of electrical pulses, acoustic pulses, X-ray energy, microwave energy, or the like, and combinations thereof. Preferably, the energy will be either pulsed electrical energy of the type which is commonly used for cellular electroporation or will be ultrasonic energy of the type commonly employed for sonoporation of cells. The energy may be applied using the same catheters or other structures which are used for delivering the toxins. Alternatively, the energy may be applied using separate external or internal sources, such as using separate external ultrasonic transducers and/or ultrasound wave guides capable of delivering focused or unfocused ultrasound into the target tissues of the nasal cavity.

[0017] In specific embodiments of the methods of the present invention, the toxin may be introduced to the target region through a catheter. For example, the catheter may carry a balloon which engages the nasal membrane in order to effect delivery of the toxin to the target cells. In a particular example, the balloon is porous over at least a portion of its area so that the toxin may be released to specific areas of the nasal membrane, typically being incorporated into a suitable liquid, gel, or other fluid or fluidizable carrier. In other embodiments, the toxin may be introduced through one or more needles carried on the catheter, and in still other embodiments the toxin may be aerosolized from a small port, nozzle, or other orifice or structure on the catheter.

[0018] While the energy may be applied from a separate external source, as generally described above, the energy will most often be applied from the same catheter or other apparatus used to deliver the toxin. For example, when ultrasonic or other acoustic energy is being applied, the transducer may be on or associated with the catheter. In a particular example, it is shown that the transducer may be located within or beneath the porous balloon which is used to deliver toxin to the nasal membrane. When electrical energy is used for poration, the electrodes may be on the catheter within or surrounding the region which delivers the energy to the nasal membrane. In other instances, the energy may be applied from a separate catheter or other device adapted for intrabronchial introduction. In still other instances, the energy application will apply energy transcutaneously, for example from the skin of the face, typically surrounding the nose over the sinus cavities.

[0019] In addition to the methods described above, the present invention further provides systems for delivering toxins to Goblet and other target cells as defined above in a nasal membrane. The systems may typically comprise a catheter adapted to introduce a toxin to a

region adjacent to the target cells. An energy applicator is further provided for applying energy to the target cells under conditions which cause a reversible poration of the cell membranes to enhance delivery of the toxin. Systems may still further comprise a source of the toxin suitable for introduction from or through the catheter. The energy applicator may be mounted on or incorporated within the catheter, or may be a separate or intrabronchial external source. In an exemplary embodiment illustrated below, an external applicator may comprise a mask or other structure which fits over the nose and/or sinus region of the patient and which is capable of delivering acoustic energy to the target cells within the target regions.

[0020] When the energy applicator is incorporated with or within the catheter, the delivery pattern of the energy will usually be at least partially overlapping with the toxin delivery pattern of the catheter. For example, when a porous balloon is used for toxin delivery, the acoustic transducer, electroporation electrodes, or the like, will usually be disposed to deliver energy which at least partly overlaps with the dispersion pattern of the toxin. In some instances, the region of applied energy will be coextensive with the region of toxin dispersion. In other instances, the two regions will only partially overlap. In the latter case, the delivery of the toxin will be enabled or enhanced principally within the regions of overlap.

BRIEF DESCRIPTION OF THE DRAWINGS

[0021] Figure 1 – depicts a schematic of the creation of neurotoxin Botulinum Toxin Type A (BoNT/A), including the light chain (LC) fragment or portion.

[0022] Figure 2A – depicts a schematic of a target cell, including the cell membrane, and inner cellular matrices.

[0023] Figure 2B – depicts a schematic of the target cell wherein LC molecule has been introduced.

[0024] Figure 3A-3B – depicts a the target cell of Figure 2 showing application of an energy field (EF) to produce permeabilization or pores (P) in the cell membrane, and introduction of the LC fragment therethrough.

[0025] Figure 4 – depicts a schematic of a cell wherein the energy field has been discontinued, and neurotransmission of the cell has been effectively blocked.

[0026] Figures 5, 5A-5B – depicts various embodiments of a delivery device of the present invention utilizing multiple energy transmission elements and an energy transmission system.

[0027] Figures 6A-D – depict various electrode catheter configurations adapted to deliver energy or energy and therapeutic agents to target tissue.

[0028] Figure 7 - depicts an embodiment of the present invention utilizing an ultrasound element on a catheter device.

[0029] Figure 8 – depicts an embodiment of the present invention utilizing an aerosolizing element.

[0030] Figure 9 - depicts use on an external hand held transducer for enhancing cellular uptake of toxin delivered from a separate nasal aerosolizer.

[0031] Figures 10 and 11- depicts depict use of balloon catheters for delivering toxin to the nasopharynx.

[0032] Figures 12A-12C - depict use of a self-expanding toxin delivery structure on a catheter.

[0033] Figures 13 and 14 - depict a protocol for limiting toxin introduction by partial filling of a porous delivery balloon.

[0034] Figure 15 and 16 - depict sizing of a delivery balloon to control distribution of toxin released into the nasal cavity.

[0035] Figure 17 - depicts the use of multiple small balloons for selective toxin delivery into the nasal cavity.

[0036] Figures 18 and 19 - depict sonoporation using an external mask placed over the sinuses and nose.

[0037] Figures 22 and 23 - depict nose plugs for occluding and optionally delivery poration energy to the nasal cavity.

[0038] Figures 24 and 25 - depict an alternate occlusion catheter system for targeted toxin delivery to the nasopharynx.

[0039] Figures 26 and 27 - depict use of a toxin delivery catheter having side holes and a distal occlusion balloon for isolating and protecting the olefactory bulb.

[0040] Figures 28 and 30- depict use of a simple catheter having a shaped distal end for aerosolizing a toxin into a target nasal sinus though an ostium open to the sinus.

[0041] Figure 29 - depicts toxin delivery using a nasal spray and energy delivery using a face mask.

[0042] Figures 31 and 32, depict use of a catheter having a shaped distal end for positioning separate infusion structures with a target sinus cavity.

DETAILED DESCRIPTION OF THE INVENTION

[0043] The present invention is directed at methods and systems for delivering toxins to target cells within a patient's nasal cavity. The toxins may be intact toxins, such as botulinum toxin, ricin, exotoxin A, diphtheria toxin, cholera toxin, tetanus toxin, other neurotoxins, and active fragments thereof. Each of these toxins comprise a heavy chain responsible for cell binding and a light chain having enzyme activity responsible for cell toxicity.

[0044] Botulinum toxin blocks acetylcholine release from cells, such as the goblet cells in the nasal membranes responsible for mucus hypersecretion, and can thus be effective even without energy-mediated delivery in accordance with the principles of the present invention. The use of energy to permeablize or porate the cell membranes of the goblet cells or other mucus-secreting cells of the nasal lining, in accordance with the present invention, allows botulinum and other toxins to be preferentially delivered to the targeted goblet and other mucus-producing cells. Additionally, it allows use of the active or light chains of these toxins (having the heavy chains removed or inactivated) for treatments in accordance with the present invention. Normally, the light chains when separated from the cell-binding heavy chains of botulinum and the other toxins are incapable of entering the cells and thus will be free from significant cell toxicity. By using the energy-mediated protocols of the present invention, the toxin light chains may be locally and specifically introduced into the target cells located within defined regions of the nasal membrane. Thus, even if the toxin fragments are accidentally dispersed beyond the desired target regions, the fragments will not generally enter cells without the additional application of cell permeablizing or porating energy. For that reason, the toxin delivery methods of the present invention are particularly safe when performed with toxin fragments, such as the light chain of botulinum and other toxins.

[0045] While the remaining portion of this disclosure will be presented with specific reference to the botulinum toxin light chain, it will be appreciated that the energy-mediated delivery protocols and systems may also be used with other intact toxins and in particular with other light chain toxin fragments as just discussed.

[0046] Generally, the botulinum toxin molecule (BoNT) is synthesized as a single polypeptide chain of 150kD molecular weight. The neurotoxin is then exposed to enzymes, either during cultivation of the *Clostridium botulinum* organism or subsequent to purification of the toxin, wherein specific peptide bonds are cleaved or "nicked" resulting in the formation of a dichain molecule referred to as BoNT. As shown in Figure 1, dichain neurotoxin is composed of a light chain region 50kD molecular weight linked by disulfide bonds to a heavy chain 100kD molecular weight (Kistner, A., Habermann, E. (1992) *Naunyn Schmiedebergs Arch. Pharmacol.* **345,** 227-334). When the light chain is separated from the heavy chains of botulinum toxin, neither chain is capable of blocking neurotransmitter release, however, the light chain alone is capable of blocking acetylcholine release if transported directly into the cell cytosol. (Ahnert-Hilger, G., Bader, M.F., Bhakdi, S., Gratzl, M. (1989) *J. Neurochem.* **52,** 1751-1758 and Simpson, L.L. (1981) *Pharmacol. Rev.* **33,** 155-188.) Focusing on the light chain, the isolation or separation process essentially renders the light chain "non-toxic" in a general environment, while still maintaining its effect or toxicity, once it is transported through the target cell membrane.

[0047] Over the past several years, the separation and purification of the light chain and heavy chain of BoNT has seen significant development activity. In the case of the heavy chain (HC), researchers are interested in its ability to bond with a target cell and deliver certain molecules into that cell. For example, various drug delivery applications have been suggested, for example, using the HC to bind to tPA so that a patient could inhale the HC-bound tPA allowing it to cross the membrane of the lungs and be transported into the bloodstream for anticoagulation. Of particular interest to the present invention are the efforts to isolate and purify the light chain (LC) of the botulinum molecule. In its isolated and purified form, all HC elements are removed, rendering the LC incapable of crossing the cell membrane without assistance. This renders the LC a non-toxic protein to the cell environment, while still maintaining its encoded toxicity by, once it is effectively delivered to its appropriate catalytic environment; the cell cytosol.

[0048] Various groups have been active in the area of isolation and purification. For example, companies such as Metabiologics, a group affiliated with the University of Wisconsin, the Center for Applied Microbiology and Research (CAMR), a division of the UK Health Protection Agency, List Biological Laboratories, Inc. of California, and other research groups throughout the world. Many of these companies provide purified preparations of

botulinum neurotoxins from *Clostridium botulinum* types A and B. List Laboratories in particular provides recombinantly produced light chains from both types A, B, C, D and E.

[0049] For purposes of this specification, the terms“poration” or "permeablization" includes various forms of electrically-medicated poration, such as the use of pulsed electric fields (PEFs), nanosecond pulsed electric fields (nsPEFs), ionophoreseis, electrophoresis, electropermeabilization, as well as other energy mediated permeabilization, including sonoporation (mediated by ultrasonic or other acoustic energy), and/or combinations thereof, to create temporary pores in a targeted cell membrane. Similarly, the term “electrode” or “energy source” used herein, encompasses the use of various types of energy producing devices, including x-ray, radiofrequency (RF), DC current, AC current, microwave, ultrasound, adapted and applied in ranges to produce membrane permeabilization in the targeted cell.

[0050] Reversible electroporation, first observed in the early 1970’s, has been used extensively in medicine and biology to transfer chemicals, drugs, genes and other molecules into targeted cells for a variety of purposes such as electrochemotherapy, gene transfer, transdermal drug delivery, vaccines, and the like.

[0051] In general, electroporation may be achieved utilizing a device adapted to activate an electrode set or series of electrodes to produce an electric field. Such a field can be generated in a bipolar or monopolar electrode configuration. When applied to cells, depending on the duration and strength of the applied pulses, this field operates to increase the permeabilization of the cell membrane and reversibly open the cell membrane for a short period of time by causing pores to form in the cell lipid bilayer allowing entry of various therapeutic elements or molecules, after which, when energy application ceases, the pores spontaneously close without killing the cell after a certain time delay. As characterized by Weaver, *Electroporation: A General Phenomenon for Manipulating Cells and* Tissues Journal of Cellular Biochemistry, 51:426-435 (1993), short(1-100μs) and longer (1-10 ms) pulses have induced electroporation in a variety of cell types. In a single cell model, most cells will exhibit electroporation in the range of 1-1.5V applied across the cell (membrane potential).

[0052] In addition, it is known in the art that macromolecules can be made to cross reversibly created pores at voltages of 120V or less applied to cells for durations of 20 microseconds to many milliseconds. For applications of electroporation to cell volumes, ranges of 10 V/cm to 10,000 V/cm and pulse durations ranging from 1 nanosecond to 0.1 seconds can be applied. In one example, a relatively narrow (μsec) high voltage (200V)pulse can be followed by a longer

(>msec) lower voltage pulse (<100V). The first pulse or pulses open the pores and the second pulse or series of pulses assist in the movement of the BoNT-LC across the cell membrane and into the cell.

[0053] Certain factors affect how a delivered electric field will affect a targeted cell, including cell size, cell shape, cell orientation with respect to the applied electric field, cell temperature, distance between cells (cell-cell separation), cell type, tissue heterogeneity, properties of the cellular membrane and the like.

[0054] Various waveforms or shapes of pulses may be applied to achieve electroporation, including sinusoidal AC pulses, DC pulses, square wave pulses, exponentially decaying waveforms or other pulse shapes such as combined AC/DC pulses, or DC shifted RF signals such as those described by Chang in *Cell Poration and Cell Fusion using an Oscillating Electric Field*, Biophysical Journal October 1989, Volume 56 pgs 641-652, depending on the pulse generator used or the effect desired. The parameters of applied energy may be varied, including all or some of the following: waveform shape, amplitude, pulse duration, interval between pulses, number of pulses, combination of waveforms and the like.

[0055] There are at least two general power categories of medical ultrasound waves. One category of medical ultrasound wave is high acoustic pressure ultrasound. Another category of medical ultrasound wave is low acoustic pressure ultrasound.

[0056] Acoustic power is expressed in a variety of ways by those skilled in the art. One method of estimating the acoustic power of an acoustic wave on tissue is the Mechanical Index. The Mechanical Index (MI) is a standard measure of the acoustic output in an ultrasound system.

[0057] High acoustic pressure ultrasound systems generally have a MI greater than 10. Low acoustic pressure systems generally have a MI lower than 5. For example, diagnostic ultrasound systems are limited by law to a Mechanical Index not to exceed 1.9.

[0058] Another measurement used by those skilled in the art is the spatial peak, peak average intensity (Isppa). The intensity of an ultrasound beam is greater at the center of its cross section than at the periphery. Similarly, the intensity varies over a given pulse of ultrasound energy. Isppa is measured at the location where intensity is maximum averaged over the pulse duration. Isppa for high acoustic pressure or high intensity focused ultrasound (HIFU)

applications ranges from approximately 1500W/cm2. to 9000 W/cm2. Diagnostic ultrasound equipment, for instance, will generally have, and an Isppa less than 700 W/cm2.

[0059] Yet another way in which ultrasound waves can be characterized is by the amplitude of their peak negative pressure. High acoustic pressure or HIFU applications employ waves with peak amplitudes in excess of 10MPa. Low acoustic pressure ultrasound will generally have peak negative pressures in the range of 0.01 to 5.0 MPa . Diagnostic ultrasound equipment, for example, will generally have a peak amplitude less than 3.0MPa.

[0060] Both high and low acoustic pressure ultrasound systems generally operate within the frequency range of 20KHz – 10.0 MHz Interventional applications (such as in blood vessels) operate clinically up to about 50 MHz. Also ophthalmologic applications up to about 15 MHz. Diagnostic imaging typically uses frequencies of about 3 to about 10 MHz. Physical therapy ultrasound systems generally operate at frequencies of either 1.0MHz or 3.3MHz.

[0061] High acoustic pressure ultrasound or high intensity focused ultrasound has been used for tissue disruption, for example for direct tumor destruction. High intensity focused ultrasound using high acoustic pressure ultrasound is most commonly focused at a point in order to concentrate the energy from the generated acoustic waves in a relatively small focus of tissue.

[0062] Systems for permeabilization of target tissue cell membranes may employ either high acoustic pressure or low acoustic pressure ultrasound. Some embodiments may preferably employ relatively low acoustic pressure, for example the systems described herein where the transducers are mounted on the delivery devices and operate inside the body. Other systems may operate at interim acoustic pressure ranges. For example, systems described herein which employ an external ultrasound generator and transducer and which conduct the ultrasound to the target tissues through the use of a wave guide. In these systems, losses due to transduction through the wave guide can be compensated for by increasing the input power to the wave guide until adequate power is delivered to the target tissue. Finally, some systems described herein may employ focused or partially focused higher pressure ultrasound, for example the systems which employ an external mask to conduct the ultrasonic power through the tissues to the target tissues. It should be appreciated that combinations of high and low acoustic pressure systems may also be employed.

[0063] A schematic example of the methods of the present invention are shown in Figures 2-4 in a simplified single cell model. A targeted cell, e.g., a goblet cell of the type which line the

nasal cavity membrane, is shown in Figure 2A. Fragmented neurotoxin such as BoNT-LC is introduced into the vicinity of the targeted cell as depicted in Figure 2B. An energy field (EF) is applied in accordance with the present invention resulting in the transfer of the BoNT-LC to the intracellular matrix (cytosol or cytoplasm) as shown in Figures 3A and 3B. Once this transfer has occurred, the release of acetylcholine from the presynaptic neurons at the neuromuscular junctions of the goblet or other target cells is then blocked or disrupted. Once energy application is discontinued, the pores in the cell membrane recover or close as depicted in Figure 4.

[0064] The terms "poration" and "permeablization" will also cover forms of cellular sonoporation. Just as pulses of high voltage electricity can open transient pores in the cell membrane, ultrasonic energy can do the same. See for example Guzman et al. "Equilibrium Loading of Cells with Macromolecules by Ultrasound: Effects of Molecular Sizing and Acoustic Energy", *Journal of Pharmaceutical Sciences*, 91:7, 1693-1701, which examines the viability of ultrasound to deliver molecules of a variety of sizes into target cells. In addition, techniques for nebulizing fluids and aqueous drugs are well known in the art, and as such, devices of the present invention may be adapted to introduce a BoNT-LC solution to a target region, such as the lung and then effect selective membrane transport of the BoNT-LC into the cell using sonoporation.

[0065] To achieve the goals of the present invention, it may be desirable to employ methods and apparatus for achieving cell membrane permeabilization via the application of an energy source, either from a catheter located directly in the vicinity of the targeted cells, or an externally focused energy system. For purposes of this specification, the term "catheter" may be used to refer to an elongate element, hollow or solid, flexible or rigid and capable of percutaneous introduction to a body (either by itself, or through a separately created incision or puncture), such as a sheath, a trocar, a needle, a lead. Further descriptions of certain electroporation catheters are described in United States Provisional Patent Application No. 60/701,747 (Attorney Docket No. 020979-003500US) filed July 22, 2005, the full disclosures of which are expressly incorporated herein by reference.

[0066] Figures 5 and 5A-5B depict a system utilizing an electroporation catheter for selective electroporation of targeted cells. In certain configurations of the present invention, voltages may be applied via the electroporation catheter to induce reversible electroporation at the same time as the catheter delivers the fragmented neurotoxin to the targeted region.

[0067] Referring to Figure 5, electroporation catheter system comprises a pulse generator such as those generators available from Cytopulse Sciences, Inc. (Columbia, MD) or the Gene Pulser Xcell (Bio-Rad, Inc.), or IGEA (Carpi, Italy), electrically connected to a catheter having a proximal end and a distal end and adapted for minimally invasive insertion into the desired region of the body as described herein. The catheter further comprises an electroporation element at the distal end thereof. The electroporation element consists of a first and second electrode operatively connected to the pulse generator for delivering the desired number, duration, amplitude and frequency of pulses to affect the targeted cells. These parameters can be modified either by the system or the user, depending on the location of the catheter within the body (intervening tissues or structures), and the timing and duration of reversible cell poration desired.

[0068] Figure 5A depicts an arrangement of electrodes that produces an electric field concentrated in a lateral direction from the catheter body whereas, Figure 5B shows a device constructed to create a more uniform electric field about the shaft of the catheter body. Further catheter device and electrode configurations are shown in Figures 6A-6D, including, Figure 6A depicting an elongate catheter having a first and second electrode near the distal tip thereof, and including a monitoring or stimulation electrode in the vicinity of the active porating electrodes for localizing the treatment area. In some embodiments, the monitoring or stimulating function may be performed by one or more of the treatment electrodes. The catheter device may have an optional sharp tip to facilitate percutaneous introduction. Figure 6B is a similar catheter device, but is further adapted to be steerable, or articulate at a region near the distal end of the device. Such steering ability enables the operator to introduce the device into tight or tortuous spaces (such as the bronchial passages, or cardiovascular vessels) so that optimal placement of the device at the target location may be achieved.

[0069] Figure 6C depicts a further embodiment of the catheter device described above, that includes an injection element to allow for the injection of a therapeutic agent such as a fragmented neurotoxin before, during or after the application of the pulsed energy or electroporation. The injection element may be a needle as shown in Figure 6C, an infusion port, or other infusion means.

[0070] Figure 6D depicts an alternative embodiment of the present invention, showing a catheter device having electrode elements that are adapted to extend laterally from the main catheter body, and in some cases, penetrate the surrounding tissue prior to application of

energy. In doing so the depth and direction of the energy field created by the electroporative process, may be further controlled.

[0071] Figure 7 depicts an embodiment of the present invention utilizing an ultrasonic element that may be particularly useful in delivery of the BoNT-LC to nasal tissue that provides a broad but targeted transport of the LC across the epithelial and goblet cell walls. In this device, ultrasound energy is delivered to the distal end of the catheter device via an ultrasonic waveguide that is operatively connected to an ultrasound energy source (U/SES). The LC fragment would be delivered via the same lumen as the waveguide, or via a separate lumen that exits the distal tip of the device. In operation, the ultrasonic energy would cause the LC solution to be nebulized, forming a mist within the lung, as shown in Figure 8. The mist itself, in the appropriate concentrations, may act as an ultrasound coupler, conveying the ultrasonic energy to the wall of the lung or other targeted cellular structures, causing sonoporation of the targeted cells whereby the LC fragment is transmitted across the cell membranes to become an effective neurotransmitter blocker. In an alternative embodiment, an ultrasonic transducer may be located directly at the tip of the delivery device, eliminating the need for a wave guide. Various catheters useful for delivering vibrational energy to tissue are described in United States Patent 6,361,554 and 6,464,680 to Brisken, the contents of which are expressly incorporated herein by reference in their entirety, for various therapeutic effects, such as enhancing cellular absorption of a substance.

[0072] Any of the catheter devices described herein, or described in the contemporaneously filed United States Provisional Patent Application No. 60/701,747 (Attorney Docket No. 020979-003500US) previously incorporated by reference in its entirety, may be adapted to include an energy delivery element such as those described herein for purposes of providing a membrane transport system for delivery of a toxin fragment of neurotoxin. In addition, certain catheter devices and methods such as those set forth in United States Patents 5,964,223 and 6,526,976 to Baran may be adapted to include energy transmission elements capable of producing a porative effect at the cellular level, including electrodes, ultrasonic elements and the like, for treatment in the respiratory tract.

[0073] Furthermore, any of the foregoing systems may include electrodes or other monitoring systems either located on the treatment catheter, or external to the patient, to determine the degree of treatment to the region, including, thermocouple, ultrasound transducers, fiberoptics, sensing or stimulating electrodes. Further, it may be desirable to

incorporate multiple pairs of electrodes that may be activated in pairs, in groups, or in a sequential manner in order to maximize the desired shape of the energy field (EF) while minimizing the field strength requirements.

[0074] It is within the scope of the present invention to deliver the toxin, the energy, or both, non-invasively. For example, as illustrated in Figure 9, the patient may draw the toxin into the nasal cavity from a hand-held dispersion device DD. After a sufficient amount of the toxin has been infused into the nasal cavity, a separate hand-held transducer TD connected to an appropriate power supply PS will be energized and applied to the nasal cavities by passing the transducer over the appropriate regions of the forehead and nose. Optionally, the transducer can have a focused output so that the acoustic energy is focused in an appropriate depth beneath the skin surface. Typically, from about 0.1 cm to 2 cm.

[0075] While the toxins and porating energy of the present invention may be delivered to the nasal cavity in a variety of ways, the following provides a number of specific examples of catheters and other structures for delivering toxins to preselected portions of the nasal cavity. For example, as shown in Figure 10, a balloon catheter 100 may be provided with a porous balloon 102 at its distal end. The balloon would be porous over at least a portion of its body so that solution delivered to inflate the balloon, which would contain desired levels of the toxin or toxin fragment, would release the solution through the balloon at a controlled rate. By further providing one or more ultrasonic transducers 104 within the balloon, optionally mounted on the catheter body, ultrasonic poration energy can be delivered to the adjacent nasal membranes which are receiving the toxin solution. As illustrated in Figure 10, the toxin is being delivered to a lower surface of the inferior meatus IM to localize and enhance cellular delivery at the balloon tissue interface. Alternatively, the balloon could carry the toxin in a releasable form over its exterior surface in order to deliver to any adjacent tissue structure. In some instances, the toxin could be carried or encapsulated in delivery vesicles which are preferentially fractured by the same acoustic energy which permeablizes the cell wall. Other coatings include hydrogels, such as those produced by Surmodics, Inc., BioCoat, Inc., or the like. In some instances, it may be desirable to provide a coupling agent over and/or within the balloon in order to enhance the delivery of ultrasonic energy from the internal transducer. In still other instances, it would be possible to place polymeric transducers on or within the balloon surface in order to directly deliver ultrasonic or other acoustic energy into the adjacent tissues.

[0076] In all the above cases, the ultrasonic transducers can be configured in order to selectively deliver the energy to desired portions of the adjacent tissues. For example, in the embodiment of Figure 10, the internal transducers 104 can be configured to focus the ultrasonic energy generally upwardly (as viewed in Figure 10) in order to preferentially deliver the toxins into the inferior meatus IM while minimizing delivery elsewhere.

[0077] As a further option, the balloon could be inflated by a coupling agent in order to enhance the transmission of the ultrasonic or other acoustic energy, while the toxin solution could be infused into the treatment area before or simultaneously using either a separate lumen in the catheter or a separate tube or other delivery catheter. In this way, it would not be necessary to inflate the balloon with a relatively large volume of the toxin solution.

[0078] The balloon catheters can be introduced by any conventional technique, for example, in some instances, it may be desirable to use a guidewire to place the catheter into a desired sinus or other location, optionally using fluoroscopic, MRI or ultrasound imaging.

[0079] Referring now to Figure 11, a side view of particular balloons placed as generally shown in Figure 10, is shown in more detail. A single balloon can be around the structures H in the inferior meatus. Alternatively, a pair of balloon structures may be placed in the same space, as shown in the right hand portion of Figure 11. Optionally, the balloons could be formed from an elastic material, such as a silicone, urethane, latex, thermoplastic elastomers, or other materials where the material is treated to be appropriately porous, for example by laser drilling. Alternatively, the balloons could be formed from non-distensible materials which are pre-formed to conform to the desired target cavities. The non-distensible balloons could also be laser drilled or otherwise made permeable in order to release the toxin solutions of the present invention. Alternatively, either type of balloon could be coated with the toxin solutions, coupling solutions, or other materials which are useful in the protocols of the present invention.

[0080] Referring now to Figure 12, as an alternative to inflatable balloons, toxin delivery structures may be made to be various shapes, for example a generally "flattened" balloons, whose profile is narrower in one axis than the other, for example by placement of an internal nitinol or other elastic frame or scaffold, or a stainless steel wire than is fed into the balloon outer structure to form such shape, within a suitable porous cover or membrane. Thus, the structure 120 may be expanded by the scaffold 122 after release from a delivery tube 124. The structures can be used to deliver energy and/or toxin in any of the ways described previously

with respect to balloons, including by carrying a transducer or electrode on or within the structure and delivering a toxin solution from the interior of the self-expanding structure through a porous portion of the structure wall.

[0081] In some instances, it will be desirable to protect the olfactory bulb of the sinuses from treatment with the toxin solutions of the present invention. Referring now to Figures 13 and 14, the porous portion of a delivery balloon can be positioned so that the remaining non-porous segment is in contact with the olfactory bulb (Figure 13). Thus, when the balloon is inflated and the toxin solution delivered, it will not be directed at the tissues of the olfactory bulb.

[0082] As shown in Figure 14, which is a cross-sectional view of Figure 13, instead of rendering the top portion of the delivery balloon non-porous, it would be possible simply refrain from filling the top portion with the toxin solution and/or a coupling solution. This can be achieved by filling the balloon with a known volume of air in addition to the toxin solution. With the patient positioned appropriately, the air will fill the portion of the balloon in proximity to olfactory bulb, excluding this tissue from toxin contact. Additionally, the air bubble may act as an ultrasound insulator to inhibit energy delivery to the non-targeted or protected tissue. Thus, delivery of the toxin to the region around the olfactory bulb and/or delivery of the energy to the region around the olfactory bulb can be partially or wholly prevented.

[0083] Referring now to Figures 15 and 16, the balloon may be sized and positioned to target an area of high goblet cell concentration, for example in the back of the nasal passages in the area of the nasopharynx. By targeting this area of the nasal membrane, a high percentage of mucus-secreting goblet cells can be treated with a device which is relatively small and which may carry a relatively low infusion volume and require less energy. Moreover, the olfactory bulb is inherently protected of this technique since the balloon is positioned well away from that area. If desired, of course, additional shielding, shaping or other protective balloons could be positioned between the olfactory bulb and the toxin and energy delivering components of the present invention.

[0084] As shown in Figure 17, direct infusion and treatment of particular sinuses may be effected using relatively small occlusion balloons which occlude and isolate natural openings into those sinuses. Once the balloon is in place and the occlusion balloon employed, the toxic solution can be delivered by infusion, dispersion, or other conventional techniques. Once the

toxin solution is present in the sinus, all or a portion of the membrane of the sinus can then be treated with an external or other ultrasonic source.

[0085] For example, as shown in Figures 18 and 19, the external transducer may comprise a mask which conforms to the nose and optionally over the sinuses, where the mask carries one or more ultrasonic or other acoustic transducer adapted to deliver energy transcutaneously into the sinuses. The mask may comprise a plurality of individual transducers, which may be made from one, two, or several generally continuous piezoelectric films which are formed over or lamented within the mask. Alternatively, multiple individual piezoelectric crystal transducers can be built into the mask.

[0086] The effect of such externally applied ultrasonic energy can be enhanced by introducing microbubbles (free air) into the isolated sinuses which have been filled with toxin solution. For example, encapsulated microbubbles, which are generally useful as echocardiographic contrast agents, or specialty perfluorocarbons, are useful as such ultrasonic enhancing agents. By encapsulating the toxin molecules in spheres or bubbles, or by simply placing the spheres or bubbles in proximity to toxin molecules, the ultrasonic or other acoustic energy can be captured and stored until it is abruptly released with fracture of the sphere or bubble. Such microspheres will also act as resonance bodies as defined below.

[0087] Referring now to Figures 20 and 21, a catheter is placed at the posterior outlet of the nasal passages in the region of the nasopharynx. The catheters configured to occlude outflow from these sinuses and passages into the throat. As shown in Figure 20, a balloon catheter or other occlusion device could be configured to block such passage. As shown in Figure 20, the catheter is delivered in through the mouth and guided into the posterior portion of the nasal cavities, typically using a guidewire. Once the nasopharynx of the posterior portion of the nasal cavity is occluded, toxin solution can be infused through the occluding catheter lumen, or through a separate infusion catheter or tube, in order to treat substantially the entire sinus cavity membrane at once. When the toxin solution is introduced through the catheter at the posterior region of the cavities, it will frequently be desirable to occlude the nostrils, for example using a nasal clip.

[0088] As shown in Figures 22 and 23, specially designed nose plugs can be provided with air bleed valves which are used to occlude the nostrils in order to evacuate or bleed air from the nasal passages while filling the passages with the toxin solution. The nose plugs could optionally include ultrasonic transducers in order to deliver ultrasonic or other acoustic energy

into the solutions entrapped within the nasal cavities using the nostril plugs. Alternatively, of course, the ultrasound or other acoustic energy could be delivered from an external transducer as described previously.

[0089] Referring now to Figures 24 and 25, an alternate occlusion catheter system for nasopharynx occlusion is illustrated. An occlusion catheter is introduced through a nostril, where the tip includes an ultrasonic transducer to provide sonoporation. A nostril plug is provided proximally on the shaft of the catheter, while the cavity is blocked with a separate occlusion balloon introduced through the mouth and into the posterior nasopharynx region. The toxin solution can be introduced into the cavity through either the catheters which pass through or reside in the nostrils or the catheter which occludes the posterior nasopharynx.

[0090] Figures 26 and 27 illustrate how a catheter with side holes can be configured to deliver toxin away from the olfactory bulb, even when used alone without separate nasopharynx occlusion catheters. The catheters preferably carry an occlusion balloon or other structure near their distal ends to prevent or inhibit toxin from reaching the olefactory bulb

[0091] Use of these or other catheter devices can deliver toxin incorporated into vesicles which may be configured as "resonance" bodies, which reduce the need to fill the nasal cavities with a liquid or other form of toxin. For example, lipid microspheres which incorporate the toxin may be sprayed or aerosolized onto target surfaces of the nasal epithelium. After the lipid or other resonance bodies are attached to the targeted epithelium membrane surface(s), the ultrasound energy can be delivered from the catheter or externally through the skin in order to selectively porate the epithelial or goblet cells to enhance introduction of the toxin *vis-à-vis* resonance bodies. A protection device at the end of the shaft can be provided to shield the olfactory bulb from the toxin.

[0092] Referring to Figure 29, the toxin may be delivered as a conventional nasal spray, as mentioned hereinbefore, and the poration energy can be delivered through a face mask. The poration energy might alternatively be delivered as ultrasound energy delivered through a mist, without direct contact to the tissues. This mist might be the same mist which contains the toxin, or it might be a different, possibly denser mist delivered at some time after the toxin has been delivered. The delivery devices for these mists might be introduced a relatively short distance into the nose. Thus the entire therapy might comprise the specialized delivery of two mists.

[0093] Referring now to Figures 28 and 30, an infusion catheter can be engaged against the ostium of a sinus cavity (Figure 28). A guidewire may then be advanced through the infusion

catheter and into the sinus cavity (Figure 30). The guidewire can be formed as a wave guide to deliver ultrasonic energy, as an electrode to deliver electroporation energy, or as an infusion wire to deliver the toxin solution itself. The wire could further be configured to perform two or more of these functions. The catheter could be configured to act as a counter electrode when the guidewire is acting as an electroporation electrode in bipolar energy delivery.

[0094] Referring now to Figure 31, the catheter advanced to the os of a sinus cavity, as illustrated, can also be used to deliver a helical or randomly shaped delivery tube which is deployed within the sinus. Preferably, the tube will expand to engage a major portion of the wall of the sinus cavity. Alternatively, the geometry could be selected to selectively engage only a particular portion of the wall of the sinus cavity. The wire can further be adapted to deliver energy, either electrical or acoustic, and/or may be configured to deliver and distribute the toxin solution within the cavity. In still other configurations, the wire could be coated to deliver the agent to the wall, and still further the wire could deliver ultrasound gels, saline, degassed water, or the like, to enhance coupling of a separate ultrasonic energy source.

[0095] Referring now to Figure 32, two or more deployment catheters can be used to advance any of the guidewires or other wire structures discussed above. As illustrated in Figure 32, an electrode basket may be deployed through the delivery catheter. Alternatively, a multiply tined catheter structure may be delivered through the delivery catheter.

[0096] Throughout this disclosure, the LC solution has typically been referred to as an infused, aerosolized or sprayed liquid. LC incorporated into coatings on devices has also been described. It should be noted, however, that other forms of LC delivery may be desirable.

[0097] For instance, commercially available botulinum toxins (such as Botox tm – Allergan) are supplied as a dry lyophilized powder, and must be reconstituted prior to delivery by the addition of saline to the packaging vial. Similarly, light chain would be most readily available and stable in a powdered form. It may be desirable to spray or blow the powdered form of the LC into the target airways directly, without any reconstitution by liquid.

[0098] The lyophilized powder could also be formed into sheets, ribbons, pellets microspheres, or any other desirable form, and introduced to the target tissues.

[0099] Instead of a saline or low viscosity carrier, it may be desirable to deliver the light chain in a gel carrier, such as the types of gels which are commonly used for ultrasound coupling. Other appropriate gel carriers include such biocompatible gels as hyaluronic acid

(HA). HA has the added benefit of being a thixotropic liquid – its viscosity drops as it begins to flow or as increasing shear stress is applied, and then returns to a higher viscosity state as it comes to rest. This would aid in delivery of the solution through catheters and the like, while allowing the gel to remain in place once delivered. HA is also extremely biocompatible, and would allow efficient ultrasonic coupling to the target tissues. The application of ultrasonic energy might also reduce the viscosity of the HA gel, possibly improving the delivery of toxin into the tissues.

[0100] It may also be desirable to incorporate the light chain into a foam, or to foam the LC solution upon or during delivery of the LC to the target tissues. Foams may better fill the entire targeted airway, and may trap water or coupling agents to allow efficient ultrasonic coupling. In a further embodiment, the foam may be energized within or as it exits the catheter shaft to further enhance the delivery of the LC to cells that are contacted by the energized foam and LC foam solution.

[0101] In addition to BTX-LC, it may be desirable to deliver additional agents to the nasal passages and sinuses prior to, coincident with, or after deliver of the LC. Adjunctive therapies may include agents designed to slow down or halt the motion of the cilia, in order to aid in delivery of the LC to the target tissues by prevention of their mobilization by the cilia. Agents known to slow or halt the motion of cilia include but are not limited to epinephrine dilutions of 1:1000 (which causes ciliary death), 1:10,000 (which causes reversible paralysis), 10% cocaine (induces paralysis) or 2.5% cocaine (slows or stops cilia).

[0102] Other adjunctive therapies may include the use of or pretreatment with mucolytics, which will thin mucus secretions within the nose and may allow better penetration of LC into the target cells.

[0103] Decongestants such as epinephrine also cause constriction of the vasculature in the nasal passages, which in addition to temporarily reducing swelling of the target tissues, may decrease the risks of LC entering the blood stream during poration and delivery. Epinephrine also constricts the blood vessels locally, which may increase the residence time of other locally delivered agents or decrease their likelihood of entering systemic circulation.

[0104] Steroids may be used to reduce swelling and inflammation prior to LC treatment in order to improve LC delivery to target tissues. In the abovementioned embodiments, it may be important to note that by far the most significant effects will be seen in areas where both the LC and the permeablizing energy are delivered. Therefore, although it may be best to deliver

both LC and energy to substantially the same area, for reasons of anatomy, ease of delivery, etc., either the LC or the energy might be delivered more broadly, or to a somewhat different area. As an extreme example, the LC might be delivered systemically or to the entire respiratory pathway, followed by very localized delivery of energy to the desired area. Alternatively, the LC could be delivered to a specific sinus, followed by energy to the entire nose and sinus using a standardized external energy delivery mask.

[0105] While the above is a complete description of the preferred embodiments of the invention, various alternatives, modifications, and equivalents may be used. Therefore, the above description should not be taken as limiting the scope of the invention which is defined by the appended claims.

WHAT IS CLAIMED IS:

1. A method for delivering toxins to target cells in a nasal membrane of a patient, said method comprising:

introducing a toxin to a region adjacent the target cells; and

applying energy to the target cells under conditions which cause poration of the cell membranes to enhance delivery of the toxin to the cells.

2. A method as in claim 1, wherein the region comprises at least one paranasal sinus or a main nasal passage.

3. A method as in claim 1, wherein the region comprises substantially the entire nasal cavity.

4. A method as in claim 1, wherein the region comprises the nasopharynx.

5. A method as in claim 1, wherein the target cells comprise goblet cells.

6. A method as in claim 1, wherein the energy is selectively applied to target cells within the region where toxin has been introduced.

7. A method as in claim 1, wherein the energy is applied non-selectively within the region where toxin has been introduced.

8. A method as in claim 1, wherein the patient suffers from or is at risk of suffering from rhinorreah.

9. A method as in claim 1, wherein the patient suffers from or is at risk of suffering from sinus headaches.

10. A method as in claim 1, wherein the patient suffers from or is at risk of suffering from migraine headaches.

11. A method as in claim 1, wherein the toxin comprises botulinum toxin.

12. A method as in claim 11, wherein the toxin is a fragment of botulinum toxin.

13. A method as in claim 12, wherein the fragment is a light chain fragment of botulinum toxin.

14. A method as in claim 13, wherein the light chain fragment is derived from at least one of botulinum toxins A, B, C, D, E, F, and G.

15. A method of claim 1, wherein the energy applied to the targeted region is an electric pulse.

16. A method of claim 15, wherein the electric pulse is applied from between 1V to 500V.

17. A method of claim 15, wherein the electric pulse is an RF signal.

18. A method of claim 15, wherein the RF signal is pulsed for durations between 5 microseconds to 100 milliseconds.

19. A method of claim 15, wherein the electric pulse is produced by a DC power source.

20. A method of claim 15, wherein the electric pulse is produced by an AC power source.

21. A method of claim 1, wherein the energy applied to the target region is ultrasonic.

22. A method of claim 1, wherein the energy applied to the targeted region is an x-ray beam.

23. A method of claim 1, wherein the energy applied to the targeted region is ultrasound.

24. A method of claim 1, wherein the energy applied to the targeted region is microwave.

25. A method as in claim 1, wherein the toxin is introduced to the target region through a catheter.

26. A method as in claim 25, wherein the toxin is introduced through a balloon on the catheter.

27. A method as in claim 26, wherein the balloon is porous and the toxin is introduced through the balloon.

28. A method as in claim 25, wherein the toxin is introduced through a needle on the catheter.

29. A method as in claim 24, wherein the toxin is aerosolized from the catheter.

30. A method as in claim 1, wherein energy is applied from a source on the catheter.

31. A method as in claim 30, wherein acoustic energy is applied from a transducer on the catheter.

32. A method as in claim 30, wherein electrical energy is applied from an electrode on the catheter.

33. A method as in claim 1, wherein the energy is delivered from a source external to the patient.

34. A method as in claim 33, wherein the source is an acoustic energy transducer.

35. A method as in claim 34, wherein the acoustic energy transducer is a focused ultrasound transducer.

36. A system for delivering toxins to target cells in a nasal membrane, said system comprising:

a catheter or other structure adapted to introduce a toxin to a region adjacent the target cells;

an energy applicator for applying energy to the target cells under conditions which cause poration of the cell membranes to enhance delivery of the toxin; and

a source of toxin suitable for introduction from the catheter.

37. A system as in claim 36, wherein the energy applicator is adapted to selectively apply energy to target cells within the region where toxin has been introduced.

38. A system as in claim 36, wherein the energy applicator is adapted to apply energy non-selectively within the region where toxin has been introduced.

39. A system as in claim 35, wherein the toxin comprises botulinum toxin.

40. A system as in claim 39, wherein the toxin is a fragment of botulinum toxin.

41. A system as in claim 40, wherein the fragment is a light chain fragment of botulinum toxin.

42. A system as in claim 41, wherein the light chain fragment is derived from at least one of botulinum toxins A, B, C, D, E, F, and G.

43. A system as in claim 1, wherein the energy applicator is adapted to apply an electric pulse of between 1v and 500V to the targeted region.

44. A system as in claim 43, wherein the electric pulse is an RF signal.

45. A system as in claim 43, wherein the RF signal is pulsed for durations between 5 microseconds to 100 milliseconds.

46. A system as in claim 43, wherein the electric pulse is produced by a DC power source.

47. A system as in claim 43, wherein the electric pulse is produced by an AC power source.

48. A system as in claim 1, wherein the energy applicator is adapted to apply ultrasonic energy to the targeted region.

49. A system as in claim 36, wherein the energy applicator is adapted to apply an x-ray beam to the targeted region.

50. A system as in claim 36, wherein the energy applicator is adapted to apply focused ultrasound to the targeted region.

51. A system as in claim 36, wherein the energy applicator is adapted to apply microwave to the targeted region.

52. A system as in claim 36, wherein the toxin is introduced through a balloon on the catheter.

53. A system as in claim 52, wherein the balloon is porous and the toxin is introduced through the balloon.

54. A system as in claim 52, wherein the toxin is introduced through a needle on the catheter.

55. A system as in claim 52, wherein the toxin is aerosolized from the catheter.

56. A system as in claim 36, wherein energy applicator is on the catheter.

57. A system as in claim 56, wherein energy applicator applies acoustic energy from a transducer on the catheter.

58. A system as in claim 56, wherein energy applicator applies electrical energy from an electrode on the catheter.

59. A system as in claim 36, wherein the applicator applies energy from a source external to the patient.

60. A system as in claim 59, wherein the source is an acoustic energy transducer.

61. A system as in claim 60, wherein the acoustic energy transducer is a focused ultrasound transducer.

62. A system as in claim 36, wherein the toxin is introduced through a membrane supported on a scaffold on the catheter.

63. A system as in claim 36, wherein the toxin is introduced by a needless injector.

64. A system as in claim 36, wherein the energy applicator is a wave guide positionable through a nostril.

ABSTRACT OF THE DISCLOSURE

Methods and systems for delivering toxin and toxin fragments to a patient's nasal cavity provide for both release of the toxin and delivery of energy which selectively porates target cells to enhance uptake of the toxin. The use of energy-mediated delivery is particularly advantageous with light chain fragment toxins which lack cell binding capacity.

Electronic Patent Application Fee Transmittal

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First Named Inventor:	Mark E. Deem
Filer:	James Mitchell Heslin/Nancy Pizzo
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