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[54] **CLASS OF PROTEINS FOR THE CONTROL OF PLANT PESTS**

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Related U.S. Application Data

[63] Continuation-in-part of Ser. No. 463,483, Jun. 5, 1995, which is a continuation-in-part of Ser. No. 314,594, Sep. 28, 1994, abandoned, which is a continuation-in-part of Ser. No. 218,018, Mar. 23, 1994, abandoned, which is a continuation-in-part of Ser. No. 37,057, Mar. 25, 1993, abandoned.

[51] **Int. Cl.⁶** **C12N 1/20; C07H 21/04**

[52] **U.S. Cl.** **435/252.3; 435/252.33; 435/252.31; 435/252.32; 435/252.34; 435/252.35; 435/235.1; 435/254.11; 435/257.2; 435/320.1; 536/23.71; 530/350**

[58] **Field of Search** **435/252.3, 252.33, 435/252.31, 252.32, 252.34, 252.35, 235.1, 254.11, 257.2, 410, 412, 419; 536/23.71; 530/350**

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[57] ABSTRACT

The present invention is drawn to a novel class of proteins, and their receptors. Novel processes, assays and methods for controlling plant pests are provided.

30 Claims, 13 Drawing Sheets

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1 20
SGSPGLQEFAAASTMYSRIFFLLVIVCAVKASLFTVN
40 60
VYDDNPETEIASSLKGCNPQECDQRCRRLKFPGGA
80 100
CVNGRCKCDNFLSVKDDVSVEEPAILKDLVSLEAEQ
120 140
AAKSRRCRNRVCDAVCRALHNTSGACVDGQCKCTN
160
KISAGDIVSDPAESLRTCNPIRCDEQCRRNGHEFGV
180 200
CFKGQCKCDYFLKEEVDEPEVTSLPKNCNPQECDQ
220 240
RCRRLKFPGGACVNGRCKCDNFFSAGDIVSDPAES
260 280
LRSCNPIRCDEQCRRNGHEFGVCFKGQCKCDYFL
300
NSEVDAVNEFPQAGSKRYCNLTQCNQTCANRFYD
320 340
SARVIHGWCKCYSKMERQDASPLNDVTEDENEVS
360 380
NDILRTVAEELSDVSPRACKSASCNQACRAFYFKG
396
GWCRFGRCQCF

Figure 1

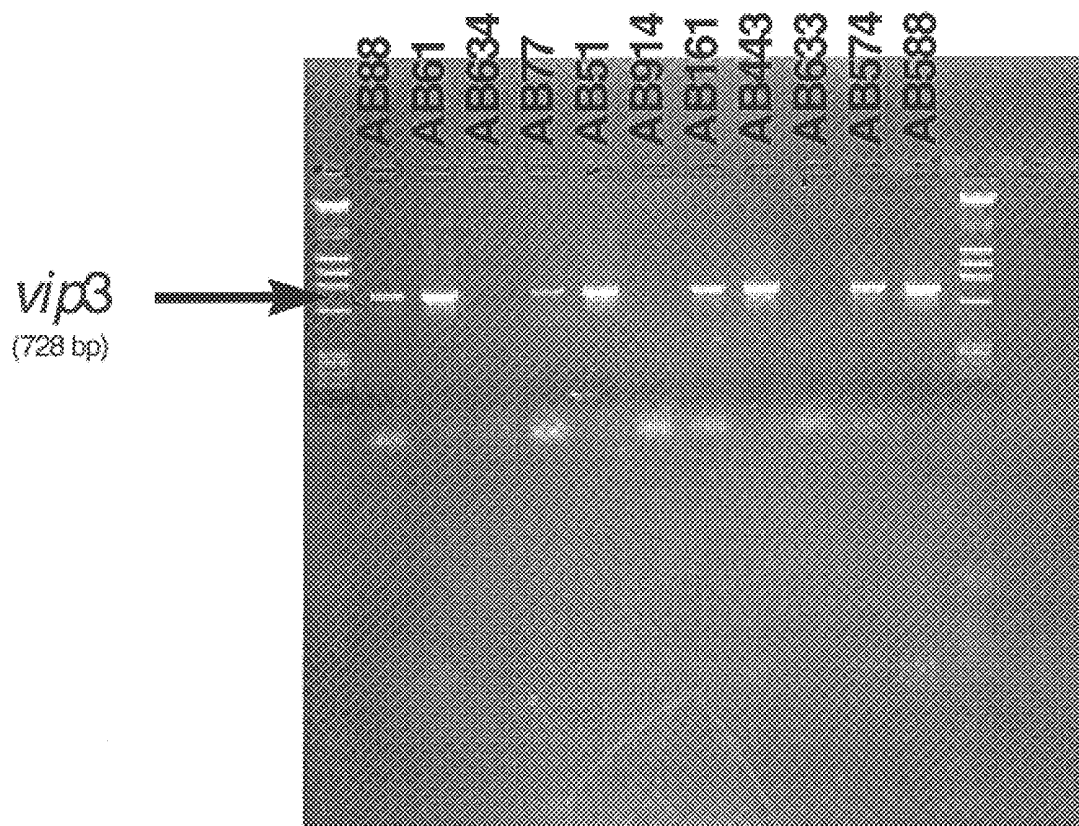


Figure 2

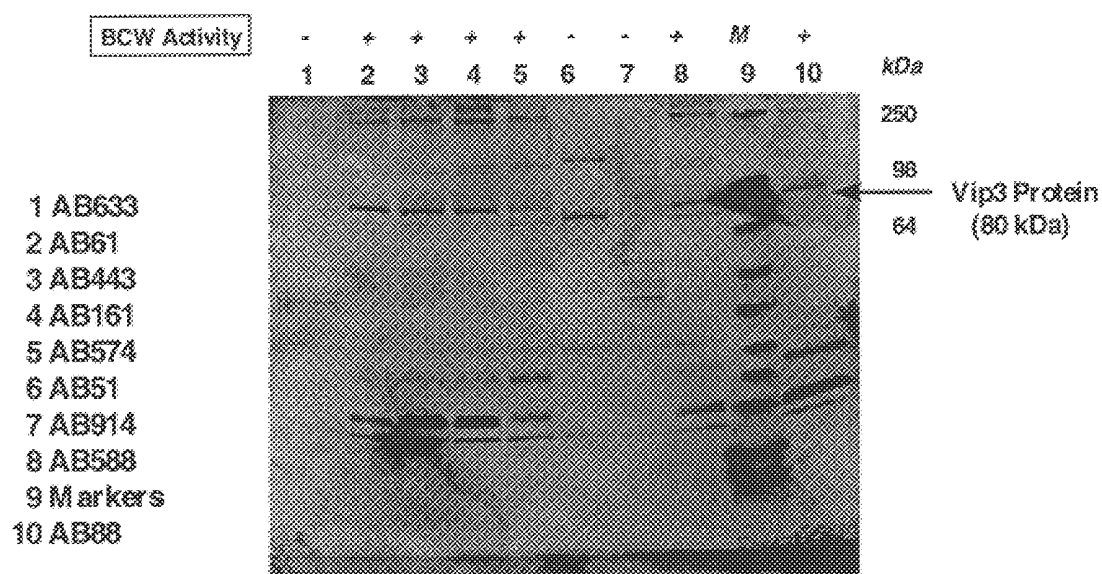


Figure 3

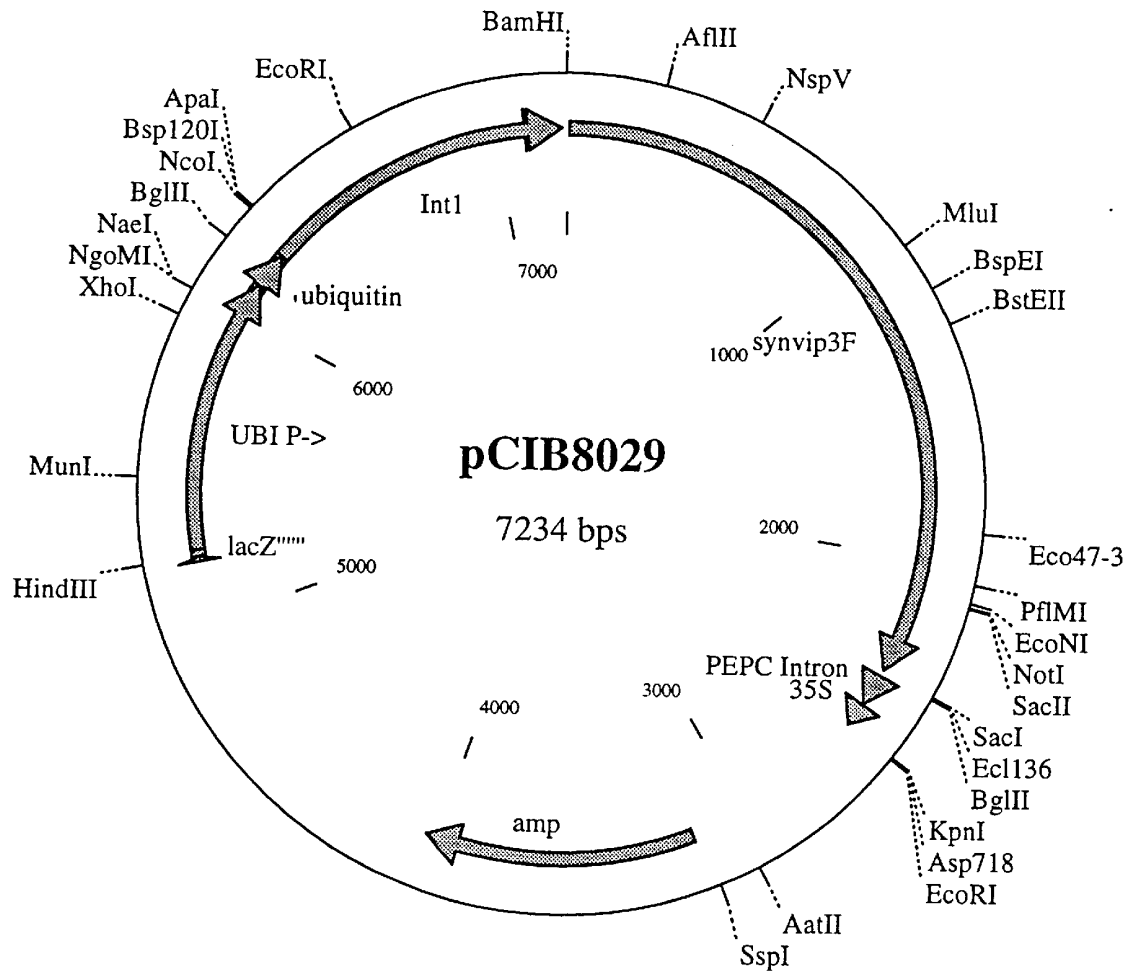
**Figure 4**

Figure 5

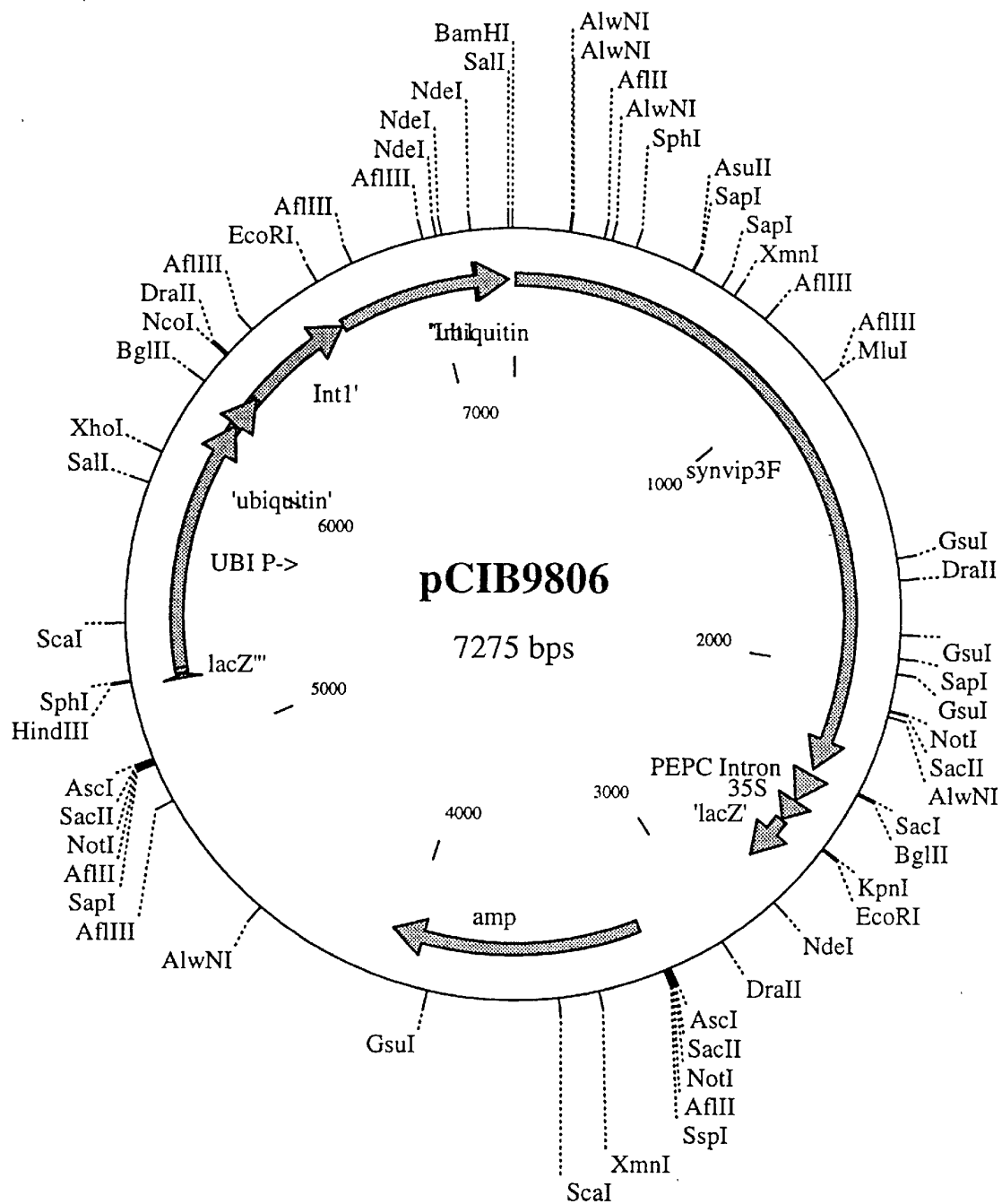
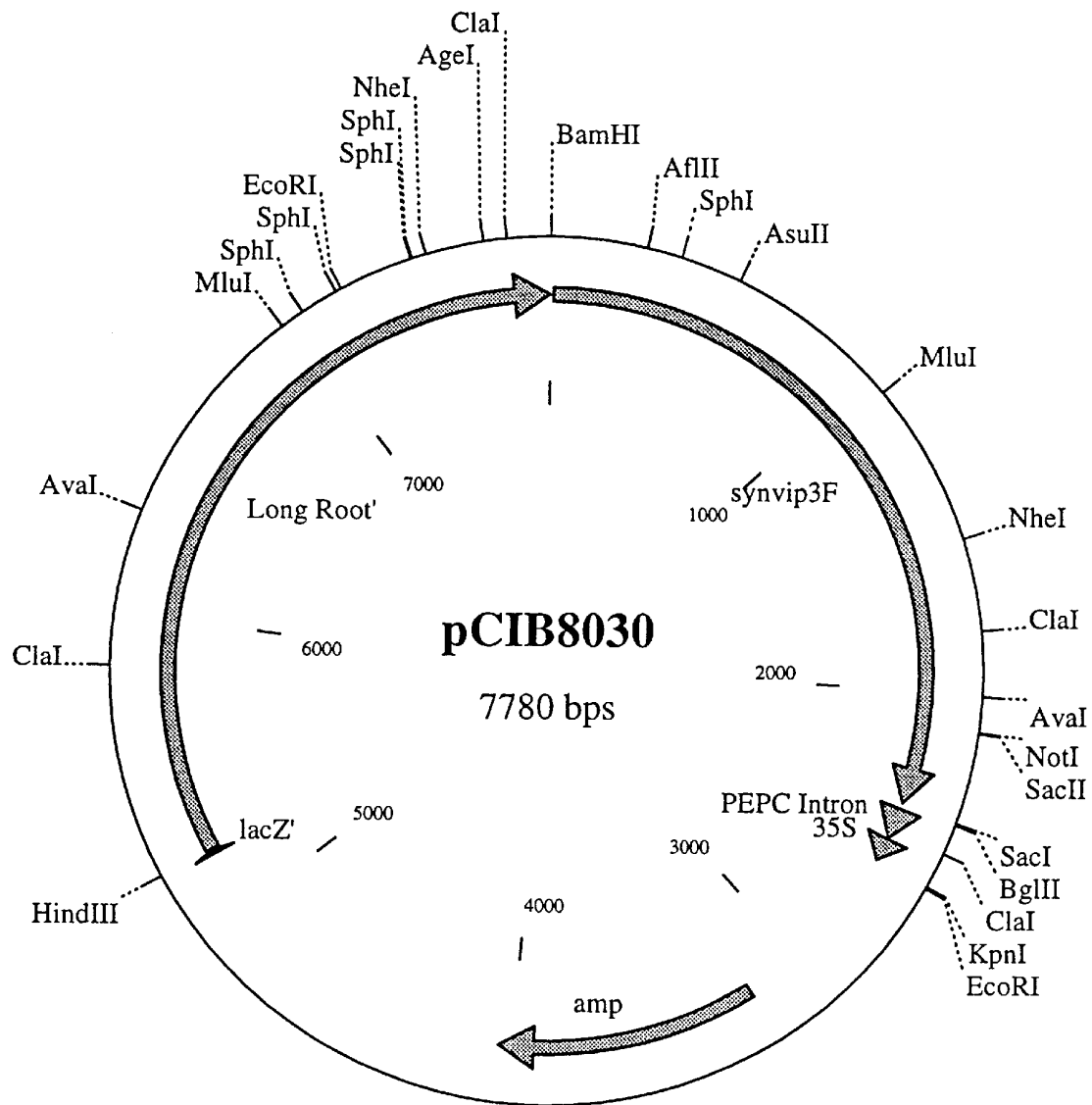
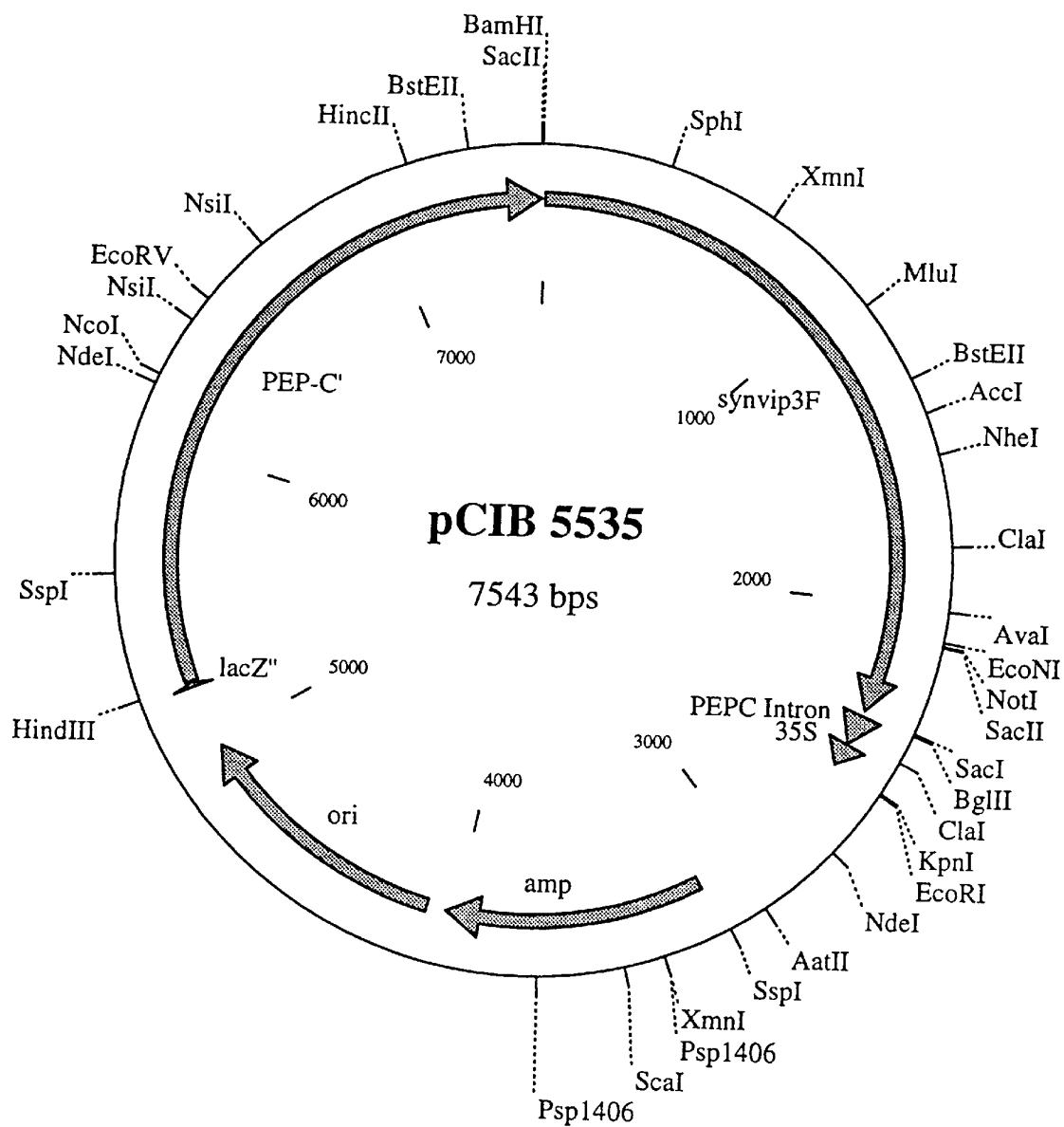
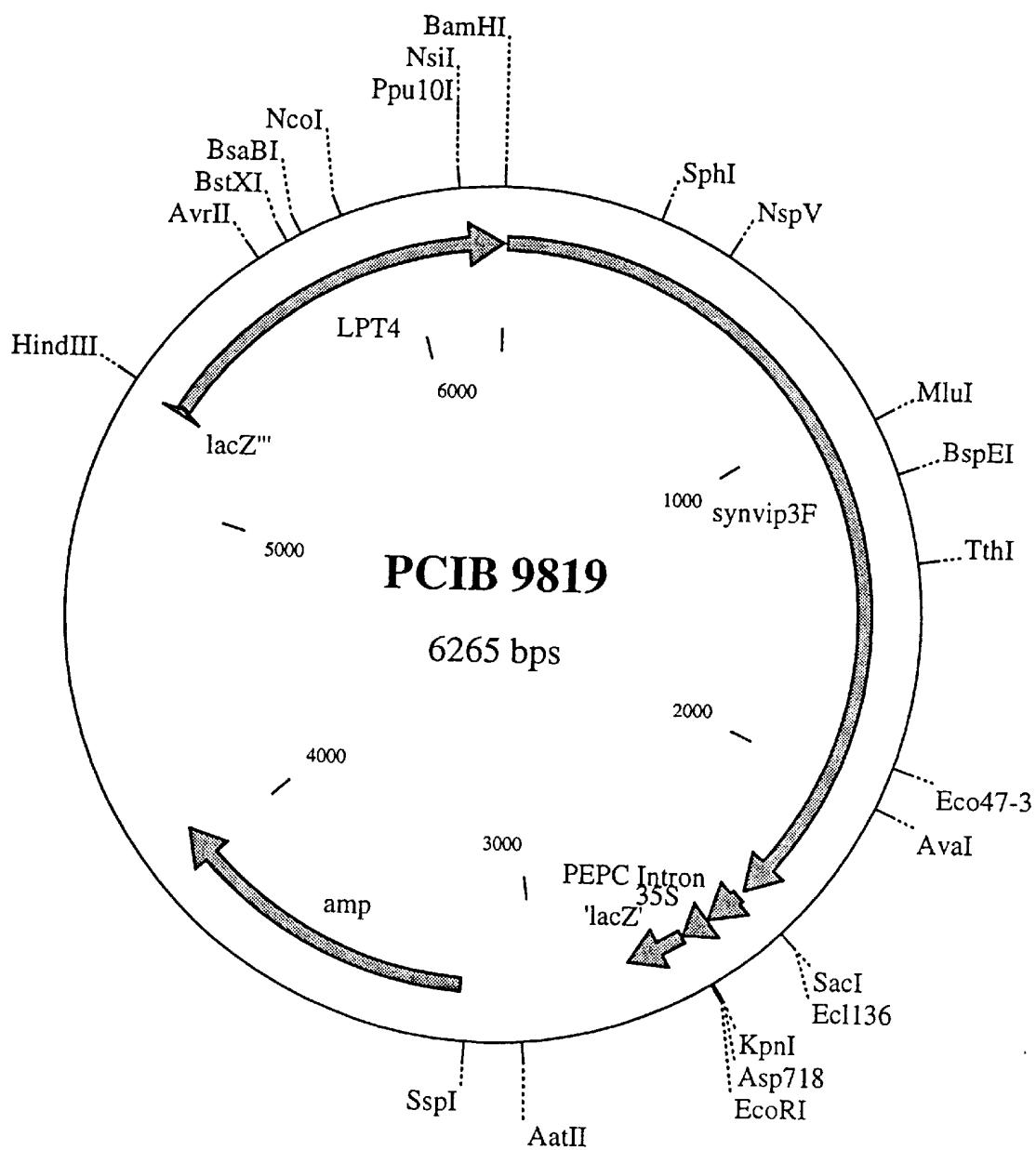


Figure 6

**Figure 7**

**Figure 8**

**Figure 9**

1	MNKNNTKLSTRALPSFIDYFNGIYGFBATGIKDIMNMIFKTDGTGGDLTLDE	50	SEQ	ID NO:2
1	MNKNNTKLSTRALPSFIDYFNGIYGFBATGIKDIMNMIFKTDGTGGDLTLDE	50	SEQ	ID NO:4
51	ILKNQQLLNDISGKLDGVNGSLNDLIAQGNLNTELSKEILKIANEQNQVL	100		
51	ILKNQQLLNDISGKLDGVNGSLNDLIAQGNLNTELSKEILKIANEQNQVL	100		
101	NDVNNKLD AINTMLRVYLPKITSMLS DVMKQNYALSLQIEYLSKQLQEIS	150		
101	NDVNNKLD AINTMLRVYLPKITSMLS DVMKQNYALSLQIEYLSKQLQEIS	150		
151	DKLDIINNVNLINSTLTEITPAYQRIKYVNEKFEELTFATETSSKVKKDG	200		
151	DKLDIINNVNLINSTLTEITPAYQRIKYVNEKFEELTFATETSSKVKKDG	200		
201	SPADILDELTELTELAKSVTKNDVDGFEFYLNTHDVMVGNNLFGRSALK	250		
201	SPADIRDELTELTELAKSVTKNDVDGFEFYLNTHDVMVGNNLFGRSALK	250		
251	TASELITKENVKTS GSEVGNVYNFLIVLTALQAQAFLLTLCRKLGLAD	300		
251	TASELITKENVKTS GSEVGNVYNFLIVLTALQAQAFLLTLCRKLGLAD	300		
301	IDYTSIMNEHLNKEKEEFVRN ILPTLSNTFSNP NYAKVKGSD EDAKMIVE	350		
301	IDYTSIMNEHLNKEKEEFVRN ILPTLSNTFSNP NYAKVKGSD EDAKMIVE	350		
351	AKPGHALIGFEISNDSITVLKVYEAKLKQNYQVDKDSLSEVIYGDMDKLL	400		
351	AKPGHALIGFEISNDSITVLKVYEAKLKQNYQVDKDSLSEVIYGDMDKLL	400		

Figure 10A

401 CPDQSEQIYYTNNIVFPNEYVITKIDFTKKMKTLRYEVTANFYDSSTGEI 450
||| |:|||||||||||||||||||||||||||||||||||||||
401 CPDQSGQIYYTNNIVFPNEYVITKIDFTKKMKTLRYEVTANFYDSSTGEI 450

451 DLNKKKVESSEAEYRTLSANDDGVYMP LGVISETFLTPINGFGLQADENS 500
|||||:|||||||||||||||||||||||||||||||||||||||
451 DLNKKKVESSEAEYRTLSANDDGVYMP LGVISETFLTPINGFGLQADENS 500

501 RLITLTCKSYLRELLLATDLSNKETKLIVPPSGFISNIVENGSI EEDNLE 550
|||||:|||||||||||||||||||||||||||||||||||||||
501 RLITLTCKSYLRELLLATDLSNKETKLIVPPSGFISNIVENGSI EEDNLE 550

551 PWKANNKNAYVDHTGGVNGTKALYVHKDGGISQFIGDKLKP KTEYVIQYT 600
|||||:|||||||||||||||||||||||||||||||||||||||
551 PWKANNKNAYVDHTGGVNGTKALYVHKDGGISQFIGDKLKP KTEYVIQYT 600

601 VKGKPSIHLK DENTGYIHYEDTNNNLEDYQTINKRFTTGTD LKGVYLILK 650
|||||:|||||||||||||||||||||||||||||||||||||||
601 VKGKPSIHLK DENTGYIHYEDTNNNLEDYQTINKRFTTGTD LKGVYLILK 650

651 SQNGDEAWGDNFIILEISPSEKLLSPELINTNNWTSTGSTN ISGNTLTLY 700
|||||:|||||||||||||||||||||||||||||||||||||||
651 SQNGDEAWGDNFIILEISPSEKLLSPELINTNNWTSTGSTN ISGNTLTLY 700

701 QGGRGILKQNLQLDSFSTYRVYFSVSGDANVRIRNSREVL FEKRYMSGAK 750
|||||:|||||||||||||||||||||||||||||||||||||||
701 QGGRGILKQNLQLDSFSTYRVYFSVSGDANVRIRNSREVL FEKRYMSGAK 750

751 DVSEMFTTKFEKDNFYIELSQGNNLYGGPIVHFYDVSIK 789
|||||:|||||||||||||||||||||||||||||||||||||||
751 DVSEMFTTKFEKDNFYIELSQGNNLYGGPIVHFYDVSIK 789

Figure 10B

[illegible]

Figure 11A

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401 CPDQSEQIYYTNNIVFPNEYVITKIDFTKKMKTLYEVTANFYDSSTGEI 450
    |||||:|||||
401 CPDQSGQIYYTNNIVFPNEYVITKIDFTKKMKTLYEVTANFYDSSTGEI 450
    |||||:|||||
451 DLNKKKVESSEAEYRTLSANDDGVYMP LGVISETFLTPINGFGLQADENS 500
    |||||:|||||
451 DLNKKKVESSEAEYRTLSANDDGVYMP LGVISETFLTPINGFGLQADENS 500
    |||||:|||||
501 RLITLTCKSYLRELLLATDLSNKETKLIVPPSGFISNIVENGSI EEDNLE 550
    |||||:|||||
501 RLITLTCKSYLRELLLATDLSNKETKLIVPPSGFISNIVENGSI EEDNLE 550
    |||||:|||||
551 PWKANNKNAYVDHTGGVNGTKALYVHKDGGISQFIGDKLKP KTEYVIQYT 600
    |||||:|||||
551 PWKANNKNAYVDHTGGVNGTKALYVHKDGGISQFIGDKLKP KTEYVIQYT 600
    |||||:|||||
601 VKGKPSIHLKDENTGYIHYEDTNNNLEDYQTINKRFTTGTD LKGVYLILK 650
    |||||:|||||
601 VKGKPSIHLKDENTGYIHYEDTNNNLEDYQTINKRFTTGTD LKGVYLILK 650
    |||||:|||||
651 SQNGDEAWGDNFIILEISPSEKLLSPELINTNNWTSTGSTN ISGNTLTLY 700
    |||||:|||||
651 SQNGDEAWGDNFIILEISPSEKLLSPELINTNNWTSTGSTN ISGNTLTLY 700
    |||||:|||||
701 QGGRGILKQNLQLDSFSTYRVYFSVSGDANVRIRNSREVL FEKRYM 746
    |||||:|||||
701 QGGRGILKQNLQLDSFSTYRVYFSVSGDANVRIRNSREVL FEKKDI 746
    |||||:|||||

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Figure 11B

CLASS OF PROTEINS FOR THE CONTROL OF PLANT PESTS

The present invention is a continuation-in-part application of U.S. Ser. No. 08/463,483 filed Jun. 5, 1995, which is a continuation-in-part application of U.S. application Ser. No. 08/314,594 filed Sep. 28, 1994, now abandoned, which is a continuation-in-part application of U.S. application Ser. No. 08/218,018 filed Mar. 23, 1994, now abandoned, which is a continuation-in-part application of U.S. application Ser. No. 08/037,057 filed Mar. 25, 1993, now abandoned, the disclosures of which are herein incorporated by reference.

FIELD OF THE INVENTION

The present invention relates to a novel class of proteins for the control of plant pests.

BACKGROUND OF THE INVENTION

Plant pests are a major factor in the loss of the world's commercially important agricultural crops resulting both in economic hardship to farmers and nutritional deprivation for local populations in many parts of the world. Broad spectrum chemical pesticides have been used extensively to control or eradicate pests of agricultural importance. There is, however, substantial interest in developing effective alternative pesticides.

Control of various pests through the use of biological molecules has been possible in only a limited number of cases. The best known examples of biological molecules with pesticidal uses are the δ -endotoxins from *Bacillus thuringiensis* (Bt), which is a gram-positive spore forming microorganism. Varieties of Bt are known that produce more than 25 different but related δ -endotoxins. Bt strains produce δ -endotoxins during sporulation.

The majority of δ -endotoxins made by Bt are toxic to larvae of certain insects in the orders Lepidoptera, Diptera and Coleoptera. Some of these δ -endotoxins have useful insecticidal activities against different insect pests. However, use of the δ -endotoxins is limited because they are active against only a very few of the many insect pests.

The limited specificity of the Bt endotoxins is dependent, at least in part, on both the activation of the toxin in the insect gut (Haider, M. Z. et al., 1986, Eur. J. Biochem. 156:531-540) and its ability to bind to specific receptors present on the insect's midgut epithelial cells (Hofmann, C. P. et al, 1988, PNAS 85:7844-7848). Among the factors which prevent activity of a particular δ -endotoxin against a specific insect is the lack of appropriate receptors in the insect gut or lack of affinity of the δ -endotoxin for the receptors which may be present, thus resulting in no binding of the δ -endotoxin to the brush border membranes. Therefore, the ability to control a specific insect pest using δ -endotoxins at present depends on the ability to find an appropriate δ -endotoxin with the desired range of activity. In many cases, no such δ -endotoxin is known, and it is not certain that one even exists.

Plants also routinely become infected by fungi and bacteria, and many microbial species have evolved to utilize the different niches provided by the growing plant. Some phytopathogens have evolved to infect foliar surfaces and are spread through the air, from plant-to-plant contact or by various vectors, whereas other phytopathogens are soil-borne and preferentially infect roots and newly germinated seedlings. In addition to infection by fungi and bacteria, many plant diseases are caused by nematodes which are soil-borne and infect roots, typically causing serious damage

when the same crop species is cultivated for successive years on the same area of ground.

The severity of the destructive process of disease depends on the aggressiveness of the phytopathogen and the response of the host, and one aim of most plant breeding programs is to increase the resistance of host plants to disease. Novel gene sources and combinations developed for resistance to disease have typically only had a limited period of successful use in many crop-pathogen systems due to the rapid evolution of phytopathogens to overcome resistance genes. In addition, there are several documented cases of the evolution of fungal strains which are resistant to particular fungicides, such as powdery mildew, wheat mildew, *Botrytis*, *Pyrenophora*, *Pseudocercospora* and *Mycosphaerella* (Jones and Clifford; Cereal Diseases, John Wiley, 1983).

It is apparent, therefore, that scientists must constantly be in search of new methods with which to protect crops against plant pests. It has been found in the present invention a novel class of proteins which can be used to control plant pests.

Programmed cell death is a process whereby developmental or environmental stimuli activate a genetic program that culminate in the death of the cell (Jacobson, M. D. et al., 1997, Cell 88: 347-354). This genetic potential exists in most, if not all, multicellular organisms. In the case of invertebrates, programmed cell death appears to play a dual role by being an integral part of both the insect development process and a response mechanism to infections particularly of viral nature (Clem, R. J. et al., 1991, Science 254: 1388-1390). Programmed cell death appears to be executed in several different manners leading to either apoptosis, atrophy or differentiation. Apoptosis is one of the best characterized types of programmed cell death encompassing cytological changes including membrane-bound apoptotic bodies and cytoplasmic blebbing as well as molecular changes such as endonucleolysis typified by the generation of oligosomal length fragments (Vaux, D. L. and Strasser, A., 1996, PNAS 93:2239-2244). Although the overall apoptotic phenomenology is rather conserved among the different organisms, it is interesting to point out that, for many insect cells, cytoplasmic vacuolization and swelling rather than condensation seem to be the cytological features associated with apoptotic processes (Bowen, I. D., et al., 1996, Microsc. Res. Techniq. 34:202-217). The present invention provides a novel class of proteins which induce programmed cell death and exert a pesticidal effect.

SUMMARY OF THE INVENTION

The present invention is drawn to proteins of the VIP3 class. Preferred proteins are VIP3A(a), VIP3A(b) and VIP3A(c). Also preferred are homologues of VIP3A(a), VIP3A(b) and VIP3A(c). Also provided by the invention are domains of the VIP3 proteins, including the toxic domain and the stabilizing domain. A preferred embodiment of the invention is the toxic domain of the VIP3A(a) protein and homologues thereof. Another preferred embodiment are antibodies to proteins of the VIP3 class.

The invention also provides hybrid toxins comprising a toxic domain of a protein of the VIP3 class. In a preferred embodiment, the hybrid toxin is a chimeric proteins having a toxic core domain operably linked to a heterologous stabilizing domain. In another preferred embodiment, the hybrid toxin comprises an antibody, or immunologically-active fragment thereof, which immunologically recognizes the VIP3 receptor operably linked to a toxic domain from other proteins, wherein the toxin domain is obtained from a

number of cytotoxic proteins including but not limited to *Bacillus* toxins, including endotoxins and vegetative insecticidal proteins.

Also encompassed by the invention are plants comprising a DNA sequence which encodes a protein of the VIP3 class. Preferred embodiments include plants selected from the group consisting of maize, sorghum, wheat, sunflower, tomato, cole crops, cotton, rice, soybean, sugarbeet, sugarcane, tobacco, barley, and oilseed rape. In a particularly preferred embodiment, the plant is a maize plant.

Also provided by the invention are plants further comprising a second DNA sequence which encodes a second insecticidal protein. Particularly preferred second DNA sequences are those which encode a δ -endotoxin, those which encode another protein of the VIP3 class, or those which encode a protein of the VIP1 or VIP2 classes. In a more preferred embodiment, the δ -endotoxin is active against an insect selected from the group consisting of Lepidoptera and Coleoptera. In a more particularly preferred embodiment the δ -endotoxin is active against Ostrinia, or Diabrotica. In another particularly preferred is a second DNA sequence which encodes a δ -endotoxin protein selected from the group consisting of Cry1, Cry3, Cry5 and Cry9. In a more particularly preferred embodiment, the δ -endotoxin is selected from the group consisting of Cry1Aa, Cry1Ab, Cry1Ac, Cry1B, Cry1C, Cry1D, Cry1Ea, Cry1Fa, Cry3A, Cry9A, Cry9C and Cry9B. Most particularly preferred are δ -endotoxins selected from the group consisting of Cry1Ab, Cry1Ba and Cry9C proteins.

The invention also provides microorganisms comprising a heterologous DNA sequence which encodes a protein of the VIP3 class. In a preferred embodiment, the microorganism is selected from the group consisting of bacteria, baculovirus, algae and fungi. In another preferred embodiment, the microorganism is selected from the group consisting of *Bacillus*, *Pseudomonas*, *Clavibacter*, and *Rhizobium*. Further encompassed by the invention are entomocidal compositions comprising microorganisms comprising a heterologous DNA sequence which encodes a protein of the VIP3 class. Also provided by the invention are microorganisms further comprising a second DNA sequence which encodes a second insecticidal protein. Particularly preferred second DNA sequences are those which encode a δ -endotoxin, those which encode another protein of the VIP3 class, or those which encode a protein of the VIP1 or VIP2 classes. In a more preferred embodiment, the δ -endotoxin is active against an insect selected from the group consisting of Lepidoptera and Coleoptera. In a more particularly preferred embodiment the δ -endotoxin is active against Ostrinia, or Diabrotica. In another particularly preferred is a second DNA sequence which encodes a δ -endotoxin endotoxin protein selected from the group consisting of Cry1, Cry3, Cry5 and Cry9. In a more particularly preferred embodiment, the δ -endotoxin is selected from the group consisting of Cry1Aa, Cry1Ab, Cry1Ac, Cry1B, Cry1C, Cry1D, Cry1Ea, Cry1Fa, Cry3A, Cry9A, Cry9C and Cry9B. Most particularly preferred are δ -endotoxins selected from the group consisting of Cry1Ab, Cry1Ba and Cry9C proteins.

The invention further provides a method of controlling insects by contacting the insects with an insecticidal amount of a protein of the VIP3 class or an insecticidal amount of a chemical ligand to a receptor of the VIP3 class of proteins. In one preferred embodiment, the insects are contacted with a transgenic plant comprising a DNA sequence which expresses a protein of the VIP3 class. In another preferred embodiment, the insects are contacted with a an entomocidal

composition comprising a protein of the VIP3 class, or comprising a DNA sequence which expresses a protein of the VIP3 class. In another preferred embodiment, the transgenic plant comprises a DNA sequence which expresses the VIP3A(a) protein. In another preferred embodiment the insect is selected from the group consisting of Coleoptera, Diptera, Hymenoptera, Lepidoptera, Mallophaga, Homoptera, Hemiptera, Orthoptera, Thysanoptera, Dermaptera, Isoptera, Anoplura, Siphonaptera, Trichoptera, and Acari. In a particularly preferred embodiment, the insect is a Coleoptera or Lepidoptera. In another particularly preferred embodiment, the insect is selected from the group consisting of black cutworm (*Agrotis ipsilon*), fall armyworm (*Spodoptera frugiperda*), beet armyworm (*S. exigua*), yellow striped armyworm (*S. ornithogalli*), southwestern corn borer (*Diatraea grandiosella*), sugarcane borer (*D. saccharalis*), corn earworm (*Helicoverpa zea*), mediterranean corn borer (*Sesamia nonagroides*), cabbage looper (*Trichoplusia ni*), velvetbean caterpillar (*Anticarsia gemmatilis*), diamondback moth (*Plutella xylostella*) and tobacco budworm (*Heliothis virescens*).

Also provided by the invention is a method of controlling insects wherein the transgenic plant or microorganism further comprises a second DNA sequence which encodes a second insecticidal protein. Particularly preferred second DNA sequences are those which encode a δ -endotoxin, those which encode another protein of the VIP3 class, or those which encode a protein of the VIP1 or VIP2 classes. In a more preferred embodiment, the δ -endotoxin is active against an insect selected from the group consisting of Lepidoptera and Coleoptera. In a more particularly preferred embodiment the δ -endotoxin is active against Ostrinia, or Diabrotica. In another particularly preferred is a second DNA sequence which encodes a δ -endotoxin protein selected from the group consisting of Cry 1, Cry3, Cry5 and Cry9. In a more particularly preferred embodiment, the δ -endotoxin is selected from the group consisting of Cry1Aa, Cry1Ab, Cry1Ac, Cry1B, Cry1C, Cry1D, Cry1Ea, Cry1Fa, Cry3A, Cry9A, Cry9C and Cry9B. Most particularly preferred are δ -endotoxins selected from the group consisting of Cry1Ab, Cry1Ba and Cry9C proteins.

The invention further provides recombinant DNA sequences which encode a protein of the VIP3 class. Preferred embodiments of the DNA sequence encode a protein selected from the group consisting of VIP3A(a), VIP3A(b), VIP3A(c). Also preferred are DNA sequences identified as homologues of vip3A(a). In another preferred embodiment, the DNA sequence is a synthetic sequence which has been altered for optimum expression in a plant, particularly where the DNA sequence has been optimized for expression in a maize plant. Also preferred are DNA sequences which comprise both a synthetic portion and a native portion. In a particularly preferred embodiment, the DNA sequence encoding the VIP3A(a) protein has been optimized for expression in a maize plant. Another preferred embodiment are DNA sequences which are homologous to a DNA sequence which encodes a protein of the VIP3 class. Particularly preferred are DNA sequences which hybridize under moderately stringent conditions to the vip3A(a) coding sequence. Yet another embodiment of the invention is a recombinant DNA sequence which expresses a protein of the VIP3 class under the control of a heterologous promoter, or wherein the coding regions is incorporated into the genome of an organism where it is not naturally expressed or is expressed at higher levels than that occurring naturally.

The invention is further drawn to a method of identifying and isolating homologues of a protein of the VIP3 class or

of a DNA sequence which encodes a protein of the VIP3 class. In a preferred embodiment, the method comprises obtaining a DNA sequence which encodes a protein of the VIP3 class, hybridizing said DNA sequence with DNA obtained from a test organism, and isolating said hybridized DNA. In another preferred embodiment, the method comprises obtaining a DNA sample from an organism, using primers to a DNA sequence encoding a protein of the VIP3 class and obtaining a reaction product, then isolating a DNA sequence which encodes a protein of the VIP3 class from said organism. In another preferred embodiment, the method comprises obtaining a protein sample from a test organism, obtaining an antibody to a protein of the VIP3 class, reacting said antibody with said protein sample, and detecting and isolating homologues by detecting the presence of an immunological reaction.

Also provided by the invention are expression cassettes comprising a promoter operably linked to a DNA sequence encoding a protein of the VIP3 class. In one preferred embodiment the promoter is selected from the group consisting of constitutive, tissue-preferred preferred and tissue-specific promoters for expression in plants. In a particularly preferred embodiment, the promoter is selected from the group consisting of the ubiquitin, PEP carboxylase, LPT and MTL promoters. In another preferred embodiment, the promoter is functional in a microorganism.

The invention further provides a receptor to a protein of the VIP3 class and DNA sequences which. In one embodiment of the invention, the receptor comprises a death domain and a repeated EGF-motif. A more preferred embodiment of the invention comprises a receptor to the VIP3A(a). A more particularly preferred embodiment is the receptor protein sequence set forth in SEQ ID NO:9, and homologues thereto. Also encompassed by the invention are DNA sequences which encode these receptor proteins, e.g., the DNA sequence set forth in SEQ ID NO:8 and homologues thereto. Antibodies to a receptor of the VIP3 class of proteins are also encompassed by the invention.

Also provided by the invention is a method of identifying a compound as a VIP3 receptor chemical ligand having pesticidal activity comprising exposing a cell, preferably an insect cell, to a test compound, and assaying said cell for apoptotic activity. In another embodiment of the invention, the method comprises measuring specific binding between VIP3 receptor and a test compound. A preferred embodiment are VIP3 receptor ligands identified by the method.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1: Amino acid sequence of the receptor for VIP3A(a) translated from the cDNA. Several features of the protein are shown: dotted line—signal peptide (amino acid 13 to 35); italic—domain spanning the putative death domain (amino acid 81–205); double underline—sequences with strong homology to sequences found in consensus death domains; bold—CKC motif repeated six times spanning the EGF-motifs; underline—sequences repeated within the EGF-motifs.

FIG. 2: Presence of vip3 genes in *Bacillus* isolates as identified by PCR assay.

FIG. 3: Presence of VIP3 proteins in *Bacillus* isolates as identified by Western assay.

FIG. 4: Plasmid pCIB8029 containing a maize ubiquitin promoter in an expression cassette.

FIG. 5: Plasmid pCIB8055 containing a maize ubiquitin promoter in an expression cassette.

FIG. 6: Plasmid pCIB9806 containing a maize ubiquitin promoter in an expression cassette.

FIG. 7: Plasmid pCIB8030 containing a promoter from a maize methalothionein-like gene in an expression cassette.

FIG. 8: Plasmid pCTB5535 containing a promoter from a maize PEPC gene in an expression cassette.

FIG. 9: Plasmid pCIB9819 containing a promoter from a barley non-specific lipid transfer protein LTP4 gene in an expression cassette.

FIGS. 10a and b: Alignment of VIP3A(a) (Upper Line) against VIP3A(b) (Lower Line)

FIGS. 11a and b: Alignment of VIP3A(a) (Upper Line) against VIP3A(c) (Lower Line).

DETAILED DESCRIPTION OF THE INVENTION

Definitions

“Plant pest” means any organism known to associate with plants and which, as a result of that association, causes a detrimental effect on the plant’s health and vigor. Plant pests include but are not limited to fungi, bacteria, insects, and nematodes. The term plant as used herein encompasses whole plants and parts of plants such as roots, stems, leaves and seed, as well as cells and tissues within the plants or plant parts.

The “VIP3 class of proteins” comprises VIP3A(a), VIP3A(b) VIP3A(c) and their homologues. “Homologue” is used throughout to mean that the indicated protein or polypeptide bears a defined relationship to other members of the VIP3 class of proteins. This defined relationship includes but is not limited to, 1) proteins which are at least 70%, more preferably 80% and most preferably 90% identical at the sequence level to another member of the VIP3 class of proteins while also retaining pesticidal activity, 2) proteins which are cross-reactive to antibodies which immunologically recognize another member of the VIP3 class of proteins, 3) proteins which are cross-reactive with a receptor to another member of the VIP3 class of proteins and retain the ability to induce programmed cell death, and 4) proteins which are at least 70%, more preferably 80% and most preferably 90% identical at the sequence level to the toxic core region of another member of the VIP3 class of proteins while also retaining pesticidal activity.

A “hybrid toxin” is used to indicate a genetic fusion, having domains operably linked so that, when translated, a functional chimeric protein is formed having, in the aggregate, the properties of the individual domains. “Domain” is used to indicate a region or portion of a protein or confers a recognizable function or structure which contributes to the overall functionality of the protein. It is recognized that a DNA sequence which encodes a protein domain is also encompassed by this definition.

“Heterologous” is used to indicate that a protein, polypeptide or nucleotide sequence has a different natural origin with respect to its current host. For example, if a vip3A(a) gene from a *Bacillus thuringiensis* is genetically transformed into a plant cell, then the gene is described as being heterologous with respect to its current host, which is the plant cell. Furthermore, if a vip3A(a) gene from *Bacillus thuringiensis* is genetically transformed into a *Pseudomonas* bacterium, then the gene is also described as being heterologous with respect to the *Pseudomonas*. “Heterologous” is also used to indicate that one or more of the domains present in a chimeric protein, polypeptide or nucleotide sequence differ in their natural origin with respect to other domains present. For example, if the toxic domain from VIP3A(a) protein is fused to the binding domain from the VIP1A(a) protein to make a functional insecticidal protein, then the chimeric fusion would have domains that are heterologous

to each other. In addition, a heterologous chimeric protein or polypeptide comprising the fusion of a toxic domain from VIP3A(a) protein to the binding domain from the VIP1A(a) protein, when expressed in a plant, would also be considered heterologous with respect to the plant host.

The term “chimeric” is used to indicate that the protein, polypeptide, or nucleotide sequence is comprised of domains at least one of which has an origin that is heterologous with respect to the other domains present. These chimeric proteins or polypeptides are encoded by chimeric nucleotide sequences which have been fused or ligated together resulting in a coding sequence which does not occur naturally. Such chimeric constructions may also be designated as “recombinant.”

“Expression cassette” as used herein means a DNA sequence capable of directing expression of a gene in plant cells, comprising a promoter operably linked to an amino

acid coding region which is operably linked to a termination region. The gene may be chimeric, meaning that at least one component of the gene is heterologous with respect to at least one other component of the gene. The gene may also be naturally occurring, but which has been obtained in a recombinant form useful for genetic transformation of a plant or microorganism.

Arthropod Pests

For purposes of the present invention, pests include insects and arachnids selected from the orders Coleoptera, Diptera, Hymenoptera, Lepidoptera, Mallophaga, Homoptera, Hemiptera, Orthoptera, Thysanoptera, Dermaptera, Isoptera, Anoplura, Siphonaptera, Trichoptera, and Acari, particularly Coleoptera and Lepidoptera.

A list of pests associated with major crop plants are provided in Tables 1–19. Such pests are included within the scope of the present invention.

TABLE 1

Lepidoptera (Butterflies and Moths)	
<u>Maize</u>	<i>Agrotis ipsilon</i> , black cutworm
<i>Ostrinia nubilalis</i> , European corn borer	<u>Tomato</u>
<i>Agrotis ipsilon</i> , black cutworm	<i>Helicoverpa zea</i> , tomato fruitworm
<i>Helicoverpa zea</i> , corn earworm	<i>Spodoptera exigua</i> , beet armyworm
<i>Spodoptera frugiperda</i> , fall armyworm	<i>Spodoptera frugiperda</i> , fall armyworm
<i>Diatraea grandiosella</i> , southwestern corn borer	<i>Spodoptera ornithogalli</i> , yellowstriped armyworm
<i>Elasmopalpus lignosellus</i> , lesser cornstalk borer	<i>Spodoptera praefica</i> , western yellowstriped armyworm
<i>Diatraea saccharalis</i> , sugarcane borer	
<i>Sesamia nonagroides</i> , Mediterranean corn borer	<i>Spodoptera eridania</i> , southern armyworm
<i>Ostrinia furnacalis</i> , Asian corn borer	<i>Agrotis ipsilon</i> , black cutworm
<u>Sorghum</u>	<i>Peridroma saucia</i> , variegated cutworm
<i>Chilo partellus</i> , sorghum borer	<i>Papaipema nebris</i> , stalk borer
<i>Spodoptera frugiperda</i> , fall armyworm	<i>Trichoplusia ni</i> , cabbage looper
<i>Helicoverpa zea</i> , corn earworm	<i>Keiferia lycopersicella</i> , tomato pinworm
<i>Elasmopalpus lignosellus</i> , lesser cornstalk borer	<i>Manduca sexta</i> , tobacco hornworm
<i>Feltia subterranea</i> , granulate cutworm	<i>Manduca quinquemaculata</i> , tomato hornworm
<u>Wheat</u>	
<i>Pseudaletia unipunctata</i> , army worm	
<i>Spodoptera frugiperda</i> , fall armyworm	
<i>Elasmopalpus lignosellus</i> , lesser cornstalk borer	
<i>Agrotis orthogonia</i> , pale western cutworm	
<i>Elasmopalpus lignosellus</i> , lesser cornstalk borer	
<u>Sunflower</u>	
<i>Suleima helianthana</i> , sunflower bud moth	
<i>Homoeosoma electellum</i> , sunflower moth	
<u>Cotton</u>	
<i>Heliothis virescens</i> , cotton boll worm	
<i>Helicoverpa zea</i> , cotton bollworm	
<i>Spodoptera exigua</i> , beet armyworm	
<i>Pectinophora gossypiella</i> , pink bollworm	
<i>Helicoverpa armigera</i> , cotton bollworm	
<u>Rice</u>	
<i>Diatraea saccharalis</i> , sugarcane borer	
<i>Spodoptera frugiperda</i> , fall armyworm	
<i>Helicoverpa zea</i> , corn earworm	
<i>Chilo suppressalis</i> , asiatic rice borer	
<i>Scirpophaga sp.</i>	
<u>Soybean</u>	
<i>Pseudoplusia includens</i> , soybean looper	
<i>Anticarsia gemmatilis</i> , velvetbean caterpillar	
<i>Plathypena scabra</i> , green cloverworm	
<i>Ostrinia nubilalis</i> , European corn borer	
<i>Agrotis ipsilon</i> , black cutworm	
<i>Spodoptera exigua</i> , beet armyworm	
<i>Heliothis virescens</i> , cotton boll worm	
<i>Helicoverpa zea</i> , cotton bollworm	

TABLE 1-continued

Lepidoptera (Butterflies and Moths)
<u>Barley</u>
<i>Ostrinia nubilalis</i> , European corn borer
Crucifers (broccoli, cabbage, cauliflower, collards)
<i>Artogeia rapae</i> , imported cabbageworm
<i>Pieris brassicae</i> , cabbage butterfly
<i>Trichoplusia ni</i> , cabbage looper
<i>Plutella xylostella</i> , diamondback moth
<i>Spodoptera exigua</i> , beet armyworm
<i>Agrotis ipsilon</i> , black cutworm
<i>Agrotis segetum</i> , common cutworm
<i>Mamestra configurata</i> , bertha army worm
<u>Grapes</u>
<i>Endopiza viteana</i> , grape berry moth
<u>Deciduous Fruits and Nuts</u>
<i>Cydia pomonella</i> , codling moth
<i>Platynota idaeusalis</i> , tufted apple bud moth
<u>Peppers</u>
<i>Ostrinia nubilalis</i> , European corn borer
<i>Spodoptera exigua</i> , beet armyworm
<i>Spodoptera eridania</i> , southern armyworm
<u>Potato</u>
<i>Ostrinia nubilalis</i> , European corn borer
<i>Phthorimaea operculella</i> , potato tuberworm
<u>Canola</u>
<i>Plutella xylostella</i> , diamondback moth
<u>Sugarcane</u>
<i>Diatraea saccharalis</i> , sugarcane borer

TABLE 2

Coleoptera (Beetles)	
<u>Maize</u>	<u>Cotton</u>
<i>Diabrotica virgifera. virgifera</i> , western corn rootworm	<i>Anthonomus grandis</i> , boll weevil
	<u>Rice</u>
<i>Diabrotica longicornis barberi</i> , northern corn rootworm	<i>Colaspis brunnea</i> , grape colaspis
<i>Diabrotica undecimpunctata howardi</i> , southern corn rootworm	<i>Lissorhoptrus oryzophilus</i> , rice water weevil
<i>Melanotus spp.</i> , wireworms	<i>Sitophilus oryzae</i> , rice weevil
	<i>Oulema oryzae</i> , rice beetle
	<u>Soybean</u>
<i>Cyclocephala borealis</i> , northern masked chafer (white grub)	<i>Epilachna varivestis</i> , Mexican bean beetle
	<u>Tomato</u>
<i>Cyclocephala immaculata</i> , southern masked chafer (white grub)	<i>Leptinotarsa decemlineata</i> , Colorado potato beetle
<i>Popillia japonica</i> , Japanese beetle	<i>Epitrix hirtipennis</i> , tobacco flea beetle
	<u>Crucifers (broccoli, cabbage, cauliflower, collards)</u>
<i>Chaetocnema pulicaria</i> , corn flea beetle	<i>Phyllotreta cruciferae</i> , crucifer flea beetle
<i>Sphenophorus maidis</i> , maize billbug	<i>Phyllotreta pusilla</i> , western black flea beetle
<u>Sorghum</u>	<u>Peppers</u>
<i>Phyllophaga crinita</i> , white grub	<i>Anthonomus eugenii</i> , pepper weevil
<i>Eleodes</i> , <i>Conoderus</i> , and <i>Aeolus spp.</i> , wireworms	<u>Potato</u>
<i>Oulema melanopus</i> , cereal leaf beetle	<i>Leptinotarsa decemlineata</i> , Colorado potato beetle
<i>Chaetocnema pulicaria</i> , corn flea beetle	<i>Epitrix cucumeris</i> , potato flea beetle
<i>Sphenophorus maidis</i> , maize billbug	<i>Hemicrepidus memnonius</i> , wireworms

TABLE 2-continued

Coleoptera (Beetles)	
<u>Wheat</u>	<i>Melanopus</i> spp., wireworms
<i>Oulema melanopus</i> , cereal leaf beetle	<u>Canola</u>
<i>Hypera punctata</i> , clover leaf weevil	<i>Ceutorhynchus assimilis</i> , cabbage seedpod weevil
<i>Diabrotica undecimpunctata howardi</i> , southern corn rootworm	<i>Phyllotreta cruciferae</i> , crucifer flea beetle
<u>Sunflower</u>	
<i>Zygogramma exclamationis</i> , sunflower beetle	
<i>Bothyrus gibbosus</i> , carrot beetle	

TABLE 3

Homoptera (Whiteflies, Aphids etc..)	
<u>Maize</u>	<i>Bemisia tabaci</i> , sweetpotato whitefly
<i>Rhopalosiphum maidis</i> , corn leaf aphid	<i>Bemisia argentifolii</i> , silverleaf whitefly
<i>Anuraphis maidiradicis</i> , corn root aphid	<u>Crucifers (Broccoli, cabbage, cauliflower, collards)</u>
<u>Sorghum</u>	<i>Brevicoryne brassicae</i> , cabbage aphid
<i>Rhopalosiphum maidis</i> , corn leaf aphid	<i>Myzus persicae</i> , green peach aphid
<i>Sipha flava</i> , yellow sugarcane aphid	<u>Peppers</u>
<u>Wheat</u>	<i>Myzus persicae</i> , green peach aphid
Russian wheat aphid	<u>Potato</u>
<i>Schizaphis graminum</i> , greenbug	<i>Empoasca fabae</i> , potato leafhopper
<i>Macrosiphum avenae</i> , English grain aphid	<i>Myzus persicae</i> , green peach aphid
<u>Cotton</u>	<i>Macrosiphum euphorbiae</i> , potato aphid
<i>Aphis gossypii</i> , cotton aphid	<i>Paratrioza cockerelli</i> , potato psyllid
<i>Pseudatomoscelis seriatus</i> , cotton fleahopper	<u>Melon</u>
<i>Trialeurodes abutilonea</i> , bandedwinged whitefly	<i>Bemisia argentifolii</i> , silverleaf whitefly
<u>Rice</u>	<i>Bemisia tabaci</i> , sweetpotato whitefly
<i>Nephotettix nigropictus</i> , rice leafhopper	<u>Carrot</u>
<i>Nilaparvata lugens</i>	<i>Cavariella aegopodii</i> , carrot aphid
<i>Sogatella furcifera</i>	<u>Canola</u>
<i>Laodelphaax striatellus</i>	<i>Brevicoryne brassicae</i> , cabbage aphid
<u>Soybean</u>	<u>Vegetables</u>
<i>Myzus persicae</i> , green peach aphid	<i>Aphis fabae</i> , bean aphid
<i>Empoasca fabae</i> , potato leafhopper	<u>Sugar Beet</u>
<u>Barley</u>	<i>Pemphigus popullivenerae</i> , sugar beet root aphid
<i>Schizaphis graminum</i> , greenbug	<u>Deciduous Fruits and Nuts</u>
<u>Oil Seed Rape</u>	<i>Dysaphis plantaginea</i> , rosy apple aphid
<i>Brevicoryne brassicae</i> , cabbage aphid	<u>Sugarcane</u>
<u>Tomato</u>	<i>Saccharosydne saccharivora</i> , West Indian canefly
<i>Myzus persicae</i> , green peach aphid	<i>Sipha flava</i> , yellow sugarcane aphid
<i>Macrosiphum euphorbiae</i> , potato aphid	
<i>Trialeurodes vaporariorum</i> , greenhouse whitefly	

TABLE 4

Hemiptera (Bugs)	
<u>Maize</u>	5
<i>Blissus leucopterus leucopterus</i> , chinch bug <u>Sorghum</u>	
<i>Blissus leucopterus leucopterus</i> , chinch bug <u>Cotton</u>	10
<i>Lygus lineolaris</i> , tarnished plant bug <u>Rice</u>	
<i>Blissus leucopterus leucopterus</i> , chinch bug <i>Acrosternum hilare</i> , green stink bug <u>Soybean</u>	15
<i>Acrosternum hilare</i> , green stink bug <u>Barley</u>	
<i>Blissus leucopterus leucopterus</i> , chinch bug <i>Acrosternum hilare</i> , green stink bug <i>Euschistus servus</i> , brown stink bug <u>Tomato</u>	20
<i>lygus bug</i> <i>Acrosternum hilare</i> , green stink bug <i>Euschistus servus</i> , brown stick bug	25

TABLE 5

Orthoptera (Grasshoppers Crickets, and Cockroaches)	
<u>Maize</u>	30
<i>Melanoplus femurrubrum</i> , redlegged grasshopper <i>Melanoplus sanguinipes</i> , migratory grasshopper <u>Wheat</u>	35
<i>Melanoplus femurrubrum</i> , redlegged grasshopper <i>Melanoplus differentialis</i> , differential grasshopper <i>Melanoplus sanguinipes</i> , migratory grasshopper <u>Cotton</u>	40
<i>Melanoplus femurrubrum</i> , redlegged grasshopper <i>Melanoplus differentialis</i> , differential grasshopper <u>Soybean</u>	
<i>Melanoplus femurrubrum</i> , redlegged grasshopper <i>Melanoplus differentialis</i> , differential grasshopper	45

TABLE 6

Diptera (Flies and Mosquitoes)	
<u>Maize</u>	50
<i>Hylemya platura</i> , seedcorn maggot <i>Agromyza parvicornis</i> , corn blotch leafminer <u>Sorghum</u>	
<i>Contarinia sorghicola</i> , sorghum midge <u>Wheat</u>	
<i>Mayetiola destructor</i> , Hessian fly <i>Sitodiplosis mosellana</i> , wheat midge <i>Meromyza americana</i> , wheat stem maggot <i>Hylemya coarctata</i> , wheat bulb fly <u>Sunflower</u>	
<i>Neolasioptera murtfeldtiana</i> , sunflower seed midge <u>Soybean</u>	
<i>Hylemya platura</i> , seedcorn maggot	

TABLE 6-continued

Diptera (Flies and Mosquitoes)	
<u>Barley</u>	
<i>Hylemya platura</i> , seedcorn maggot <i>Mayetiola destructor</i> , Hessian fly <u>Tomato</u>	
<i>Liriomyza trifolii</i> , leafminer <i>Liriomyza sativae</i> , vegetable leafminer <i>Scrobipalpula absoluta</i> , tomato leafminer <u>Crucifers (broccoli, cabbage, cauliflower, collards)</u>	
<i>Delia brassicae</i> , cabbage maggot <i>Delia radicum</i> , cabbage root fly <u>Carrot</u>	
<i>Psilia rosae</i> , carrot rust fly <u>Sugarbeet</u>	
<i>Tetanops myopaeformis</i> , sugarbeet root maggot <u>Vegetables</u>	
<i>Liriomyza sativae</i> , vegetable leaf miner	

TABLE 7

Thysanoptera (Thrips)	
<u>Maize</u>	
<i>Anaphothrips obscurus</i> , grass thrips <u>Wheat</u>	
<i>Frankliniella fusca</i> , tobacco thrips <u>Cotton</u>	
<i>Thrips tabaci</i> , onion thrips <i>Frankliniella fusca</i> , tobacco thrips <u>Soybean</u>	
<i>Sericothrips vanabilis</i> , soybean thrips <i>Thrips tabaci</i> , onion thrips <u>Tomato</u>	
<i>Frankliniella occidentalis</i> , western flower thrips <i>Frankliniella fusca</i> , tobacco thrips <i>Thrips tabaci</i> , onion thrips <u>Crucifers (broccoli, cabbage, cauliflower, collards)</u>	
<i>Thrips tabaci</i> , onion thrips <u>Peppers</u>	
<i>Thrips palmi</i> , melon thrips <u>Potato</u>	
<i>Thrips palmi</i> , melon thrips	55

TABLE 8

Hymenoptera (Sawflies, Ants, Wasps etc)	
<u>Maize</u>	
<i>Solenopsis milesta</i> , thief ant <u>Wheat</u>	
<i>Cephus cinctus</i> , wheat stem sawfly	65

TABLE 9

Acari (Mites and Ticks)	
<u>Maize</u>	
<i>Tetranychus urticae</i> , twospotted spider mite	
<u>Sorghum</u>	
<i>Tetranychus cinnabarinus</i> , carmine spider mite	
<i>Tetranychus urticae</i> , twospotted spider mite	
<u>Wheat</u>	
<i>Aceria tulipae</i> , wheat curl mite	
<u>Cotton</u>	
<i>Tetranychus cinnabarinus</i> , carmine spider mite	
<i>Tetranychus urticae</i> , twospotted spider mite	
<u>Soybean</u>	
<i>Tetranychus turkestanii</i> , strawberry spider mite.	
<i>Tetranychus urticae</i> , twospotted spider mite	
<u>Barley</u>	
<i>Petrobia latens</i> , brown wheat mite	
<u>Tomato</u>	
<i>Tetranychus urticae</i> , two spotted spider mite	
<i>Aculops lycopersici</i> , tomato russet mite	
<i>Steneotarsonemus pallidus</i> , cyclamen mite	
<u>Citrus</u>	
<i>Panonychus citri</i> , citrus red mite	
<i>Brevipalpus lewisi</i> , citrus flat mite	
<i>Phyllocoptruta oleivora</i> , citrus rust mite	
<u>Deciduous Fruits and Nuts</u>	
<i>Panonychus ulmi</i> , European red mite	
<i>Tetranychus sp.</i> , spider mite	

For purposes of the present invention, pests also include fungal phytopathogens of plants. A list of fungal pests associated with major crop plants is provided in Table 10. Such pests are included within the scope of the present invention.

TABLE 10

<u>Fungal Diseases of Plants</u>	
<u>Ear Molds</u>	
Gibberella ear mold	<i>Gibberella zeae</i> <i>G. saubinetii</i>
Aspergillus ear rot	<i>Aspergillus flavus</i> <i>A. parasiticus</i>
Diplodia ear rot	<i>Diplodia maydis</i> <i>D. macrospora</i>
Fusarium ear rot	<i>Fusarium moniliforme</i> <i>F. monilif. var. subglutinans</i>
<u>Stalk Rots</u>	
Pythium stalk rot	<i>Pythium aphanidermata</i>
Anthracnose stalk rot	<i>Colletotrichum graminicola</i> <i>C. tucumanensis</i> <i>Glomerella graminicola</i>
Diplodia stalk rot	<i>Diplodia maydis</i> <i>D. zeae-maydis</i> <i>Stenocarpella maydis</i> <i>Macrodiplodia zeae</i> <i>Sphaeria maydis</i> <i>S. zeae</i> <i>D. macrospora</i>
Fusarium stalk rot	<i>Fusarium moniliforme</i>
Gibberella stalk rot	<i>G. zeae</i> <i>G. saubinetii</i>
Stewart's wilt & leaf blight	<i>Erwinia stewartii</i>

TABLE 10-continued

<u>Fungal Diseases of Plants</u>	
<u>Ear Molds</u>	
<u>Leaf Diseases</u>	
Northern corn leaf blight	<i>Exserohilum turcicum</i>
Southern corn leaf blight	<i>Bipolaris maydis</i>
10 Gray leaf spot	<i>Cercospora zeae-maydis</i> <i>C. sorghi var. maydis</i> <i>Colletotrichum graminicola</i>
Anthracnose leaf blight	<i>Puccinia sorghi</i>
Common rust	<i>P. maydis</i> <i>Puccinia polysora</i> <i>Dicaeoma polysorum</i>
Southern rust	<i>Sphaelotheca reiliana</i>
15 Head smut	<i>Ustilago maydis</i>
Common smut	<i>Helminthosporium carbonum</i>
Carbonum leaf spot	<i>Kabatiella zeae</i>
Eye spot	
<u>Downy Mildews</u>	
20 Sorghum downy mildew	<i>Peronosclerospora sorghi</i>
Brown stripe downy mildew	<i>Sclerophthora rayssiae</i>
Sugarcane downy mildew	<i>Peronosclerospora sacchari</i>
Phillipine downy mildew	<i>Peronoscler. philippinensis</i>
Java downy mildew	<i>Peronosclerospora maydis</i>
Spontaneum downy mildew	<i>Peronosclerospora spontanea</i>
25 Rajasthan downy mildew	<i>Peronosclerospora heteropogoni</i>
Graminicola downy mildew	<i>Sclerospora graminicola</i>
Rusts	<i>Puccinia graminis f.sp. tritici</i> <i>Puccinia recondita f.sp. tritici</i> <i>Puccinia striiformis</i> <i>Tilletia tritici</i>
Smuts	<i>Tilletia controversa</i> <i>Tilletia indica</i> <i>Ustilago tritici</i> <i>Urocystis tritici</i>
30 Root rots, Foot rots and Blights	<i>Gaeumannomyces graminis</i> <i>Pythium spp.</i> <i>Fusarium culmorum</i> <i>Fusarium graminearum</i> <i>Fusarium avenaceum</i> <i>Drechslera tritici-repentis</i> <i>Rhizoctonia spp.</i> <i>Colletotrichum graminicola</i> <i>Helminthosporium spp.</i> <i>Microdochium nivale</i> <i>Pseudocercospora herpotrichoides</i>
35 Mildews	<i>Erysiphe graminis f.sp. tritici</i> <i>Sclerophthora macrospora</i>
Miscellaneous Fungal Diseases	<i>Septoria tritici</i> <i>Septoria nodorum</i>
45	

VIP3 is a Novel Class of Proteins

The proteins of the VIP3 class are secreted to the media by *Bacillus* spp. in vegetative stages of growth. VIP3A(a) is a member of a newly discovered class of proteins displaying insecticidal activity against a broad spectrum of lepidopteran insects including black cutworm (*Agrotis ipsilon*), fall armyworm (*Spodoptera frugiperda*), beet armyworm (*S. exigua*), yellow striped armyworm (*S. ornithogalli*), southwestern corn borer (*Diatraea grandiosella*), sugarcane borer (*D. saccharalis*), corn earworm (*Helicoverpa zea*), Mediterranean corn borer (*Sesamia nonagroides*), cabbage looper (*Trichoplusia ni*), velvetbean caterpillar (*Anticarsia gemmatilis*), diamondback moth (*Plutella xylostella*) and tobacco budworm (*Heliothis virescens*). Some of these lepidopteran insects have been shown to be very resistant to other insecticidal proteins such as δ -endotoxin. For example, the reported LC₅₀ for Cry1A(c), which is one of the most effective δ -endotoxin against black cutworm, is greater than 6000 ng/cm² (MacIntosh et al., J. Invertebr. Pathol. 56:258-266 (1990)). In contrast, it takes 260-fold less of VIP3A(a) protein to kill 50% of the black cutworm larvae. Thus, the VIP3A(a) protein displays a unique spectrum of insecticidal activities.

Both the DNA (SEQ ID NO:1) and the protein sequence of VIP3A(a) (SEQ ID NO:2) were used to search existing publicly available databases. The search was performed by using FastA (for nucleic acid), TFASTA (for protein) and BLAST (for protein) (Wisconsin Package, Programme Manual, version 9 UNIX, 1997). FastA and TFASTA use the method of Pearson and Lipman (Proc. Natl. Acad. Sci. U.S.A. 85:2444-2448 (1988)) to search for similarities between one sequence (query) and any group of sequences of the same type. BLAST (Basic Local Alignment Search Tool) uses the method of Altschul et al. (J. Biol. Mol. 215:403-41, 1990) to search the similarities between a query sequence and all sequences in a database. The three methods combined constitute a very powerful tool to search for relationships between a query sequence and sequences in databases. The search using vip3A(a) gene sequence and VIP3A(a) protein sequence as query sequences resulted in no significant homology with any other gene or protein of the databases (GenBank, EMBL and the SWISS-PROT). Therefore, the proteins of the VIP3 class, and the genes which encode them, are novel.

HOMOLOGUES TO VIP3A(a)

It is recognized that there are multiple approaches to identifying and isolating homologues within the VIP3 class of proteins and the DNA sequences which encode them. For example, to obtain the nucleotide sequence encoding a protein which is a member of the VIP3 class, cosmid clones, which express the protein, can be isolated from a genomic library. From larger active cosmid clones, smaller subclones can be made and tested for activity. In this manner, clones which express an active VIP3 protein can be sequenced to determine the nucleotide sequence of the gene. Then, an amino acid sequence can be deduced for the protein. For general molecular methods, see, for example, Molecular Cloning, A Laboratory Manual, Second Edition, Vols. 1-3, Sambrook et al. (eds.) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989), and the references cited therein.

The present invention also encompasses nucleotide sequences from organisms other than *Bacillus*, where the nucleotide sequences can be isolated by hybridization with the nucleotide sequences of vip3A(a). Proteins encoded by such nucleotide sequences can be tested for pesticidal activity. The invention also encompasses the proteins encoded by the nucleotide sequences. Furthermore, the invention encompasses proteins obtained from organisms other than *Bacillus* wherein the protein cross-reacts with antibodies raised against the proteins of the invention. Again the isolated proteins can be assayed for pesticidal activity by the methods disclosed herein or by other methods well-known in the art.

Genes homologous to vip3A(a) can also be identified by means of Southern analysis of DNA isolated from different biological sources. Total DNA can be isolated from any organism (see Ausubel, F. et al Current Protocols in Molecular Biology, 1988), restriction digested, run in agarose gels and blotted onto either nitrocellulose or nylon filters. These filters can be probed with full or partial-length coding sequences of the vip3A(a) gene. At high stringent hybridization and washing conditions only the genes with a similarity to the vip3A(a) higher than 80% will be identified. These high stringent conditions consist of an overnight hybridization at 68° C. in a variety of buffers (Ausubel, F. et al. Current Protocols in Molecular Biology, 1988), followed by 2 washes of 10 min each at 68° C. in 2×standard saline citrate, SSC, 0.1% SDS, one wash of 10 min at 68° C. in

1×SSC/0.1% SDS, and one wash of 5 min at 68° C. in 0.1×SSC/0.1% SDS. At low stringent hybridization and washing conditions, genes with a degree of similarity to the vip3A(a) gene as low as 30% can be identified. These low stringent conditions consist of an overnight hybridization at 52° C. followed by 2 washes of 5 min at 42° C. in 2×SSC/0.1% SDS.

Once the nucleotide sequences encoding the proteins of the VIP3 class have been isolated, they can be manipulated and used to express the protein in a variety of hosts, including microorganisms and plants.

Another means by which homologues to vip3A(a) can be identified and isolated is through the use of PCR technology. Primer sequences can be made which recognize either conserved or variable regions of the coding sequence, and then used to screen DNA samples obtained from either known or unknown strains.

Homologues to the VIP3A(a) protein can also be identified and isolated through the use of antibody cross-reaction. Either monoclonal or polyclonal antibodies can be raised against the protein and then used to screen protein preparations obtained from the strains themselves or from the medium in which they are grown. Useful methods of screening protein samples obtained in this way include but are not limited to Western analysis and ELISA analysis.

Members of the proteins of the VIP3 class include but are not limited to VIP3A(a) isolated from strain AB88 (deposited as Accession No. NRRL B-21225) as disclosed in SEQ ID NO:1-2; VIP3A(b) isolated from strain AB424 (deposited as Accession No. NRRL B-21439) as disclosed in SEQ ID NO:3-4; and VIP3A(c) isolated from strain AB51 (deposited as Accession No. NRRL B-21675) as disclosed in SEQ ID NO:5-6. All deposits were made in accordance with the Budapest Treaty by submission to the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Ill. 61604, U.S.A. The DNA and protein sequences for the above three proteins are aligned in Tables 11-12.

Polypeptide Domains of the VIP3 Class of Proteins

It has been shown that the VIP3A(a) protein undergoes proteolytic processing when mixed with the gut fluids of insect larvae. When gut fluids isolated from black cutworm are mixed with purified VIP3A(a), four major proteolytic products derived from VIP3A(a) can be identified having a molecular weight of approximately 66, 45, 33 and 22 kDa. The 22 kDa band comprises the N-terminal portion of the VIP3A(a) protein from amino acid 1 to amino acid 198 of SEQ ID NO:2. The 66 kDa band comprises the rest of the VIP3A(a) protein from amino acid 200 to amino acid 789 of SEQ ID NO:2. Both the 45 and 33 kDa bands are derived by proteolysis from the 66 kDa band and constitute amino acid 412 to amino acid 789, and from amino acid 200 to amino acid 455, respectively, of SEQ ID NO:2. The 33 kDa band is the main component of the VIP3A(a) protein that remains after an incubation period of more than two hours. This 33 kDa "toxic core" domain (amino acids 200 to 455 of SEQ ID NO:2) of the VIP3A(a) protein retains full insecticidal properties against a broad spectrum of lepidopteran insects. Similar results are obtained when VIP3A(a) is incubated with gut fluids isolated from fall armyworm, another insect sensitive to VIP3A(a).

In addition to the toxic core domain, the VIP3A(a) protein also possesses a stabilizing domain at the C-terminus. The role of the stabilizing domain was explored using mutants of the VIP3A(a) protein and the VIP3A(c) protein, neither of which display insecticidal properties when ingested by

insects known to be sensitive to VIP3A(a). When similar studies addressing the stability in black cutworm gut fluid was conducted with VIP3A(a)-mutants, in particular with a mutant of the VIP3A(a) protein that contains three point mutations located at the carboxy-terminal domain (amino acid 742(E→D); amino acid 770 (S→P); and amino acid 784 (Y→H)), it was found that the protein was completely hydrolyzed. Similar results were obtained for the VIP3A(c) (SEQ ID NO:6) protein isolated from AB51, which shares an overall identity of 96% with the VIP3A(a) protein but lacks the carboxy-terminal domain of VIP3A(a). Both the mutant and VIP3A(c) protein, however, are active against the insect cell line SF-9. These results indicate that the function of the carboxy-terminal domain of proteins of the VIP3 class is to provide stability to the protein in the gut environment of susceptible insects.

Hybrid Toxins Comprising a VIP3 Region and a Heterologous Region

Toxins, enzymes, transcription factors, antibodies, cell binding moieties or other protein domains can be operably linked to the novel proteins of the present invention by producing in frame genetic fusions which, when translated by ribosomes, would produce a fusion protein with the combined attributes of the VIP and the other component used in the fusion. Furthermore, if the protein domain fused to the VIP has an affinity for another protein, nucleic acid, carbohydrate, lipid, or other chemical or factor, then a three-component complex can be formed. This complex will have the attributes of all of its components. A similar rationale can be used for producing four or more component complexes. These complexes are useful as insecticidal toxins, pharmaceuticals, laboratory reagents, and diagnostic reagents, etc. Examples where such complexes are currently used are fusion toxins for potential cancer therapies, reagents in ELISA assays and immunoblot analysis.

The hybrid toxins of the invention include chimeric proteins having a toxic core domain which is heterologous to the stabilizing domain. Hybrid toxins are also created by combining an antibody, or immunologically-active fragment thereof, which immunologically recognizes the VIP3 receptor with a toxic domain from other proteins. The toxin domain is obtained from a number of cytotoxic proteins. These include but are not limited to *Bacillus* toxins, including endotoxins and vegetative insecticidal proteins. See for example U.S. application Ser. No. 08/037,057, filed Mar. 25, 1993 and U.S. application Ser. No. 07/951,715 filed Sep. 25, 1992, herein incorporated by reference. Other toxins include catalytic ribosome inactivators such as gelonin, *Pseudomonas* exotoxin A or phytolaccin, (the structure of *Pseudomonas* exotoxin has been well characterized in Chaudhary et al., J. Biol. Chem. 265:16303–16310 (1990)); cell metabolism disrupters, such as ribonucleases, (see, for example, Mariani et al. Nature 347:737–741 (1990)); Barnase toxin (or PE-Bar), a chimeric toxin derived from *Pseudomonas* exotoxin A and a ribonuclease, (see, Prior et al Cell 64:1017–1023 (1991)); hydrophilic peptides that create pores in membranes (see, Frohlich and Wells, Int. J. Peptide Protein Res. 37:2–6 (1991)).

Mode of Action of VIP3A(a)

The VIP3A(a) protein has been shown to be active against a broad spectrum of plant pests. For example, histopathological observations indicate that VIP3A(a) ingestion by susceptible insects such as black cutworm (*Agrotis epsilon*) and fall armyworm (*Spodoptera frugiperda*) causes gut paralysis at concentrations as low as 4 ng/cm² of diet, with complete lysis of the gut epithelial cells resulting in larval death at concentrations above 40 ng/cm². Less susceptible

insects like European corn borer (*Ostrinia nubilalis*) do not develop any pathology upon ingesting VIP3A(a). While the proteolytic processing of the VIP3A(a) protein by midgut fluids obtained from susceptible and non-susceptible insects is comparable, in vivo immuno-localization studies show that VIP3A(a) binding is restricted to gut cells of susceptible insects. Therefore, the insect host range for VIP3A(a) seems to be determined by its binding ability to gut cells. Histopathological observations indicate that midgut epithelial cells of susceptible insects are the primary target for the VIP3A(a) insecticidal protein and their subsequent lysis is the primary mechanism of lethality.

Programmed cell death is an active process of self-destruction that seems to be important for development and maintenance of multicellular organisms (Clem, R. J. et al. Science 254: 1388–1390 (1991)). Cells undergoing apoptosis, which is a form of programmed cell death, generate membrane-bound apoptotic bodies and activate endogenous nucleases that cleaves the chromatin into discrete fragments. SF-9 insect cells derived from *S. frugiperda* exposed to the VIP3A(a) protein undergo a series of cytological and molecular changes including membrane protrusions, profuse vacuolization and endonucleolysis which are indicative of an apoptotic-type of programmed cell death. Histological studies have shown that the VIP3A(a) protein targets midgut epithelial cells of susceptible insects initiating a series of cytological changes comprising profuse vacuolization and swelling prior to cell lysis and larval death. These midgut cells also experienced an endonucleolysis process when exposed to the VIP3A(a) protein as revealed by in situ detection of DNA fragmentation. These results indicate that VIP3A(a) exerts its insecticidal properties on susceptible insect cells by triggering an apoptotic-type of programmed cell death.

The Receptor for VIP3A(a) has been Isolated

The immunohistochemistry results provided above indicate that VIP3A(a) has the ability to bind to the apical membranes of midgut epithelial cells and that this binding triggers the process that will eventually end with cell lysis. This indicates that there exists one or more proteins located in the apical membrane that recognize and bind to VIP3A(a) acting as a receptor. This receptor signals the interaction with VIP3A(a) and triggers the process of apoptosis. Thus, the receptor will mediate the response of the insect cell to VIP3A(a).

To isolate this receptor, a cDNA library was screened which was made from mRNA isolated from midgut tissue of black cutworm. The objective of the screen was to identify and isolate cDNA sequences which encode proteins that will interact with VIP3A(a) in the two hybrid system (see Fields, S. and Song, O.-K. Nature 340:245–246 (1989)). This approach resulted in the identification and isolation of one cDNA whose encoded protein strongly interacted with the VIP3A(a) protein. This 1.75 Kb-long cDNA (SEQ ID NO:8) encodes a protein of approximately 48 kDa (396 amino acids; see SEQ ID NO:9). The cloned cDNA is similar in size to the mRNA encoding the cDNA as analyzed by Northern. A portion of the DNA sequence which encodes the first 5 to 20 amino acids may be missing. The following features can be identified in the cDNA encoded protein (see FIG. 1): 1) it contains a signal peptide; 2) it contains a domain with homology to the so-called death domain (Feinstein, E. et al. Trends in Biochem. 20:342–344 (1995)); and 3) it contains EGF-like motifs or repeats (Fantl, W. J. et al. Annu. Rev. Biochem. 62:453–481 (1993)). A search of protein databases using the receptor of VIP3A(a) showed homology with a family of extracellular glycoproteins

known as Tenascins (Pearson, C. A. et al EMBO J. 7:2677-2681 (1988)) or Hexabrachion (Nies, D. E. et al. J. Biol. Chem. 266:2818-2823 (1991)). This family of proteins contains EGF-like repeats, interacts with multiple ligands, and performs a role in cell adhesion and/or signaling. The combination of a death domain and repeated EGF-motifs as observed in the VIP3 receptor is unique among programmed cell death receptors.

In addition, a portion of the VIP3A(a) receptor shares homology with the so-called "death domain." The death domain is a 60 to 70 amino acid long motif which is involved in protein to protein interaction and is shared by proteins with diverse cellular functions (Feinstein, E. et al. Trends in Biochem. 20:342-344 (1995)). Some of the protein members containing death domain motifs include receptors known to be associated with apoptotic processes. Some examples include the Fas receptor (Brakebush, C. et al. EMBO J. 11:943-950 (1992)) and the tumor necrosis factor (TNF) (Tartaglia, L. A. et al. Cell 74:845-853 (1993)).

Homologues to the VIP3A(a) receptor can be identified and isolated by various means, for example, by nucleic acid hybridization. Southern blot analysis can be performed on DNA samples taken from insect cells or fungal cells that has been enzyme restricted, run in agarose and blotted onto nitrocellulose and/or nylon filters. The Southern blot can be probed with the full-or partial length of the nucleic acid encoding the receptor of the VIP3A(a) protein under low stringency hybridization and washing conditions. The genes can be readily cloned and sequenced from a cDNA or genomic library. A size-selected genomic library can also be obtained to facilitate cloning of the gene(s) of interests. The technical protocols to perform the experiments outlined above are readily available (see for instance Molecular Cloning, A Laboratory Manual, Second Edition, Vols. 1-3, Sambrook et al. (eds.) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989) and reference therein).

Antibodies to VIP3A(a) and its Receptor

Polyclonal and monoclonal antibodies to a VIP3 protein or its receptor, including fragments thereof which immunologically recognize a portion of either protein, are provided. The antibody and monoclonal antibodies of the present invention can be prepared by utilizing a VIP3 protein or its receptor as the antigen.

The antibodies of the invention include polyclonal and monoclonal antibodies as well as fragments thereof which retain their ability to bind a VIP3 protein or its receptor. An antibody, monoclonal antibody, or fragment thereof is said to be capable of binding a molecule if it is capable of specifically reacting with the molecule to thereby bind the molecule to the antibody, monoclonal antibody, or fragment thereof. The term "antibody" (Ab) or "monoclonal antibody" (Mab) is meant to include intact molecules as well as fragments or binding regions or domains thereof (such as, for example, Fab and F(ab)₂ fragments) which are capable of binding hapten. Such fragments are typically produced by proteolytic cleavage, such as papain or pepsin. Alternatively, hapten-binding fragments can be produced through the application of recombinant DNA technology or through synthetic chemistry.

Methods for the preparation of the antibodies of the present invention are generally known in the art. For example, see Antibodies, A Laboratory Manual, Ed Harlow and David Lane (eds.) Cold Spring Harbor Laboratory, New York (1988), as well as the references cited therein. Standard reference works setting forth the general principles of immunology include: Klein, J. Immunology: The Science of Cell-Noncell Discrimination, John Wiley & Sons, New York

(1982); Dennett, R., et al. Monoclonal Antibodies, Mybri-doma: A New Dimension in Biological Analyses, Plenum Press, New York (1980); and Campbell, A. "Monoclonal Antibody Technology," In Laboratory Techniques in Biochemistry and Molecular Biology, Vol. 13, Burdon et al. (eds.), Elsevier, Amsterdam (1984). See also, U.S. Pat. Nos: 4,609,893; 4,713,325; 4,714,681; 4,716,111; 4,716,117; and 4,720,459.

It is recognized that following the methods described herein, antibodies specific for a particular VIP3 protein or its receptor can be generated. The subset of MAb lines which possess the desired binding specificity can be used as a source of messenger RNA for cloning of the cDNA for the particular monoclonal antibody.

The cloned DNA can then be sequenced by methods known in the art. See, for example, Sambrook et al, Molecular Cloning: A Laboratory Manual, 2nd. Edition, Cold Spring Harbor Laboratory Press, New York (1989) vol. 1-3, and the references cited therein. From the nucleic acid sequence, the protein sequence of the binding region from the selected MAb can be deduced.

One use of the antibodies and monoclonal antibodies of the invention includes but is not limited to the production of hybrid toxin molecules. That is, when linked, the monoclonal antibody or antibody fragment retains its binding properties and the toxin moiety retains its cytotoxic properties.

Various methods are known for obtaining antibody genes. One method is to clone a random library of antibody genes in a phage and screen the library for ability to bind to a VIP3 protein or its receptor. Another available approach is to generate monoclonal antibodies which bind to a VIP3 protein or its receptor and then clone the antibody genes from such lines. For the present example, the second method is used. Antibody genes can be cloned from hybridoma cells using primers to conserved DNA sequences within the constant regions and the framework regions of the variable regions and amplified for cloning using the polymerase chain reaction (PCR). See generally, Mullis et al, Meth. Enzymol., 155:335-350 (1987); Erlich, (ed.), PCR Technology, Stockton Press (New York 1989). A database of mouse heavy chain and light chain sequences compiled by Kabat et al, U.S. Dept Health and Human Services, U.S. Government Printing Offices (1991) has been successfully used to generate both isotype specific and degenerate primers for cloning antibody genes. (Jones et al Bio/technology 9:88-89 (1991)). Additionally, techniques are well known for cloning of smaller fragments of antibodies (Fab) which possess the binding properties of the original antibody. Complete antibodies are large molecules (150 kDa), but much smaller Fab and Fv antigen-binding fragments (12 kDa-50 kDa) have been shown to retain full binding affinity. Single chain Fv fragments (scFv) in which Vh and Vl domains are linked by a hydrophilic and flexible peptide have been used successfully to target enzymes and toxins to specific cells (Bird, Science 242:423-426 (1988); Huston, PNAS 85:5879-5883 (1988)). Single Vh domains (Dabs) and single complementary determining regions as small as 20 amino acids in length, called minimal recognition units (m.r.u.), have also been used for antigen binding (Ward, Nature 341:544-546 (1989); Taub, J. Biol. Chem 264:259-265 (1989); Williams, PNAS 86:5537-5541 (1989)). Thus, it is possible to reduce the binding domain specific for a VIP3 or its receptor to a very small size.

Polymerase chain reaction technology and specific oligo-nucleotide primers are used to clone immunoglobulin genes or regions from immunoglobulin genes. PCR primers specific

for both the heavy and light chains of IgM and the three IgG isotypes were selected from the Kabat database described above. Primers for the region encoding the NH₂-terminal end of the mature variable region were designed to initiate at the first framework region and were made with some degeneracy to allow these to be used as "universal primers". The 3' primers used for the specific PCR amplification of the variable regions were designed from conserved sequences of the first constant domain (CH1) of both the light and heavy chains. A different 3' primer is used for immunoglobulin isotypes IgG1, IgG3, and IgM. Isotypes IgG2A and IgG2B can be amplified with the same primers used for IgG1. Antibody variable regions are cloned into a light and heavy chain expression vector containing an endoplasmic reticulum signal peptide and the constant regions of IgG1 light and heavy chains, respectively.

Primer sequences used for the PCR cloning of the mouse immunoglobulin light and heavy variable regions are available in the published literature (Coloma et al. *Bio/Techniques* 11: 152-156 (1991); Jones et al *Bio/Technology* 9:88-89 (1991)). Oligonucleotides were made on an Applied Biosystems DNA synthesizer 380B (Applied Biosystems, Foster City, Calif.) using standard conditions as described below. The PCR primers incorporate restriction sites and, after amplification and digestion, can be cloned into a plant expression vector under the control of a plant-expressible promoter. Restriction sites were chosen that were known to be absent in sequenced antibody genes.

Another use of the polyclonal and/or monoclonal antibodies of the invention includes the stimulation of apoptosis by targeting the receptor to Vip3A with antibodies. The interaction of antibodies raised against cell surface-located proteins that are involved in controlling the cell growth result in the induction of apoptosis by means of preventing the said receptor from binding to its natural ligand(s). For instance, the anti-APO-1 antibody completely blocks proliferation of leukemia cells bearing the APO-1 protein and triggers apoptosis in these cells (Trauth, B. C. et al *Science* 245:301-305 (1989)). Also, the activity resulting from the interaction between a given receptor and a ligand is mimicked by substituting the ligand for antibodies raised against the receptor. For instance, the addition of certain anti-Fas antibodies to cells bearing the Fas receptor in their cell surfaces will mediate apoptosis in a similar fashion as when the ligand of the Fas receptor is added (Itoh, N. et al. *Cell* 66:233-243 (1991)).

The receptor to Vip3A(a) isolated from black cutworm shares homology with a family of extracellular glycoproteins known as Tenascins, and in particular with Tenascin-X (Bristow, J. et al. *J. Cell Biol.* 122:265-278 (1993)). Tenascin-Xs are known to be involved in cell-to-cell adhesion and signaling. Lack of functionality of Tenascin-X either by mutation or by removal of the gene leads to lethality. Therefore, antibodies raised against different domains of the receptor to Vip3A(a) either effectively block the receptor from binding to its ligand(s) or mimic the interaction of the Vip3A(a) protein triggering apoptosis. This approach is extended to different receptors with similar biological functions. In this sense, antibodies raised against insect cell receptors involved in crucial cell growth and interaction processes lead to induction of apoptosis and are used as an strategy to control insects.

Screening for Novel Insecticidal Activities whose Mode of Action is Apoptosis

The materials described in this invention are used to screen for chemical ligands that have pesticidal properties triggering apoptotic responses. Chemical ligands include

small organic molecules, peptides, and proteins. In one embodiment of the invention, insect cell lines are used as model organisms for insects to screen for compounds that are insecticidal as a consequence of their ability to induce apoptosis. These cell lines are handled in a high-throughput screening format where the cells are grown in multi-well plates and are exposed to a variety of compounds. Yeast is also used as a model organism. Using procedures described herein or known in the art, determining whether a compound is pesticidal as a consequence of inducing apoptosis is accomplished.

One means by which to identify compounds that trigger apoptotic responses through interaction with a known receptor is to resort to identified receptors involved in the signal transduction pathway triggered in apoptotic insect cell lines. These receptors are transformed into heterologous cell lines creating isogenic lines with one of them containing a gene for expression of a specific receptor and another one which does not either possess, or express, such a gene. These cell lines are handled in a high-throughput screening format whereby the transformed cell lines expressing the receptor have a differential response against compounds that trigger apoptosis through their specific interaction with said receptor.

Also encompassed by the present invention is the characterization of biochemical and/or molecular markers that specifically identify an insect cell line undergoing apoptosis. For example, it is possible to isolate specific cDNAs induced during an apoptotic process in specific insect cell lines. Although the death core pathway seems to be phylogenetically conserved (Nagata, S. *Cell* 88:355-365 (1997)), the signal transduction pathway from the receptor to the death core pathway is subject to variation across organisms. Messenger RNAs differentially expressed in insect cells undergoing apoptosis are identified by a number of techniques readily available such as differential display (Bauer, D. et al. *Nucleic Acid Res.* 21:4272-4280 (1993)) or subtractive libraries (Sommer, H. et al *EMBO J.* 9:605-613 (1990)). The differentially expressed cDNA-encoded proteins are used as markers for apoptosis in specific insect cell lines. Transgenic Plants Comprising a DNA Sequence Encoding a Protein of the VIP3 Class

A host plant expressing at least one of the sequences of the invention has enhanced resistance to attack by plant pests and is thus better equipped to withstand crop losses associated with such attack. By plant is meant any plant species which can be genetically transformed by methods known in the art. Methods known in the art for plant transformation are discussed below. Host plants include, but are not limited to, those species previously listed as target crops.

PLANT EXPRESSION CASSETTES

Methodologies for the construction of plant expression cassettes as well as the introduction of foreign DNA into plants are described in the art. Such expression cassettes may include promoters, terminators, enhancers, leader sequences, introns and other regulatory sequences operably linked to the pesticidal protein coding sequence. It is further recognized that promoters or terminators of the VIP3 genes can be used in expression cassettes.

Toxin genes derived from microorganisms may also differ from plant genes. Plant genes differ from genes found in microorganisms in that their transcribed RNA does not possess defined ribosome binding site sequence adjacent to the initiating methionine. Consequently, microbial genes can be enhanced by the inclusion of a eukaryotic consensus translation initiator at the ATG (Kozak, *Cell* 44:283-292

(1986)). Clontech (1993/1994 catalog, page 210) has suggested the sequence GTCGACCATGGTC as a consensus translation initiator for the expression of the *E. coli* uidA gene in plants. Further, Joshi (Nucleic Acids Res. 15: 6643–6653 (1987)) has compared many plant sequences adjacent to the ATG and suggests the consensus TAAACA ATGGCT. In situations where difficulties are encountered in the expression of microbial ORFs in plants, inclusion of one of these sequences at the initiating ATG may improve translation. In such cases the last three nucleotides of the consensus may not be appropriate for inclusion in the modified sequence due to their modification of the second AA residue. Preferred sequences adjacent to the initiating methionine may differ between different plant species. By surveying the sequence of maize genes present in the GenBank/EMBL database it can be discerned which nucleotides adjacent to the ATG should be modified to enhance translation of the toxin gene introduced into maize.

In addition, it has been shown that removal of illegitimate splice sites can enhance expression and stability of introduced genes. Genes cloned from non-plant sources and not optimized for expression in plants may contain motifs which can be recognized in plants as 5' or 3' splice sites. Consequently, the transcription process can be prematurely terminated, generating truncated or deleted mRNA. The toxin genes can be engineered to remove these illegitimate splice sites using multiple techniques. For example, several available methods can be utilized to identify potential splice sites in a DNA sequence. First, potential splice sites may be identified by computer analysis of the DNA sequence. Consensus sequences which identify splice sites are known in the art. See, for example, Goodall, G. J. and Filipowicz, W., EMBO J. 10, 2635–2644 (1991) and Brown, J. W. S., Nucleic Acids Research 14, 9549–9559 (1986). Alternately, one can identify splice sites actually processed by a plant by comparing PCR analysis of cDNA derived from the gene with actual gene products. Shorter than expected products are indicative of splicing. Such smaller products are then cloned and sequenced and the exact location of the splice determined. It is also recognized that a combination of computer searching and PCR analysis can be utilized.

The novel toxin genes of the present invention, either as their native sequence or as optimized synthetic sequences as described above, can be operably fused to a variety of promoters for expression in plants including constitutive, inducible, temporally regulated, developmentally regulated, chemically regulated, tissue-preferred and tissue-specific promoters to prepare recombinant DNA molecules, i.e., chimeric genes. Preferred constitutive promoters include the CaMV 35S and 19S promoters (Fraley et al., U.S. Pat. No. 5,352,605, issued Oct. 4, 1994). An additionally preferred promoter is derived from any one of several of the actin genes, which are known to be expressed in most cell types. The promoter expression cassettes described by McElroy et al (Mol. Gen. Genet. 231: 150–160 (1991)) can be easily modified for the expression of the novel toxin gene and are particularly suitable for use in monocotyledonous hosts.

Yet another preferred constitutive promoter is derived from ubiquitin, which is another gene product known to accumulate in many cell types. The ubiquitin promoter has been cloned from several species for use in transgenic plants (e.g. sunflower—Binet et al Plant Science 79: 87–94 (1991), maize—Christensen et al Plant Molec. Biol. 12: 619–632 (1989)). The maize ubiquitin promoter has been developed in transgenic monocot systems and its sequence and vectors constructed for monocot transformation are disclosed in the patent publication EP 0 342 926. The ubiquitin promoter is

suitable for the expression of the novel toxin gene in transgenic plants, especially monocotyledons.

Tissue-specific or tissue-preferential promoters useful for the expression of the novel toxin gene in plants, particularly maize, are those which direct expression in root, pith, leaf or pollen. Such promoters are disclosed in U.S. Ser. No. 07/951,715 and published as WO 93/07278, herein incorporated by reference in its entirety. Chemically inducible promoters useful for directing the expression of the novel toxin gene in plants are disclosed in U.S. Ser. No. 08/181,271, herein incorporated by reference in its entirety.

In addition to promoters, a variety of transcriptional terminators are also available for use in chimeric gene construction using the novel toxin gene of the present invention. Transcriptional terminators are responsible for the termination of transcription beyond the transgene and its correct polyadenylation. Appropriate transcriptional terminators and those which are known to function in plants include the CaMV 35S terminator, the tml terminator, the nopaline synthase terminator, the pea rbcS E9 terminator and others known in the art. These can be used in both monocotyledons and dicotyledons.

Numerous sequences have also been found to enhance gene expression from within the transcriptional unit and these sequences can be used in conjunction with the novel toxin gene of this invention to increase their expression in transgenic plants. Various intron sequences have been shown to enhance expression, particularly in monocotyledonous cells. For example, the introns of the maize Adh1 gene have been found to significantly enhance the expression of the wild-type gene under its cognate promoter when introduced into maize cells (Callis et al, Genes Develop. 1: 1183–1200 (1987)). Intron sequences have been routinely incorporated into plant transformation vectors, typically within the non-translated leader.

A number of non-translated leader sequences derived from viruses are also known to enhance expression, and these are particularly effective in dicotyledonous cells. Specifically, leader sequences from Tobacco Mosaic Virus (TMV, the “Ω-sequence”), Maize Chlorotic Mottle Virus (MCMV), and Alfalfa Mosaic Virus (AMV) have been shown to be effective in enhancing expression (e.g. Gallie et al. Nucl. Acids Res. 15: 8693–8711 (1987); Skuzeski et al. Plant Molec. Biol. 15: 65–79 (1990)).

The 5' regulatory region of the expression cassette may also include other enhancing sequences. Numerous sequences have been found to enhance gene expression in transgenic plants. For example, a number of non-translated leader sequences derived from viruses are known to enhance expression. Specifically, leader sequences from Tobacco Mosaic Virus (TMV, the “W-sequence”), Maize Chlorotic Mottle Virus (MCMV), and Alfalfa Mosaic Virus (AMV) have been shown to be effective in enhancing expression (e.g. Gallie et al. Nucl. Acids Res. 15: 8693–8711 (1987); Skuzeski et al. Plant Molec. Biol. 15: 65–79 (1990)). Other leaders known in the art include but are not limited to:

Picornavirus leaders, for example, EMCV leader (Encephalomyocarditis 5' noncoding region) (Elroy-Stein, O., Fuerst, T. R., and Moss, B. PNAS USA 86: 6126–6130 (1989));

Potyvirus leaders, for example, TEV leader (Tobacco Etch Virus) (Allison et al., (1986); MDMV leader (Maize Dwarf Mosaic Virus); Virology, 154: 9–20);

Human immunoglobulin heavy-chain binding protein (BiP) leader, (Macejak, D. G., and Sarnow, P., Nature, 353: 90–94 (1991);

Untranslated leader from the coat protein mRNA of alfalfa mosaic virus (AMV RNA 4), (Jobling, S. A., and Gehrke, L., *Nature*, 325:622-625 (1987);

Tobacco mosaic virus leader (TMV), (Gallie, D. R. et al., *Molecular Biology of RNA*, pages 237-256 (1989); and

Maize Chlorotic Mottle Virus leader (MCMV) (Lommel, S. A. et al., *Virology*, 81:382-385 (1991). See also, Della-Cioppa et al., *Plant Physiology*, 84:965-968 (1987).

Various intron sequences have been shown to enhance expression when added to the 5' regulatory region, particularly in monocotyledonous cells. For example, the introns of the maize Adh1 gene have been found to significantly enhance the expression of the wild-type gene under its cognate promoter when introduced into maize cells (Callis et al., *Genes Develop.* 1:1183-1200(1987)).

In addition to promoters, a variety of 3' transcriptional terminators are also available for use in the present invention. Transcriptional terminators are responsible for the termination of transcription and correct mRNA polyadenylation. Appropriate transcriptional terminators and those which are known to function in plants include the CaMV 35S terminator, the tml terminator, the nopaline synthase terminator, the pea rbcS E9 terminator and others known in the art. These can be used in both monocotyledons and dicotyledons.

OPTIMIZING vip3 GENES FOR PLANT EXPRESSION

The pesticidal genes of the invention can be optimized for enhanced expression in plants. See, for example, EPA 0359472; EPA 0385962; WO 91/16432; and, Perlak et al., *Proc. Natl. Acad. Sci.* 88:3324-3328 (1991). In this manner, the coding sequences can be synthesized which are optimized for plant expression.

In one embodiment of the invention the vip3A(a) is made according to the procedure disclosed in U.S. Ser. No. 07/951,715, herein incorporated by reference. In this procedure, maize preferred codons, i.e., the single codon which most frequently encodes that amino acid in maize, are used. The maize preferred codon for a particular amino acid may be derived, for example, from known gene sequences from maize. Maize codon usage for 28 genes from maize plants is found in Murray et al., *Nucleic Acids Research* 17: 477-498 (1989), the disclosure of which is incorporated herein by reference. Examples of synthetic sequences made with maize optimized codons are set forth in SEQ ID NO:7 (VIP3A(a)), in SEQ ID NO:19 (VIP3A(b)), and in SEQ ID NO:20 (VIP3A(c)).

In this manner, the nucleotide sequences can be optimized for expression in any plant.

It is recognized that all or any part of the gene sequence may be optimized or synthetic. That is, synthetic or partially optimized sequences may also be used.

PLANT TRANSFORMATION

The recombinant DNA molecules can be introduced into the plant cell in a number of art-recognized ways. Those skilled in the art will appreciate that the choice of method might depend on the type of plant, i.e. monocot or dicot, targeted for transformation. Suitable methods of transforming plant cells include microinjection (Crossway et al., *BioTechniques* 4:320-334 (1986)), electroporation (Riggs et al, *Proc. Natl. Acad. Sci. U.S.A.* 83:5602-5606 (1986), *Agrobacterium*-mediated transformation (Hinchee et al., *Biotechnology* 6:915-921 (1988)), direct gene transfer

(Paszkowski et al., *EMBO J.* 3:2717-2722 (1984)), and ballistic particle acceleration using devices available from Agracetus, Inc., Madison, Wis. and Dupont, Inc., Wilmington, Del. (see, for example, Sanford et al., U.S. Pat. 4,945,050; and McCabe et al., *Biotechnology* 6:923-926 (1988); see also Weissinger et al., *Annual Rev. Genet.* 22:421-477 (1988); Sanford et al., *Particulate Science and Technology* 5:27-37 (1987)(onion); Christou et al., *Plant Physiol.* 87:671-674 (1988)(soybean); McCabe et al., *Bio/Technology* 6:923-926 (1988)(soybean); Datta et al., *Bio/Technology* 8:736-740 (1990)(rice); Klein et al., *Proc. Natl. Acad. Sci. U.S.A.*, 85:4305-4309 (1988)(maize); Klein et al., *Bio/Technology* 6:559-563 (1988)(maize); Klein et al., *Plant Physiol.* 91:440-444 (1988)(maize); Fromm et al., *Bio/Technology* 8:833-839 (1990); and Gordon-Kamm et al., *Plant Cell* 2:603-618 (1990)(maize); Svab et al. *Proc. Natl. Acad. Sci. U.S.A.* 87: 8526-8530 (1990) (tobacco chloroplast); Koziel et al. (*Biotechnology* 11: 194-200 (1993)) (maize); Shimamoto et al *Nature* 338: 274-277 (1989) (rice); Christou et al. *Biotechnology* 9: 957-962 (1991) (rice); European Patent Application EP 0 332 581 (orchardgrass and other Pooideae); Vasil et al (*Biotechnology* 11: 1553-1558 (1993) (wheat); Weeks et al. (*Plant Physiol.* 102: 1077-1084 (1993) (wheat); Wan et al (*Plant Physiol.* 104: 37-48 (1994)(barley)); Umbeck et al., (*Bio/Technology* 5: 263-266 (1987)(cotton); Casas, A. M. et al. *Proc. Natl. Acad. Sci. U.S.A.* 90: 11212-11216 (1991) (sorghum).

One particularly preferred set of embodiments for the introduction of the expression cassettes of the present invention into maize by microprojectile bombardment is described in U.S. Ser. No. 08/008,374, herein incorporated by reference in its entirety. An additional preferred embodiment is the protoplast transformation method for maize as disclosed in European Patent Application EP 0 292 435, as well as in U.S. Pat. No. 5,350,689, hereby incorporated by reference in its entirety. One particularly preferred set of embodiments for the introduction of the expression cassettes of the present invention into wheat by microprojectile bombardment can be found in U.S. Pat. No. 5,610,042 herein incorporated by reference in its entirety.

Transformation of plants can be undertaken with a single DNA molecule or multiple DNA molecules (i.e. co-transformation), and both these techniques are suitable for use with the expression cassettes of the present invention. Numerous transformation vectors are available for plant transformation, and the expression cassettes of this invention can be used in conjunction with any such vectors. The selection of vector will depend upon the preferred transformation technique and the target species for transformation.

Numerous transformation vectors are available for plant transformation, and the genes of this invention can be used in conjunction with any such vectors. The selection of vector for use will depend upon the preferred transformation technique and the target species for transformation.

Many vectors are available for transformation using *Agrobacterium tumefaciens*. These typically carry at least one T-DNA border sequence and include vectors such as pBIN19 (Bevan, *Nucl. Acids Res.* (1984)). In one preferred embodiment, the novel toxin gene of the present invention may be inserted into either of the binary vectors pCIB200 and pCIB2001 for use with *Agrobacterium*. These vector cassettes for *Agrobacterium*-mediated transformation can be constructed in the following manner. pTJS75kan was created by NarI digestion of pTJS75 (Schmidhauser & Helinski, *J Bacteriol.* 164: 446-455 (1985)) allowing exci-

sion of the tetracycline-resistance gene, followed by insertion of an *AccI* fragment from pUC4K carrying an NPTII (Messing & Vierra, *Gene* 19: 259–268 (1982); Bevan et al., *Nature* 304: 184–187 (1983); McBride et al., *Plant Molecular Biology* 14: 266–276 (1990)). *XhoI* linkers were ligated to the *EcoRV* fragment of pCIB7 which contains the left and right T-DNA borders, a plant selectable nos/*nptII* chimeric gene and the pUC polylinker (Rothstein et al., *Gene* 53: 153–161 (1987)), and the *XhoI*-digested fragment was cloned into *Sall*-digested pTJS75kan to create pCIB200 (see also EP 0 332 104, example 19). pCIB200 contains the following unique polylinker restriction sites: *EcoRI*, *SstI*, *KpnI*, *BglII*, *XbaI*, and *Sall*. pCIB2001 is a derivative of pCIB200 which created by the insertion into the polylinker of additional restriction sites. Unique restriction sites in the polylinker of pCIB2001 are *EcoRI*, *SstI*, *KpnI*, *BglII*, *XbaI*, *Sall*, *MluI*, *BclI*, *AvrII*, *ApaI*, *HpaI*, and *StuI*. pCIB2001, in addition to containing these unique restriction sites also has plant and bacterial kanamycin selection, left and right T-DNA borders for *Agrobacterium*-mediated transformation, the RK2-derived *trfA* function for mobilization between *E. coli* and other hosts, and the *OriT* and *OriV* functions also from RK2. The pCIB2001 polylinker is suitable for the cloning of plant expression cassettes containing their own regulatory signals.

An additional vector useful for *Agrobacterium*-mediated transformation is the binary vector pCIB10 contains a gene encoding kanamycin resistance for selection in plants, T-DNA right and left border sequences and incorporates sequences from the wide host-range plasmid pRK252 allowing it to replicate in both *E. coli* and *Agrobacterium*. Its construction is described by Rothstein et al. (*Gene* 53: 153–161 (1987)). Various derivatives of pCIB10 have been constructed which incorporate the gene for hygromycin B phosphotransferase described by Gritz et al (*Gene* 25: 179–188 (1983)). These derivatives enable selection of transgenic plant cells on hygromycin only (pCIB743), or hygromycin and kanamycin (pCIB715, pCIB717).

Methods using either a form of direct gene transfer or *Agrobacterium*-mediated transfer usually, but not necessarily, are undertaken with a selectable marker which may provide resistance to an antibiotic (e.g., kanamycin, hygromycin or methotrexate) or a herbicide (e.g., phosphinothricin). The choice of selectable marker for plant transformation is not, however, critical to the invention unless the expression of this resistance and its biochemical activity interferes with the choice of protoxin to toxin conversion chosen for use in creating conditional fertility.

For certain plant species, different antibiotic or herbicide selection markers may be preferred. Selection markers used routinely in transformation include the *nptII* gene which confers resistance to kanamycin and related antibiotics (Messing & Vierra, *Gene* 19: 259–268 (1982); Bevan et al., *Nature* 304:184–187 (1983)), the *bar* gene which confers resistance to the herbicide phosphinothricin (White et al., *Nucl Acids Res* 18:1062 (1990), Spencer et al., *Theor Appl Genet* 79:625–631(1990)), the *hph* gene which confers resistance to the antibiotic hygromycin (Blochinger & Diggelmann, *Mol Cell Biol* 4: 2929–2931), the *dhfr* gene, which confers resistance to methotrexate (Bourouis et al, *EMBO J.* 2: 1099–1104 (1983)), the mannose phosphate isomerase gene, which allows selection on mannose as a carbon source (EP 530 129,WO 94/20627).

One such vector useful for direct gene transfer techniques in combination with selection by the herbicide Basta (or phosphinothricin) is pCIB3064. This vector is based on the plasmid pCIB246, which comprises the CaMV 35S pro-

motor in operational fusion to the *E. coli* GUS gene and the CaMV 35S transcriptional terminator and is described in the PCT published application WO 93/07278, herein incorporated by reference. The gene providing resistance to phosphinothricin is the *bar* gene from *Streptomyces hygroscopicus* (Thompson et al. *EMBO J.* 6: 2519–2523 (1987)). This vector is suitable for the cloning of plant expression cassettes containing their own regulatory signals. Another useful selectable marker is obtained by operably linking a ubiquitin promoter, a synthetic PAT gene and a nos terminator. Once example of a vector comprising this marker is the plasmid pCIB9804.

An additional transformation vector is pSOG35 which utilizes the *E. coli* gene dihydrofolate reductase (DHFR) as a selectable marker conferring resistance to methotrexate. PCR was used to amplify the 35S promoter (~800 bp), intron 6 from the maize *Adh1* gene (~550 bp; see Dennis et al., *Nucleic Acid Res.* 12:3983–4000 (1984)) and 18 bp of the GUS untranslated leader sequence (see Jefferson et al., *Proc. Nat. Acad. Sci. U.S.A.* 83: 8447–8451 (1986)). A 250 bp fragment encoding the *E. coli* dihydrofolate reductase type II gene was also amplified by PCR and these two PCR fragments were assembled with a *SacI*-*PstI* fragment from pBI221 (Clontech) which comprised the pUC19 vector backbone and the nopaline synthase terminator. Assembly of these fragments generated pSOG19 which contains the 35S promoter in fusion with the intron 6 sequence, the GUS leader, the DHFR gene and the nopaline synthase terminator. Replacement of the GUS leader in pSOG19 with the leader sequence from Maize Chlorotic Mottle Virus check (MCMV) generated the vector pSOG35. pSOG19 and pSOG35 carry the pUC gene for ampicillin resistance and have *HindIII*, *SphI*, *PstI* and *EcoRI* sites available for the cloning of foreign sequences.

Another transformation vector is the vector pGL2 (Shimamoto et al. *Nature* 338, 274–276 (1989)) which contains the *Streptomyces hygromycin* phosphotransferase gene (*hpt*) operably linked to the 35S promoter and 35S terminator sequences.

Transgenic plants can also be identified through the use of a scorable marker. Examples of scorable markers useful in the invention are β -glucuronidase, green fluorescent protein, and the C1 and B-peru regulatory genes of the maize anthocyanin pathway. In addition, transgenic plants expressing a VIP3 protein can be identified by screening them for insecticidal activity without the need for either scorable or selectable markers.

Transformation of maize with a DNA sequence encoding a protein of the VIP3 class according to any of the above methods can be readily achieved by microprojectile bombardment of either immature zygotic embryos or serially-propagatable Type I embryogenic callus.

For transformation using immature zygotic embryos, ears are self-pollinated and immature zygotic embryos are obtained approximately 10 days later. Approximately eight hundred immature zygotic embryos are divided among different target plates containing a medium capable of inducing and supporting the formation of embryogenic callus. The immature zygotic embryos are transferred immediately to the same medium but containing 12% sucrose. After 5 hours, the immature zygotic embryos are bombarded with a plasmid or plasmids using the PDS-1000/He device from Bio-Rad. The plasmid or plasmids comprise a selectable marker, such as a gene conferring resistance to phosphinothricin, or a scorable marker, such as green fluorescent protein, and a gene encoding a protein of the VIP3 class prepared for

delivery to and expression in maize according to the above description. The plasmid or plasmids are precipitated onto 1 μ m gold particles essentially according to the published procedure from BioRad. The particles are delivered using a burst pressure of 1550 psi of helium. Each target plate is shot twice with the plasmid and gold particle preparation. Since in one embodiment of the invention the plasmid or plasmids comprise a chimeric gene coding for resistance to phosphinothricin this substance could be used to select transformed cells in vitro. If used, the selection agent is applied at 10 mg/L on the day of gene delivery and increased to 40 mg/L after approximately one month. The embryogenic callus so obtained may be regenerated in the presence of the selection agent phosphinothricin if the selectable marker is used. Plants are obtained from the selected embryogenic callus lines. The regenerated plants are assayed for resistance to a susceptible insect. All the plants that are resistant to the insect also express the introduced chimeric gene encoding a protein or proteins of the VIP3 class as evidenced by the detection of VIP3 protein in the plant using an ELISA assay. Plants resistant to the insect and expressing the VIP3 protein are transformed.

For transformation of maize using Type I embryogenic callus, the callus is obtained from immature zygotic embryos using standard culture techniques. For gene delivery, approximately 300 mg of the Type I callus is prepared by either chopping with a scalpel blade or by subculturing 3–5 days prior to gene delivery. Prior to gene delivery, the prepared callus is placed onto semi-solid culture medium again containing 12% sucrose. After approximately 4 hours, the tissue is bombarded using the PDS-1000/He Biolistic device from BioRad. The plasmid or plasmids comprise a selectable marker, such as a gene conferring resistance to phosphinothricin, or a scorable marker, such as green fluorescent protein, and a gene encoding a protein of the VIP3 class prepared for delivery to and expression in maize according to the above description. The plasmids are precipitated onto 1 μ m gold particles using essentially the standard protocol from BioRad. Approximately 16 hours after gene delivery the callus is transferred to standard culture medium containing 2% sucrose and, if the selectable marker is used, to 1 mg/L phosphinothricin. The callus is subcultured on selection for 8 weeks, after which surviving and growing callus is transferred to standard regeneration medium for the production of plants. The regenerated plants are assayed for resistance to a susceptible insect. All the plants that are resistant to the insect also express the introduced chimeric gene encoding a protein of the VIP3 class as evidenced by the detection of VIP3 protein in the plant using an ELISA assay. Plants resistant to the insect and expressing a protein of the VIP3 class are transformed.

SUPPLEMENTAL INSECT CONTROL PRINCIPLES

The pesticidal proteins of the invention can be used in combination with Bt δ -endotoxins or other insecticidal proteins to increase insect target range. Furthermore, the use of the VIPs of the present invention in combination with Bt δ -endotoxins or other insecticidal principles of a distinct nature has particular utility for the prevention and/or management of insect resistance.

The various insecticidal crystal proteins from *Bacillus thuringiensis* have been classified based upon their spectrum of activity and sequence similarity. The classification put forth by Höfte and Whiteley, Microbiol. Rev. 53: 242–255 (1989) placed the then known insecticidal crystal proteins

into four major classes. Generally, the major classes are defined by the spectrum of activity, with the Cry1 proteins active against Lepidoptera, Cry2 proteins active against both Lepidoptera and Diptera, Cry3 proteins active against Coleoptera, and Cry4 proteins active against Diptera.

Within each major class, the δ -endotoxins are grouped according to sequence similarity. The Cry1 proteins are typically produced as 130–140 kDa protoxin proteins which are proteolytically cleaved to produce active toxin proteins about 60–70 kDa. The active portion of the δ -endotoxin resides in the NH₂-terminal portion of the full-length molecule. Höfte and Whiteley, supra, classified the then known Cry1 proteins into six groups, 1Aa, 1Ab, 1Ac, 1B, 1C, and 1D. Since then, proteins classified as Cry1Ea, Cry1Fa, Cry9A, Cry9C and Cry9B have also been characterized.

The spectrum of insecticidal activity of an individual δ -endotoxin from *Bacillus thuringiensis* tends to be quite narrow, with a given δ -endotoxin being active against only a few insects. Specificity is the result of the efficiency of the various steps involved in producing an active toxin protein and its subsequent ability to interact with the epithelial cells in the insect digestive tract. In one preferred embodiment, expression of VIPs in a transgenic plant is accompanied by the expression of one or more Bt δ -endotoxins. Particularly preferred Bt δ -endotoxins are those disclosed in U.S. application Ser. No. 07/951,715, herein incorporated by reference.

It is well known that many δ -endotoxin proteins from *Bacillus thuringiensis* are actually expressed as protoxins. These protoxins are solubilized in the alkaline environment of the insect gut and are proteolytically converted by proteases into a toxic core fragment (Höfte and Whiteley, Microbiol. Rev. 53: 242–255 (1989)). For δ -endotoxin proteins of the CryI class, the toxic core fragment is localized in the N-terminal half of the protoxin. It is within the scope of the present invention that genes encoding either the full-length protoxin form or the truncated toxic core fragment of the novel toxin proteins can be used in plant transformation vectors to confer insecticidal properties upon the host plant.

Other insecticidal principles include protease inhibitors (both serine and cysteine types), lectins, α -amylase, peroxidase and cholesterol oxidase. Other VIP genes, such as vip 1A(a) and vip2A(a) as disclosed in U.S. Ser. No. 08/463,483 and herein incorporated by reference, are also useful in the present invention.

This co-expression of more than one insecticidal principle in the same transgenic plant can be achieved by genetically engineering a plant to contain and express all the genes necessary. Alternatively, a plant, Parent 1, can be genetically engineered for the expression of VIPs. A second plant, Parent 2, can be genetically engineered for the expression of a supplemental insect control principle. By crossing Parent 1 with Parent 2, progeny plants are obtained which express all the genes introduced into Parents 1 and 2. Recombinant Microorganisms Comprising Genes and Proteins of the VIP3 Class

It is recognized that the isolated genes of the present invention which encode a protein of the VIP3 class can be transferred into any microbial host and confer their insecticidal properties upon that host. Alternate hosts for the novel genes of the present invention can be selected as suitable for cloning purposes, for purposes of characterizing the form and function of the gene or encoded protein, for use as a fermentation host to increase production of the toxin protein, for purposes of delivering at least one of the toxin proteins

more effectively to the target insect pest, or introduction of the novel toxin gene into insect pathogens such as baculovirus (a nuclear polyhedrosis virus, e.g. *Autographa californica*) to improve their effectiveness.

It is envisioned that said alternate host would be applied to the environment or plants or animals for insect control. Microorganism hosts may be selected which are known to occupy the "phytosphere" (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the pesticide from environmental degradation and inactivation.

Such microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., *Bacillus*, *Caulobacter*, *Agmenellum*, *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Xanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylius*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*, *Arthrobacter*, *Azotobacter*, *Leuconostoc*, and *Alcaligenes*; fungi, particularly yeast, e.g., *Saccharomyces*, *Cryptococcus*, *Kluyveromyces*, *Sporobolomyces*, *Rhodotorula*, and *Aureobasidium*. Of particular interest are such phytosphere bacterial species as *Bacillus* spp., *Pseudomonas syringae*, *Pseudomonas fluorescens*, *Serratia marcescens*, *Acetobacter xylinum*, *Agrobacteria*, *Rhodopseudomonas spheroides*, *Xanthomonas campestris*, *Rhizobium melioli*, *Alcaligenes entrophus*, *Clavibacter xyli* and *Azotobacter vinlandii*; and phytosphere yeast species such as *Rhodotorula rubra*, *R. glutinis*, *R. marina*, *R. aurantiaca*, *Cryptococcus albidus*, *C. diftuens*, *C. laurentii*, *Saccharomyces rosei*, *S. pretoriensis*, *S. cerevisiae*, *Sporobolomyces rosues*, *S. odoratus*, *Kluyveromyces veronae*, and *Aureobasidium pullulans*. Of particular interest are the pigmented microorganisms.

Suitable host cells, where the pesticide-containing cells will be treated to prolong the activity of the toxin in the cell when the then treated cell is applied to the environment of the target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the toxin is unstable or the level of application sufficiently low as to avoid any possibility of toxicity to a mammalian host. As hosts, of particular interest will be the prokaryotes and the lower eukaryotes, such as fungi. Illustrative prokaryotes, both Gram-negative and -positive, include Enterobacteriaceae, such as *Escherichia*, *Erwinia*, *Shigella*, *Salmonella*, and *Proteus*; Bacillaceae; Rhizobiaceae, such as *Rhizobium*; Spirillaceae, such as photobacterium, *Zymomonas*, *Serratia*, *Aeromonas*, *Vibrio*, *Desulfovibrio*, *Spirillum*; Lactobacillaceae; Pseudomonadaceae, such as *Pseudomonas* and *Acetobacter*; Azotobacteraceae and Nitrobacteraceae. Among eukaryotes are fungi, such as *Phycomycetes* and *Ascomycetes*, which includes yeast, such as *Saccharomyces* and *Schizosaccharomyces*; and Basidiomycetes yeast, such as *Rhodotorula*, *Aureobasidium*, *Sporobolomyces*, and the like.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the protein gene into the host, availability of expression systems, efficiency of expression, stability of the protein in the host, and the presence of auxiliary genetic capabilities.

Characteristics of interest for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the toxin; and the like. Other considerations include ease of formulation and handling, economics, storage stability, and the like.

Host organisms of particular interest include yeast, such as *Rhodotorula* sp., *Aureobasidium* sp., *Saccharomyces* sp., and *Sporobolomyces* sp.; phylloplane organisms such as *Pseudomonas*, *Erwinia* sp. and *Flavobacterium* sp.; or such other organisms as *Escherichia*, *Lactobacillus* sp., *Bacillus* sp., and the like. Specific organisms include *Pseudomonas aeruginosa*, *Pseudomonas fluorescens*, *Saccharomyces cerevisiae*, *Bacillus thuringiensis*, *Escherichia coli*, *Bacillus subtilis*, and the like.

A number of ways are available for introducing a gene expressing the pesticidal protein into the microorganism host under conditions which allow for stable maintenance and expression of the gene. For example, expression cassettes can be constructed which include the DNA constructs of interest operably linked with the transcriptional and translational regulatory signals for expression of the DNA constructs, and a DNA sequence homologous with a sequence in the host organism, whereby integration will occur, and/or a replication system which is functional in the host, whereby integration or stable maintenance will occur.

Transcriptional and translational regulatory signals include but are not limited to promoter, transcriptional initiation start site, operators, activators, enhancers, other regulatory elements, ribosomal binding sites, an initiation codon, termination signals, and the like. See, for example, U.S. Pat. No. 5,039,523; U.S. Pat. No. 4,853,331; EPO 0480762A2; Sambrook et al. supra; Molecular Cloning, a Laboratory Manual, Maniatis et al. (eds) Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1982); Advanced Bacterial Genetics, Davis et al. (eds.) Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1980); and the references cited therein.

The novel genes or recombinant forms thereof can be transformed into such alternate hosts using a variety of art recognized methods. One such preferred method is electroporation of microbial cells, as described, for example, by the method of Dower (U.S. Pat. No. 5,186,800). Another preferred method is that of Schurter et al. (Mol. Gen. Genet. 218: 177-181 (1989)), which is also disclosed in U.S. Ser. No. 07/353,565 which is incorporated herein in its entirety.

Genes encoding the VIP3 class of proteins can be introduced into microorganisms that multiply on plants (epiphytes) or in plants (endophytes) to deliver proteins of the VIP3 class to potential target pests. Many bacterial species are capable of living in the vascular tissues of plants. Most of these endophytes and epiphytes appear to have little physiological impact on plant growth and productivity.

Root colonizing bacteria, for example, can be isolated from the plant of interest by methods known in the art. Specifically, a *Bacillus cereus* strain which colonizes roots could be isolated from roots of a plant (for example see J. Handelsman, S. Raffel, E. Mester, L. Wunderlich and C. Grau, Appl. Environ. Microbiol. 56:713-718, (1990)). Vip3 genes can also be introduced into a root colonizing *Bacillus cereus* by standard methods known in the art. Specifically, a gene encoding a protein of the VIP3 class derived from strain AB88 can be introduced into a root colonizing *Bacillus cereus* by means of conjugation using standard methods (J. Gonzalez, B. Brown and B. Carlton, Proc. Natl. Acad. Sci. 79:6951-6955, (1982)).

Also, the novel genes of the invention can be introduced into the root colonizing *Bacillus* by means of electrotransformation. For example, vip3A(a) can be cloned into a shuttle vector, for example, pHT3101 (D. Lereclus et al., FEMS Microbiol. Letts., 60:211–218 (1989)). The shuttle vector pHT3101 containing the coding sequence can then be transformed into the root colonizing *Bacillus* by means of electroporation (D. Lereclus et al. 1989, FEMS Microbiol. Letts. 60:211–218). It is also possible to use the cotton colonizing *Bacillus megaterium*.

Another example is afforded by the endophyte *Clavibacter xyli*, which is from a genus/species known contain phytopathogenic bacteria which cause plant stunting. This bacterium can grow to very high levels in the vascular system of plants. A δ -endotoxin was introduced into this endophyte, which when inoculated into a plant, provided good control of corn borer. Other endophytes are also known.

Expression systems can be designed so that VIP3 proteins are secreted outside the cytoplasm of gram negative bacteria, *E. coli*, for example. Advantages of having VIP3 proteins secreted are (1) it can increase the level of VIP3 protein expressed and (2) can aid in efficient purification of VIP3 protein.

VIP3 proteins can be made to be secreted in *E. coli*, for example, by fusing an appropriate *E. coli* signal peptide to the amino-terminal end of the VIP3 signal peptide or replacing the VIP3 signal peptide with the *E. coli* signal peptide. Signal peptides recognized by *E. coli* can be found in proteins already known to be secreted in *E. coli*, for example the OmpA protein (J. Ghayeb, H. Kimura, M. Takahara, Y. Masui and M. Inouye, EMBO J., 3:2437–2442 (1984)). OmpA is a major protein of the *E. coli* outer membrane and thus its signal peptide is thought to be efficient in the translocation process. Also, the OmpA signal peptide does not need to be modified before processing as may be the case for other signal peptides, for example lipoprotein signal peptide (G. Duffaud, P. March and M. Inouye, Methods in Enzymology. 153:492 (1987)).

Specifically, unique BamHI restriction sites can be introduced at the amino-terminal and carboxy-terminal ends of the VIP coding sequences using standard methods known in the art. These BamHI fragments can be cloned, in frame, into the vector pIN-III-ompA1, A2 or A3 (J. Ghayeb, H. Kimura, M. Takahara, H. Hsiung, Y. Masui and M. Inouye, EMBO J., 3:2437–2442 (1984)) thereby creating ompA:VIP fusion gene which is secreted into the periplasmic space. The other restriction sites in the polylinker of pIN-III-ompA can be eliminated by standard methods known in the art so that the VIP3 amino-terminal amino acid coding sequence is directly after the ompA signal peptide cleavage site. Thus, the secreted VIP3 sequence in *E. coli* would then be identical to the native VIP3 sequence.

When the VIP3 native signal peptide is not needed for proper folding of the mature protein, such signal sequences can be removed and replaced with the ompA signal sequence. Unique BamHI restriction sites can be introduced at the amino-termini of the proprotein coding sequences directly after the signal peptide coding sequences of VIP3 and at the carboxy-termini of VIP3 coding sequence. These BamHI fragments can then be cloned into the pIN-III-ompA vectors as described above.

General methods for employing the strains of the invention in pesticide control or in engineering other organisms as pesticidal agents are known in the art. See, for example U.S. Pat. No. 5,039,523 and EP 0480762A2.

VIP3 can be fermented in a bacterial host and the resulting bacteria processed and used as a microbial spray in the same

manner that *Bacillus thuringiensis* strains have been used as insecticidal sprays. In the case of a VIP3 which is secreted from *Bacillus*, the secretion signal is removed or mutated using procedures known in the art. Such mutations and/or deletions prevent secretion of the VIP3 protein(s) into the growth medium during the fermentation process. The VIP3 proteins are retained within the cell and the cells are then processed to yield the encapsulated VIP3 protein. Any suitable microorganism can be used for this purpose. *Pseudomonas* has been used to express *Bacillus thuringiensis* endotoxins as encapsulated proteins and the resulting cells processed and sprayed as an insecticide. (H. Gaertner et al. 1993, In Advanced Engineered Pesticides, L. Kim ed.)

Various strains of *Bacillus thuringiensis* are used in this manner. Such Bt strains produce endotoxin protein(s) as well as VIP3. Alternatively, such strains can produce only VIP3. A sporulation deficient strain of *Bacillus subtilis* has been shown to produce high levels of the Cry3A endotoxin from *Bacillus thuringiensis* (Agaisse, H. and Lereclus, D., "Expression in *Bacillus subtilis* of the *Bacillus thuringiensis* CryIIIA toxin gene is not dependent on a sporulation-specific sigma factor and is increased in a spoOA mutant", J. Bacteriol., 176:4734–4741 (1994)). A similar spoOA mutant can be prepared in *Bacillus thuringiensis* and used to produce encapsulated VIP3 which are not secreted into the medium but are retained within the cell.

Target crops to be protected within the scope of the present invention comprise, e.g., the following species of plants:

cereals (wheat, barley, rye, oats, rice, sorghum and related crops), beet (sugar beet and fodder beet), forage grasses (orchardgrass, fescue, and the like), drupes, pomes and soft fruit (apples, pears, plums, peaches, almonds, cherries, strawberries, raspberries and blackberries), leguminous plants (beans, lentils, peas, soybeans), oil plants (rape, mustard, poppy, olives, sunflowers, coconuts, castor oil plants, cocoa beans, groundnuts), cucumber plants (cucumber, marrows, melons) fiber plants (cotton, flax, hemp, jute), citrus fruit (oranges, lemons, grapefruit, mandarins), vegetables (spinach, lettuce, asparagus, cabbages and other Brassicae, onions, tomatoes, potatoes, paprika), lauraceae (avocados, carrots, cinnamon, camphor), deciduous trees and conifers (e.g. linden-trees, yew-trees, oak-trees, alders, poplars, birch-trees, firs, larches, pines), or plants such as maize, tobacco, nuts, coffee, sugar cane, tea, vines, hops, bananas and natural rubber plants, as well as ornamentals (including composites).

The microorganisms which have been genetically altered to contain the pesticidal gene and protein may be used for protecting agricultural crops and products from pests. In one aspect of the invention, whole, i.e., unlysed, cells of a toxin (pesticide)-producing organism are treated with reagents that prolong the activity of the toxin produced in the cell when the cell is applied to the environment of target pest(s).

Alternatively, the pesticides are produced by introducing a heterologous gene into a cellular host. Expression of the heterologous gene results, directly or indirectly, in the intracellular production and maintenance of the pesticide. These cells are then treated under conditions that prolong the activity of the toxin produced in the cell when the cell is applied to the environment of target pest(s). The resulting product retains the toxicity of the toxin. These naturally encapsulated pesticides may then be formulated in accordance with conventional techniques for application to the environment hosting a target pest, e.g., soil, water, and foliage of plants. See, for example EPA 0192319, and the references cited therein.

The active ingredients of the present invention are normally applied in the form of compositions and can be applied to the crop area or plant to be treated, simultaneously or in succession, with other compounds. These compounds can be both fertilizers or micronutrient donors or other preparations that influence plant growth. They can also be selective herbicides, insecticides, fungicides, bactericides, nematocides, molluscicides or mixtures of several of these preparations, if desired, together with further agriculturally acceptable carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation. Suitable carriers and adjuvants can be solid or liquid and correspond to the substances ordinarily employed in formulation technology, e.g. natural or regenerated mineral substances, solvents, dispersants, wetting agents, tackifiers, binders or fertilizers.

Preferred methods of applying an active ingredient of the present invention or an agrochemical composition of the present invention which contains at least one of the pesticidal proteins produced by the bacterial strains of the present invention are leaf application, seed coating and soil application. The number of applications and the rate of application depend on the intensity of infestation by the corresponding pest.

Entomocidal Compositions Comprising a Recombinant *Bacillus thuringiensis* Strain

The present invention further provides an entomocidal composition comprising a recombinant *Bacillus thuringiensis* strain containing at least one of the novel toxin genes in recombinant form, or derivatives or mutants thereof, together with an agricultural adjuvant such as a carrier, diluent, surfactant or application-promoting adjuvant. The composition may also contain a further biologically active compound selected from fertilizers, micronutrient donors, plant growth preparations, herbicides, insecticides, fungicides, bactericides, nematocides and molluscicides and mixtures thereof. The composition may comprise from 0.1 to 99% by weight of a recombinant *Bacillus thuringiensis* strain containing at least one of the novel genes in recombinant form, or the derivatives or mutants thereof, from 1 to 99.9% by weight of a solid or liquid adjuvant, and from 0 to 25% by weight of a surfactant. The recombinant *Bacillus thuringiensis* strain containing at least one of the novel genes in recombinant form, or the composition containing it, may be administered to the plants or crops to be protected together with certain other insecticides or chemicals (1993 Crop Protection Chemicals Reference, Chemical and Pharmaceutical Press, Canada) without loss of potency. It is compatible with most other commonly used agricultural spray materials but should not be used in extremely alkaline spray solutions. It may be administered as a dust, a suspension, a wettable powder or in any other material form suitable for agricultural application.

A recombinant *Bacillus thuringiensis* strain containing at least one of the novel genes in recombinant form is normally applied in the form of entomocidal compositions and can be applied to the crop area or plant to be treated, simultaneously or in succession, with further biologically active compounds. These compounds may be both fertilizers or micronutrient donors or other preparations that influence plant growth. They may also be selective herbicides, insecticides, fungicides, bactericides, nematocides, molluscicides or mixtures of several of these preparations, if desired together with further carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation. Suitable carriers and adjuvants can be solid or liquid and correspond to the substances ordinarily employed in formu-

lation technology, e.g. natural or regenerated mineral substances, solvents, dispersants, wetting agents, tackifiers, binders or fertilizers. The formulations, i.e. the entomocidal compositions, preparations or mixtures containing the recombinant *Bacillus thuringiensis* strain containing the novel gene in recombinant form as an active ingredient or combinations thereof with other active ingredients, and, where appropriate, a solid or liquid adjuvant, are prepared in known manner, e.g., by homogeneously mixing and/or grinding the active ingredients with extenders, e.g., solvents, solid carriers, and in some cases surface-active compounds (surfactants).

Suitable solvents are: aromatic hydrocarbons, preferably the fractions containing 8 to 12 carbon atoms, e.g. xylene mixtures or substituted naphthalenes, phthalates such as dibutyl phthalate or dioctyl phthalate, aliphatic hydrocarbons such as cyclohexane or paraffins, alcohols and glycols and their ethers and esters, such as ethanol, ethylene glycol monomethyl or monoethyl ether, ketones such as cyclohexanone, strongly polar solvents such as N-methyl-2-pyrrolidone, dimethylsulfoxide or dimethylformamide, as well as vegetable oils such as epoxidised coconut oil or soybean oil; or water.

The solid carriers used, e.g., for dusts and dispersible powders, are normally natural mineral fillers such as calcite, talcum, kaolin, montmorillonite or attapulgite. In order to improve the physical properties it is also possible to add highly dispersed silicic acid or highly dispersed absorbent polymers. Suitable granulated adsorptive carriers are porous types, for example pumice, broken brick, sepiolite or bentonite; and suitable nonsorbent carriers are materials such as calcite or sand. In addition, a great number of pregranulated materials of inorganic or organic nature can be used, e.g. especially dolomite or pulverized plant residues.

Depending on the nature of the active ingredients to be formulated, suitable surface-active compounds are non-ionic, cationic and/or anionic surfactants having good emulsifying, dispersing and wetting properties. The term "surfactants" will also be understood as comprising mixtures of surfactants. Suitable anionic surfactants can be both water-soluble soaps and water-soluble synthetic surface-active compounds. Suitable soaps are the alkali metal salts, alkaline earth metal salts or unsubstituted or substituted ammonium salts of higher fatty acids (C sub 10-C sub 22), e.g. the sodium or potassium salts of oleic or stearic acid, or of natural fatty acid mixtures which can be obtained, e.g. from coconut oil or tallow oil. Further suitable surfactants are also the fatty acid methyltaurin salts as well as modified and unmodified phospholipids.

More frequently, however, so-called synthetic surfactants are used, especially fatty sulfonates, fatty sulfates, sulfonated benzimidazole derivatives or alkylarylsulfonates. The fatty sulfonates or sulfates are usually in the forms of alkali metal salts, alkaline earth metal salts or unsubstituted or substituted ammonium salts and generally contain a C sub 8-C sub 22 alkyl radical which also includes the alkyl moiety of acyl radicals, e.g. the sodium or calcium salt of lignosulfonic acid, of dodecylsulfate, or of a mixture of fatty alcohol sulfates obtained from natural fatty acids. These compounds also comprise the salts of sulfuric acid esters and sulfonic acids of fatty alcohol/ethylene oxide adducts. The sulfonated benzimidazole derivatives preferably contain 2 sulfonic acid groups and one fatty acid radical containing about 8 to 22 carbon atoms. Examples of alkylarylsulfonates are the sodium, calcium or triethanolamine salts of dodecylbenzenesulfonic acid, dibutyl-naphthalenesulfonic acid, or of a naphthalenesulfonic acid/formaldehyde condensation

product. Also suitable are corresponding phosphates, e.g. salts of the phosphoric acid ester of an adduct of p-nonylphenol with 4 to 14 moles of ethylene oxide. Non-ionic surfactant are preferably polyglycol ether derivatives of aliphatic or cycloaliphatic alcohols, or saturated or unsaturated fatty acids and alkylphenols, said derivatives containing 3 to 30 glycol ether groups and 8 to 20 carbon atoms in the (aliphatic) hydrocarbon moiety and 6 to 18 carbon atoms in the alkyl moiety of the alkylphenols.

Further suitable non-ionic surfactants are the water-soluble adducts of polyethylene oxide with polypropylene glycol, ethylenediaminopolypropylene glycol and alkylpolypropylene glycol containing 1 to 10 carbon atoms in the alkyl chain, which adducts contain 20 to 250 ethylene glycol ether groups and 10 to 100 propylene glycol ether groups. These compounds usually contain 1 to 5 ethylene glycol units per propylene glycol unit. Representative examples of non-ionic surfactants are nonylphenolpolyethoxyethanols, castor oil polyglycol ethers, polypropylene/polyethylene oxide adducts, tributylphenoxypolyethoxyethanol, polyethylene glycol and octylphenoxypolyethoxyethanol. Fatty acid esters of polyoxyethylene sorbitan, such as polyoxyethylene sorbitan trioleate, are also suitable non-ionic surfactants.

Cationic surfactants are preferably quaternary ammonium salts which contain, as N-substituent, at least one C₈-C₂₂ alkyl radical and, as further substituents, lower unsubstituted or halogenated alkyl, benzyl or hydroxyl-lower alkyl radicals. The salts are preferably in the form of halides, methylsulfates or ethylsulfates, e.g., stearyltrimethylammonium chloride or benzyldi-(2-chloroethyl)ethylammonium bromide.

The surfactants customarily employed in the art of formulation are described, e.g., in "McCutcheon's Detergents and Emulsifiers Annual", MC Publishing Corp. Ridgewood, N.J., 1979; Dr. Helmut Stache, "Tensid Taschenbuch" (Handbook of Surfactants), Carl Hanser Verlag, Munich/Vienna.

Another particularly preferred characteristic of an entomocidal composition of the present invention is the persistence of the active ingredient when applied to plants and soil. Possible causes for loss of activity include inactivation by ultra-violet light, heat, leaf exudates and pH. For example, at high pH, particularly in the presence of reductant, δ -endotoxin crystals are solubilized and thus become more accessible to proteolytic inactivation. High leaf pH might also be important, particularly where the leaf surface can be in the range of pH 8–10. Formulation of an entomocidal composition of the present invention can address these problems by either including additives to help prevent loss of the active ingredient or encapsulating the material in such a way that the active ingredient is protected from inactivation. Encapsulation can be accomplished chemically (McGuire and Shasha, 1992) or biologically (Barnes and Cummings, 1986). Chemical encapsulation involves a process in which the active ingredient is coated with a polymer while biological encapsulation involves the expression of the δ -endotoxin genes in a microbe. For biological encapsulation, the intact microbe containing the δ -endotoxin protein is used as the active ingredient in the formulation. The addition of UV protectants might effectively reduce irradiation damage. Inactivation due to heat could also be controlled by including an appropriate additive.

The entomocidal compositions usually contain 0.1 to 99%, preferably 0.1 to 95%, of a recombinant *Bacillus thuringiensis* strain containing at least one of the novel genes in recombinant form, or combination thereof with

other active ingredients, 1 to 99.9% of a solid or liquid adjuvant, and 0 to 25%, preferably 0.1 to 20%, of a surfactant. Whereas commercial products are preferably formulated as concentrates, the end user will normally employ dilute formulations of substantially lower concentration. The entomocidal compositions may also contain further ingredients, such as stabilizers, antifoams, viscosity regulators, binders, tackifiers as well as fertilizers or other active ingredients in order to obtain special effects.

Methods of Controlling Insects

In view of the above description of the invention, it is apparent that there are several methods by which insects may be controlled using proteins of the VIP3 class as an insecticidal principle, either alone or in combination with supplementary insect control principles such as δ -endotoxins. Any method of delivering a VIP3 protein for ingestion by a susceptible insect will result in the control of that insect.

In one embodiment of the invention, plants are transformed with a gene encoding a protein of the VIP3 class. Expression of the protein may occur at any time during growth and development of the plant, depending on the nature of the insect to be controlled. For example, a protein of the VIP3 class can, according to the invention, be expressed in roots, stems, leaves, seeds, pollen, etc. This provides the advantage of expressing the protein only in those cells or tissues upon which the target insect feeds. Feeding the cells or tissues of a plant expressing VIP3 protein to a susceptible insect will result in the control of that insect. In one embodiment of the invention, a VIP3 protein is expressed in the stem or stalk of a plant in order to control black cutworm. The plants may be grown under either field or greenhouse conditions. Seed containing a VIP3 protein can also be protected against insect damage when in storage.

EXAMPLES

The following examples further describe the materials and methods used in carrying out the invention and the subsequent results. They are offered by way of illustration, and their recitation should not be considered as a limitation of the claimed invention.

Example 1

Isolation and Biological Characterization of *Bacillus thuringiensis* Strain AB88

A Bt strain, designated AB88, was isolated from grain bin dust samples by standard methodologies. A subculture of AB88 was grown for 24–48 hrs. and cell-free culture supernatant was tested for insecticidal activity as follows. For European corn borer (*Ostirina nubilalis*), black cutworm (*Agrotis ipsilon*), fall armyworm (*Spodoptera frugiperda*), corn earworm (*Helicoverpa zea*) and tobacco budworm (*Heliothis virescens*) 100 μ l of culture supernatant was pipetted onto the surface of 18 cm² of solidified artificial diet (Bioserv #F9240) and allowed to air dry. Neonates were then placed onto the surface of the diet and held at 30° C. Mortality was scored after 36–72 hrs.

For Colorado potato beetle (*Leptinotarsa decemlineata*) five cm² potato leaf pieces were dipped into culture supernatant, air dried, and placed on moistened filter paper in 50x9 mm petri dishes. Neonates were then placed on the leaf pieces and held at 30° C. Mortality was scored after 36–72 hrs.

For western corn rootworm (*Diabrotica virgifera*) culture supernatant was mixed with molten artificial diet (Marrone

et al. (1985) J. of Economic Entomology 78:290–293) and allowed to solidify. Solidified diet was cut into pieces and placed in plastic dishes. Neonates were then placed on the diet pieces and held at 30° C. Mortality was scored after 6 days.

For all bioassays half of the culture supernatant sample was autoclaved 15 minutes to test for the presence of β -exotoxin. The results are as follows:

TABLE 11

Insect species tested	Order	Percent mortality of culture supernatant	
		Non-autoclaved	Autoclaved
<i>Agrotis ipsilon</i>	Lepidoptera	100	5
<i>Ostrinia nubilalis</i>	Lepidoptera	100	0
<i>Spodoptera frugiperda</i>	Lepidoptera	100	4
<i>Helicoverpa zea</i>	Lepidoptera	100	12
<i>Heliothis virescens</i>	Lepidoptera	100	12
<i>Leptinotarsa decemlineata</i>	Coleoptera	0	0
<i>Diabrotica virgifera virgifera</i>	Coleoptera	0	5

The reduction of insecticidal activity of the culture supernatant to insignificant levels by autoclaving indicates that the active principle is not β -exotoxin.

Delta-endotoxin crystals were purified from strain AB88 by standard methodologies. No activity from pure crystals was observed when bioassayed against *Agrotis ipsilon*.

Example 2

Purification of the VIP3A(a) Protein from Strain AB88

Bacterial liquid culture was grown overnight at 30° C. in TB media and the supernatant retained. Proteins were precipitated with ammonium sulfate (70% saturation), centrifuged and the pellet retained. The pellet was resuspended in the original volume of 20 mM Tris pH 7.5 and dialyzed against the same buffer. AB88 proteins have been separated by several different methods following clarification including isoelectric focusing (Rotofor, BioRad, Hercules, Calif.), precipitation at pH 4.5, ion-exchange chromatography, size exclusion chromatography and ultrafiltration.

European corn borer (ECB)-active protein remained in the pellet obtained by pH 4.5 precipitation of dialysate. When preparative IEF was done on the dialysate using pH 3–10 ampholytes, ECB insecticidal activity was found in all fractions with pH of 7 or greater. SDS-PAGE analysis of these fractions showed protein bands of MW ~60 kDa and ~80 kDa. The 60 kDa and 80 kDa bands were separated by anion exchange HPLC on a Poros-Q column (PerSeptive Biosystems, Cambridge, Mass.). N-terminal sequence was obtained from two fractions containing proteins of slightly differing MW, but both of approximately 60 kDa in size. The sequences obtained were similar to each other and to some δ -endotoxins.

anion exchange fraction 23 (smaller): xEPFVSAxxxQxxx (SEQ ID NO:10)

anion exchange fraction 28 (larger): xEYENVEPFVSAx (SEQ ID NO:11)

When the ECB-active pH 4.5 pellet was further separated by anion exchange on a Poros-Q column, activity was found only in fractions containing a major band of ~60 kDa.

Black cutworm-active protein also remained in the pellet when AB88 dialysate was brought down to pH 4.5. In

preparative IEF using pH 3–10 ampholytes, activity was not found in the ECB-active IEF fractions; instead, it was highest in a fraction of pH 4.5–5.0. Its major components have molecular weights of ~35 and ~80 kDa.

The pH 4.5 pellet was separated by anion exchange HPLC to yield fractions containing only the 35 kDa material and fractions containing both 35 kDa and 80 kDa bands.

Example 3

Characterization of VIP3A(a) from AB88

Fractions containing the various lepidopteran active vegetative proteins were generated as described in Example 2. Biological analysis of fractions demonstrated that different VIPs were responsible for the different lepidopteran species activity.

The *Agrotis ipsilon* activity is due to an 80 kDa and/or a 35 kDa protein, either delivered singly or in combination. These proteins are not related to any δ -endotoxins from Bt as evidenced by the lack of sequence homology of known Bt δ -endotoxin sequences. Also, these proteins are not found in the AB88 δ -endotoxin crystal. N-terminal sequences of the major δ -endotoxin proteins were compared with the N-terminal sequences of the 80 kDa and 35 kDa VIP and revealed no sequence homology. A summary of the results follows:

TABLE 12

Agrotis VIP N-terminal sequences	N-terminal sequence of major δ -endotoxin proteins
	130 kDa
	MDNNPNINE (SEQ ID NO:14)
80 kDa	80 kDa
MKNKNTKLPTRALP (SEQ ID NO:12)	MDNNPNINE (SEQ ID NO:15)
	60 kDa
	MNVLNSGRITI (SEQ ID NO:16)
35 kDa	
ALSENTGKDGGYIVP (SEQ ID NO:13)	

The *Ostrinia nubilalis* activity is due to a 60 kDa VIP and the *Spodoptera frugiperda* activity is due to a VIP of unknown size.

Bacillus thuringiensis strain AB88 has been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Ill. 61604, U.S.A. and given the Accession No. NRRL B-21225.

Example 4

Isolation and Biological Activity of *Bacillus thuringiensis* AB424

A *B. thuringiensis* strain, designated AB424, was isolated from a moss covered pine cone sample by standard methods known in the art. A subculture of AB424 was grown and prepared for bioassay.

Biological activity was evaluated as described in Example 1. The results are as follows:

TABLE 13

Insect species tested	Percent mortality
<i>Ostrinia nubilalis</i>	100
<i>Agrotis ipsilon</i>	100
<i>Diabrotica virgifera virgifera</i>	0

Strain AB424 has been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Ill. 61604, U.S.A., and given Accession No. NRRL B-21439.

Example 5

Cloning of the vip3A(a) and vip3A(b) Genes Which Encode Proteins Active Against Black Cutworm

DNA from isolates AB88 and AB424 was digested with the restriction enzymes XbaI and EcoRI respectively, ligated into pBluescript vector previously linearized with the same enzymes and dephosphorylated, and transformed into *E. coli* DH5 α strain. Recombinant clones were blotted onto nitrocellulose filters which were subsequently probed with a 33-base long oligonucleotide corresponding to the 11-N terminal amino acids of the 80 kDa protein active against *Agrotis ipsilon* (black cutworm). Four out of 400 recombinant clones were positive. Insect bioassays of the positive recombinants exhibited toxicity to black cutworm larvae comparable to that of AB88 or AB424 supernatants.

The nucleotide sequence of pCIB7104, a positive recombinant clone from AB88, and of pCIB7107, a positive recombinant clone from AB424, was determined by the dideoxy termination method of Sanger et al., Proc. Natl. Acad. Sci. U.S.A., 74: 5463–5467 (1977), using PRISM Ready Reaction Dye Deoxy Terminator Cycle Sequencing Kits and PRISM Sequenase® Terminator Double-Stranded DNA Sequencing Kit and analysed on an ABI 373 automatic sequencer.

The clone pCIB7104 contains the vip3A(a) gene whose coding region is disclosed in SEQ ID NO:1 and the encoded protein sequence is disclosed in SEQ ID NO:2. A synthetic version of the coding region designed to be highly expressed in maize is given in SEQ ID NO:7. Any number of synthetic genes can be designed based on the amino acid sequence given in SEQ ID NO:2.

The clone pCIB7107 contains the VIP3A(b) gene whose coding region is disclosed in SEQ ID NO:3 and the encoded protein is disclosed in SEQ ID NO:4. Both pCIB7104 and pCIB7107 have been deposited with the Agricultural Research Service Patent Culture Collection (NRRL) and given Accession Nos. NRRL B-21422 and B-21423, respectively.

Example 6

Identification of Novel vip3-like Genes by Hybridization

To identify *Bacillus* containing genes related to the vip3A(a) gene from isolate AB88, a collection of *Bacillus* isolates was screened by hybridization. Cultures of 463 *Bacillus* strains were grown in microtiter wells until sporulation. A 96-pin colony stampel was used to transfer the cultures to 150 mm plates containing L-agar. Inoculated plates were kept at 30° C. for 10 hours, then at 4° C. overnight. Colonies

were blotted onto nylon filters and probed with a 1.2 Kb HindIII VIP3A(a) derived fragment. Hybridization was performed overnight at 62° C. using hybridization conditions of Maniatis et al. Molecular Cloning: A Laboratory Manual (1982). Filters were washed with 2 $\times$ SSC/0.1% SDS at 62° C. and exposed to X-ray film.

Of the 463 *Bacillus* strains screened, 60 contain vip3-like genes that could be detected by hybridization.

Example 7

Presence of vip3-like genes and VIP3-like proteins in *Bacillus* isolates

Bacillus isolates other than AB88 have demonstrated insecticidal activity against Lepidopteran larvae when spent culture supernatants were tested. Some isolates which were active against black cutworm were analyzed for the presence of vip3-like genes and for the production of VIP3-like proteins.

A standard PCR analysis was used to determine whether the black cutworm-active *Bacillus* isolates contained a vip3-like gene. Using the PCR primer pair GW110 (5'-CGA TTA ATG TTG GCC TC-3'; SEQ ID NO:17) and GW 111 (5'-CAT TAG CAT CTC CGG ACA CAG-3'; SEQ ID NO:18) it was determined that all of the black cutworm active isolates produced a 728 bp vip3 gene product which was equal to the size produced by the type strain, AB88 (FIG. 3). One *Bacillus* isolate, AB51, which was not active against black cutworm, produced the same size vip3 product. None of the other non-black cutworm active *Bacillus* isolates produced a vip3 PCR product.

Analysis of VIP3 protein production was done using a standard western blot procedure. Antibodies raised against the VIP3A(a) protein described in the above example were used to detect immunoreactive proteins. Aliquots of cell free culture supernatants from sporulated cultures were run on SDS-PAGE gels using standard methods. Standard western blotting procedures were then carried out to determine the presence of VIP3-like proteins. All of the *Bacillus* isolates which had a 728 bp PCR product and were active against black cutworm produced an 80 kDa protein which was immunoreactive to the VIP3A(a) antibody (FIG. 3). The AB51 isolate which had the correct size vip3 PCR product but was not active against black cutworm produced an immunoreactive protein which was truncated suggesting this may be the reason no biological activity against black cutworm was observed.

Example 8

Characterization of *Bacillus thuringiensis* Strain AB51 Containing a vip3-like Gene

A. B. thuringiensis strain, designated AB51, was shown to contain proteins of the VIP3 class by western analysis using rabbit polyclonal anti-Vip3A(a) antibodies. The vip3-like gene was cloned into pKS which created pCIB7112. This gene was given the designation vip3A(c). The DNA sequence for vip3A(c) is disclosed in SEQ ID NO:5 and the encoded protein sequence is disclosed in SEQ ID NO:6. The VIP3A(c) protein is 746 amino acids long, 43 amino acids shorter than its VIP3A(a) and VIP3A(b) homologues.

Example 9

Characterization of *Bacillus thuringiensis* Strain Containing a Cryptic vip3-like Gene

A. B. thuringiensis strain, designated M2194, was shown to contain vip3-like gene(s) by colony hybridization as

described in Example 6. The M2194 vip3-like gene is considered cryptic since no expression can be detected throughout the bacterial growth phases either by immunoblot analysis using polyclonal antibodies raised against the VIP3A(a) protein isolated from AB88 or by bioassay as described in Example 1.

The M2194 vip3-like gene was cloned into pKS, which created pCIB7108. *E. coli* containing pCIB7108 which comprises the M2194 vip3 gene was active against black cutworm demonstrating that the gene encodes a functional protein with insecticidal activity. The plasmid pCIB7108 has been deposited with the Agricultural Research Service Patent Culture Collection (NRRL) and given Accession No. NRRL B-21438.

Example 10

Development of Antibodies to VIP3A(a) Protein

Antiserum against purified Vip3A(a) insecticidal protein was produced in rabbits and goats. For rabbits, nitrocellulose-bound protein (50 μ g) was dissolved in DMSO, emulsified with Freund's complete adjuvant (Difco) and injected subcutaneously twice a month for three months. For goats, active soluble pure Vip3A protein (300 μ g) was injected intramuscularly twice a month for three months. They were bled 10 days after the second and third injection and the serum was recovered from the blood sample (Harlow, E. and Lane, D. *Antibodies: A Manual Laboratory*, Cold Spring Harbor Lab. Press, N.Y., 1988). The antisera were then fractionated by affinity chromatography utilizing staphylococcal protein A, and the resulting IgG fraction was further purified by filtering through a column containing immobilized-*E. coli* lysate (Yu, C. G. et al. *Appl. Environ. Microbiol.* 63:532-536 (1997)).

The rabbit and goat antisera were characterized analyzing the Vip3A(a) protein by western blot. Proteins were separated by SDS/PAGE and transferred to nitrocellulose. Nitrocellulose blots were blocked in 20 mM Tris-HCl, pH 7.5/0.15M NaCl/0.02% Na₂S₂O₅/5% nonfat dry milk. Blots were developed by using either rabbit raised or goat-raised anti-Vip3A(a) antibodies at a concentration of 200 ng/ml or 100 ng/ml respectively. Alkaline phosphatase-conjugated goat antirabbit IgG or rabbit antigot antiserum were used as secondary antibodies at a concentration of 1 μ g/ml (Kirkegaard & Perry Laboratories, Inc.). Bromochloroindolyl-phosphate and nitroblue tetrazolium were used as substrate for the alkaline phosphatase reaction. Both anti-Vip3A(a) antibodies, the rabbit and the goat raised, are polyclonal. The anti-Vip3A(a) antibodies obtained from goat have a higher titer than the ones obtained from rabbits. In the experimental approach, anti-Vip3A(a) antibodies from rabbit should be used at a dilution 1/500 from the original serum (200 ng/ml). By comparison, the anti-Vip3A(a) antibodies obtained from goat can be diluted up to 1/2000 (100 ng/ml) from the original serum. While the rabbit raised antibodies only recognize the N-terminal portion of the Vip3A(a) protein, the antibodies obtained from goats react with epitopes present throughout the full length of the Vip3A(a) protein.

Example 11

Construction of Plant Expression Cassettes

Plant expression cassettes consist of promoters that can drive the expression of a coding sequence either constitutively or in a tissue-specific manner, the coding sequences to

be expressed and the termination sequences which allow the polyadenylation of the mRNA and its proper translation.

The promoters selected in the DNA constructs of the present invention includes constitutive promoters such as the one from the maize ubiquitin gene (Christensen et al. *Plant Mol. Biol.* 12:619-632,1989) (pCIB8029, FIG. 4; pCIB8055, FIG. 5; pCIB9806, FIG. 6), and tissue specific promoters such as those from the maize methalothionein-like gene (de Framond, A. *FEBS* 290:103-106,1991) (pCIB8030, FIG. 7; pCIB8056, pCIB9805) which provides a root-preferred expression, from the maize PEPC gene (Hudspeth, R. L. and Grula, J. W. *Plant Mol. Biol.* 12:579-589, 1989) (pCIB5535, FIG. 8; pCIB9807) which provides a green-tissue specific expression, and from the barley non-specific lipid transfer protein LTP4 (pCIB9819, FIG. 9)(Molina, A. and Garcia-Olmedo, F. *Plant J.* 4:983-991, 1993) which provides a stem-preferred expression. All constructs used in the present invention contain the terminator sequence derived from the 35S CaMV and the intron 9 derived from the maize PEPC gene for enhancing gene expression purposes. The plasmids pCIB8029, pCIB8055, and pCIB9806 contain the intron#1 of the maize ubiquitin gene placed between the maize ubiquitin promoter and the vip3A(a) gene. The construct comprising the encoding sequence of the vip3A(a) gene, the intron#9 and the 35S terminator sequence was engineered into the recipient plasmid bearing the different promoters as double digests BamHI-EcoRI.

The plant expression cassettes were used as such in the plant transformation experiments, or they were linearised by using restriction enzymes that cut in the Amp^R gene of the backbone plasmid. In some experiments, fragments comprising the promoter, gene of interest, intron and terminator were isolated from the rest of the plasmid backbone by restriction digestion and fragment purification. In these cases fragment purification proceeded as follows: 500 μ g of DNA is digested with the appropriate enzyme and separated on a 0.8% agarose gel. The fragment of interest is identified, cut out from the gel and purified using a Durapore Millipore filter (0.45 micron). The filtrate containing the fragment is precipitated with sodium acetate and ethanol. The fragment is resuspended in TE and used in transformation experiments.

Example 12

Insecticidal activity of maize plants expressing VIP3A(a)

Maize plants expressing VIP3A(a) protein were tested for insecticidal effects on the insect species listed in the table below by the following procedure. One to four 4 cm sections were cut from leaves of transgenic and control maize plants. Each leaf piece was placed on a moistened filter disc in a 50x9 mm petri dish. Five neonates of the species being tested were placed on each leaf piece giving a total of 5-20 larvae tested for each plant. The Petri dishes were incubated at 30° C. in the dark. Mortality was scored after 48-72 hours. Results are shown in Table 16.

TABLE 14

Insect species tested	Percent mortality	
	VIP3A(a)	Control
Maize Pests		
Black cutworm (<i>Agrotis ipsilon</i>)	100	0
Fall armyworm (<i>Spodoptera frugiperda</i>)	100	0
Sugarcane borer (<i>Diatraea saccharalis</i>)	100	0
Southwestern corn borer (<i>Diatraea grandiosella</i>)	100	0
Corn earworm (<i>Helicoverpa zea</i>)	100	10
Mediterranean corn borer (<i>Sesamia nonagroides</i>)	100	15
Other Lepidopteran Pests		
Beet armyworm (<i>S. exigua</i>)	100	0
Yellow striped armyworm (<i>S. ornithogalli</i>)	100	0
Cabbage looper (<i>Trichoplusia ni</i>)	100	20

Example 13

Expression of vip3A(a) in Maize Plants

Transformation of maize elite Ciba inbred lines CG00526 and 2154 with the Vip3 gene was achieved using particle bombardment of Type I callus tissue. For transformation using Type I embryogenic callus, the callus was obtained from zygotic embryos using standard culture techniques and subcultured 1–2 days prior to bombardment. Callus tissue was prepared for bombardment by placing ~20, 3–5 mm diameter pieces arranged in a ring shape onto culture medium containing 12% sucrose. Callus tissue was placed onto this media for four hours prior to bombardment. DNA used for transformation of maize callus was either circular plasmid DNA, linear plasmid DNA, or purified DNA fragments containing the Vip3 gene under control of various plant promoters. In experiments where a selectable agent was used, the gene allowed resistance to phosphinothricin or allowed for growth in the presence of mannose. Plasmids or DNA fragments isolated by filtration were precipitated onto 0.3 μ m gold particles according to published procedures from BioRad Laboratories, Hercules, Calif. Gold particles were delivered using a burst pressure of 650 psi of helium. Each target plate was shot twice with the DNA coated particles. Sixteen to twenty hours after bombardment the CG00526 callus was transferred to standard culture maintenance media. Seven days post-bombardment the tissue was transferred to media containing the selection agent, Basta at a concentration of 100 mg/L. Basta is a commercial formulation of glufosinate ammonium produced by Hoechst. Callus of 2154 was kept on 12% sucrose for 1–7 days after bombardment and transferred to standard culture media containing 20–30 mg/L Basta at day 7. The 2154 and CG00526 callus was subcultured in the presence of 30 or 100 mg/L Basta, respectively, for eight weeks. Tissue surviving selection was subcultured onto lower levels of Basta (5–40 mg/L) for a period of approximately five to ten weeks to allow for tissue bulk-up and then transferred to a standard regeneration media with no selection for the production of plants. Commonly, 12% of the callus pieces bombarded produced transformed callus that survived Basta selection. Individual transformed calli would typically be regenerated to produce 20–30 plants.

Events were generated from experiments where no selection was used. In these experiments the callus was grown for a period of 9–10 weeks on maintenance media prior to transferring to regeneration media. Event 1337 is an example of a transformed VIP3 event derived from a trans-

formation experiment with no selectable or scorable marker by screening plants for insecticidal activity.

Transformed calli were also generated from experiments where mannose selection was used. In these transformations the phosphomannose isomerase gene under control of the maize ubiquitin promoter of pCIB9818 was bombarded with the Vip3 gene. Mannose at 0.5–1.5% was included in the maintenance media for a period of twelve weeks and not included in the regeneration media.

Transgenic plants were evaluated for VIP3A(a) protein expression by insect bioassay and ELISA assay. Leaf pieces were removed from 2–4 leaf stage plants for evaluation using both black cutworm and fall army worm bioassays. Bioassays were done using ten newly hatched larvae placed in dishes with leaf pieces. Percent mortality was calculated at 72 hours. Tissues from transgenic plants were also assayed by ELISA using standard protocols to quantitate Vip3 protein levels in different plant tissues. Plant tissue was extracted and Table 17 provides representative events generated and their corresponding of insect bioassay results.

Transgenic maize plants were transformed with various plasmids containing the Vip3 gene under control of various promoters such as the maize PEP-carboxylase promoter (PEPC), the maize ubiquitin promoter (Ubi), and the maize metallothionein-like promoter (MTL). The selectable marker gene was the PAT gene under control of the maize ubiquitin promoter in pUBIAC. Representative events listed in Table 17 show the events produced with different plasmids or DNA fragments derived from plasmids. DNA fragments were generated using restriction enzyme digestions and size fractionated using electrophoresis in 0.8% agarose gels. The DNA fragments were excised from the gels, frozen, crushed and purified by filtration through 0.45 micron DuraPore Millipore filters followed by ethanol precipitation. Transformed maize events were generated with circular plasmid DNA of pCIB5535 containing the Vip3 gene under control of the maize PEPC promoter. Events were also transformed with linear plasmid DNA of pCIB5535 and pCIB8029 containing the Vip3 gene under control of the maize ubiquitin promoter. Additional events were produced by bombarding purified DNA restriction enzyme fragments containing just the Vip3 gene with promoter. Fragments corresponding to the Vip3 gene include: a 4906 bp EcoRI/HindIII fragment from pCIB5535 with the maize PEPC promoter; a 5142 bp KpnI/HindIII fragment from pCIB8030 with the MTL promoter; a 4597 bp KpnI/HindIII fragment of pCIB8029 with the maize ubiquitin promoter; a 4818 bp HindIII fragment of pCIB8055 with the maize ubiquitin promoter; a 5364 HindIII fragment of pCIB8056 with the MTL promoter; a 5964 AscI fragment of pCIB9805 with the MTL promoter; a 5418 bp AscI fragment of pCIB9806 with the maize ubiquitin promoter; and a 5727 bp AscI fragment of pCIB9807 with the maize PEPC promoter.

TABLE 15

Event No.	Plasmid Used	Chimeric Gene	Mortality (%)	
			Fall Armyworm	Black Cutworm
891	pCIB5535	PEPC:vip3A(a)	100	100
906	pCIB5535 and pCIB8029	PEPC:vip3A(a) and Ubi:vip3A(a)	100	100

TABLE 15-continued

Event No.	Plasmid Used	Chimeric Gene	Mortality (%)	
			Fall Armyworm	Black Cutworm
946	pCIB5535 and pCIB8030	PEPC:vip3A(a) and MTL:vip3A(a)	100	100

Example 14

Insecticidal activity of maize plants containing Vip3 and Bt δ -endotoxins

VIP3A(a) has little activity against European corn borer (ECB). To make plants with broad spectrum lepidopteran control, maize plants containing a vip3A(a) gene were crossed with maize plants containing a cry1B, which is active against ECB. Progeny from the crosses were bioassayed against ECB and fall armyworm (FAW) as described in Example 1. Results are shown in Table 18. Approximately 34% of the progeny were not active against either species, 15.4% were active only on ECB, 23.1% were active only on FAW and 27.9% were active against both species. Plants active against both species contained both VIP3A(a) and Cry1B protein. Similar results are obtained using other Bt δ -endotoxins, particularly Cry1Ab or Cry9C.

TABLE 16

Cross	% ECB active	% FAW active	% ECB & FAW active	% not active
VIP3A(a) X Cry1B	15.4	23.1	27.9	34.6

Example 15

VIP3A(a) Lyses the Midgut Epithelial Cells of Susceptible Insects

Feeding and gut clearance studies. The temporal sequence of symptoms following the ingestion of VIP3A(a)-containing diet by second-instar black cutworm (BCW) larvae, a susceptible insect, were recorded from the time of initial administration until larval death. Larvae exposed to control diet showed active feeding followed by uninterrupted gut parastalsis. In contrast, the addition of VIP3A(a) protein in the diet had a significant effect on feeding behavior. When added at concentrations as low as 4 ng per cm², the larvae fed on and off during periods of 10–20 min. The presence of blue color in their guts indicated feeding but the clearance of the gut content was dramatically affected as judged by the deceased number of frass. With 4 ng of VIP3A(a) per cm² added to the diet, larval development was significantly impaired after a 48 h incubation period but no mortality was observed. At concentrations of 40 ng of Vip3A(a) per cm², the larvae suffered gut paralysis upon ingestion of minute amounts of diet and no frass could be seen indicating an almost complete lack of gut clearance. Under this condition, ca. 50% mortality was recorded after 48 hr. When concentrations higher than 40 ng of VIP3A(a) per cm were used, the larvae were moribund after only a few bites, with no frass and mortality rates approaching 100%. When similar experiments were conducted with fall armyworm, also a susceptible insect, similar behavioral patterns were observed. In contrast, European corn borer did

not alter its feeding behavior when VIP3A(a) protein was added to the diet even at concentrations as high as 400 ng of VIP3A(a) per cm².

Histological observations of the effects of the Vip3A(a) protein. Histopathological observations on the effects of the VIP3A(a) protein on BCW were conducted on second and third instar larvae which had been fed a diet containing VIP3A(a). Analysis of BCW gut cross-sections showed extensive damage to the midgut epithelium indicating that the midgut tissue is a primary site of action of the Vip3A(a) protein. No damage was discernible in the foregut and hindgut. Midgut epithelial cells from untreated larvae were closely associated with one another showing no evidence of damage. Sections from larvae that had been fed for 24 h with diet containing Vip3A(a) showed that distal ends of the epithelium columnar cells had become distended and bulbous. Although the goblet cells exhibited some morphological alterations, they did not show signs of damage at this stage. Degeneration of the epithelium columnar cells continued such that, after 48 h of ingesting Vip3A(a)-containing diet, the lumen was filled with debris of disrupted cells. The goblet cells also exhibited signs of damage after 48 h, but both types of cells were still attached to the basement membrane. Black cutworm larvae were dead at 72 h and desquamation of the epithelial layer was complete. While a similar histopathology was observed for fall armyworm, European corn borer did not exhibit any tissue damage under similar experimental conditions.

In vivo immunolocalization of the Vip3A(a) protein. Third instar larvae of black cutworm and European corn borer fed on artificial diet supplemented with 100–200 ng of VIP3A(a) per cm² were used for immunocytochemical characterization of the VIP3A(a) binding to midgut sections. The bound VIP3A(a) was visualized using rabbit anti-VIP3A(a) antibodies previously purified through protein A sepharose and *E. coli* immobilized columns (Yu, C. G. et al. Appl. Environ. Microbiol. 63:532–536, 1997). VIP3A(a) binding was detected in midgut epithelium of black cutworm, while showing no binding to European corn borer midguts. Midgut sections from black cutworm larvae fed with control diet showed no VIP3A(a) binding. The VIP3A(a) binding seems to be specifically associated to the apical microvilli and it is mostly associated to the columnar cells, with no detectable signal in the goblet cells.

Example 16

VIP3A(a) and VIP3A(b) Induce Apoptosis in Insect Cells

VIP3A(a) and VIP3A(b) were shown to be a apoptosis inducing protein arose by the characterization of its insecticidal effects towards an insect cell line (Sf-9) derived from *Spodoptera frugiperda*, an insect susceptible to VIP3A(a). VIP3A(a) showed insecticidal activity towards the insect cell line when kept present throughout the experiment. When Sf-9 insect cells are transiently exposed to VIP3A(a) and VIP3A(b), their cell viability was significantly reduced even with exposure times as short as 5 min. Once the incubation time exceeded 10 min, the effects of the VIP3A(a) and VIP3A(b) on insect cell viability over a period of 6 hours were maximal showing a reduction of 90% in cell viability. The cytological changes occurring in Sf-9 cells transiently exposed to VIP3A(a) were monitored by microscopy. Small protrusions appeared on the surface of the treated cells some time between 10 and 15 min after their exposure to the VIP3A(a) protein. At this stage, the mitochondria of the cells remained functionally intact as revealed

by staining with rhodamine 123, a dye that accumulates in mitochondria with active membrane potential (Johnson, L. V. et al Proc. Natl. Acad. Sci. U.S.A. 77:990-994, 1980). These protrusions eventually disappeared and the cells entered a phase of profuse vacuolization lasting an additional 30 to 60 min. During the final stages, the insect cells are seen to swell before disintegration. For an individual cell, the entire process required 1 to 2 hours. All these cellular events are consistent with previous studies on cells undergoing apoptosis particularly considering that programmed cell death during metamorphosis of certain insects is accompanied by cellular vacuolization and swelling (Schwartz, L. M. et al Proc. Natl. Acad. Sci. U.S.A. 90:980-984 (1993)).

Recent studies have shown that the distribution of phospholipids in the plasma membrane is affected in very early stages of animal cells undergoing apoptosis (Martin, S. J., et al. J. Exp. Med.: 182, 1545-1556, 1995) particularly the externalization of the phosphatidylserine (PS). This process can be visualized by using Annexin V, an anticoagulant protein with high affinity for phosphatidylserine (PS). When VIP3A(a)-treated SF-9 cells were incubated with Annexin V, an externalization of PS was revealed in insect cell membranes as early as 5-10 min after the exposure to VIP3A(a) probably marking the onset of apoptosis.

One of the key molecular events that is the hallmark of apoptosis is endonucleolysis resulting in a double strand DNA break freeing oligonucleosome-sized fragments of 200 base pair and multiples. We examined the occurrence of endonucleolysis in SF-9 cells treated with VIP3A(a) using an in situ detection method and analysing the DNA by agarose gel electrophoresis. Based on the ability of the Klenow enzyme to incorporate modified nucleotides using the DNA ends generated by DNA fragmentation, SF-9 insect cells showed signs of endonucleolysis as early as 30 min upon their exposure to the VIP3A(a) protein. This stage will coincide with the appearance of membrane-bound subcellular apoptotic bodies visualized in the microscopical observations. These early indications of endonucleolytic activity were confirmed by the detection of DNA fragments in agarose gels characteristic of a chromatin ladder slightly later in the process. These results corroborated the indications obtained from cytological observations, that the SF-9 cells initiate an apoptotic-type of programmed cell death upon their exposure to the VIP3A(a) protein.

The VIP3A(a) and VIP3A(b) proteins were discovered on the basis of their insecticidal properties against some lepidopteran insects. Therefore, we were interested in knowing whether the VIP3A(a) protein would induce an apoptotic pathway in gut cells of susceptible insects upon its ingestion and thus, it could exert its insecticidal properties by triggering an active process of cell death. Histological and histochemical studies have shown that the VIP3A(a) protein specifically targets the columnar cells of the midgut epithelium of susceptible insects provoking cell changes characterized by membrane protrusions and extensive vacuolization leading to cell death. These cytological changes induced by VIP3A(a) in insect gut cells resemble those described above for SF-9 cells. We then examined whether midgut epithelium cells of susceptible insects undergo endonucleolysis upon ingesting diet containing VIP3A(a) by in situ detection (Cuvillier, O., et al Nature 381:800-803 (1996)) of DNA fragmentation. When sections of midgut tissue from black cutworm larvae fed with diet either containing VIP3A(a) or control diet, nuclei staining indicative of DNA fragmentation was only detectable in the columnar cells of the midgut epithelium exposed to the VIP3A(a) protein. This

result indicates that the VIP3A(a) protein induces an endonucleolysis process in the midgut epithelium cells concurrently with the cytological changes reported previously. It is our conclusion that the VIP3A(a) protein likely exerts its insecticidal properties by activating an apoptosis-type of programmed cell death of the midgut epithelium cells of susceptible insects.

Example 17

Isolation of the Receptor for VIP3A(a) from black cutworm

Black cutworm is sensitive to VIP3A(a) and therefore this insect was used for the isolation of the VIP3A(a) receptor. Midgut of third instar black cutworm larvae were collected by dissection and immediately frozen in liquid nitrogen. One gram of midgut tissue was used to isolate mRNA by following the protocol described in the two-hybrid cDNA library construction kit provided by Clontech (1997). Ten micrograms of poly A⁺ RNA were used as starting material. In first strand synthesis, both random and lock-docking oligo(dT)₂₅d(A/C/G) primers are used in separate synthesis with MML reverse transcriptase. The second strand cDNA was achieved by an optimal ratio of DNA polymerase to Rnase H activity in the second-strand enzyme cocktail. The newly synthesized double stranded cDNA is then ligated to EcoRI-NotI-SalI adaptors. The cDNAs were ligated into pGAD10 (Vijaychander, S. et al. CLONTECHniques IX-3:8-10, 1994) which provides the activation domain. The vip3A(a) gene was engineered into the polylinker site of the plasmid pGBT9 in frame with the GAL4-DNA binding domain (Bartel, P. L. et al. Cellular Interactions in Development: A Practical Approach, pp. 153-179, Oxford University Press, 1993). The recombinant pGBT9-vip3A(a) was transformed into the yeast strain GGY1::171 (Gill, G. and Ptashne, M. Cell51:121-126, 1987) by electroporation (Estruch, J. J. et al BioTechniques 16:610-612, 1994). The transformed yeast was selected in minimal media without tryptophan (Bartel, P. L. et al. Cellular Interactions in Development: A Practical Approach, pp. 153-179, Oxford University Press, 1993). The expression of the VIP3A(a) protein in the recombinant yeast was confirmed by western analysis. The yeast strain GGY1::171- VIP3A(a) was transformed with the black cutworm cDNA library represented in pGAD10. GGY1::171 possess the HIS3 marker under the control of GAL4 recognition sites. The HIS3 gene allows a positive growth selection for clones that are transformed by two interacting hybrid constructs. After plating more than 200,000 recombinant clones, only one was able to grow in minimal media without histidine. The plasmid DNA of the positive yeast colony was isolated by the yeast lysis buffer method (Kaiser, P. and Auer, B. BioTechniques 14:552 (1993)) and electroporated into *E. coli*. The insert containing the cDNA was subcloned into the EcoRI site of the pBluescript (Stratagene) and sequenced by the dideoxy termination method of Sanger et al., Proc. Natl. Acad. Sci. U.S.A., 74: 5463-5467 (1977), using PRISM Ready Reaction Dye Deoxy Terminator Cycle Sequencing Kits and PRISM Sequenase® Terminator Double-Stranded DNA Sequencing Kit and analysed on an ABI 373 automatic sequencer.

Example 18

Insect Cells Transformed with the Gene for the Receptor Exhibit Apoptosis when Exposed to the VIP3A(a) protein

The receptor in black cutworm midgut cells for the VIP3A(a) protein was cloned into the XhoI-BamHI site of

the Smart 2 cosmid vector (Speck, M. et al Gene 64: 173-177 (1988)), and the recombinant construct was used to transform the Schneider 2 (S2) *Drosophila* cell line using the calcium phosphate co-precipitation method (Clem, R. J. and Miller, L. K. Mol. Cel. Biol. 14: 5212-5222 (1994)). Smart 2 carries the selectable marker tet (tetracycline) for bacterial transformation and the neo (neomycin) for *Drosophila* cell transformation. The neo selectable marker is expressed under the control of the *Drosophila* hsp70 promoter. The transformed S2 cells were selected in S2 *Drosophila* medium supplemented with 10% of Fetal Seroalbumin and with G418 (1 mg/ml) at 30° C. (see GIBCO catalogue 1997). Several stably transformed S2 cell lines were established after 45 days of selection in the medium described above.

The sensitivity of the S2 transformed cells to the VIP3A(a) was tested by adding VIP3A(a) protein (at a final concentration of 1,7 micrograms per ml) to the media containing the transformed S2 cells that have been previously heat shocked at 42° C. 30 min. The induction of apoptosis in transformed S2 cells was confirmed by both microscopical observations and by the TACS Kit, and in situ Apoptosis Detection kit (for detailed description, see Trevigen catalogue 1996).

Example 19

Isolating Homologues to the Receptor from Other Insects

The cells of the midgut epithelium of black cutworm larvae possess a receptor that is recognized by the VIP3A(a) protein. Receptors from other insects known to be susceptible to VIP3A(a) are isolated by identifying the DNA sequences in Southern analysis. DNA is prepared, enzyme restricted, run in agarose gels and blotted onto nitrocellulose and/or nylon filters. These filters are probed with the cDNA encoding the receptor from black cutworm using low stringency conditions of hybridization and washing. Genes with a similarity to the black cutworm receptor to VIP3A(a) lower than 50% were identified. The Southern analysis can also be probed against partial sequences of the cDNA which encode specific domains such as death domain or EGF-like motifs with the intention of isolating genes that contain similar domains even though they are functionally different to the black cutworm receptor to VIP3A(a).

The isolation of homologues to the black cutworm receptor to VIP3A(a) is accomplished by the two hybrid system described in Fields, S. and Song, O.-K. Nature 340:245-246 (1989). Isolated mRNA is obtained from an organisms of interest, synthesize cDNAs and clone them into pGAD10 or equivalent plasmids. The cDNA library is co-transformed with the pGB9-bearing the vip3A(a) gene (or homologues of this gene) and rescued putative receptors in yeast by means of activating a marker based upon protein-protein interaction between the VIP3A(a) protein (or homologues) and the putative receptor.

Homologues to the black cutworm receptor to VIP3A(a) are isolated by expressing cDNAs libraries isolated from organisms of interests, cloned into appropriate expression vectors and transformed into host cells such as yeast or

insect cells which are known not to have the ability to bind and/or be sensitive to VIP3A(a). The transformed cells are screened based on their gained property of binding VIP3A(a) or undergoing apoptotic responses when incubated with VIP3A(a). In this case, the protein VIP3A(a) is used as probe and its binding will be monitored either by antibodies against VIP3A(a) or by labels such as biotin attached to VIP3A(a).

Example 20

Screening for Novel Compounds that Induce Apoptosis in Insect Cells

Model cell lines for different orders of insects (some examples include Sf-9 cells for lepidopteran, Colorado potato beetle for coleopterans, S2 from *Drosophila* for dipterans) is used to screen for novel compounds whose mode of action is induction of apoptosis. The cells are grown in multi-well plates which are used for a high-throughput assay screening for thousands of compounds (both of large and small molecular weight). The compound(s) are added as single component or as mixtures. Compound(s) inducing apoptosis are identified as follows: 1) membrane protrusions are visible in the cell membrane, 2) a reorganization of the phosphatidylserine containing membrane lipids is detectable by using specific proteins with high affinity for phosphatidylserine such as Annexin-V linked to a visual marker, 3) cytoplasmic blebbing is visible in the cell cytoplasm, 4) active mitochondria can be visualized by using vital dyes such as rhodamine 123 that accumulate in mitochondria, 5) DNA fragmentation is detected either by DNA analysis in agarose gels, by ELISA detection of nucleosomal release or by in vivo detection of DNA nicking. All these cytological and molecular features are indicative of apoptosis.

The black cutworm receptor to VIP3A(a) is transformed into S2 cell line. Therefore, isogenic S2 lines are available with and without the said receptor. These cell lines are used to screen compounds that provide a differential response due to the presence of the said receptor. Transformed S2 cells undergoing apoptosis upon exposure to certain compounds are identified as indicated above. The differential response of the transformed versus the non-transformed cell is indicative that the action of the compound is mediated by the cloned receptor. Similar approaches are undertaken with insect cells transformed with receptors homologue to the black cutworm receptor to VIP3A(a).

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i i i) NUMBER OF SEQUENCES: 20

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2378 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA (genomic)

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 9..2375
- (D) OTHER INFORMATION: /note= "Native DNA sequence
encoding VIP3A(a) protein from AB88 as contained in
pCIB7104"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 789 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Asn 6 2 5	Leu	Glu	Asp	Tyr	Gln 6 3 0	Thr	Ile	Asn	Lys	Arg 6 3 5	Phe	Thr	Thr	Gly	Thr 6 4 0		
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(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2612 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

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(i i) MOLECULE TYPE: DNA (genomic)

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 118..2484

(D) OTHER INFORMATION: /note= "Native DNA sequence
encoding VIP3A(b) from AB424"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATTGAAATTG	ATAAAAAGTT	ATGAGTGT	TTT	AATAATCAGT	AATTACCAAT	AAAGAATTAA	60
GAATACAAGT	TTACAAGAAA	TAAGTGT	TAC	AAAAAATAGC	TGAAAAGGAA	GATGAAC	117
ATG AAC AAG AAT AAT ACT AAA TTA AGC ACA AGA GCC TTA CCA AGT TTT	165						
Met Asn Lys Asn Asn Thr Lys Leu Ser Thr Arg Ala Leu Pro Ser Phe	790 795 800 805						
ATT GAT TAT TTC AAT GGC ATT TAT GGA TTT GCC ACT GGT ATC AAA GAC	213						
Ile Asp Tyr Phe Asn Gln Gly Ile Tyr Gly Phe Ala Thr Gly Ile Lys Asp	810 815 820						
ATT ATG AAC ATG ATT TTT AAA ACG GAT ACA GGT GGT GAT CTA ACC CTA	261						
Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly Gly Asp Leu Thr Leu	825 830 835						
GAC GAA ATT TTA AAG AAT CAG CAG CTA CTA AAT GAT ATT TCT GGT AAA	309						
Asp Glu Ile Leu Lys Asn Gln Gln Leu Leu Asn Asp Ile Ser Gly Lys	840 845 850						
TTG GAT GGG GTG AAT GGA AGC TTA AAT GAT CTT ATC GCA CAG GGA AAC	357						
Leu Asp Gly Val Asn Gly Ser Leu Asn Asp Leu Ile Ala Gln Gly Asn	855 860 865						
TTA AAT ACA GAA TTA TCT AAG GAA ATA TTA AAA ATT GCA AAT GAA CAA	405						
Leu Asn Thr Glu Leu Ser Lys Glu Ile Leu Lys Ile Ala Asn Glu Gln	870 875 880 885						
AAT CAA GTT TTA AAT GAT GTT AAT AAC AAA CTC GAT GCG ATA AAT ACG	453						
Asn Gln Val Leu Asn Asp Val Asn Asn Lys Leu Asp Ala Ile Asn Thr	890 895 900						
ATG CTT CGG GTA TAT CTA CCT AAA ATT ACC TCT ATG TTG AGT GAT GTA	501						
Met Leu Arg Val Tyr Leu Pro Lys Ile Thr Ser Met Leu Ser Asp Val	905 910 915						
ATG AAA CAA AAT TAT GCG CTA AGT CTG CAA ATA GAA TAC TTA AGT AAA	549						
Met Lys Glu Asn Tyr Ala Leu Ser Leu Gln Ile Glu Tyr Leu Ser Lys	920 925 930						
CAA TTG CAA GAG ATT TCT GAT AAG TTG GAT ATT ATT AAT GTA AAT GTA	597						
Gln Leu Gln Glu Ile Ser Asp Lys Leu Asp Ile Ile Asn Val Asn Val	935 940 945						
CTT ATT AAC TCT ACA CTT ACT GAA ATT ACA CCT GCG TAT CAA AGG ATT	645						
Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro Ala Tyr Gln Arg Ile	950 955 960 965						
AAA TAT GTG AAC GAA AAA TTT GAG GAA TTA ACT TTT GCT ACA GAA ACT	693						
Lys Tyr Val Asn Glu Lys Phe Glu Glu Leu Thr Phe Ala Thr Glu Thr	970 975 980						
AGT TCA AAA GTA AAA AAG GAT GGC TCT CCT GCA GAT ATT CGT GAT GAG	741						
Ser Ser Lys Val Lys Lys Asp Gly Ser Pro Ala Asp Ile Arg Asp Glu	985 990 995						
TTA ACT GAG TTA ACT GAA CTA GCG AAA AGT GTA ACA AAA AAT GAT GTG	789						
Leu Thr Glu Leu Thr Glu Leu Ala Lys Ser Val Thr Lys Asn Asp Val	1000 1005 1010						
GAT GGT TTT GAA TTT TAC CTT AAT ACA TTC CAC GAT GTA ATG GTA GGA	837						
Asp Gly Phe Glu Phe Tyr Leu Asn Thr Phe His Asp Val Met Val Gly	1015 1020 1025						
AAT AAT TTA TTC GGG CGT TCA GCT TTA AAA ACT GCA TCG GAA TTA ATT	885						
Asn Asn Leu Phe Gly Arg Ser Ala Leu Lys Thr Ala Ser Glu Leu Ile	1030 1035 1040 1045						

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ACT Thr	AAA Lys	GAA Glu	AAT Asn	GTG Val	AAA Lys	ACA Thr	AGT Ser	GGC Gly	AGT Ser	GAG Glu	GTC Val	GGA Gly	AAT Asn	GTT Val	TAT Tyr	933
				1050					1055						1060	
AAC Asn	TTC Phe	CTA Leu	ATT Ile	GTA Val	TTA Leu	ACA Thr	GCT Ala	CTG Leu	CAA Gln	GCA Ala	AAA Lys	GCT Ala	TTT Phe	CTT Leu	ACT Thr	981
			1065					1070					1075			
TTA Leu	ACA Thr	CCA Pro	TGC Cys	CGA Arg	AAA Lys	TTA Leu	TTA Leu	GGC Gly	TTA Leu	GCA Ala	GAT Asp	ATT Ile	GAT Asp	TAT Tyr	ACT Thr	1029
		1080					1085					1090				
TCT Ser	ATT Ile	ATG Met	AAT Asn	GAA Glu	CAT His	TTA Leu	AAT Asn	AAG Lys	GAA Glu	AAA Lys	GAG Glu	GAA Glu	TTT Phe	AGA Arg	GTA Val	1077
	1095					1100					1105					
AAC Asn	ATC Ile	CTC Leu	CCT Pro	ACA Thr	CTT Leu	TCT Ser	AAT Asn	ACT Thr	TTT Phe	TCT Ser	AAT Asn	CCT Pro	AAT Asn	TAT Tyr	GCA Ala	1125
	1110				1115					1120					1125	
AAA Lys	GTT Val	AAA Lys	GGA Gly	AGT Asp	GAT Glu	GAA Asp	GAT Asp	GCA Ala	AAG Lys	ATG Met	ATT Ile	GTG Val	GAA Glu	GCT Ala	AAA Lys	1173
				1130					1135					1140		
CCA Pro	GGA Gly	CAT His	GCA Ala	TTG Leu	ATT Ile	GGG Gly	TTT Phe	GAA Glu	ATT Ile	AGT Ser	AAT Asn	GAT Asp	TCA Ser	ATT Ile	ACA Thr	1221
			1145					1150					1155			
GTA Val	TTA Leu	AAA Lys	GTA Val	TAT Tyr	GAG Glu	GCT Ala	AAG Lys	CTA Leu	AAA Lys	CAA Gln	AAT Asn	TAT Tyr	CAA Gln	GTC Val	GAT Asp	1269
		1160					1165					1170				
AAG Lys	GAT Asp	TCC Ser	TTA Leu	TCG Ser	GAA Glu	GTT Val	ATT Ile	TAT Tyr	GGC Gly	GAT Asp	ATG Met	GAT Asp	AAA Lys	TTA Leu	TTG Leu	1317
	1175					1180					1185					
TGC Cys	CCA Pro	GAT Asp	CAA Gln	TCT Ser	GGA Gly	CAA Gln	ATC Ile	TAT Tyr	TAT Tyr	ACA Thr	AAT Asn	AAC Asn	ATA Ile	GTA Val	TTT Phe	1365
	1190				1195					1200					1205	
CCA Pro	AAT Asn	GAA Glu	TAT Tyr	GTA Val	ATT Ile	ACT Thr	AAA Lys	ATT Ile	GAT Asp	TTC Phe	ACT Thr	AAA Lys	AAA Lys	ATG Met	AAA Lys	1413
				1210					1215					1220		
ACT Thr	TTA Leu	AGA Arg	TAT Tyr	GAG Glu	GTA Val	ACA Thr	GCG Ala	AAT Asn	TTT Phe	TAT Tyr	GAT Asp	TCT Ser	TCT Ser	ACA Thr	GGA Gly	1461
			1225					1230					1235			
GAA Glu	ATT Ile	GAC Asp	TTA Leu	AAT Asn	AAG Lys	AAA Lys	AAA Lys	GTA Val	GAA Glu	TCA Ser	AGT Ser	GAA Glu	GCG Ala	GAG Glu	TAT Tyr	1509
		1240				1245						1250				
AGA Arg	ACG Thr	TTA Leu	AGT Ser	GCT Ala	AAT Asn	GAT Asp	GAT Asp	GGG Gly	GTG Val	TAT Tyr	ATG Met	CCG Pro	TTA Leu	GGT Gly	GTC Val	1557
	1255					1260					1265					
ATC Ile	AGT Ser	GAA Glu	ACA Thr	TTT Phe	TTG Leu	ACT Thr	CCG Pro	ATT Ile	AAT Asn	GGG Gly	TTT Phe	GGC Gly	CTC Leu	CAA Gln	GCT Ala	1605
	1270				1275					1280					1285	
GAT Asp	GAA Glu	AAT Asn	TCA Ser	AGA Arg	TTA Leu	ATT Ile	ACT Thr	TTA Leu	ACA Thr	TGT Cys	AAA Lys	TCA Ser	TAT Tyr	TTA Leu	AGA Arg	1653
				1290					1295					1300		
GAA Glu	CTA Leu	CTG Leu	CTA Leu	GCA Ala	ACA Thr	GAC Asp	TTA Leu	AGC Ser	AAT Asn	AAA Lys	GAA Glu	ACT Thr	AAA Lys	TTG Leu	ATC Ile	1701
			1305					1310					1315			
GTC Val	CCG Pro	CCA Ser	AGT Gly	GGT Phe	TTT Ile	ATT Ile	AGC Asn	AAT Ile	GTA Val	GAG Glu	AAC Asn	GGG Gly	TCC Ser	ATA Ile		1749
		1320					1325				1330					
GAA Glu	GAG Glu	GAC Asp	AAT Asn	TTA Leu	GAG Glu	CCG Pro	TGG Trp	AAA Lys	GCA Ala	AAT Asn	AAT Asn	AAG Lys	AAT Asn	GCG Ala	TAT Tyr	1797
	1335					1340					1345					
GTA Val	GAT Asp	CAT His	ACA Thr	GGC Gly	GGA Val	GTG Val	AAT Asn	GGA Gly	ACT Thr	AAA Lys	GCT Ala	TTA Leu	TAT Tyr	GTT Val	CAT His	1845
	1350				1355					1360					1365	

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AAG Lys	GAC Asp	GGA Gly	GGA Gly	ATT Ile	TCA Ser	CAA Gln	TTT Phe	ATT Ile	GGA Gly	GAT Asp	AAG Lys	TTA Leu	AAA Lys	CCG Pro	AAA Lys	1893
				1370					1375					1380		
ACT Thr	GAG Glu	TAT Tyr	GTA Val	ATC Ile	CAA Gln	TAT Tyr	ACT Thr	GTT Val	AAA Lys	GGA Gly	AAA Lys	CCT Pro	TCT Ser	ATT Ile	CAT His	1941
			1385					1390					1395			
TTA Leu	AAA Lys	GAT Asp	GAA Glu	AAT Asn	ACT Thr	GGA Gly	TAT Tyr	ATT Ile	CAT His	TAT Tyr	GAA Glu	GAT Asp	ACA Thr	AAT Asn	AAT Asn	1989
		1400					1405					1410				
AAT Asn	TTA Leu	GAA Glu	GAT Asp	TAT Tyr	CAA Gln	ACT Thr	ATT Ile	AAT Asn	AAA Lys	CGT Arg	TTT Phe	ACT Thr	ACA Thr	GGA Gly	ACT Thr	2037
		1415				1420					1425					
GAT Asp	TTA Leu	AAG Lys	GGA Gly	GTG Val	TAT Tyr	TTA Leu	ATT Ile	TTA Leu	AAA Lys	AGT Ser	CAA Gln	AAT Asn	GGA Gly	GAT Asp	GAA Glu	2085
		1430			1435					1440					1445	
GCT Ala	TGG Trp	GGA Gly	GAT Asp	AAC Phe	TTT Phe	ATT Ile	ATT Ile	TTG Leu	GAA Glu	ATT Ile	AGT Ser	CCT Pro	TCT Ser	GAA Glu	AAG Lys	2133
				1450					1455					1460		
TTA Leu	TTA Leu	AGT Ser	CCA Pro	GAA Glu	TTA Leu	ATT Ile	AAT Asn	ACA Thr	AAT Asn	AAT Asn	TGG Trp	ACG Thr	AGT Ser	ACG Thr	GGA Gly	2181
			1465					1470					1475			
TCA Ser	ACT Thr	AAT Asn	ATT Ile	AGC Ser	GGT Gly	AAT Asn	ACA Thr	CTC Leu	ACT Thr	CTT Leu	TAT Tyr	CAG Gln	GGA Gly	GGA Gly	CGA Arg	2229
		1480					1485					1490				
GGG Gly	ATT Ile	CTA Leu	AAA Lys	CAA Gln	AAC Asn	CTT Leu	CAA Gln	TTA Leu	GAT Asp	AGT Ser	TTT Phe	TCA Ser	ACT Thr	TAT Tyr	AGA Arg	2277
		1495				1500					1505					
GTG Val	TAT Tyr	TTC Phe	TCT Ser	GTG Val	TCC Ser	GGA Gly	GAT Asp	GCT Ala	AAT Asn	GTA Val	AGG Arg	ATT Ile	AGA Arg	AAT Asn	TCT Ser	2325
		1510			1515				1520						1525	
AGG Arg	GAA Glu	GTG Val	TTA Leu	TTT Phe	GAA Glu	AAA Lys	AGA Arg	TAT Tyr	ATG Met	AGC Ser	GGT Gly	GCT Ala	AAA Lys	GAT Asp	GTT Val	2373
				1530					1535					1540		
TCT Ser	GAA Glu	ATG Met	TTC Phe	ACT Thr	ACA Thr	AAA Lys	TTT Phe	GAG Glu	AAA Lys	GAT Asp	AAC Asn	TTC Phe	TAT Tyr	ATA Ile	GAG Glu	2421
			1545					1550					1555			
CTT Leu	TCT Ser	CAA Gln	GGG Gly	AAT Asn	AAT Asn	TTA Leu	TAT Tyr	GGT Gly	GGT Gly	CCT Pro	ATT Ile	GTA Val	CAT His	TTT Phe	TAC Tyr	2469
		1560					1565				1570					
GAT Asp	GTC Val	TCT Ser	ATT Ile	AAG Lys	TAAGATCGGG	ATCTAATATT	AACAGTTTTT	AGAAGCTAAT								2524
		1575														
TCTTGATAA	TGTCCTTGAT	TATGGAAAAA	CACAATTTTG	TTTGCTAAGA	TGTATATATA											2584
GCTCACTCAT	TAAAAGGCAA	TCAAGCTT														2612

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 789 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Asn Lys Asn Asn Thr Lys Leu Ser Thr Arg Ala Leu Pro Ser Phe
1 5 10 15

Ile Asp Tyr Phe Asn Gly Ile Tyr Gly Phe Ala Thr Gly Ile Lys Asp
20 25 30

Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly Gly Asp Leu Thr Leu
35 40 45

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Asp	Glu	Ile	Leu	Lys	Asn	Gln	Gln	Leu	Leu	Asn	Asp	Ile	Ser	Gly	Lys
	50					55					60				
Leu	Asp	Gly	Val	Asn	Gly	Ser	Leu	Asn	Asp	Leu	Ile	Ala	Gln	Gly	Asn
65					70				75						80
Leu	Asn	Thr	Glu	Leu	Ser	Lys	Glu	Ile	Leu	Lys	Ile	Ala	Asn	Glu	Gln
				85					90					95	
Asn	Gln	Val	Leu	Asn	Asp	Val	Asn	Asn	Lys	Leu	Asp	Ala	Ile	Asn	Thr
			100					105					110		
Met	Leu	Arg	Val	Tyr	Leu	Pro	Lys	Ile	Thr	Ser	Met	Leu	Ser	Asp	Val
		115					120					125			
Met	Lys	Gln	Asn	Tyr	Ala	Leu	Ser	Leu	Gln	Ile	Glu	Tyr	Leu	Ser	Lys
	130					135					140				
Gln	Leu	Gln	Glu	Ile	Ser	Asp	Lys	Leu	Asp	Ile	Ile	Asn	Val	Asn	Val
145					150					155					160
Leu	Ile	Asn	Ser	Thr	Leu	Thr	Glu	Ile	Thr	Pro	Ala	Tyr	Gln	Arg	Ile
				165					170					175	
Lys	Tyr	Val	Asn	Glu	Lys	Phe	Glu	Glu	Leu	Thr	Phe	Ala	Thr	Glu	Thr
			180					185					190		
Ser	Ser	Lys	Val	Lys	Lys	Asp	Gly	Ser	Pro	Ala	Asp	Ile	Arg	Asp	Glu
		195					200					205			
Leu	Thr	Glu	Leu	Thr	Glu	Leu	Ala	Lys	Ser	Val	Thr	Lys	Asn	Asp	Val
	210					215					220				
Asp	Gly	Phe	Glu	Phe	Tyr	Leu	Asn	Thr	Phe	His	Asp	Val	Met	Val	Gly
225					230					235					240
Asn	Asn	Leu	Phe	Gly	Arg	Ser	Ala	Leu	Lys	Thr	Ala	Ser	Glu	Leu	Ile
			245						250					255	
Thr	Lys	Glu	Asn	Val	Lys	Thr	Ser	Gly	Ser	Glu	Val	Gly	Asn	Val	Tyr
			260					265					270		
Asn	Phe	Leu	Ile	Val	Leu	Thr	Ala	Leu	Gln	Ala	Lys	Ala	Phe	Leu	Thr
	275						280					285			
Leu	Thr	Pro	Cys	Arg	Lys	Leu	Leu	Gly	Leu	Ala	Asp	Ile	Asp	Tyr	Thr
	290					295					300				
Ser	Ile	Met	Asn	Glu	His	Leu	Asn	Lys	Glu	Lys	Glu	Glu	Phe	Arg	Val
305					310					315					320
Asn	Ile	Leu	Pro	Thr	Leu	Ser	Asn	Thr	Phe	Ser	Asn	Pro	Asn	Tyr	Ala
			325						330					335	
Lys	Val	Lys	Gly	Ser	Asp	Glu	Asp	Ala	Lys	Met	Ile	Val	Glu	Ala	Lys
			340					345					350		
Pro	Gly	His	Ala	Leu	Ile	Gly	Phe	Glu	Ile	Ser	Asn	Asp	Ser	Ile	Thr
		355					360					365			
Val	Leu	Lys	Val	Tyr	Glu	Ala	Lys	Leu	Lys	Gln	Asn	Tyr	Gln	Val	Asp
	370					375					380				
Lys	Asp	Ser	Leu	Ser	Glu	Val	Ile	Tyr	Gly	Asp	Met	Asp	Lys	Leu	Leu
385					390					395					400
Cys	Pro	Asp	Gln	Ser	Gly	Gln	Ile	Tyr	Tyr	Thr	Asn	Asn	Ile	Val	Phe
			405						410					415	
Pro	Asn	Glu	Tyr	Val	Ile	Thr	Lys	Ile	Asp	Phe	Thr	Lys	Lys	Met	Lys
			420					425					430		
Thr	Leu	Arg	Tyr	Glu	Val	Thr	Ala	Asn	Phe	Tyr	Asp	Ser	Ser	Thr	Gly
		435					440					445			
Glu	Ile	Asp	Leu	Asn	Lys	Lys	Lys	Val	Glu	Ser	Ser	Glu	Ala	Glu	Tyr
	450				455						460				
Arg	Thr	Leu	Ser	Ala	Asn	Asp	Asp	Gly	Val	Tyr	Met	Pro	Leu	Gly	Val

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4 6 5				4 7 0				4 7 5				4 8 0			
I l e	S e r	G l u	T h r	P h e 4 8 5	L e u	T h r	P r o	I l e	A s n 4 9 0	G l y	P h e	G l y	L e u	G l n 4 9 5	A l a
A s p	G l u	A s n 5 0 0	S e r	A r g	L e u	I l e	T h r	L e u 5 0 5	T h r	C y s	L y s	S e r	T y r 5 1 0	L e u	A r g
G l u	L e u 5 1 5	L e u	L e u	A l a	T h r	A s p	L e u 5 2 0	S e r	A s n	L y s	G l u	T h r 5 2 5	L y s	L e u	I l e
V a l 5 3 0	P r o	P r o	S e r	G l y	P h e	I l e 5 3 5	S e r	A s n	I l e	V a l	G l u 5 4 0	A s n	G l y	S e r	I l e
G l u 5 4 5	G l u	A s p	A s n	L e u	G l u 5 5 0	P r o	T r p	L y s	A l a	A s n 5 5 5	A s n	L y s	A s n	A l a	T y r 5 6 0
V a l	A s p	H i s	T h r	G l y 5 6 5	G l y	V a l	A s n	G l y	T h r 5 7 0	L y s	A l a	L e u	T y r	V a l 5 7 5	H i s
L y s	A s p	G l y	G l y 5 8 0	I l e	S e r	G l n	P h e	I l e 5 8 5	G l y	A s p	L y s	L e u	L y s 5 9 0	P r o	L y s
T h r	G l u	T y r 5 9 5	V a l	I l e	G l n	T y r	T h r 6 0 0	V a l	L y s	G l y	L y s	P r o 6 0 5	S e r	I l e	H i s
L e u 6 1 0	L y s	A s p	G l u	A s n	T h r	G l y 6 1 5	T y r	I l e	H i s	T y r	G l u 6 2 0	A s p	T h r	A s n	A s n
A s n 6 2 5	L e u	G l u	A s p	T y r	G l n 6 3 0	T h r	I l e	A s n	L y s	A r g 6 3 5	P h e	T h r	T h r	G l y	T h r 6 4 0
A s p	L e u	L y s	G l y	V a l 6 4 5	T y r	L e u	I l e	L e u	L y s 6 5 0	S e r	G l n	A s n	G l y	A s p 6 5 5	G l u
A l a	T r p	G l y	A s p 6 6 0	A s n	P h e	I l e	I l e	L e u 6 6 5	G l u	I l e	S e r	P r o	S e r 6 7 0	G l u	L y s
L e u	L e u 6 7 5	S e r	P r o	G l u	L e u	I l e	A s n 6 8 0	T h r	A s n	A s n	T r p	T h r 6 8 5	S e r	T h r	G l y
S e r 6 9 0	T h r	A s n	I l e	S e r	G l y	A s n 6 9 5	T h r	L e u	T h r	L e u	T y r 7 0 0	G l n	G l y	G l y	A r g
G l y 7 0 5	I l e	L e u	L y s	G l n 7 1 0	A s n	L e u	G l n	L e u	A s p	S e r 7 1 5	P h e	S e r	T h r	T y r	A r g 7 2 0
V a l	T y r	P h e	S e r	V a l 7 2 5	S e r	G l y	A s p	A l a	A s n 7 3 0	V a l	A r g	I l e	A r g	A s n 7 3 5	S e r
A r g	G l u	V a l 7 4 0	L e u	P h e	G l u	L y s	A r g	T y r 7 4 5	M e t	S e r	G l y	A l a	L y s 7 5 0	A s p	V a l
S e r	G l u	M e t 7 5 5	P h e	T h r	T h r	L y s	P h e 7 6 0	G l u	L y s	A s p	A s n	P h e 7 6 5	T y r	I l e	G l u
L e u 7 7 0	S e r	G l n	G l y	A s n	A s n	L e u 7 7 5	T y r	G l y	G l y	P r o	I l e 7 8 0	V a l	H i s	P h e	T y r
A s p 7 8 5	V a l	S e r	I l e	L y s											

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2364 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA (genomic)

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 56..2292
- (D) OTHER INFORMATION: /product="VIP3A(c) protein"

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(x i) SEQUENCE DESCRIPTION: SEQ ID NO:5:

AATACAATTT	ACGAGGGATA	AGTGTTACAA	AGAATAGCTG	AGGAGGGAGA	TGAAC	ATG	58
						Met 1	
AAC	AAG	AAT	AAT	GCT	AAA	TTA	106
Asn	Lys	Asn	Asn	Ala	Lys	Leu	
			5				
AGC	ACA	AGA	GCC	TTA	CCA	AGT	
Ser	Thr	Arg	Ala	Leu	Pro	Ser	
	10				15		
GAT	TAT	TTC	AAT	GGC	ATT	TAT	154
Asp	Tyr	Phe	Asn	Gly	Ile	Tyr	
		20				25	
GGT	TTT	GCC	ACT	GGT	ATC	AAA	
Gly	Phe	Ala	Thr	Gly	Ile	Lys	
					30		
ATG	AAC	ATG	ATT	TTT	AAA	ACG	202
Met	Asn	Met	Ile	Phe	Lys	Thr	
	35					40	
GAT	ACA	GGT	GGT	GAT	CTA	GCC	
Thr	Gly	Gly	Asp	Leu	Ala	Leu	
			45				
GAA	ATT	TTA	GAG	AAT	CAG	CTA	250
Glu	Ile	Leu	Glu	Asn	Gln	Leu	
				55			
GAT	CTA	AAT	GAT	ATT	TCT	GGT	
Asp	Leu	Asn	Asp	Ile	Ser	Gly	
			60				
GAT	GGG	GTG	AAT	GGA	AGC	TTA	298
Asp	Gly	Val	Asn	Gly	Ser	Leu	
			70				
GAT	CTT	ATC	GCA	CAG	GGA	AAC	
Leu	Ile	Ala	Gln	Gly	Asn	Leu	
			75		80		
AAT	ACA	GAA	TTA	TCT	AAG	GAA	346
Asn	Thr	Glu	Leu	Ser	Lys	Glu	
			85				
AAT	ATA	TTA	AAA	ATT	GCA	AAT	
Leu	Ile	Leu	Lys	Ile	Ala	Asn	
		90				95	
CAA	GTT	TTA	AAT	GAT	GTT	AAT	394
Gln	Val	Leu	Asn	Asp	Val	Asn	
		100				105	
AAA	CTC	GAT	GCG	ATA	AAT	ACG	
Lys	Leu	Asp	Ala	Ile	Asn	Thr	
				110			
CTT	CGG	GTA	TAT	CTA	CCT	AAA	442
Leu	Arg	Val	Tyr	Leu	Pro	Lys	
		115				120	
ATT	ACC	TCT	ATG	TTG	AGT	GAT	
Ile	Thr	Ser	Met	Leu	Ser	Asp	
				125			
AAA	CAA	AAT	TAT	GCG	CTA	AGT	490
Lys	Gln	Asn	Tyr	Ala	Leu	Ser	
				135			
GAT	CTG	CAA	ATA	GAA	TAC	TTA	
Leu	Gln	Ile	Glu	Tyr	Leu	Ser	
			140				
TTG	CAA	GAG	ATT	TCT	GAT	AAG	538
Leu	Gln	Glu	Ile	Ser	Asp	Lys	
			150				
GAT	ATT	ATT	AAT	GTA	AAT	GTA	
Asp	Ile	Ile	Asn	Val	Asn	Val	
	155			160			
ATT	AAC	TCT	ACA	CTT	ACT	GAA	586
Ile	Asn	Ser	Leu	Thr	Glu	Ile	
			165				
ACA	CCT	GCG	TAT	CAA	AGG	ATT	
Thr	Pro	Ala	Tyr	Gln	Arg	Ile	
				175			
TAT	GTG	AAC	GAA	AAA	TTT	GAG	634
Tyr	Val	Asn	Glu	Lys	Phe	Glu	
		180				185	
GAT	TTA	ACT	TTT	GCT	ACA	GAA	
Leu	Thr	Phe	Ala	Thr	Glu	Thr	
				190			
TCA	AAA	GTA	AAA	AAG	GAT	GGC	682
Ser	Lys	Val	Lys	Lys	Asp	Ser	
		195				200	
TCT	CCT	GCA	GAT	ATT	CGT	GAT	
Pro	Ala	Asp	Ile	Arg	Asp	Glu	
				205			
AGT	GAG	TTA	ACT	GAA	CTA	GCG	730
Ser	Glu	Leu	Thr	Glu	Leu	Ala	
				215			
GAT	GTA	ACA	CAA	AAT	GAT	GTG	
Val	Val	Thr	Gln	Asn	Asp	Val	
		220				225	
GGT	TTT	GAA	TTT	TAC	CTT	AAT	778
Gly	Phe	Glu	Phe	Tyr	Leu	Asn	
			230				
ACA	TTC	CAC	GAT	GTA	ATG	GTA	
Thr	Phe	His	Asp	Val	Met	Val	
		235				240	
AAT	TTA	TTC	GGG	CGT	TCA	GCT	826
Asn	Leu	Phe	Gly	Arg	Ser	Ala	
			245				
AAA	ACT	GCA	TCG	GAA	TTA	ATT	
Lys	Thr	Ala	Ser	Glu	Leu	Ile	
		250				255	
AAA	GAA	AAT	GTG	AAA	ACA	AGT	874
Lys	Glu	Asn	Val	Lys	Thr	Ser	
		260					
GGC	AGT	GAG	GTC	GGA	AAT	GTT	
Gly	Ser	Glu	Val	Gly	Asn	Val	
		265				270	
TTC	CTA	ATT	GTA	TTA	ACA	GCT	922
Phe	Leu	Ile	Val	Leu	Thr	Ala	
		275				280	
CTG	CAA	GCA	CAA	GCT	TTT	CTT	
Leu	Gln	Ala	Gln	Phe	Leu	Thr	
			285				
ACA	CCA	TGC	CGA	AAA	TTA	TTA	970
Thr	Pro	Cys	Arg	Lys	Leu	Leu	
GGC	TTA	GCA	GAT	ATT	GAT	TAT	
Gly	Leu	Ala	Asp	Ile	Asp	Tyr	
ACT	TCT						
Thr	Ser						

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290			295			300			305							
ATT Ile	ATG Met	AAT Asn	GAA Glu	CAT His 310	TTA Leu	AAT Asn	AAG Lys	GAA Glu	AAA Lys 315	GAG Glu	GAA Glu	TTT Phe	AGA Arg	GTA Val 320	AAC Asn	1018
ATC Ile	CTC Leu	CCT Pro	ACA Thr 325	CTT Leu	TCT Ser	AAT Asn	ACT Thr	TTT Phe 330	TCT Ser	AAT Asn	CCT Pro	AAT Asn	TAT Tyr 335	GCA Ala	AAA Lys	1066
GTT Val	AAA Lys	GGA Gly 340	AGT Ser	GAT Asp	GAA Glu	GAT Asp	GCA Ala 345	AAG Lys	ATG Met	ATT Ile	GTG Val	GAA Glu 350	GCT Ala	AAA Lys	CCA Pro	1114
GGA Gly	CAT His 355	GCA Ala	TTG Leu	ATT Ile	GGG Gly	TTT Phe 360	GAA Glu	ATT Ile	AGT Ser	AAT Asn	GAT Asp 365	TCA Ser	ATT Ile	ACA Thr	GTA Val	1162
TTA Leu 370	AAA Lys	GTA Val	TAT Tyr	GAG Glu	GCT Ala 375	AAG Lys	CTA Leu	AAA Lys	CAA Gln	AAT Asn 380	TAT Tyr	CAA Gln	GTC Val	GAT Asp	AAG Lys 385	1210
GAT Asp	TCC Ser	TTA Leu	TCG Ser	GAA Glu 390	GTT Val	ATT Ile	TAT Tyr	GGC Gly	GAT Asp 395	ATG Met	GAT Asp	AAA Lys	TTA Leu	TTG Leu 400	TGC Cys	1258
CCA Pro	GAT Asp	CAA Gln	TCT Ser 405	GGA Gly	CAA Gln	ATC Ile	TAT Tyr	TAT Tyr 410	ACA Thr	AAT Asn	AAC Asn	ATA Ile 415	GTA Val	TTT Phe	CCA Pro	1306
AAT Asn	GAA Glu	TAT Tyr 420	GTA Val	ATT Ile	ACT Thr	AAA Lys	ATT Ile 425	GAT Asp	TTC Phe	ACT Thr	AAA Lys	AAA Lys 430	ATG Met	AAA Lys	ACT Thr	1354
TTA Leu	AGA Arg 435	TAT Tyr	GAG Glu	GTA Val	ACA Thr	GCG Ala 440	AAT Asn	TTT Phe	TAT Tyr	GAT Asp	TCT Ser 445	TCT Ser	ACA Thr	GGA Gly	GAA Glu	1402
ATT Ile 450	GAC Asp	TTA Leu	AAT Asn	AAG Lys 455	AAA Lys	AAA Lys	GTA Val	GAA Glu	TCA Ser	AGT Ser 460	GAA Glu	GCG Ala	GAG Glu	TAT Tyr	AGA Arg 465	1450
ACG Thr	TTA Leu	AGT Ser	GCT Ala	AAT Asn 470	GAT Asp	GAT Asp	GGG Gly	GTG Val	TAT Tyr 475	ATG Met	CCG Pro	TTA Leu	GGT Gly	GTC Val 480	ATC Ile	1498
AGT Ser	GAA Glu	ACA Thr	TTT Phe 485	TTG Leu	ACT Thr	CCG Pro	ATT Ile	AAT Asn 490	GGG Gly	TTT Phe	GGC Gly	CTC Leu	CAA Gln 495	GCT Ala	GAT Asp	1546
GAA Glu	AAT Asn	TCA Ser 500	AGA Arg	TTA Leu	ATT Ile	ACT Thr	TTA Leu 505	ACA Thr	TGT Cys	AAA Lys	TCA Ser	TAT Tyr 510	TTA Leu	AGA Arg	GAA Glu	1594
CTA Leu	CTG Leu 515	CTA Leu	GCA Ala	ACA Thr	GAC Asp	TTA Leu 520	AGC Ser	AAT Asn	AAA Lys	GAA Glu	ACT Thr 525	AAA Lys	TTG Leu	ATC Ile	GTC Val	1642
CCG Pro 530	CCA Pro	AGT Ser	GGT Gly	TTT Phe 535	ATT Ile	AGC Ser	AAT Asn	ATT Ile	GTA Val	GAG Glu 540	AAC Asn	GGG Gly	TCC Ser	ATA Ile	GAA Glu 545	1690
GAG Glu	GAC Asp	AAT Asn	TTA Leu	GAG Glu 550	CCG Pro	TGG Trp	AAA Lys	GCA Ala	AAT Asn 555	AAT Asn	AAG Lys	AAT Asn	GCG Ala	TAT Tyr 560	GTA Val	1738
GAT Asp	CAT His	ACA Thr	GGC Gly 565	GGA Gly	GTG Val	AAT Asn	GGA Gly	ACT Thr 570	AAA Lys	GCT Ala	TTA Leu	TAT Tyr	GTT Val 575	CAT His	AAG Lys	1786
GAC Asp	GGA Gly	GGA Gly 580	ATT Ile	TCA Ser	CAA Gln	TTT Phe 585	ATT Ile 585	GGA Gly	GAT Asp	AAG Lys	TTA Leu	AAA Lys 590	CCG Pro	AAA Lys	ACT Thr	1834
GAG Glu	TAT Tyr 595	GTA Val	ATC Ile	CAA Gln	TAT Tyr	ACT Thr 600	GTT Val	AAA Lys	GGA Gly	AAA Lys	CCT Pro 605	TCT Ser	ATT Ile	CAT His	TTA Leu	1882
AAA Lys	GAT Asp	GAA Glu	AAT Asn	ACT Thr	GGA Gly	TAT Tyr	ATT Ile	CAT His	TAT Tyr	GAA Glu	GAT Asp	ACA Thr	AAT Asn	AAT Asn	AAT Asn	1930

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6 1 0					6 1 5					6 2 0					6 2 5					
TTA Leu	GAA Glu	GAT Asp	TAT Tyr	CAA Gln 6 3 0	ACT Thr	ATT Ile	AAT Asn	AAA Lys	CGT Arg 6 3 5	TTT Phe	ACT Thr	ACA Thr	GGA Gly	ACT Thr 6 4 0	GAT Asp	1978				
TTA Leu	AAG Lys	GGA Gly	GTG Val 6 4 5	TAT Tyr	TTA Leu	ATT Ile	TTA Leu	AAA Lys 6 5 0	AGT Ser	CAA Gln	AAT Asn	GGA Gly	GAT Asp 6 5 5	GAA Glu	GCT Ala	2026				
TGG Trp	GGA Gly	GAT Asp 6 6 0	AAC Asn	TTT Phe	ATT Ile	ATT Ile	TTG Leu 6 6 5	GAA Glu	ATT Ile	AGT Ser	CCT Pro	TCT Ser 6 7 0	GAA Glu	AAG Lys	TTA Leu	2074				
TTA Leu	AGT Ser 6 7 5	CCA Pro	GAA Glu	TTA Leu	ATT Ile	AAT Asn 6 8 0	ACA Thr	AAT Asn	AAT Asn	TGG Trp	ACG Thr 6 8 5	AGT Ser	ACG Thr	GGA Gly	TCA Ser	2122				
ACT Thr 6 9 0	AAT Asn	ATT Ile	AGC Ser	GGT Gly	AAT Asn 6 9 5	ACA Thr	CTC Leu	ACT Thr	CTT Leu	TAT Tyr 7 0 0	CAG Gln	GGA Gly	GGA Gly	CGA Arg	GGG Gly 7 0 5	2170				
ATT Ile	CTA Leu	AAA Lys	CAA Gln	AAC Asn 7 1 0	CTT Leu	CAA Gln	TTA Leu	GAT Asp	AGT Ser 7 1 5	TTT Phe	TCA Ser	ACT Thr	TAT Tyr	AGA Arg 7 2 0	GTG Val	2218				
TAT Tyr	TTC Phe	TCT Ser	GTG Val 7 2 5	TCC Ser	GGA Gly	GAT Asp	GCT Ala	AAT Asn 7 3 0	GTA Val	AGG Arg	ATT Ile	AGA Arg	AAT Asn 7 3 5	TCT Ser	AGG Arg	2266				
GAA Glu	GTG Val 7 4 0	TTA Leu	TTT Phe	GAA Glu	AAA Lys	AAG Lys	GAT Asp 7 4 5	ATA Ile	TGA	GC	GGCGCTAAAG	ATGTTTCTGA				2318				
AATGTTCACT ACAAAATTGA AAGATAACTT CTATATAGAG CTTTCT																2364				

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 746 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Asn Lys Asn Asn Ala Lys Leu Ser Thr Arg Ala Leu Pro Ser Phe
 1 5 10 15

Ile Asp Tyr Phe Asn Gly Ile Tyr Gly Phe Ala Thr Gly Ile Lys Asp
 20 25 30

Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly Gly Asp Leu Ala Leu
 35 40 45

Asp Glu Ile Leu Glu Asn Gln Gln Leu Leu Asn Asp Ile Ser Gly Lys
 50 55 60

Leu Asp Gly Val Asn Gly Ser Leu Asn Asp Leu Ile Ala Gln Gly Asn
 65 70 75 80

Leu Asn Thr Glu Leu Ser Lys Glu Ile Leu Lys Ile Ala Asn Glu Gln
 85 90 95

Asn Gln Val Leu Asn Asp Val Asn Asn Lys Leu Asp Ala Ile Asn Thr
 100 105 110

Met Leu Arg Val Tyr Leu Pro Lys Ile Thr Ser Met Leu Ser Asp Val
 115 120 125

Met Lys Gln Asn Tyr Ala Leu Ser Leu Gln Ile Glu Tyr Leu Ser Lys
 130 135 140

Gln Leu Gln Glu Ile Ser Asp Lys Leu Asp Ile Ile Asn Val Asn Val
 145 150 155 160

Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro Ala Tyr Gln Arg Ile
 165 170 175

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81

82

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Lys	Tyr	Val	Asn	Glu	Lys	Phe	Glu	Glu	Leu	Thr	Phe	Ala	Thr	Glu	Thr
			180					185					190		
Ser	Ser	Lys	Val	Lys	Lys	Asp	Gly	Ser	Pro	Ala	Asp	Ile	Arg	Asp	Glu
		195					200					205			
Leu	Ser	Glu	Leu	Thr	Glu	Leu	Ala	Lys	Ser	Val	Thr	Gln	Asn	Asp	Val
	210					215					220				
Asp	Gly	Phe	Glu	Phe	Tyr	Leu	Asn	Thr	Phe	His	Asp	Val	Met	Val	Gly
225					230					235					240
Asn	Asn	Leu	Phe	Gly	Arg	Ser	Ala	Leu	Lys	Thr	Ala	Ser	Glu	Leu	Ile
				245					250					255	
Thr	Lys	Glu	Asn	Val	Lys	Thr	Ser	Gly	Ser	Glu	Val	Gly	Asn	Val	Tyr
			260					265					270		
Asn	Phe	Leu	Ile	Val	Leu	Thr	Ala	Leu	Gln	Ala	Gln	Ala	Phe	Leu	Thr
		275					280					285			
Leu	Thr	Pro	Cys	Arg	Lys	Leu	Leu	Gly	Leu	Ala	Asp	Ile	Asp	Tyr	Thr
	290					295					300				
Ser	Ile	Met	Asn	Glu	His	Leu	Asn	Lys	Glu	Lys	Glu	Glu	Phe	Arg	Val
305					310					315					320
Asn	Ile	Leu	Pro	Thr	Leu	Ser	Asn	Thr	Phe	Ser	Asn	Pro	Asn	Tyr	Ala
				325					330					335	
Lys	Val	Lys	Gly	Ser	Asp	Glu	Asp	Ala	Lys	Met	Ile	Val	Glu	Ala	Lys
			340					345					350		
Pro	Gly	His	Ala	Leu	Ile	Gly	Phe	Glu	Ile	Ser	Asn	Asp	Ser	Ile	Thr
		355					360					365			
Val	Leu	Lys	Val	Tyr	Glu	Ala	Lys	Leu	Lys	Gln	Asn	Tyr	Gln	Val	Asp
	370					375					380				
Lys	Asp	Ser	Leu	Ser	Glu	Val	Ile	Tyr	Gly	Asp	Met	Asp	Lys	Leu	Leu
385					390					395					400
Cys	Pro	Asp	Gln	Ser	Gly	Gln	Ile	Tyr	Tyr	Thr	Asn	Asn	Ile	Val	Phe
				405					410					415	
Pro	Asn	Glu	Tyr	Val	Ile	Thr	Lys	Ile	Asp	Phe	Thr	Lys	Lys	Met	Lys
			420					425					430		
Thr	Leu	Arg	Tyr	Glu	Val	Thr	Ala	Asn	Phe	Tyr	Asp	Ser	Ser	Thr	Gly
		435					440					445			
Glu	Ile	Asp	Leu	Asn	Lys	Lys	Lys	Val	Glu	Ser	Ser	Glu	Ala	Glu	Tyr
	450					455					460				
Arg	Thr	Leu	Ser	Ala	Asn	Asp	Asp	Gly	Val	Tyr	Met	Pro	Leu	Gly	Val
465					470					475					480
Ile	Ser	Glu	Thr	Phe	Leu	Thr	Pro	Ile	Asn	Gly	Phe	Gly	Leu	Gln	Ala
				485					490					495	
Asp	Glu	Asn	Ser	Arg	Leu	Ile	Thr	Leu	Thr	Cys	Lys	Ser	Tyr	Leu	Arg
			500					505					510		
Glu	Leu	Leu	Leu	Ala	Thr	Asp	Leu	Ser	Asn	Lys	Glu	Thr	Lys	Leu	Ile
		515					520					525			
Val	Pro	Pro	Ser	Gly	Phe	Ile	Ser	Asn	Ile	Val	Glu	Asn	Gly	Ser	Ile
	530					535					540				
Glu	Glu	Asp	Asn	Leu	Glu	Pro	Trp	Lys	Ala	Asn	Asn	Lys	Asn	Ala	Tyr
545					550					555					560
Val	Asp	His	Thr	Gly	Gly	Val	Asn	Gly	Thr	Lys	Ala	Leu	Tyr	Val	His
				565					570					575	
Lys	Asp	Gly	Gly	Ile	Ser	Gln	Phe	Ile	Gly	Asp	Lys	Leu	Lys	Pro	Lys
			580					585					590		
Thr	Glu	Tyr	Val	Ile	Gln	Tyr	Thr	Val	Lys	Gly	Lys	Pro	Ser	Ile	His

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5 9 5						6 0 0						6 0 5					
Leu	Lys 610	Asp	Glu	Asn	Thr	Gly 615	Tyr	Ile	His	Tyr	Glu 620	Asp	Thr	Asn	Asn		
Asn 625	Leu	Glu	Asp	Tyr	Gln 630	Thr	Ile	Asn	Lys	Arg 635	Phe	Thr	Thr	Gly	Thr 640		
Asp	Leu	Lys	Gly	Val 645	Tyr	Leu	Ile	Leu	Lys 650	Ser	Gln	Asn	Gly	Asp 655	Glu		
Ala	Trp	Gly	Asp 660	Asn	Phe	Ile	Ile	Leu 665	Glu	Ile	Ser	Pro	Ser 670	Glu	Lys		
Leu	Leu	Ser 675	Pro	Glu	Leu	Ile	Asn 680	Thr	Asn	Asn	Trp	Thr 685	Ser	Thr	Gly		
Ser	Thr 690	Asn	Ile	Ser	Gly	Asn 695	Thr	Leu	Thr	Leu	Tyr 700	Gln	Gly	Gly	Arg		
Gly 705	Ile	Leu	Lys	Gln	Asn 710	Leu	Gln	Leu	Asp	Ser 715	Phe	Ser	Thr	Tyr	Arg 720		
Val	Tyr	Phe	Ser	Val 725	Ser	Gly	Asp	Ala	Asn 730	Val	Arg	Ile	Arg	Asn 735	Ser		
Arg	Glu	Val	Leu 740	Phe	Glu	Lys	Lys	Asp 745	Ile								

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2403 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 11..2389
- (D) OTHER INFORMATION: /note= "maize optimized DNA
sequence encoding VIP3A(a)"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:7:

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GGATCCACCA ATGAACATGA ACAAGAACAA CACCAAGCTG AGCACCCGCG CCCTGCCGAG      60
CTTCATCGAC TACTTCAACG GCATCTACGG CTTGCCACC GGCATCAAGG ACATCATGAA      120
CATGATCTTC AAGACCGACA CCGGCGGCGA CCTGACCCTG GACGAGATCC TGAAGAACCA      180
GCAGCTGCTG AACGACATCA GCGGCAAGCT GGACGGCGTG AACGGCAGCC TGAACGACCT      240
GATCGCCCAG GGCAACCTGA ACACCGAGCT GAGCAAGGAG ATCCTTAAGA TCGCCAACGA      300
GCAGAACCAG GTGCTGAACG ACGTGAACAA CAAGCTGGAC GCCATCAACA CCATGCTGCG      360
CGTGTACCTG CCGAAGATCA CCAGCATGCT GAGCGACGTG ATGAAGCAGA ACTACGCCCT      420
GAGCCTGCAG ATCGAGTACC TGAGCAAGCA GCTGCAGGAG ATCAGCGACA AGCTGGACAT      480
CATCAACGTG AACGTCCTGA TCAACAGCAC CCTGACCGAG ATCACCCCGG CCTACCAGCG      540
CATCAAGTAC GTGAACGAGA AGTTCGAAGA GCTGACCTTC GCCACCGAGA CCAGCAGCAA      600
GGTGAAGAAG GACGGCAGCC CGGCCGACAT CCTGGACGAG CTGACCGAGC TGACCGAGCT      660
GGCCAAGAGC GTGACCAAGA ACGACGTGGA CGGCTTCGAG TTCTACCTGA ACACCTTCCA      720
CGACGTGATG GTGGGCAACA ACCTGTTCTGG CCGCAGCGCC CTGAAGACCG CCAGCGAGCT      780
GATCACCAAG GAGAACGTGA AGACCAGCGG CAGCGAGGTG GGCAACGTGT ACAACTTCCT      840
GATCGTGCTG ACCGCCCTGC AGGCCCAGGC CTTCTGTACC CTGACCACCT GTCGCAAGCT      900

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GCTGGGCCTG	GCCGACATCG	ACTACACCAG	CATCATGAAC	GAGCACTTGA	ACAAGGAGAA	9 6 0
GGAGGAGTTC	CGCGTGAACA	TCCTGCCGAC	CCTGAGCAAC	ACCTTCAGCA	ACCCGAAC TA	1 0 2 0
CGCCAAGGTG	AAGGGCAGCG	ACGAGGACGC	CAAGATGATC	GTGGAGGCTA	AGCCGGGGCCA	1 0 8 0
CGCGTTGATC	GGCTTCGAGA	TCAGCAACGA	CAGCATCACC	GTGCTGAAGG	TGTACGAGGC	1 1 4 0
CAAGCTGAAG	CAGAACTACC	AGGTGGACAA	GGACAGCTTG	AGCGAGGTGA	TCTACGGCGA	1 2 0 0
CATGGACAAAG	CTGCTGTGTC	CGGACCAGAG	CGAGCAAATC	TACTACACCA	ACAACATCGT	1 2 6 0
GTTCCCGAAC	GAGTACGTGA	TCACCAAGAT	CGACTTCACC	AAGAAGATGA	AGACCCTGCG	1 3 2 0
CTACGAGGTG	ACCGCCAAC T	TCTACGACAG	CAGCACCGGC	GAGATCGACC	TGAACAAGAA	1 3 8 0
GAAGGTGGAG	AGCAGCGAGG	CCGAGTACCG	CACCCTGAGC	GCGAACGACG	ACGGCGTCTA	1 4 4 0
CATGCCACTG	GGCGTGATCA	GCGAGACCTT	CCTGACCCCG	ATCAACGGCT	TTGGCCTGCA	1 5 0 0
GGCCGACGAG	AACAGCCGCC	TGATCACCCCT	GACCTGTAAG	AGCTACCTGC	GCGAGCTGCT	1 5 6 0
GCTAGCCACC	GACCTGAGCA	ACAAGGAGAC	CAAGCTGATC	GTGCCACCGA	GCGGCTTCAT	1 6 2 0
CAGCAACATC	GTGGAGAACG	GCAGCATCGA	GGAGGACAAC	CTGGAGCCGT	GGAAGGCCAA	1 6 8 0
CAACAAGAAC	GCCTACGTGG	ACCACACCGG	CGGCGTGAAC	GGCACCAAGG	CCCTGTACGT	1 7 4 0
GCACAAGGAC	GGCGGCATCA	GCCAGTTCAT	CGGCGACAAG	CTGAAGCCGA	AGACCGAGTA	1 8 0 0
CGTGATCCAG	TACACCGTGA	AGGGCAAGCC	ATCGATTAC	CTGAAGGACG	AGAACACCGG	1 8 6 0
CTACATCCAC	TACGAGGACA	CCAACAACAA	CCTGGAGGAC	TACCAGACCA	TCAACAAGCG	1 9 2 0
CTTCAACCACC	GGCACCGACC	TGAAGGGCGT	GTACCTGATC	CTGAAGAGCC	AGAACGGCGA	1 9 8 0
CGAGGCCTGG	GGCGACAAC T	TCATCATCCT	GGAGATCAGC	CCGAGCGAGA	AGCTGCTGAG	2 0 4 0
CCCGGAGCTG	ATCAACACCA	ACAAC TGGAC	CAGCACCGGC	AGCACCAACA	TCAGCGGCAA	2 1 0 0
CACCCTGACC	CTGTACCAGG	GCGGCCGCGG	CATCCTGAAG	CAGAACCTGC	AGCTGGACAG	2 1 6 0
CTTCAGCACC	TACCGCGTGT	ACTTCAGCGT	GAGCGGCGAC	GCCAACGTGC	GCATCCGCAA	2 2 2 0
CAGCCGCGAG	GTGCTGTTCG	AGAAGAGGTA	CATGAGCGGC	GCCAAGGACG	TGAGCGAGAT	2 2 8 0
GTTCAACCACC	AAGTTTCGAGA	AGGACAAC TT	CTACATCGAG	CTGAGCCAGG	GCAACAACCT	2 3 4 0
GTACGGCGGC	CCGATCGTGC	ACTTCTACGA	CGTGAGCATC	AAGTTAACGT	AGAGCTCAGA	2 4 0 0
TCT						2 4 0 3

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1638 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: cDNA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 2..1191
- (D) OTHER INFORMATION: /product="Translation of cDNA
encoding VIP3A(a) receptor"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:8:

T	AGT	GGA	TCC	CCC	GGG	CTG	CAG	GAA	TTC	GCG	GCC	GCG	TCG	ACC	ATG	4 6
	Ser	Gly	Ser	Pro	Gly	Leu	Gln	Glu	Phe	Ala	Ala	Ala	Ser	Thr	Met	
	1				5					10					15	
TAC	TCT	AGA	ATA	TTT	TTC	CTC	CTT	GTG	ATA	GTG	TGT	GCT	GTT	AAG	GCT	9 4
Tyr	Ser	Arg	Ile	Phe	Phe	Leu	Leu	Val	Ile	Val	Cys	Ala	Val	Lys	Ala	

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20				25				30											
TCT Ser	CTG Leu	TTT Phe	ACT Thr 35	GTA Val	AAT Asn	GTG Val	TAT Tyr	GAT Asp 40	GAT Asp	AAC Asn	CCC Pro	GAA Glu	ACT Thr 45	GAA Glu	ATT Ile	142			
GCG Ala	AGT Ser	AGT Ser 50	CTA Leu	AAA Lys	GGC Gly	TGT Cys	AAC Asn 55	CCC Pro	CAA Gln	GAG Glu	TGT Cys	GAC Asp 60	CAG Gln	CGG Arg	TGT Cys	190			
CGT Arg 65	AGA Arg	CTG Leu	AAG Lys	TTT Phe	CCC Pro	GGT Gly 70	GGC Gly	GCC Ala	TGT Cys	GTC Val	AAT Asn 75	GGT Gly	CGC Arg	TGC Cys	AAG Lys	238			
TGT Cys 80	GAC Asp	AAC Asn	TTC Phe	CTC Leu	AGT Ser 85	GTA Val	AAA Lys	GAT Asp	GAC Asp	GTG Val 90	TCT Ser	GTT Val	GAA Glu	GAG Glu	CCT Pro 95	286			
GCG Ala	ATT Ile	CTC Leu	AAA Lys 100	GAT Asp	TTG Leu	GTG Val	TCA Ser	TTA Leu	GAA Glu 105	GCT Ala	GAA Glu	CAG Gln	GCA Ala	GCG Ala 110	AAA Lys	334			
AGT Ser	AGA Arg	TGC Cys	AGA Arg 115	AAC Asn	AGA Arg	GTG Val	TGT Cys	GAC Asp 120	GCG Ala	GTG Val	TGC Cys	CGT Arg	GCC Ala 125	CTA Leu	CAC His	382			
AAC Asn	ACC Thr	AGT Ser 130	GGT Gly	GCC Ala	TGT Cys	GTT Val	GAT Asp 135	GGA Gly	CAA Gln	TGC Cys	AAG Lys	TGT Cys 140	ACT Thr	AAT Asn	AAG Lys	430			
ATC Ile 145	AGT Ser	GCA Ala	GGA Gly	GAT Asp	ATT Ile	GTG Val 150	TCT Ser	GAT Asp	CCT Pro	GCT Ala	GAA Glu 155	TCG Ser	CTA Leu	CGC Arg	ACT Thr	478			
TGT Cys 160	AAC Asn	CCT Pro	ATA Ile	AGG Arg	TGT Cys 165	GAC Asp	GAA Glu	CAA Gln	TGT Cys	AGA Arg 170	AGA Arg	AAT Asn	GGC Gly	CAT His	GAA Glu 175	526			
TTT Phe	GGT Gly	GTT Val	TGC Cys	TTC Phe 180	AAA Lys	GGA Gly	CAA Gln	TGC Cys	AAG Lys 185	TGT Cys	GAT Asp	TAC Tyr	TTC Phe	CTC Leu 190	AAG Lys	574			
GAA Glu	GAA Glu	GTC Val	GAT Asp 195	GAA Glu	CCT Pro	GAA Glu	GTT Val	ACA Thr 200	AGC Ser	CTT Leu	CCA Pro	AAA Lys	AAC Asn 205	TGC Cys	AAC Asn	622			
CCC Pro	CAA Gln	GAG Glu 210	TGT Cys	GAC Asp	CAG Gln	CGT Arg	TGT Cys 215	CGT Arg	AGA Arg	CTG Leu	AAG Lys	TTC Phe 220	CCC Pro	GGT Gly	GGC Gly	670			
GCC Ala 225	TGT Cys	GTC Val	AAC Asn	GGG Gly	CGC Arg	TGC Cys 230	AAG Lys	TGT Cys	GAC Asp	AAC Asn	TTC Phe 235	TTC Phe	AGT Ser	GCA Ala	GGA Gly	718			
GAT Asp 240	ATT Ile	GTG Val	TCT Ser	GAT Asp	CCT Pro 245	GCC Ala	GAA Glu	TCG Ser	CTA Leu	CGC Arg 250	TCT Ser	TGT Cys	AAC Asn	CCT Pro	ATA Ile 255	766			
AGG Arg	TGT Cys	GAC Asp	GAA Glu	CAA Gln 260	TGT Cys	AGA Arg	AGA Arg	AAT Asn	GGC Gly 265	CAT His	GAA Glu	TTT Phe	GGT Gly	GTT Val 270	TGC Cys	814			
TTC Phe	AAA Lys	GGA Gly	CAA Gln 275	TGC Cys	AAG Lys	TGT Cys	GAT Asp	TAC Tyr 280	TTC Phe	CTC Leu	AAC Asn	TCA Ser	GAA Glu 285	GTA Val	GAC Asp	862			
GCT Ala	GTT Val	AAT Asn 290	GAG Glu	TTT Phe	CCT Pro	CAA Gln	GCG Ala 295	GGC Gly	TCA Ser	AAA Lys	CGC Arg	TAC Tyr 300	TGC Cys	AAC Asn	TTA Leu	910			
ACG Thr	CAA Gln 305	TGC Cys	AAC Asn	CAG Gln	ACG Thr	TGC Cys 310	GCC Ala	AAT Asn	CGT Arg	TTC Phe	TAT Tyr 315	GAT Asp	AGT Ser	GCT Ala	AGA Arg	958			
GTG Val 320	ATC Ile	CAC His	GGC Gly	TGG Trp	TGC Cys 325	AAA Lys	TGC Cys	TAC Tyr	AGT Ser	AAG Lys 330	ATG Met	GAA Glu	AGA Arg	CAG Gln	GAT Asp 335	1006			
GCA Ala	TCT Ser	CCA Pro	TTA Leu	AAC Asn	GAT Asp	GTG Val	ACT Thr	GAG Glu	GAT Asp	GAA Glu	AAT Asn	GAA Glu	GTT Val	TCT Ser	AAC Asn	1054			

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3 4 0								3 4 5				3 5 0					
GAT	ATC	CTG	AGG	ACT	GTT	GCA	GAG	GAG	CTG	TCT	GAT	GTG	TCA	CCT	AGG	1 1 0 2	
Asp	Ile	Leu	Arg	Thr	Val	Ala	Glu	Glu	Leu	Ser	Asp	Val	Ser	Pro	Arg		
			3 5 5					3 6 0					3 6 5				
GCC	TGC	AAA	TCA	GCG	AGC	TGC	AAT	CAA	GCA	TGT	CGC	GCC	TTC	TAC	TTT	1 1 5 0	
Ala	Cys	Lys	Ser	Ala	Ser	Cys	Asn	Gln	Ala	Cys	Arg	Ala	Phe	Tyr	Phe		
		3 7 0					3 7 5					3 8 0					
AAA	GGA	GGG	TGG	TGT	CGC	TTT	GGA	CGA	TGC	CAA	TGC	TTC	TA			1 1 9 1	
Lys	Gly	Gly	Trp	Cys	Arg	Phe	Gly	Arg	Cys	Gln	Cys	Phe					
	3 8 5					3 9 0					3 9 5						
AAATTAGTAT			GATATATGAA			TTTTGTATTA		TTCGGTTAAT			TGTGTTATGT		TTAAAAAACA			1 2 5 1	
TAATGTCTTC			ATTTTAGAAA			AAAGTACCTT		CACTAAAGCG			CAACAATTAA		CTAGTAGTTA			1 3 1 1	
ATTATTAAC			AGTAGTTAAA			TTATTGATGA		TTATGATTAT			CTTAGTAGTA		GTAAATTATA			1 3 7 1	
ATCATCAACT			ATTAAGTAGT			AGTTAATTAT		TAAGTAGTAG			TTAAATTATT		GATGATTATG			1 4 3 1	
ATTATCTTAG			TAGTAGTTAA			TTATTGTTTC		TTATAATAAT			CTAGTATGTT		GGTAGGTACT			1 4 9 1	
TAATAATAAC			GCTTCTGACA			AAAAATTTAA		AATTAAATAA			TTCTATCAAA		CATAAATAAT			1 5 5 1	
AACTGAAATA			AAAATTTATA			AGAGAAAAAA		AAAAAGTCGA			CGCGGCCGCG		AATTTCGATAT			1 6 1 1	
CAAGCTTATC			GATACCGTCG			ACCTCGA											1 6 3 8

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 396 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Ser	Gly	Ser	Pro	Gly	Leu	Gln	Glu	Phe	Ala	Ala	Ala	Ser	Thr	Met	Tyr	
1				5					10					15		
Ser	Arg	Ile	Phe	Phe	Leu	Leu	Val	Ile	Val	Cys	Ala	Val	Lys	Ala	Ser	
			20					25					30			
Leu	Phe	Thr	Val	Asn	Val	Tyr	Asp	Asp	Asn	Pro	Glu	Thr	Glu	Ile	Ala	
		35					40					45				
Ser	Ser	Leu	Lys	Gly	Cys	Asn	Pro	Gln	Glu	Cys	Asp	Gln	Arg	Cys	Arg	
	50					55					60					
Arg	Leu	Lys	Phe	Pro	Gly	Gly	Ala	Cys	Val	Asn	Gly	Arg	Cys	Lys	Cys	
65					70					75					80	
Asp	Asn	Phe	Leu	Ser	Val	Lys	Asp	Asp	Val	Ser	Val	Glu	Glu	Pro	Ala	
			85						90					95		
Ile	Leu	Lys	Asp	Leu	Val	Ser	Leu	Glu	Ala	Glu	Gln	Ala	Ala	Lys	Ser	
		100						105					110			
Arg	Cys	Arg	Asn	Arg	Val	Cys	Asp	Ala	Val	Cys	Arg	Ala	Leu	His	Asn	
		115					120					125				
Thr	Ser	Gly	Ala	Cys	Val	Asp	Gly	Gln	Cys	Lys	Cys	Thr	Asn	Lys	Ile	
	130					135					140					
Ser	Ala	Gly	Asp	Ile	Val	Ser	Asp	Pro	Ala	Glu	Ser	Leu	Arg	Thr	Cys	
145					150					155					160	
Asn	Pro	Ile	Arg	Cys	Asp	Glu	Gln	Cys	Arg	Arg	Asn	Gly	His	Glu	Phe	
			165						170					175		
Gly	Val	Cys	Phe	Lys	Gly	Gln	Cys	Lys	Cys	Asp	Tyr	Phe	Leu	Lys	Glu	
			180					185					190			
Glu	Val	Asp	Glu	Pro	Glu	Val	Thr	Ser	Leu	Pro	Lys	Asn	Cys	Asn	Pro	
		195					200					205				

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Gln	Glu	Cys	Asp	Gln	Arg	Cys	Arg	Arg	Leu	Lys	Phe	Pro	Gly	Gly	Ala
	210					215					220				
Cys	Val	Asn	Gly	Arg	Cys	Lys	Cys	Asp	Asn	Phe	Phe	Ser	Ala	Gly	Asp
225					230					235					240
Ile	Val	Ser	Asp	Pro	Ala	Glu	Ser	Leu	Arg	Ser	Cys	Asn	Pro	Ile	Arg
				245					250					255	
Cys	Asp	Glu	Gln	Cys	Arg	Arg	Asn	Gly	His	Glu	Phe	Gly	Val	Cys	Phe
			260					265					270		
Lys	Gly	Gln	Cys	Lys	Cys	Asp	Tyr	Phe	Leu	Asn	Ser	Glu	Val	Asp	Ala
		275					280					285			
Val	Asn	Glu	Phe	Pro	Gln	Ala	Gly	Ser	Lys	Arg	Tyr	Cys	Asn	Leu	Thr
	290					295					300				
Gln	Cys	Asn	Gln	Thr	Cys	Ala	Asn	Arg	Phe	Tyr	Asp	Ser	Ala	Arg	Val
305					310					315					320
Ile	His	Gly	Trp	Cys	Lys	Cys	Tyr	Ser	Lys	Met	Glu	Arg	Gln	Asp	Ala
				325					330					335	
Ser	Pro	Leu	Asn	Asp	Val	Thr	Glu	Asp	Glu	Asn	Glu	Val	Ser	Asn	Asp
			340					345					350		
Ile	Leu	Arg	Thr	Val	Ala	Glu	Glu	Leu	Ser	Asp	Val	Ser	Pro	Arg	Ala
		355					360					365			
Cys	Lys	Ser	Ala	Ser	Cys	Asn	Gln	Ala	Cys	Arg	Ala	Phe	Tyr	Phe	Lys
	370					375					380				
Gly	Gly	Trp	Cys	Arg	Phe	Gly	Arg	Cys	Gln	Cys	Phe				
385					390					395					

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 14 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(i i i) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(v i) ORIGINAL SOURCE:

- (A) ORGANISM: Bacillus thuringiensis
- (B) STRAIN: AB88

(i x) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..14
- (D) OTHER INFORMATION: /note= "N-terminal amino acid
sequence of protein known as anion exchange fraction 23
(s m a l l e r)"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Xaa	Glu	Pro	Phe	Val	Ser	Ala	Xaa	Xaa	Xaa	Gln	Xaa	Xaa	Xaa
1				5						10			

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 13 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(v i) ORIGINAL SOURCE:

- (A) ORGANISM: Bacillus thuringiensis

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(x i) SEQUENCE DESCRIPTION: SEQ ID NO:11:

X a a	G l u	T y r	G l u	A s n	V a l	G l u	P r o	P h e	V a l	S e r	A l a	X a a
1				5					10			

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 14 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(v i) ORIGINAL SOURCE:

- (A) ORGANISM: Bacillus thuringiensis

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:12:

M e t	A s n	L y s	A s n	A s n	T h r	L y s	L e u	P r o	T h r	A r g	A l a	L e u	P r o
1				5					10				

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(i i i) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(v i) ORIGINAL SOURCE:

- (A) ORGANISM: Bacillus thuringiensis
- (B) STRAIN: AB88

(i x) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..15
- (D) OTHER INFORMATION: /note= "N-terminal amino acid sequence of 35 kDa VIP active against Agrotis ipsilon"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:13:

A l a	L e u	S e r	G l u	A s n	T h r	G l y	L y s	A s p	G l y	G l y	T y r	I l e	V a l	P r o
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(v i) ORIGINAL SOURCE:

- (A) ORGANISM: Bacillus thuringiensis

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:14:

M e t	A s p	A s n	A s n	P r o	A s n	I l e	A s n	G l u
1				5				

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(i x) FEATURE:

- (A) NAME/KEY: Peptide
(B) LOCATION: 1..9
(D) OTHER INFORMATION: /note= "N-terminal sequence of 80 kDa delta- endotoxin"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Met Asp Asn Asn Pro Asn Ile Asn Glu
1 5

(2) INFORMATION FOR SEO ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 11 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(i i i) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(v i) ORIGINAL SOURCE:

- (A) ORGANISM: *Bacillus thuringiensis*

(i x) FEATURE:

- (A) NAME/KEY: Peptide
(B) LOCATION: 1..11
(D) OTHER INFORMATION: /note= "N-terminal sequence from 60 kDa delta- endotoxin"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Asn Val Leu Asn Ser Gly Arg Thr Thr Ile
1 5 10

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "primer sequence"

(i i i) HYPOTHETICAL: NO

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CGATT AATGT TGG CCTC

17

(2) INFORMATION FOR SEO ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "primer sequence"

(i i i) HYPOTHETICAL: NO

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CATTAGCATC TCCGGACACA G

21

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(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2370 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "synthetic DNA encoding VIP3A(b)"

(i i i) HYPOTHETICAL: NO

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:19:

ATGAACAAGA	ACAACACCAA	GCTGAGCACC	CGCGCCCTGC	CGAGCTTCAT	CGACTACTTC	60
AACGGCATCT	ACGGCTTCGC	CACCGGCATC	AAGGACATCA	TGAACATGAT	CTTCAAGACC	120
GACACCGGCG	GCGACCTGAC	CCTGGACGAG	ATCCTGAAGA	ACCAGCAGCT	GCTGAACGAC	180
ATCAGCGGCA	AGCTGGACGG	CGTGAACGGC	AGCCTGAACG	ACCTGATCGC	CCAGGGCAAC	240
CTGAACACCG	AGCTGAGCAA	GGAGATCCTT	AAGATCGCCA	ACGAGCAGAA	CCAGGTGCTG	300
AACGACGTGA	ACAACAAGCT	GGACGCCATC	AACACCATGC	TGCGCGTGTA	CCTGCCGAAG	360
ATCACCAGCA	TGCTGAGCGA	CGTGATGAAG	CAGAACTACG	CCCTGAGCCT	GCAGATCGAG	420
TACCTGAGCA	AGCAGCTGCA	GGAGATCAGC	GACAAAGCTGG	ACATCATCAA	CGTGAACGTC	480
CTGATCAACA	GCACCCTGAC	CGAGATCACC	CCGGCCTACC	AGCGCATCAA	GTACGTGAAC	540
GAGAAAGTTCG	AAGAGCTGAC	CTTCGCCACC	GAGACCAGCA	GCAAGGTGAA	GAAGGACGGC	600
AGCCCGGCCG	ACATCCTGGA	CGAGCTGACC	GAGCTGACCG	AGCTGGCGAA	GAGCGTGACC	660
AAGAACGACG	TGGACGGCTT	CGAGTTCTAC	CTGAACACCT	TCCACGACGT	GATGGTGCGC	720
AACAACCTGT	TCGGCCGCAG	CGCCCTGAAG	ACCGCCAGCG	AGCTGATCAC	CAAGGAGAAC	780
GTGAAGACCA	GCGGCAGCGA	GGTGGGCAAC	GTGTACAAC	TCCTGATCGT	GCTGACCGCC	840
CTGCAGGCCA	AGGCCTTCCT	GACCCTGACC	CCCTGTGCGA	AGCTGCTGGG	CCTGGCCGAC	900
ATCGACTACA	CCAGCATCAT	GAACGAGCAC	TTGAACAAGG	AGAAGGAGGA	GTTCCGCGTG	960
AACATCCTGC	CGACCCTGAG	CAACACCTTC	AGCAACCCGA	ACTACGCCAA	GGTGAAGGGC	1020
AGCGACGAGG	ACGCCAAGAT	GATCGTGAG	GCTAAGCCGG	GCCACGCGTT	GATCGGCTTC	1080
GAGATCAGCA	ACGACAGCAT	CACCGTGCTG	AAGGTGTACG	AGGCCAAGCT	GAAGCAGAAC	1140
TACCAGGTGG	ACAAGGACAG	CTTGAGCGAG	GTGATCTACG	GCGACATGGA	CAAGCTGCTG	1200
TGTCCGGACC	AGAGCGGGCA	AATCTACTAC	ACCAACAACA	TCGTGTTCCC	GAACGAGTAC	1260
GTGATCACCA	AGATCGACTT	CACCAAGAAG	ATGAAGACCC	TGCGCTACGA	GGTGACCGCC	1320
AACTTCTACG	ACAGCAGCAC	CGGCGAGATC	GACCTGAACA	AGAAGAAGGT	GGAGAGCAGC	1380
GAGGCCGAGT	ACCGCACCTT	GAGCGCGAAC	GACGACGGCG	TCTACATGCC	ACTGGGCGTG	1440
ATCAGCGAGA	CCTTCCTGAC	CCCGATCAAC	GGCTTTGGCC	TGCAGGCCGA	CGAGAACAGC	1500
CGCCTGATCA	CCCTGACCTG	TAAGAGCTAC	CTGCGCGAGC	TGCTGCTAGC	CACCGACCTG	1560
AGCAACAAGG	AGACCAAGCT	GATCGTGCCA	CCGAGCGGCT	TCATCAGCAA	CATCGTGGAG	1620
AACGGCAGCA	TCGAGGAGGA	CAACCTGGAG	CCGTGGAAGG	CCAACAACAA	GAACGCCTAC	1680
GTGGACCACA	CCGGCGGGCT	GAACGGCACC	AAGGCCCTGT	ACGTGCACAA	GGACGGCGGC	1740
ATCAGCCAGT	TCATCGGCGA	CAAGCTGAAG	CCGAAGACCG	AGTACGTGAT	CCAGTACACC	1800
GTGAAGGGCA	AGCCATCGAT	TCACCTGAAG	GACGAGAACA	CCGGCTACAT	CCACTACGAG	1860
GACACCAACA	ACAACCTGGA	GGACTACCAG	ACCATCAACA	AGCGCTTCAC	CACCGGCACC	1920

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GACCTGAAGG	GCGTGACCT	GATCCTGAAG	AGCCAGAACG	GCGACGAGGC	CTGGGGCGAC	1980
AACTTCATCA	TCCTGGAGAT	CAGCCCGAGC	GAGAAGCTGC	TGAGCCCGGA	GCTGATCAAC	2040
ACCAACAAC	GGACCAGCAC	CGGCAGCACC	AACATCAGCG	GCAACACCCT	GACCCTGTAC	2100
CAGGGCGGCC	GCGGCATCCT	GAAGCAGAAC	CTGCAGCTGG	ACAGCTTCAG	CACCTACCGC	2160
GTGTACTTCA	GCGTGAGCGG	CGACGCCAAC	GTGCGCATCC	GCAACTCCCG	CGAGGTGCTG	2220
TTCAAGAAGA	GGTACATGAG	CGGCGCCAAG	GACGTGAGCG	AGATGTTTAC	CACCAAGTTC	2280
GAGAAGGACA	ACTTCTACAT	CGAGCTGAGC	CAGGGCAACA	ACCTGTACGG	CGGCCCGATC	2340
GTGCACTTCT	ACGACGTGAG	CATCAAGTAG				2370

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2241 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "synthetic DNA encoding VIP3A(c)"

(i i i) HYPOTHETICAL: NO

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:20:

ATGAACAAGA	ACAACGCCAA	GCTGAGCACC	CGCGCCCTGC	CGAGCTTCAT	CGACTACTTC	60
AACGGCATCT	ACGGCTTCGC	CACCGGCATC	AAGGACATCA	TGAACATGAT	CTTCAAGACC	120
GACACCGGCG	GCGACCTGGC	CCTGGACGAG	ATCCTGGAGA	ACCAGCAGCT	GCTGAACGAC	180
ATCAGCGGCA	AGCTGGACGG	CGTGAACGGC	AGCCTGAACG	ACCTGATCGC	CCAGGGCAAC	240
CTGAACACCG	AGCTGAGCAA	GGAGATCCTT	AAGATCGCCA	ACGAGCAGAA	CCAGGTGCTG	300
AACGACGTGA	ACAACAAGCT	GGACGCCATC	AACACCATGC	TGCGCGTGTA	CCTGCCGAAG	360
ATCACCAAGCA	TGCTGAGCGA	CGTGATGAAG	CAGAACTACG	CCCTGAGCCT	GCAGATCGAG	420
TACCTGAGCA	AGCAGCTGCA	GGAGATCAGC	GACAAAGCTGG	ACATCATCAA	CGTGAACGTC	480
CTGATCAACA	GCACCCTGAC	CGAGATCACC	CCGGCCTACC	AGCGCATCAA	GTACGTGAAC	540
GAGAAGTTCTG	AAGAGCTGAC	CTTCGCCACC	GAGACCAGCA	GCAAGGTGAA	GAAGGACGGC	600
AGCCCGGGCCG	ACATCCGGGA	CGAGCTGAGC	GAGCTGACCG	AGCTGGCGAA	GAGCGTGACC	660
CAGAACGACG	TGGACGGCTT	CGAGTTCTAC	CTGAACACCT	TCCACGACGT	GATGGTGGGC	720
AACAACCTGT	TCGGCCGCAG	CGCCCTGAAG	ACCGCCAGCG	AGCTGATCAC	CAAGGAGAAC	780
GTGAAGACCA	GCGGCAGCGA	GGTGGGCAAC	GTGTACAAC	TCCTGATCGT	GCTGACCGCC	840
CTGCAGGCCC	AGGCCTTCCT	GACCCTGACC	CCCTGTCGCA	AGCTGCTGGG	CCTGGCCGAC	900
ATCGACTACA	CCAGCATCAT	GAACGAGCAC	TTGAACAAGG	AGAAGGAGGA	GTTCCGCGTG	960
AACATCCTGC	CGACCCTGAG	CAACACCTTC	AGCAACCCGA	ACTACGCCAA	GGTGAAGGGC	1020
AGCGACGAGG	ACGCCAAGAT	GATCGTGAG	GCTAAGCCGG	GCCACGCGTT	GATCGGCTTC	1080
GAGATCAGCA	ACGACAGCAT	CACCGTGCTG	AAGGTGTACG	AGGCCAAGCT	GAAGCAGAAC	1140
TACCAGGTGG	ACAAGGACAG	CTTGAGCGAG	GTGATCTACG	GCGACATGGA	CAAGCTGCTG	1200
TGTCCGGACC	AGAGCGGGCA	AATCTACTAC	ACCAACAACA	TCGTGTTCCC	GAACGAGTAC	1260
GTGATCACCA	AGATCGACTT	CACCAAGAAG	ATGAAGACCC	TGCGCTACGA	GGTGACCGCC	1320
AACTTCTACG	ACAGCAGCAC	CGGCGAGATC	GACCTGAACA	AGAAGAAGGT	GGAGAGCAGC	1380

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GAGGCCGAGT	ACCGCACCT	GAGCGCGAAC	GACGACGGCG	TCTACATGCC	ACTGGGCGTG	1440
ATCAGCGAGA	CCTTCCTGAC	CCCATCAAC	GGCTTTGGCC	TGCAGGCCGA	CGAGAACAGC	1500
CGCCTGATCA	CCCTGACCTG	TAAGAGCTAC	CTGCGCGAGC	TGCTGCTAGC	CACCGACCTG	1560
AGCAACAAGG	AGACCAAGCT	GATCGTGCCA	CCGAGCGGGT	TCATCAGCAA	CATCGTGAG	1620
AACGGCAGCA	TCGAGGAGGA	CAACCTGGAG	CCGTGGAAGG	CCAACAACAA	GAACGCCTAC	1680
GTGGACCACA	CCGGCGGGCT	GAACGGCACC	AAGGCCCTGT	ACGTGCACAA	GGACGGCGGC	1740
ATCAGCCAGT	TCATCGGCGA	CAAGCTGAAG	CCGAAGACCG	AGTACGTGAT	CCAGTACACC	1800
GTGAAGGGCA	AGCCATCGAT	TCACCTGAAG	GACGAGAACA	CCGGCTACAT	CCACTACGAG	1860
GACACCAACA	ACAACCTGGA	GGACTACCAG	ACCATCAACA	AGCGCTTCAC	CACCGGCACC	1920
GACCTGAAGG	GCGTGTACCT	GATCCTGAAG	AGCCAGAACG	GCGACGAGGC	CTGGGGCGAC	1980
AACTTCATCA	TCCTGGAGAT	CAGCCCGAGC	GAGAAAGCTGC	TGAGCCCGGA	GCTGATCAAC	2040
ACCAACAAC	GGACCAGCAC	CGGCAGCACC	AACATCAGCG	GCAACACCCT	GACCCTGTAC	2100
CAGGGCGGGC	GCGGCATCCT	GAAGCAGAAC	CTGCAGCTGG	ACAGCTTCAG	CACCTACCGC	2160
GTGTACTTCA	GCGTGAGCGG	CGACGCCAAC	GTGCGCATCC	GCAACTCCCG	CGAGGTGCTG	2220
TTCGAGAAGA	AGGACAAGTA	G				2241

What is claimed is:

1. A microorganism comprising a recombinant DNA sequence which encodes a protein of the VIP3 class, wherein said DNA sequence is selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 and SEQ ID NO:5.
2. The microorganism of claim 1 wherein the DNA sequence is SEQ ID NO:1.
3. The microorganism of claim 1 wherein said microorganism is selected from the group consisting of bacteria, baculovirus, algae and fungi.
4. The microorganism of claim 3 wherein said microorganism is selected from the group consisting of Bacillus, Pseudomonas, Clavibacter, and Rhizobium.
5. The microorganism of claim 1 further comprising a second DNA sequence which encodes a second insecticidal protein.
6. The microorganism of claim 5 wherein said second DNA sequence encodes a δ -endotoxin or a vegetative insecticidal protein.
7. The microorganism of claim 6 wherein said second DNA sequence is a δ -endotoxin.
8. An entomocidal composition comprising microorganisms according to claim 1.
9. A recombinant DNA sequence which encodes a protein of the VIP3 class, wherein said DNA sequence is selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 and SEQ ID NO:5.
10. The recombinant DNA sequence of claim 9 wherein said DNA sequence is SEQ ID No:1.
11. The recombinant DNA sequence of claim 9 wherein the DNA sequence is a synthetic sequence which has been designed for optimum expression in a plant.
12. The recombinant DNA sequence of claim 11 wherein the plant is a maize plant.
13. An expression cassette comprising a heterologous promoter operably linked to the DNA sequence of claim 9.
14. The expression cassette of claim 13 wherein said promoter functions in plants is selected from the group consisting of inducible, constitutive, tissue-preferred and tissue-specific promoters.
15. The expression cassette of claim 13 wherein the promoter is selected from the group consisting of the ubiquitin, PEP carboxylase, LPT and MTL promoters.
16. A recombinant DNA sequence which encodes a protein of the VIP3 class wherein said sequence has a complement which hybridizes to a coding sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 and SEQ ID NO:5 under hybridization conditions of 68° C. followed by washing at 68° C. in 2xSSC containing 0.1% SDS.
17. The recombinant DNA sequence of claim 16 wherein said DNA sequence has a complement which hybridizes to SEQ ID NO:1 under hybridization conditions of 68° C. followed by washing at 68° C. in 2xSSC containing 0.1% SDS.
18. The recombinant DNA sequence of claim 16 wherein said DNA sequence has a complement which hybridizes to SEQ ID NO:3 under hybridization conditions of 68° C. followed by washing at 68° C. in 2xSSC containing 0.1% SDS.
19. The recombinant DNA sequence of claim 16 wherein said DNA sequence has a complement which hybridizes to SEQ ID NO:5 under hybridization conditions of 68° C. followed by washing at 68° C. in 2xSSC containing 0.1% SDS.
20. The recombinant DNA sequence of claim 16 wherein the DNA sequence is a synthetic sequence which has been designed for optimum expression in a plant.
21. The recombinant DNA sequence of claim 20 wherein the plant is a maize plant.
22. An expression cassette comprising a heterologous promoter operably linked to the DNA sequence of claim 16.
23. The expression cassette of claim 22 wherein said promoter functions in plants is selected from the group consisting of inducible, constitutive, tissue-preferred and tissue-specific promoters.
24. The expression cassette of claim 22 wherein the promoter is selected from the group consisting of the ubiquitin, PEP carboxylase, LPT and MTL promoters.
25. A microorganism comprising a recombinant DNA sequence which encodes a protein of the VIP3 class wherein

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said sequence has a complement which hybridizes to a coding sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 and SEQ ID NO:5 under hybridization conditions, of 68° C. followed by washing at 68° C. in 2×SSC containing 0.1% SDS.

26. The microorganism of claim **25** wherein said microorganism is selected from the group consisting of bacteria, baculovirus, algae and fungi.

27. The microorganism of claim **26** wherein said microorganism is selected from the group consisting of Bacillus, Pseudomonas, Clavibacter, and Rhizobium.

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28. The microorganism of claim **25** further comprising a second DNA sequence which encodes a second insecticidal protein.

29. The microorganism of claim **28** wherein said second DNA sequence encodes a δ -endotoxin or a vegetative insecticidal protein.

30. The microorganism of claim **29** wherein said second DNA sequence is a δ -endotoxin.

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