



US011896675B2

(12) **United States Patent**
Chung et al.

(10) **Patent No.:** **US 11,896,675 B2**

(45) **Date of Patent:** **Feb. 13, 2024**

(54) **SITE-SPECIFIC ANTIBODY CONJUGATION
AND ANTIBODY-DRUG CONJUGATE AS
SPECIFIC EMBODIMENT THEREOF**

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Young Geun Lee, Gyeonggi-do (KR);
Tae Jin Lee, Gyeonggi-do (KR); **Jin
Woo Seo**, Gyeonggi-do (KR)

(73) Assignee: **AbTis Co., Ltd.**, Gyeonggi-do (KR)

(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 0 days.

(21) Appl. No.: **18/154,998**

(22) Filed: **Jan. 16, 2023**

(65) **Prior Publication Data**

US 2023/0248842 A1 Aug. 10, 2023

Related U.S. Application Data

(63) Continuation of application No. 16/980,503, filed on
Sep. 14, 2020.

(60) Provisional application No. 62/815,557, filed on Mar.
8, 2019.

(51) **Int. Cl.**
A61P 35/00 (2006.01)
C07D 207/46 (2006.01)
C07K 16/32 (2006.01)
A61K 47/68 (2017.01)

(52) **U.S. Cl.**
CPC **A61K 47/6803** (2017.08); **A61K 47/6855**
(2017.08); **A61K 47/6889** (2017.08); **A61P**
35/00 (2018.01); **C07D 207/46** (2013.01);
C07K 16/32 (2013.01)

(58) **Field of Classification Search**
CPC A61K 47/6803; A61K 47/6855; A61K
47/6889; A61P 35/00; C07D 207/46;
C07K 16/32
USPC 424/133.1
See application file for complete search history.

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Primary Examiner — Yan Xiao

(74) *Attorney, Agent, or Firm* — Harness, Dickey &
Pierce, P.L.C.

(57) **ABSTRACT**

The present invention relates to technology capable of
labeling a certain site of an antibody with a certain number
of chemical functional groups or cargo moieties. The present
invention may provide an antibody product having high
uniformity. The present invention may provide an antibody
product whose antibody functions are not degraded. That is,
the present invention may provide an antibody product
whose antibody binding affinity and half-life are not
degraded. The present invention is of great significance as
being the first technology allowing site-specific labeling of
an antibody without any complicated processes.

13 Claims, 73 Drawing Sheets

Specification includes a Sequence Listing.

Exhibit 1001
AJINOMOTO CO., INC.
IPR2025-00283

amino acid sequence :

G—P—S—V—F—L—F—P—P—K—P—K—D—T—L—M—I

EU numbering
for heavy chain :

237	239	241	243	245	247	249	251	253
238	240	242	244	246	248	250	252	

FIG. 1

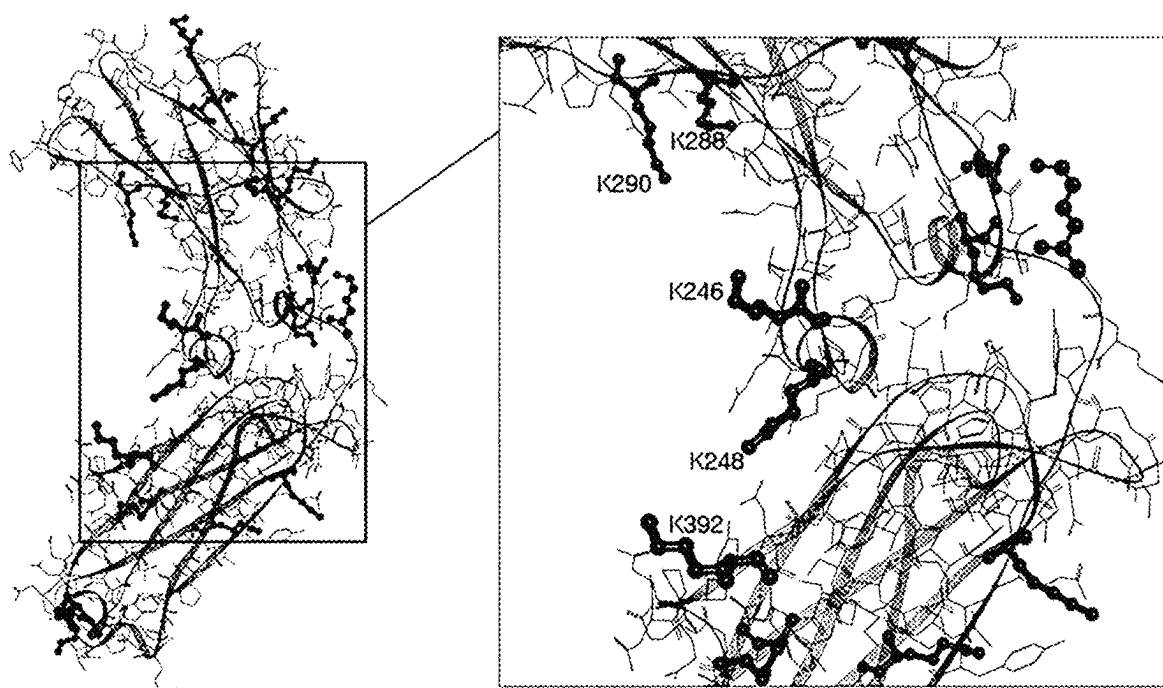


FIG. 2

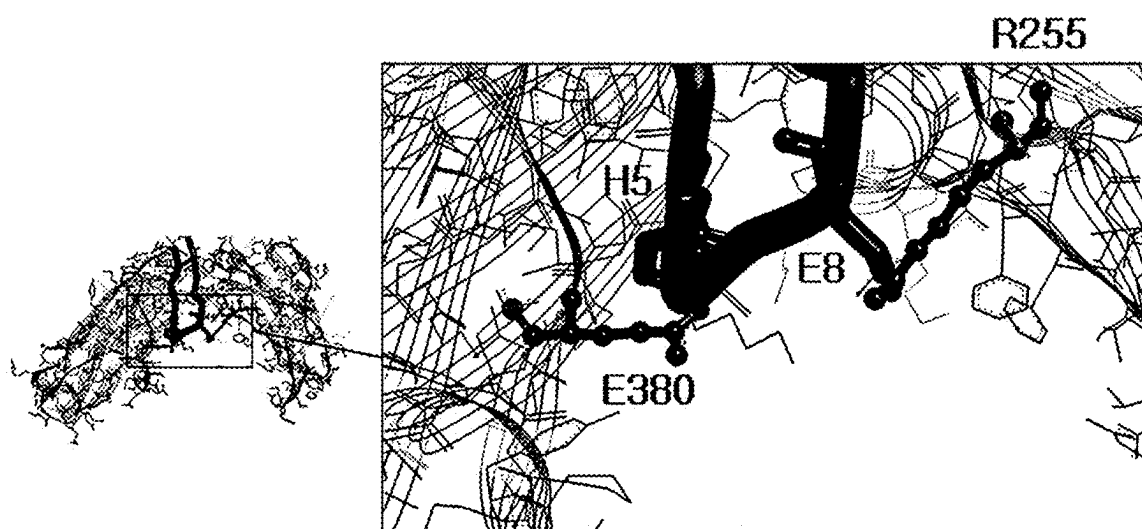


FIG. 3

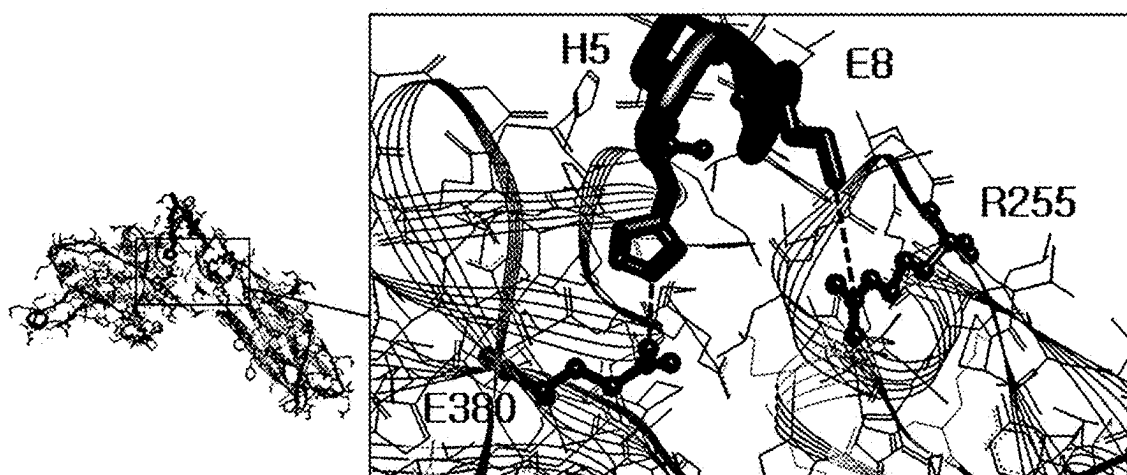


FIG. 4

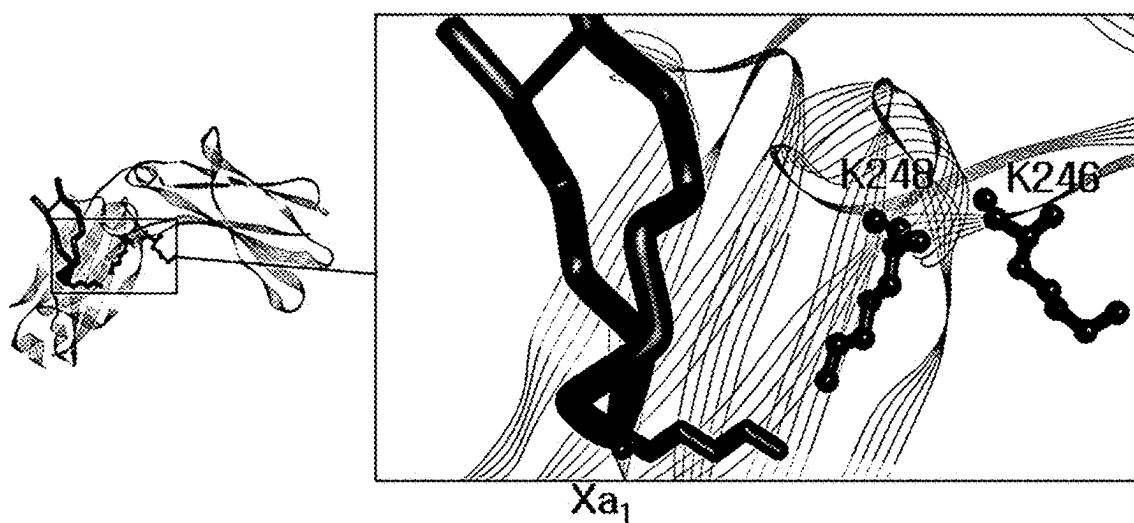


FIG. 5

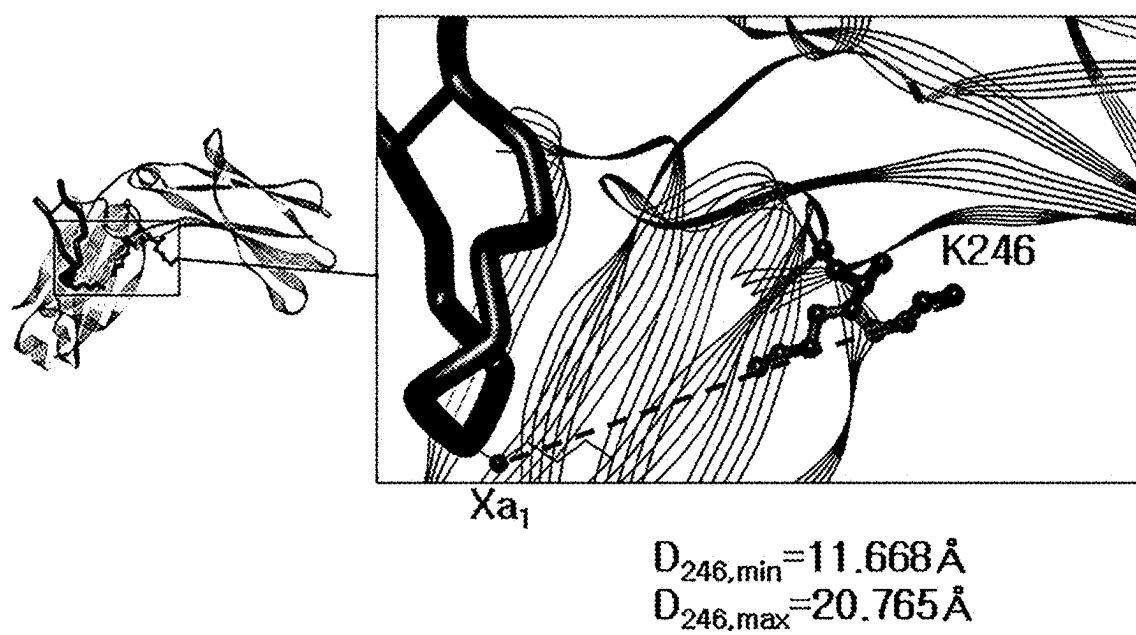


FIG. 6

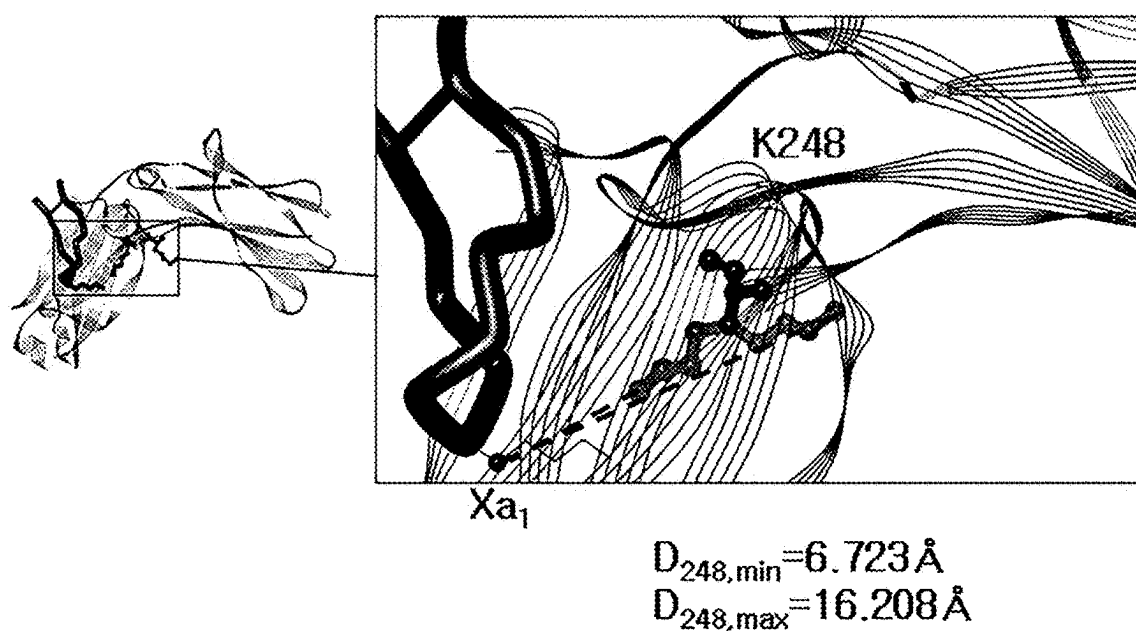


FIG. 7

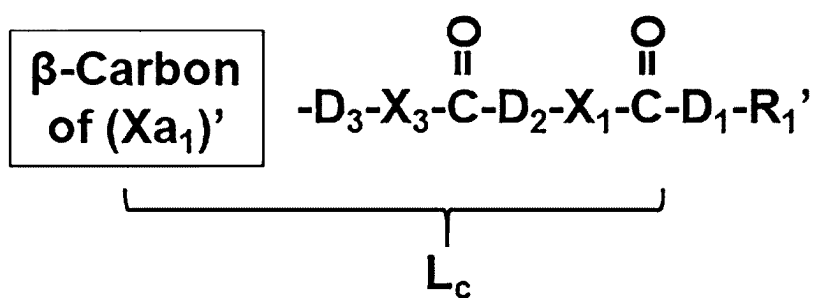
$R_1'-L2-SSAI$ 

FIG. 8

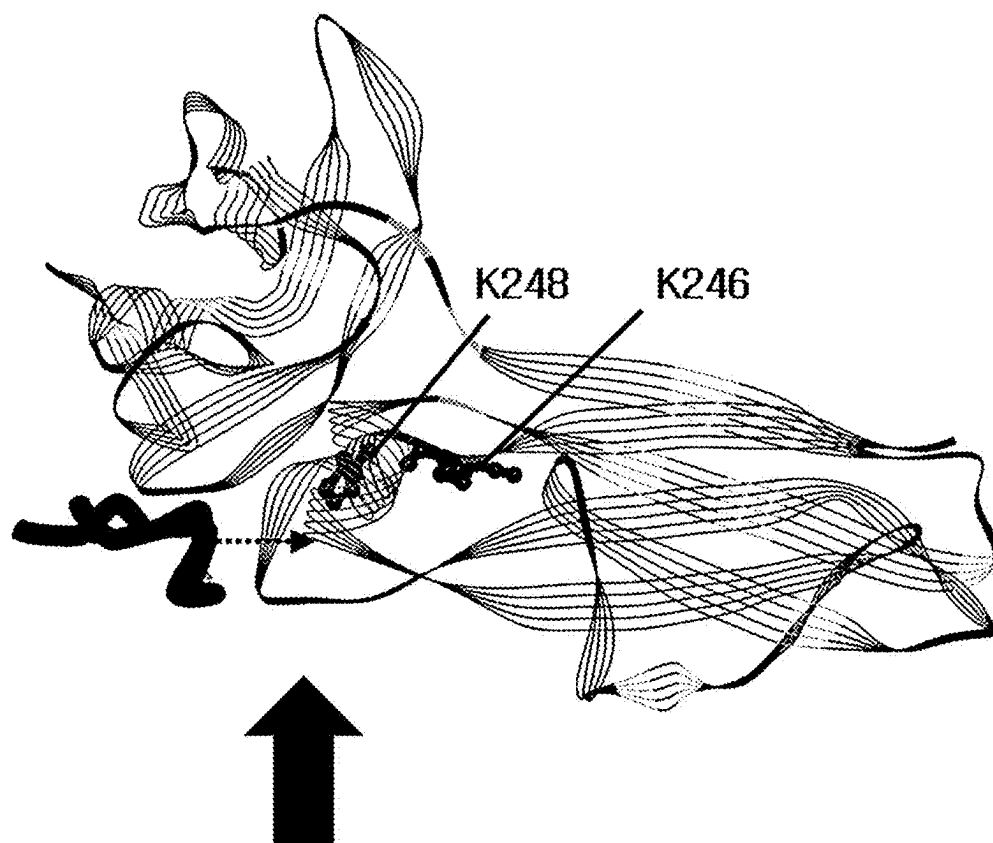


FIG. 9

If L_c is shorter than $D_{246, \min}$



FIG. 10

If L_c is longer than $D_{248, \max}$



FIG. 11

If L_c is between about $D_{246,\min}$ and about $D_{248,\max}$



FIG. 12

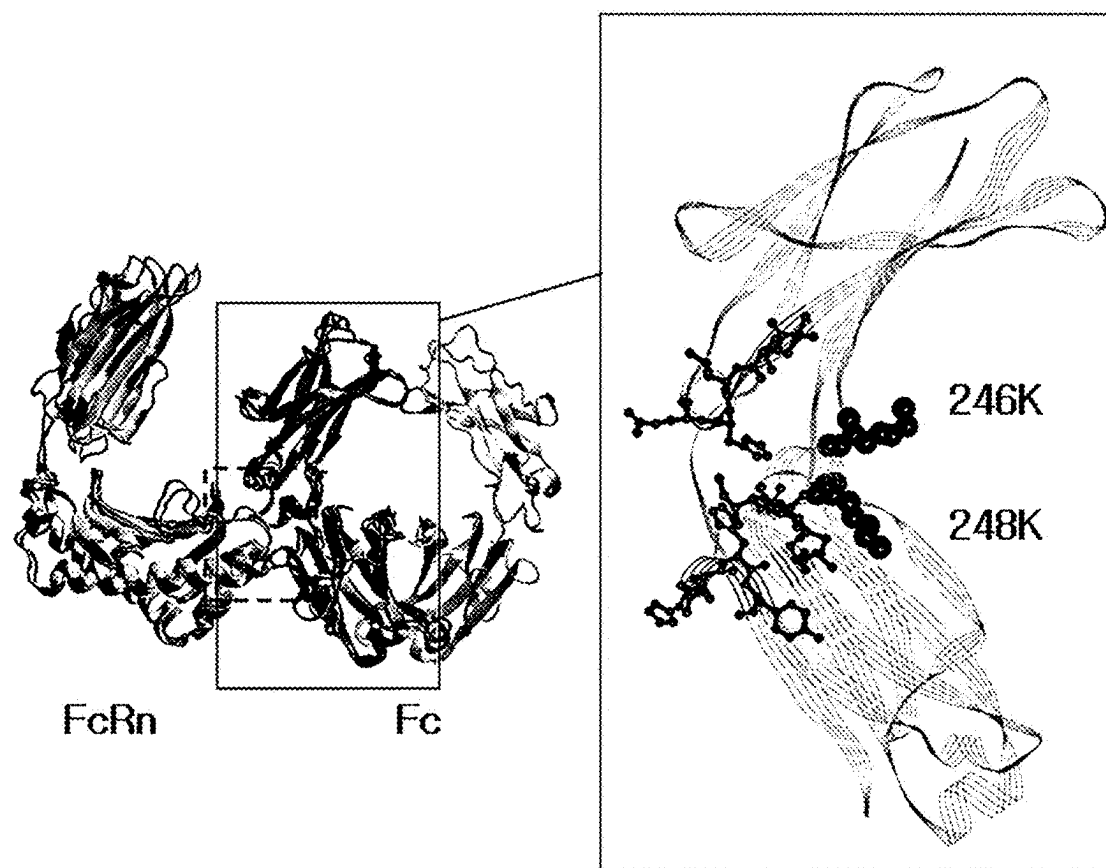


FIG. 13

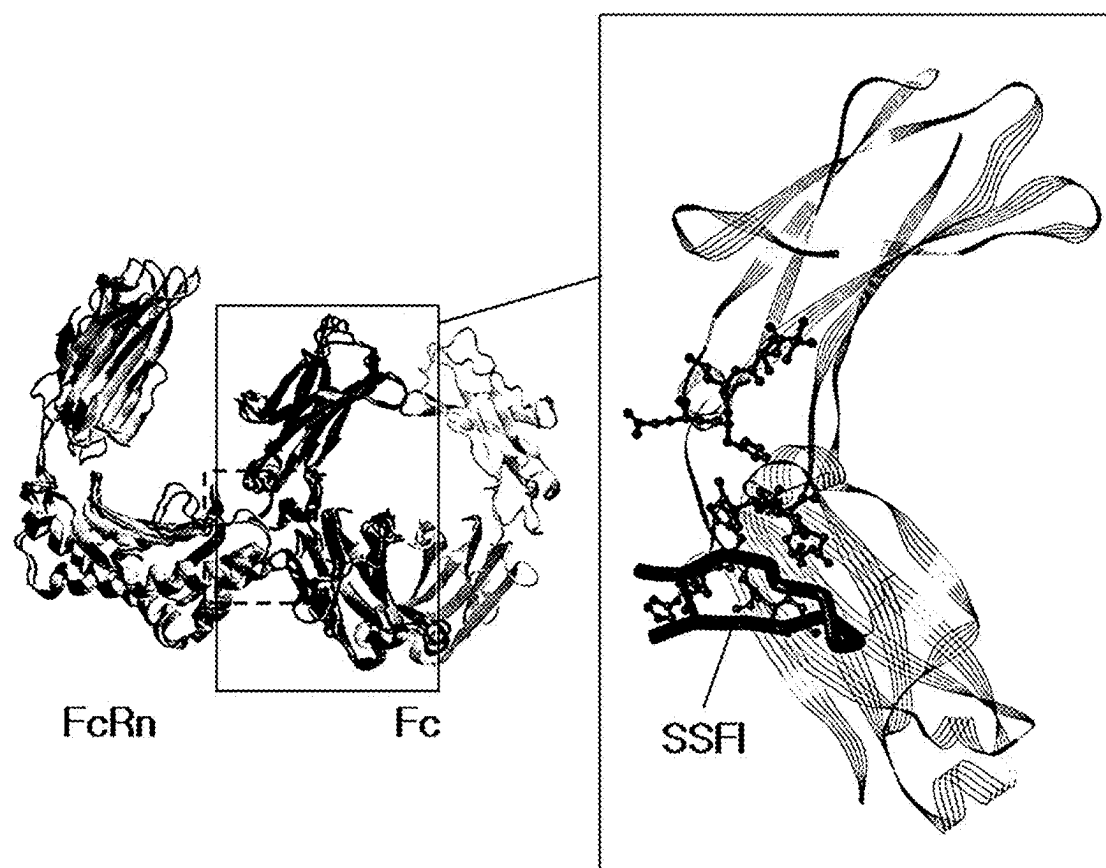


FIG. 14

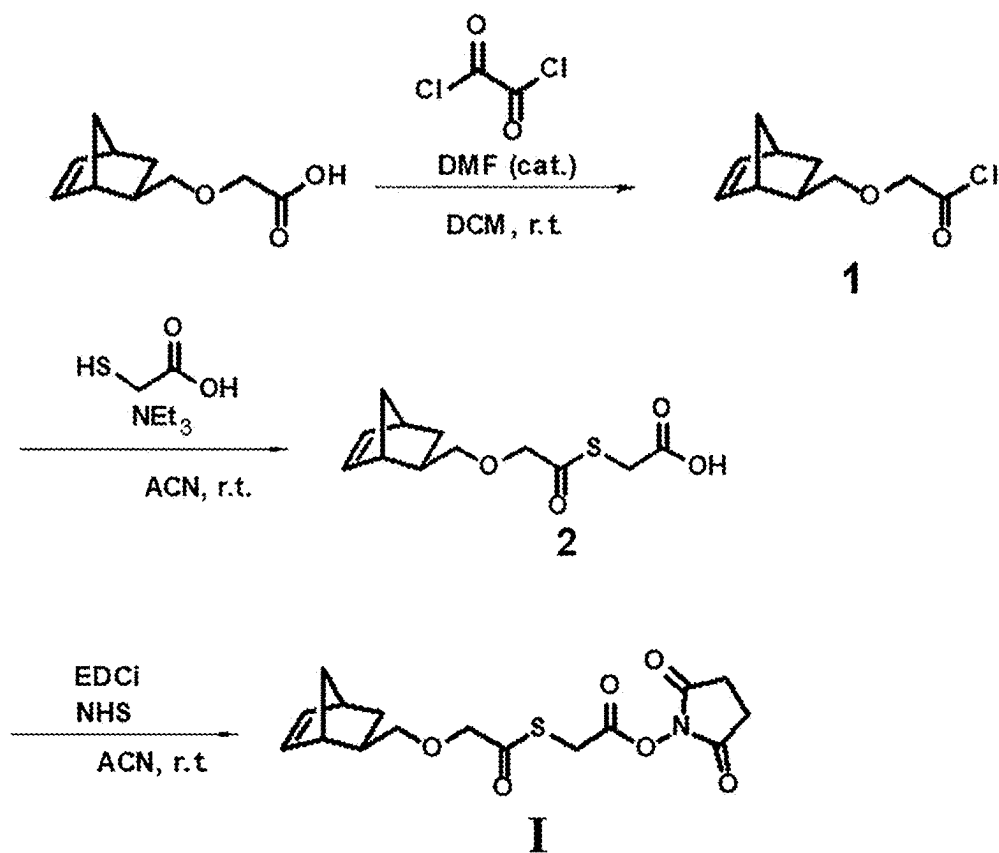


FIG. 15

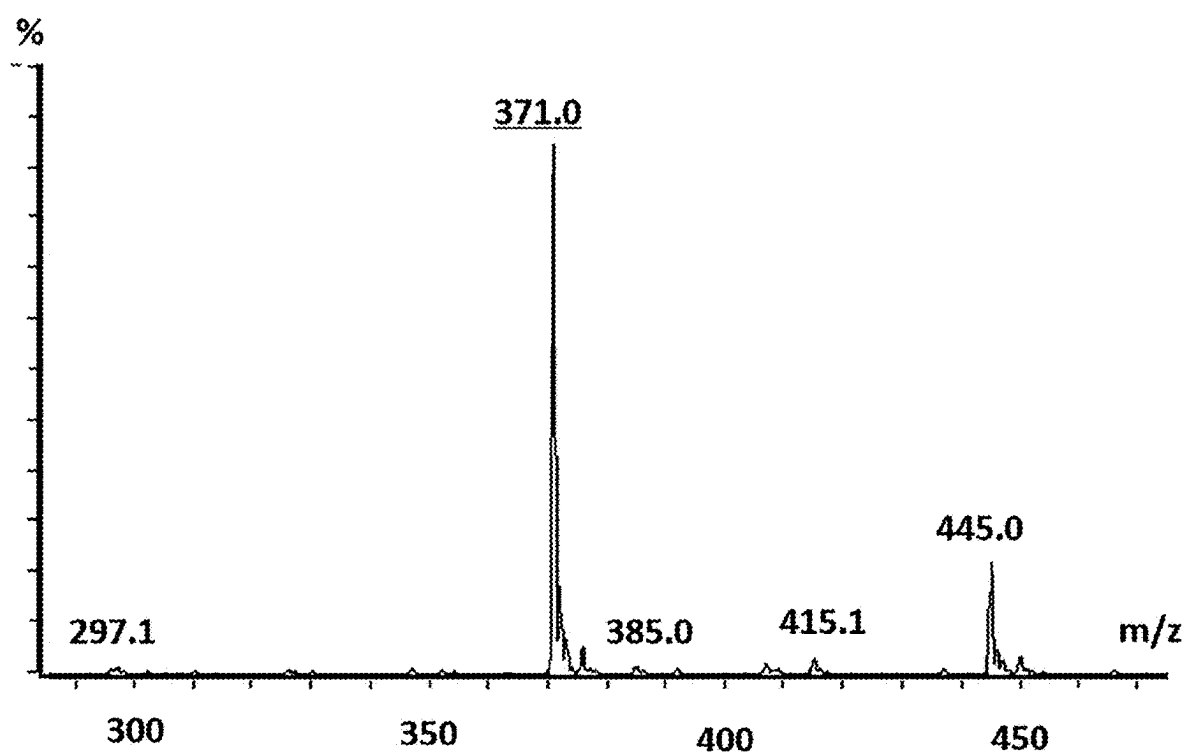


FIG. 16

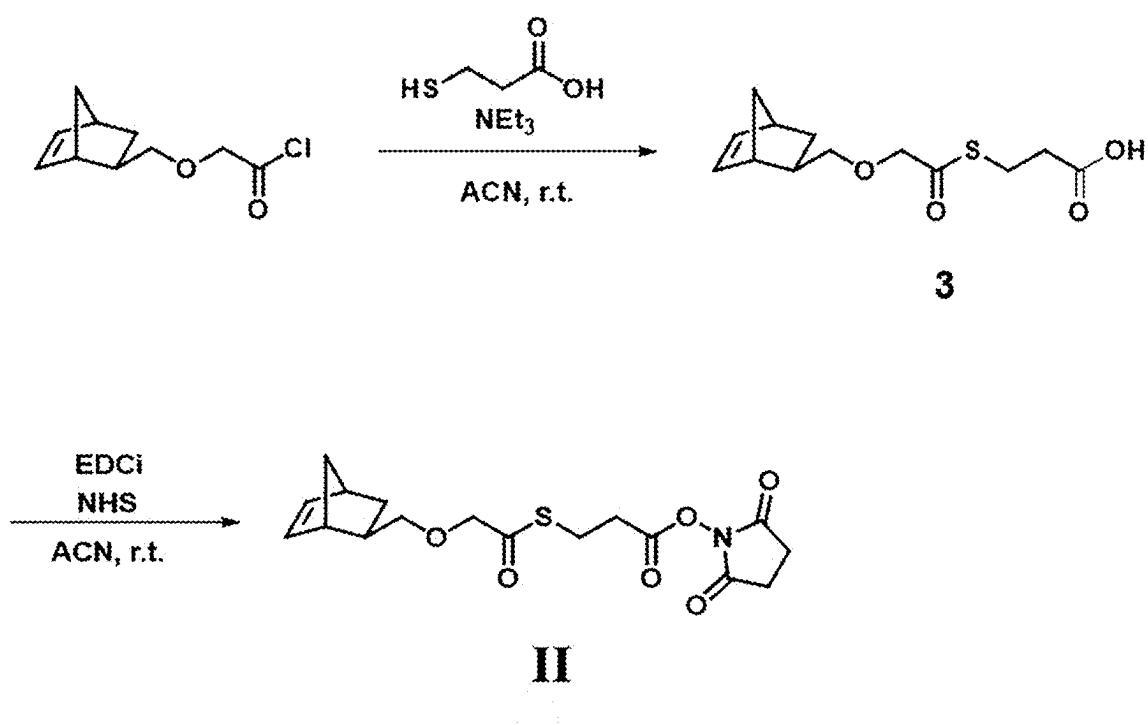


FIG. 17

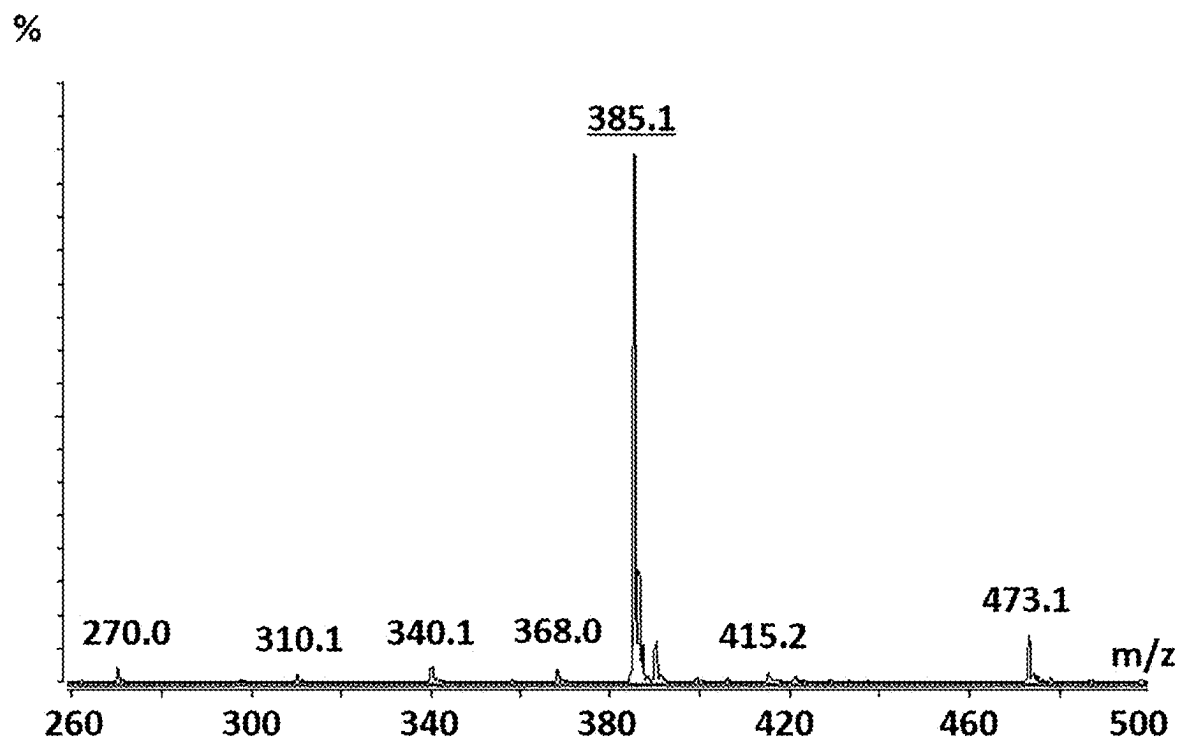


FIG. 18

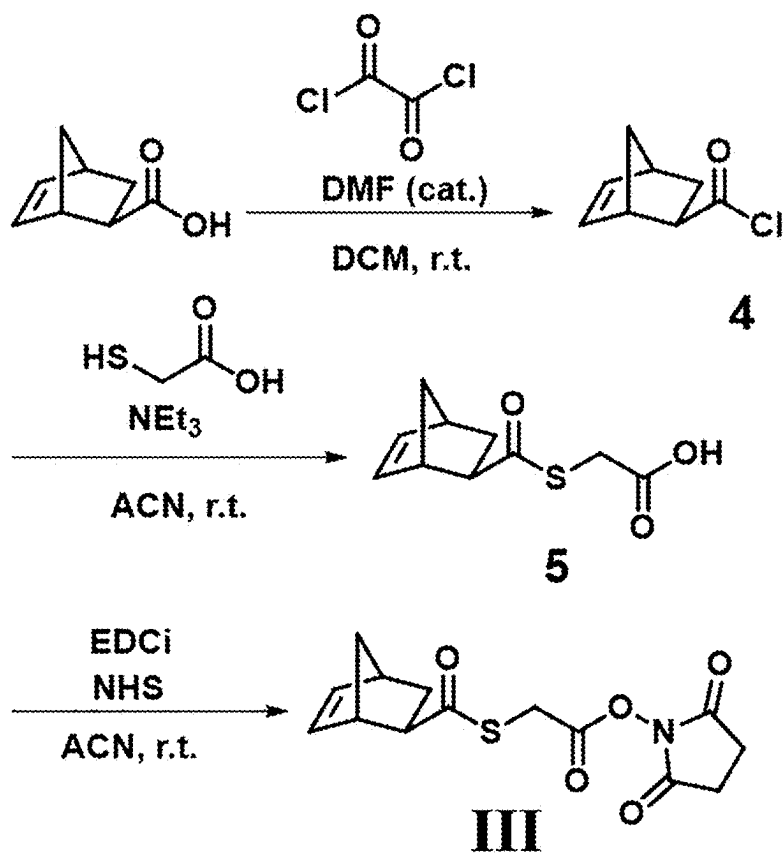


FIG. 19

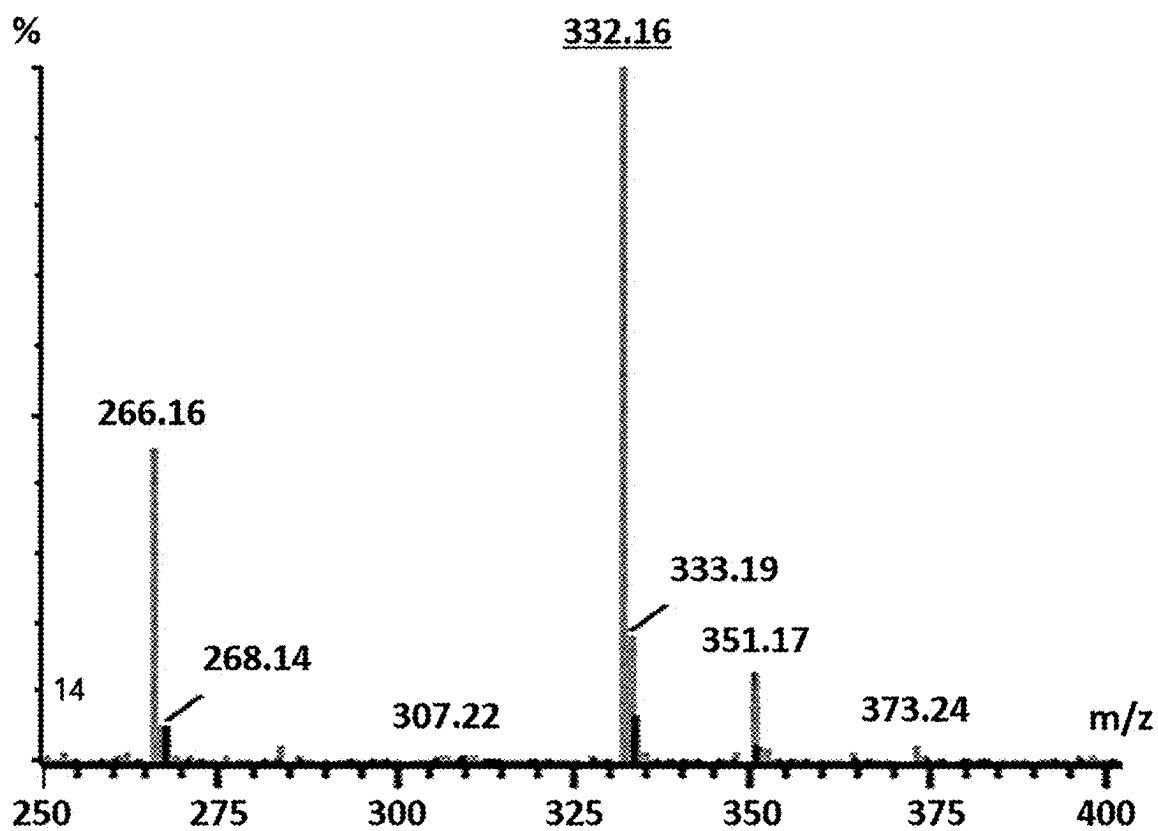


FIG. 20

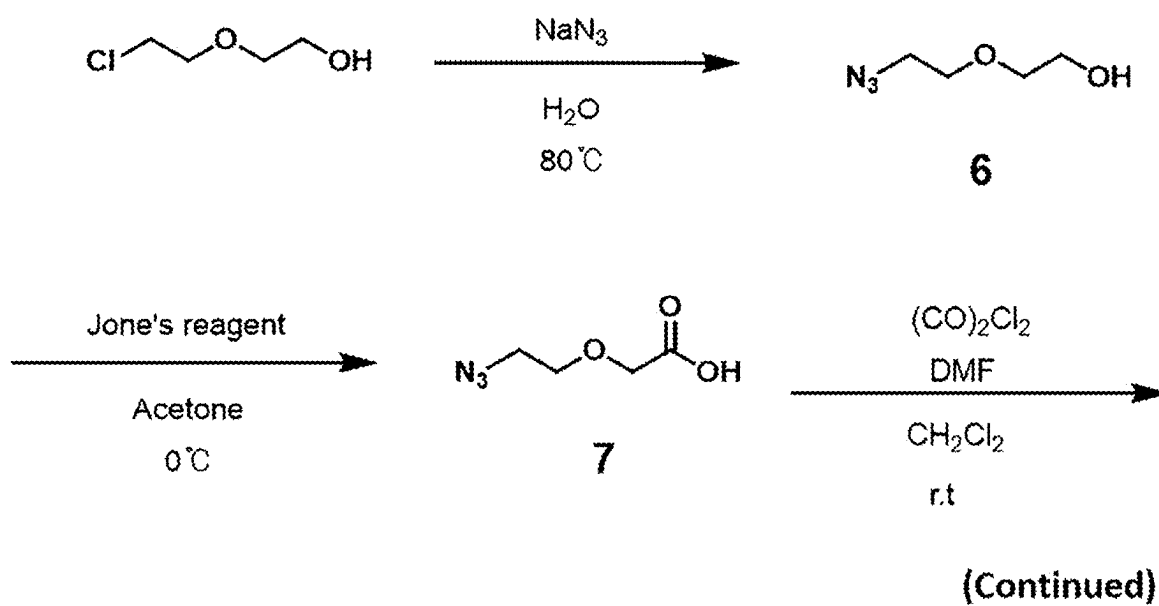


FIG. 21

(Continued)

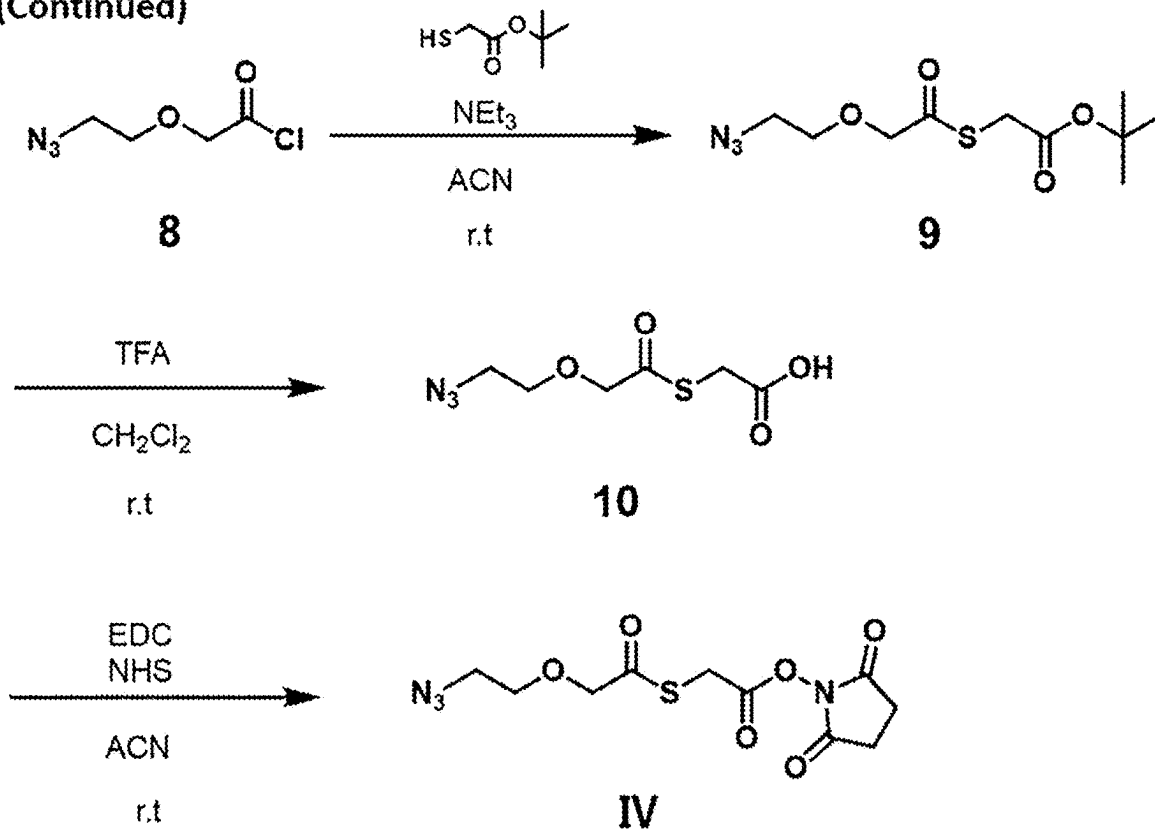


FIG. 22

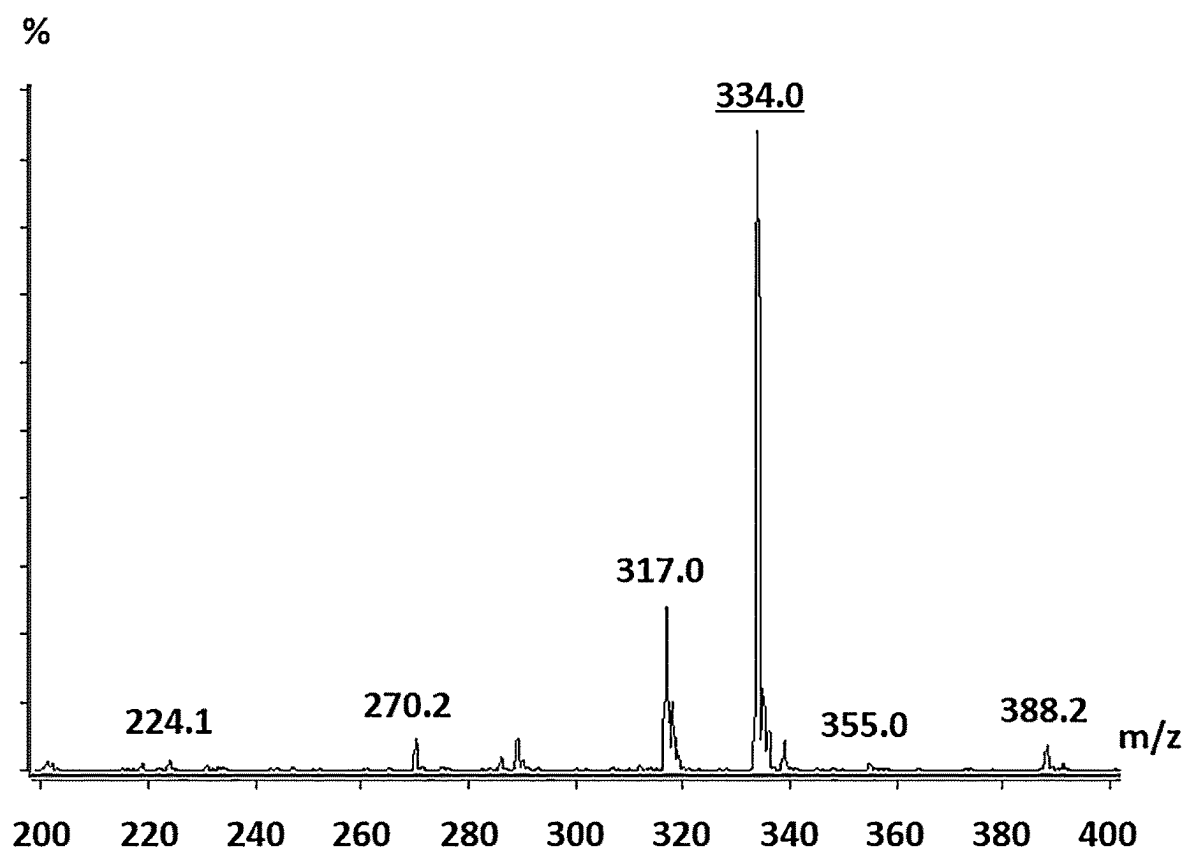


FIG. 23

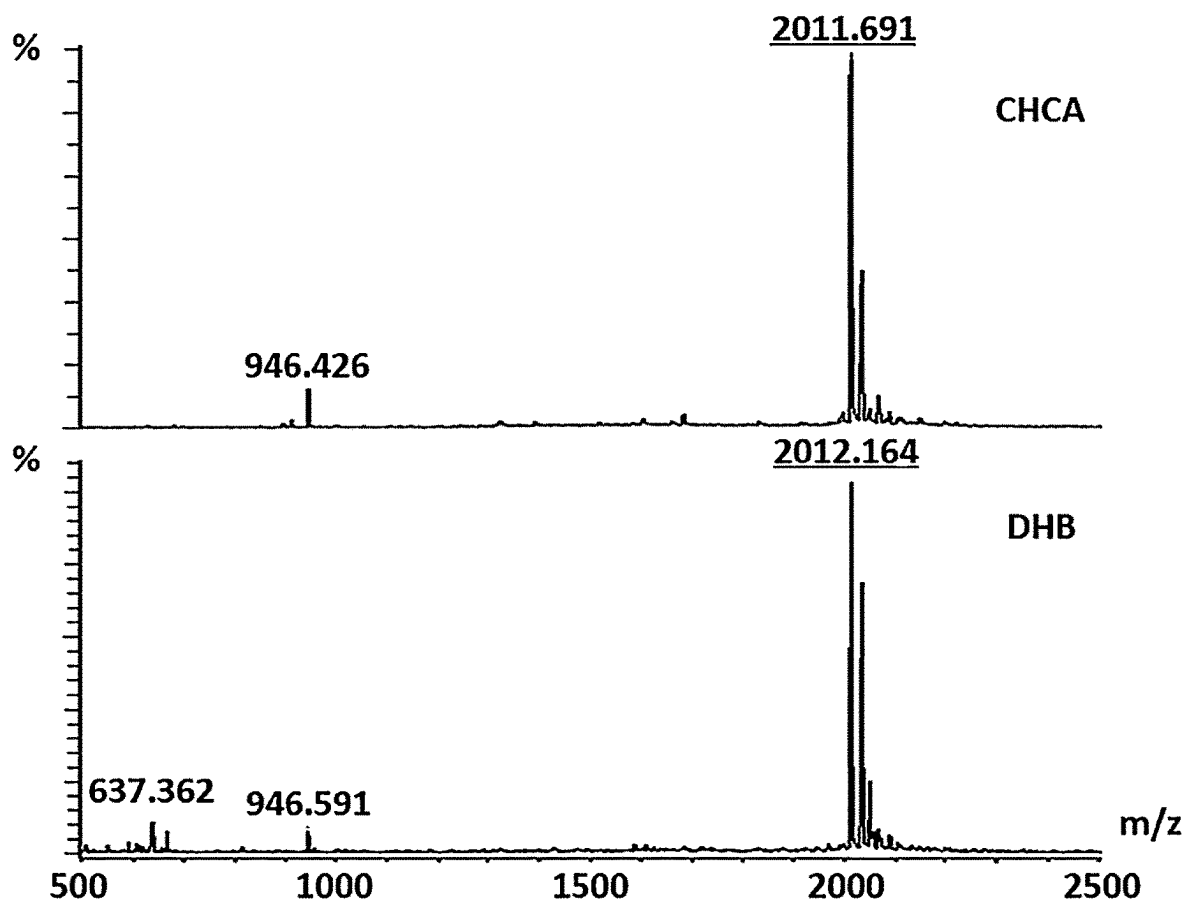


FIG. 24

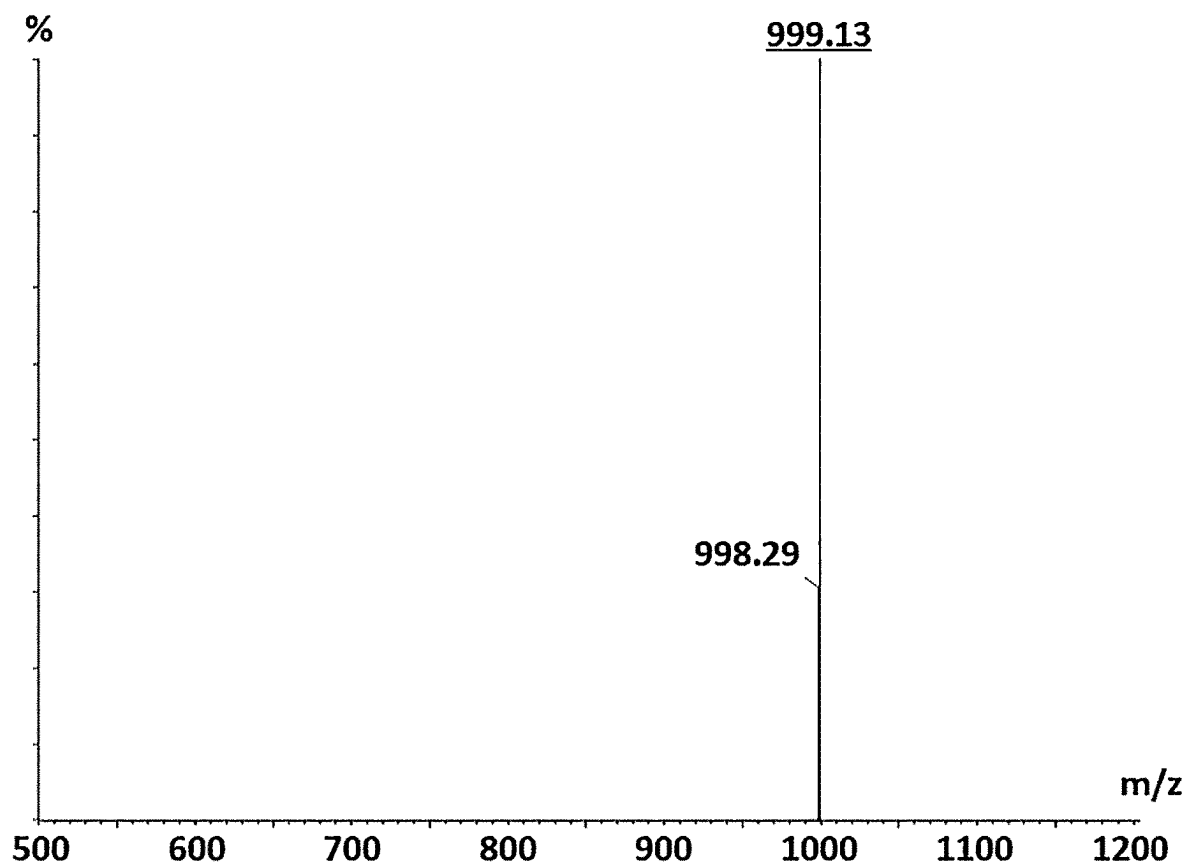


FIG. 25

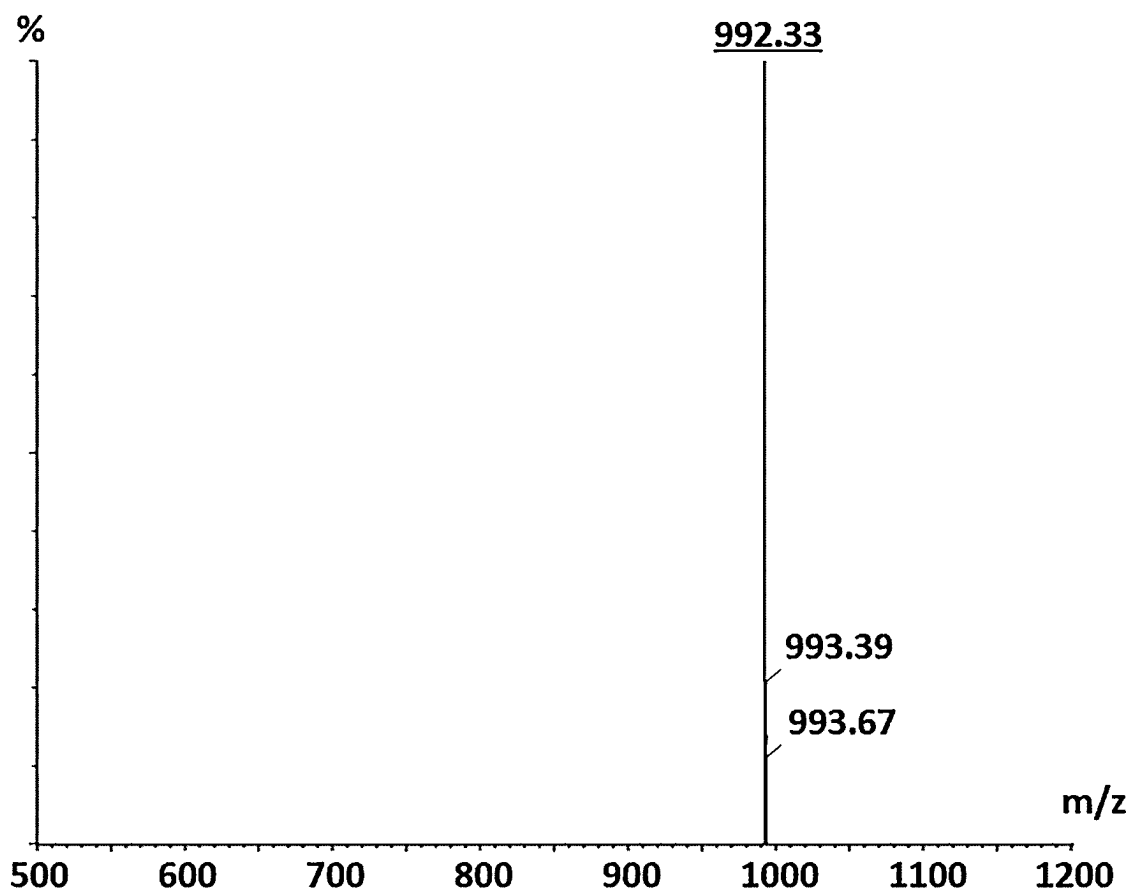


FIG. 26

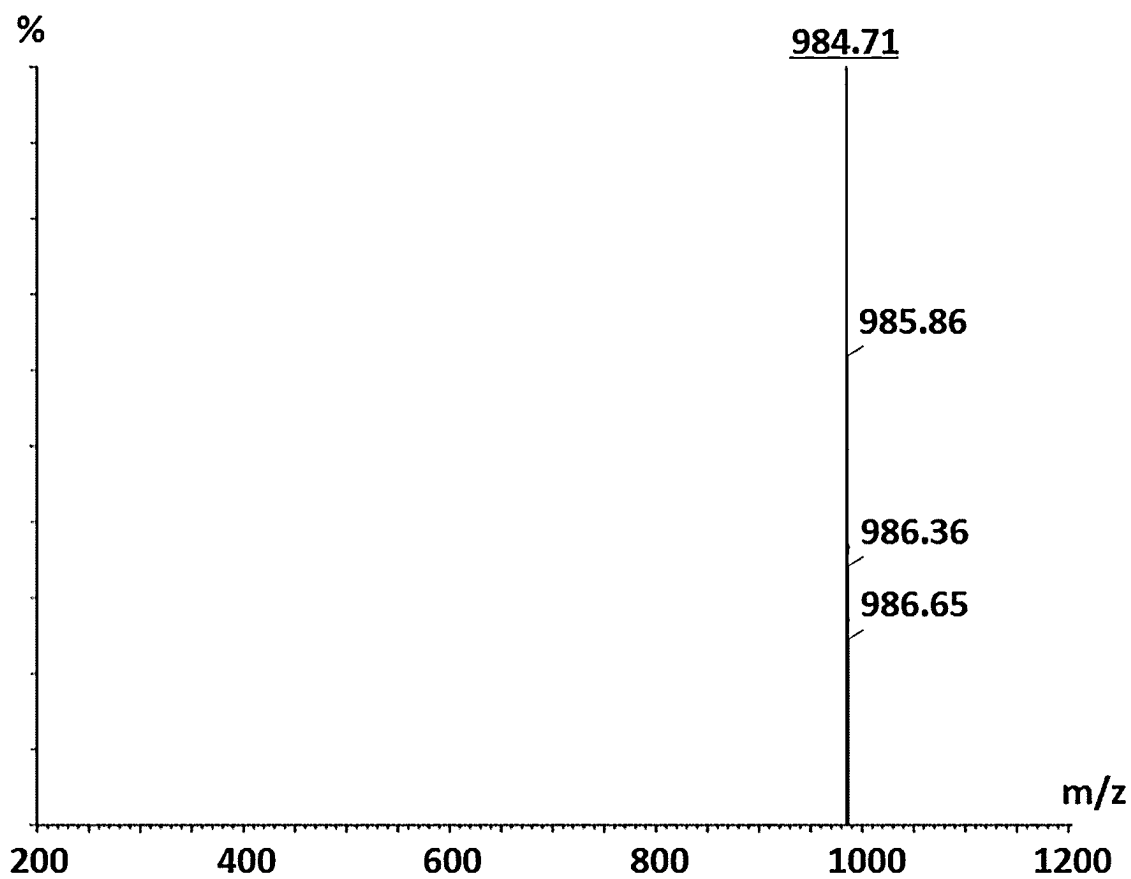


FIG. 27

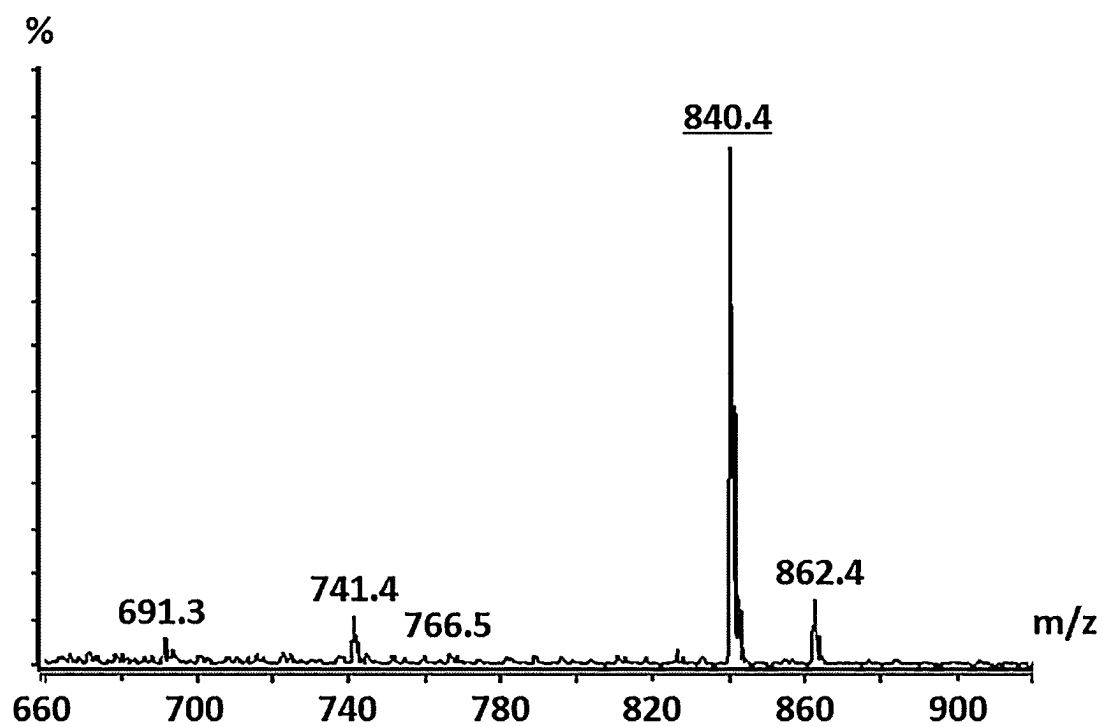


FIG. 28

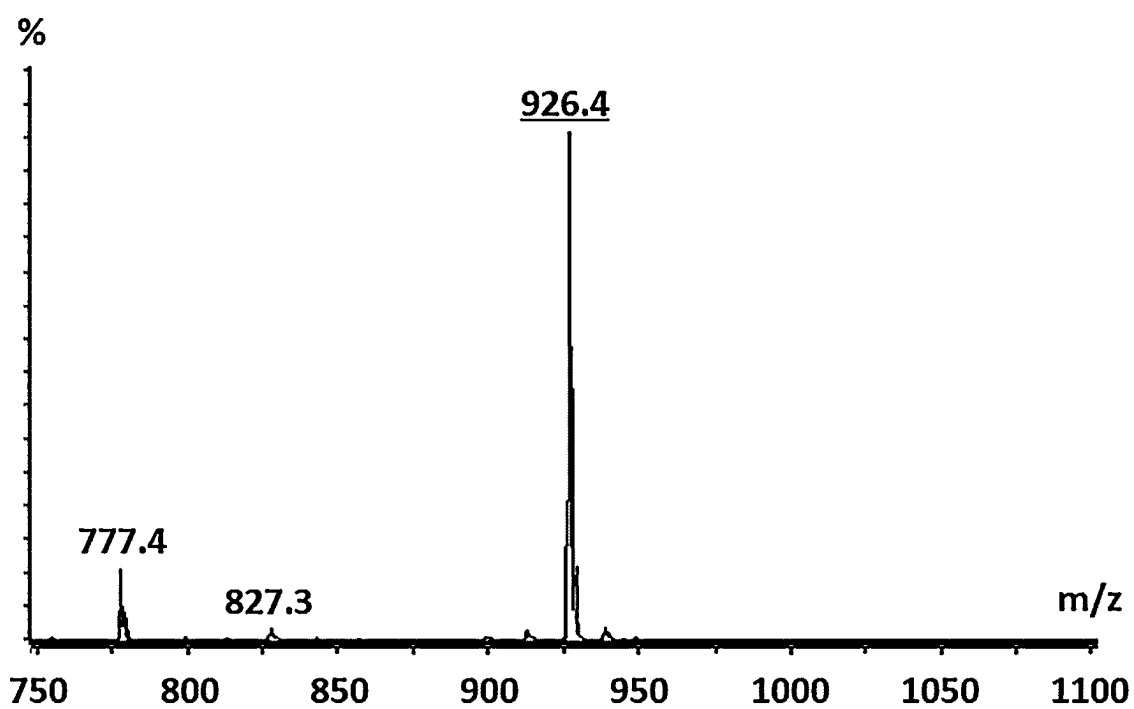


FIG. 29

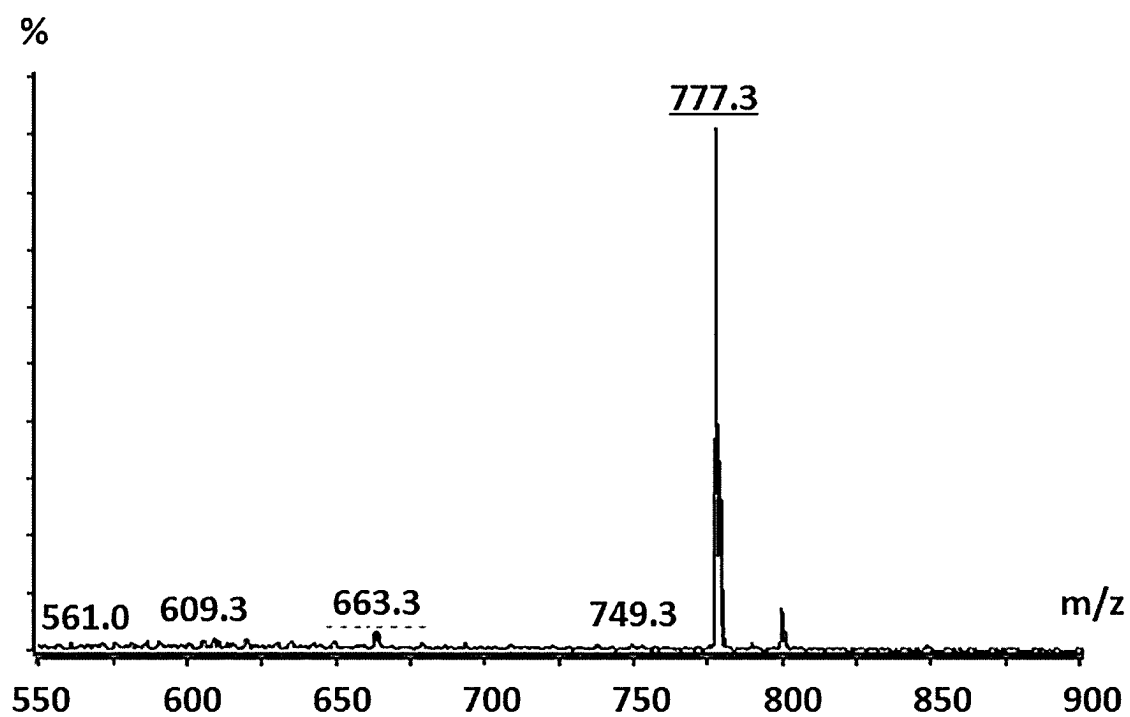


FIG. 30

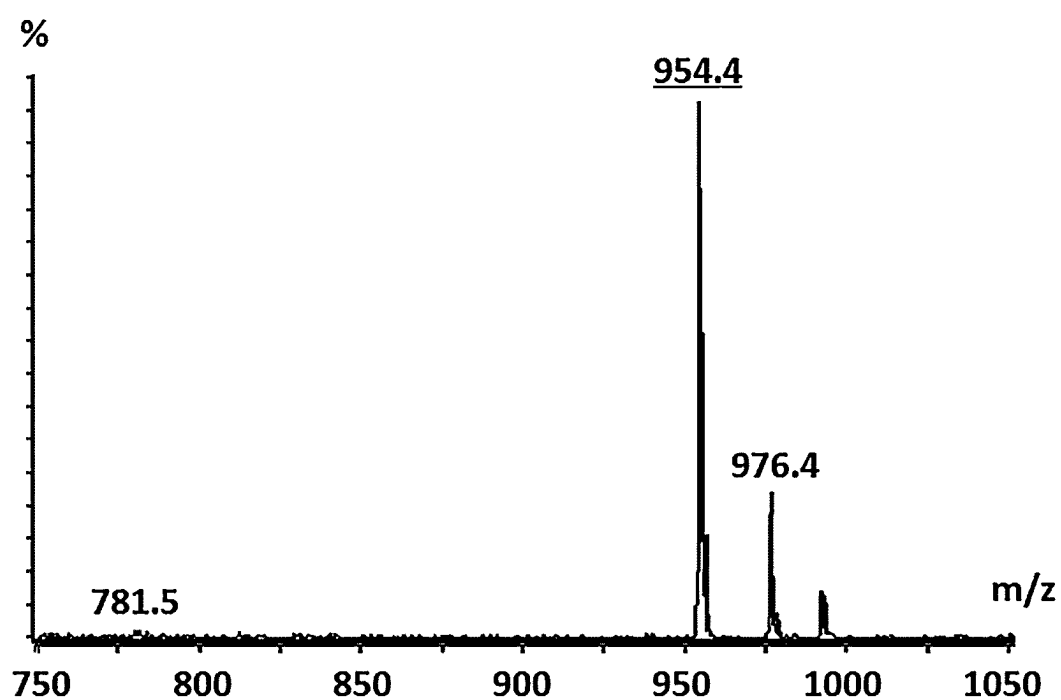


FIG. 31

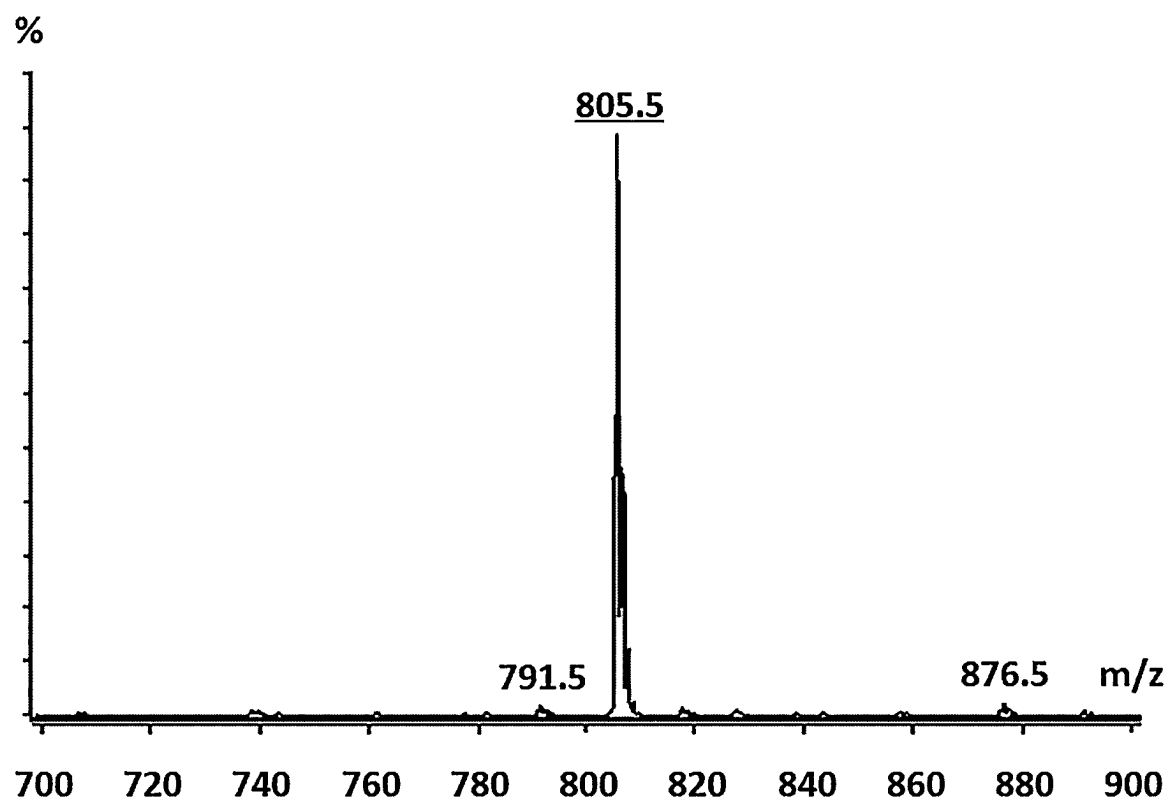


FIG. 32

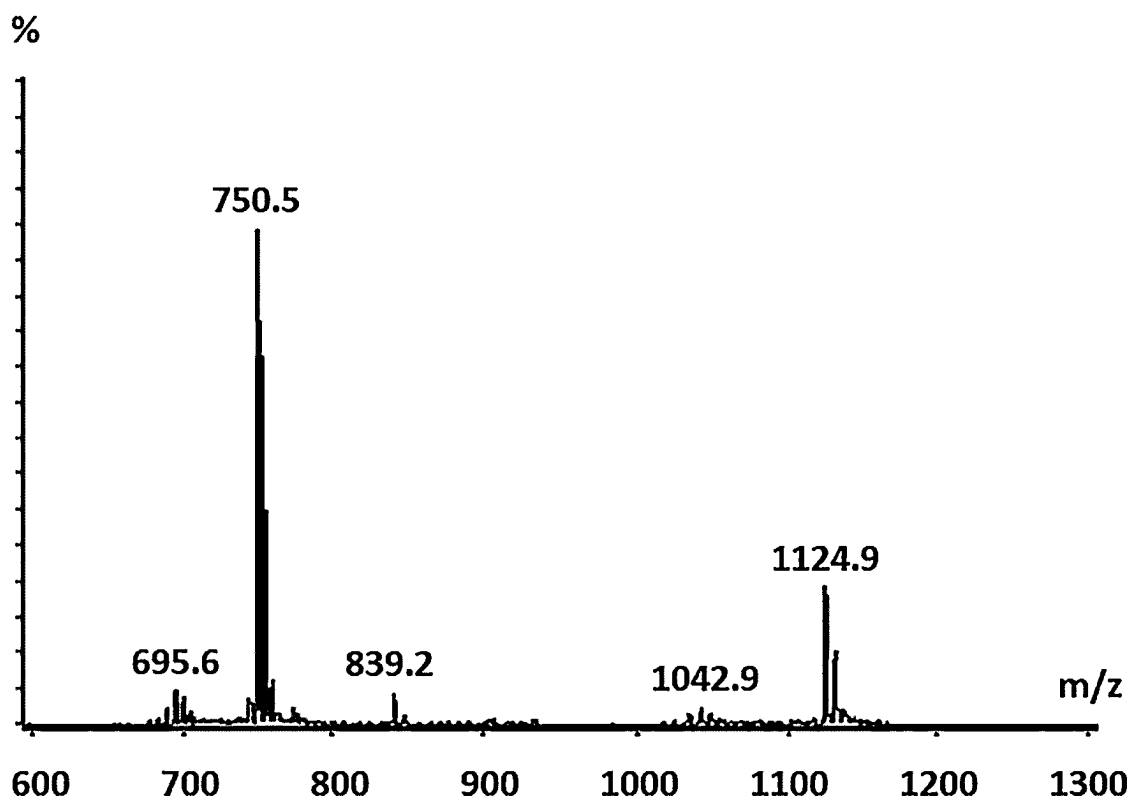


FIG. 33

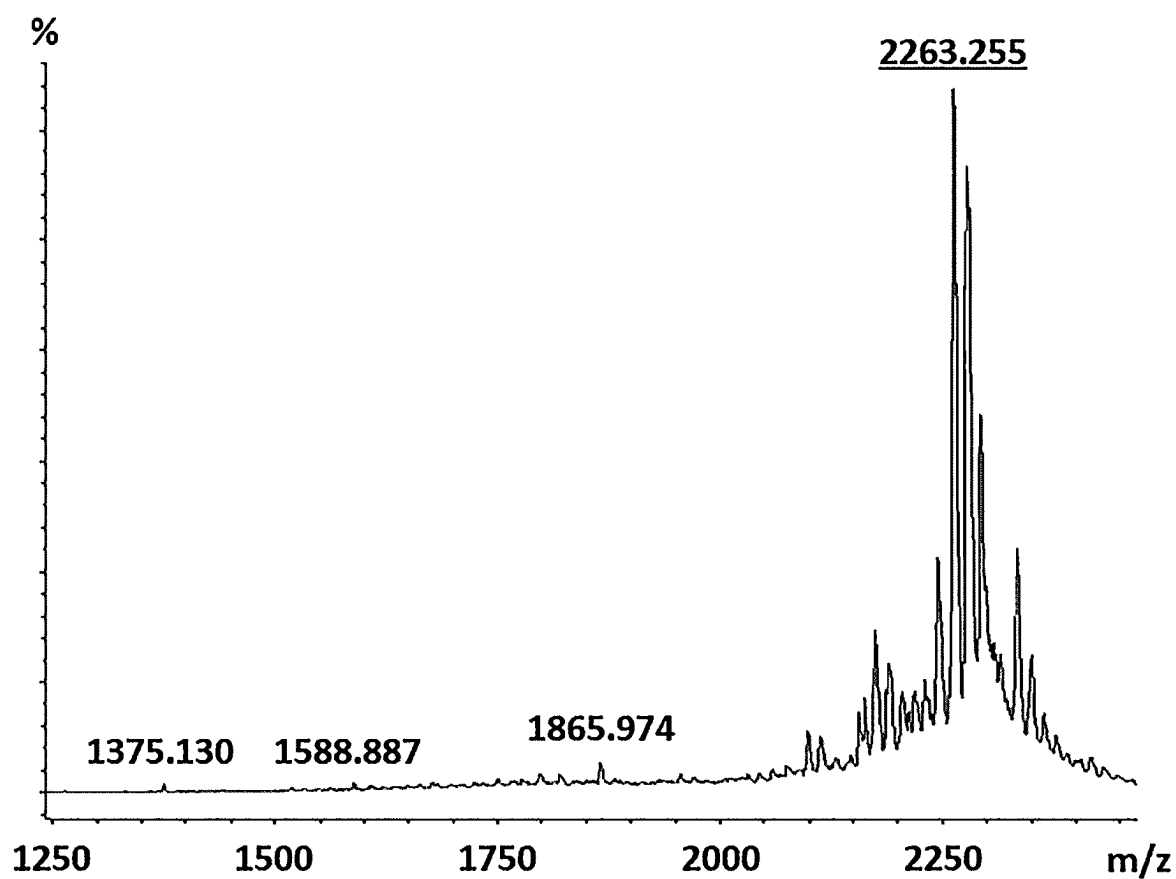


FIG. 34

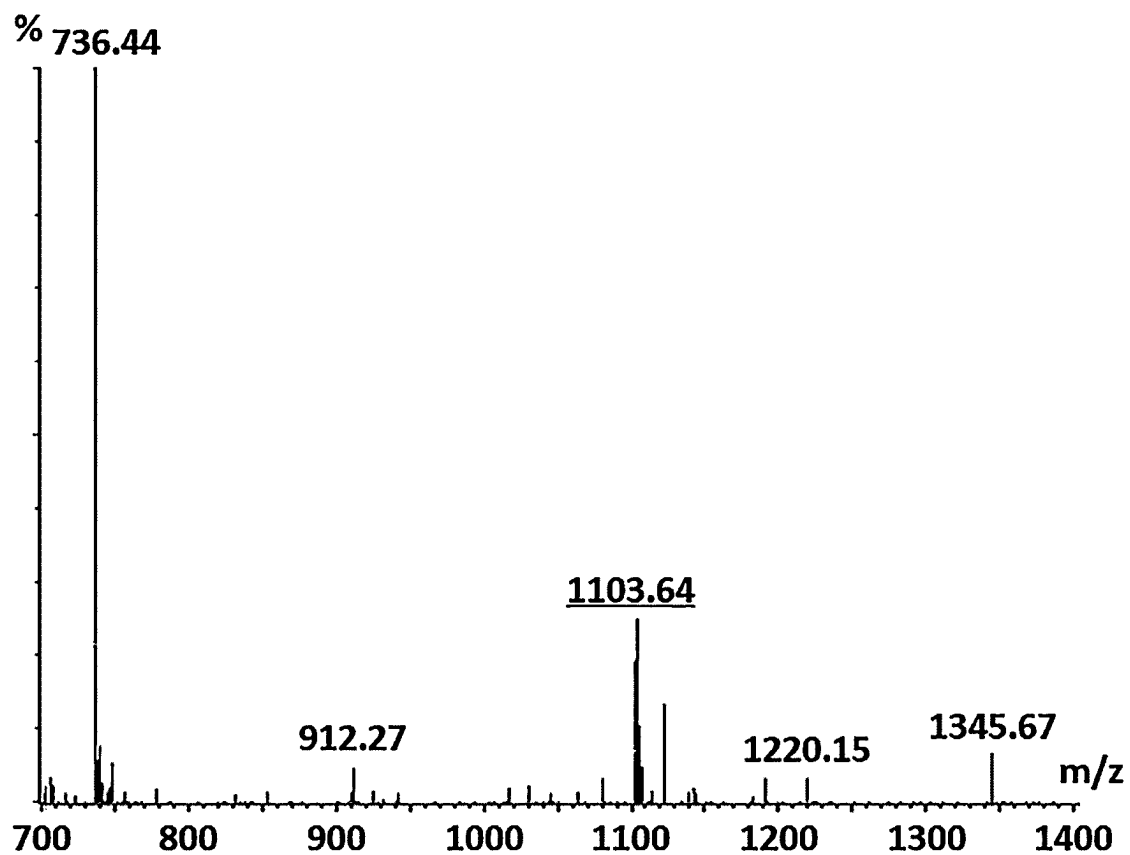


FIG. 35

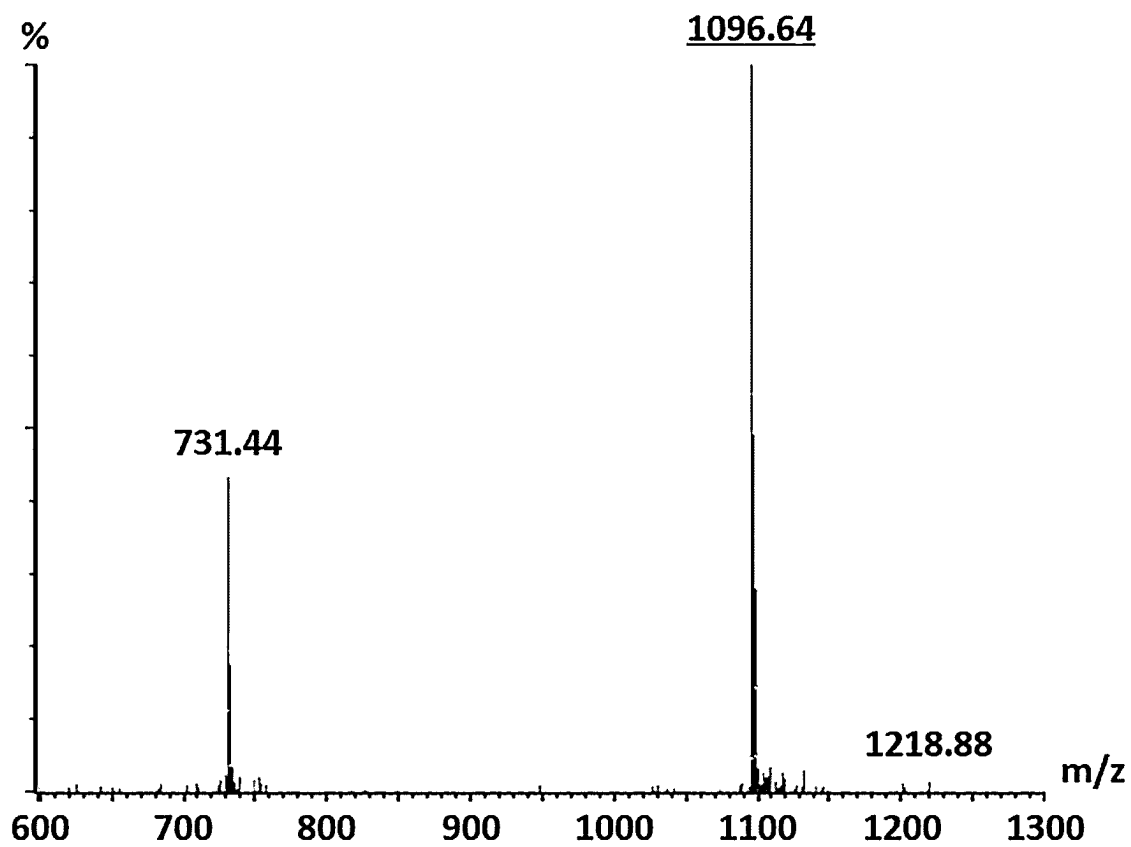


FIG. 36

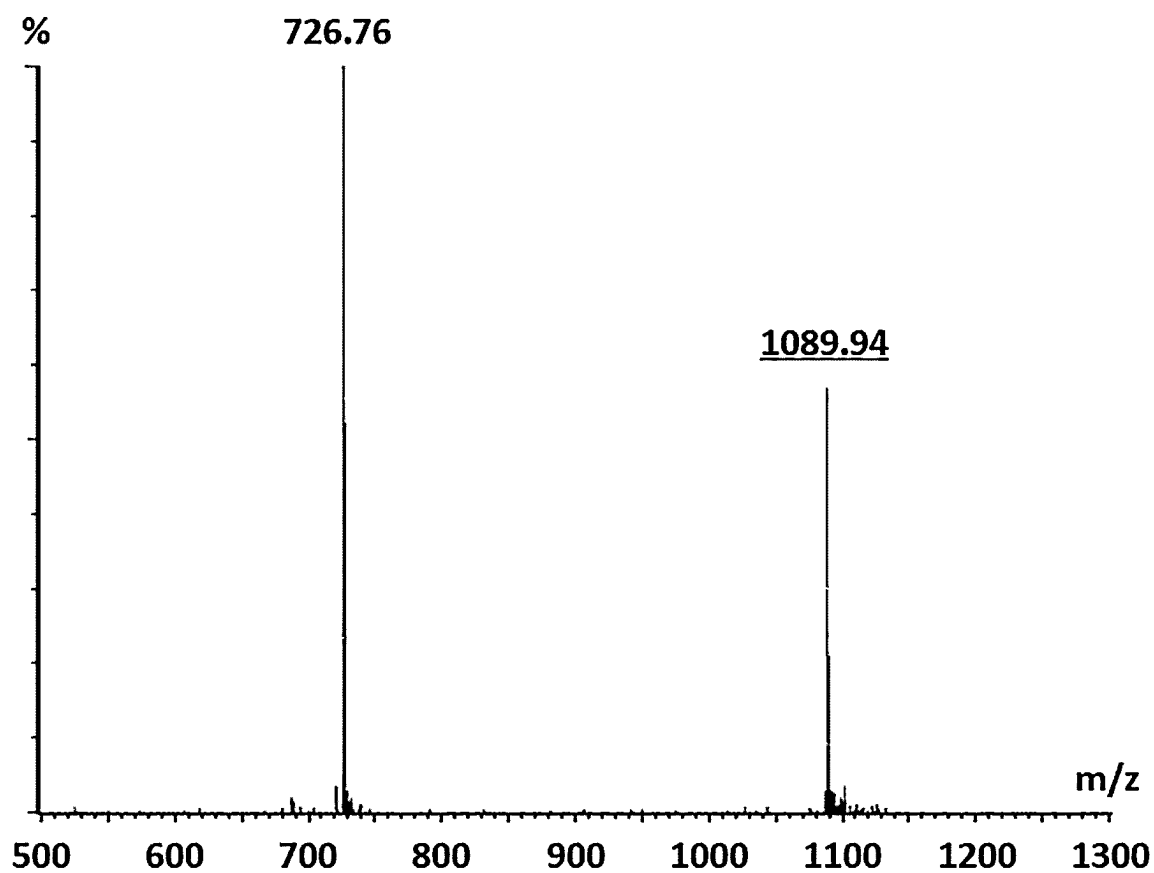


FIG. 37

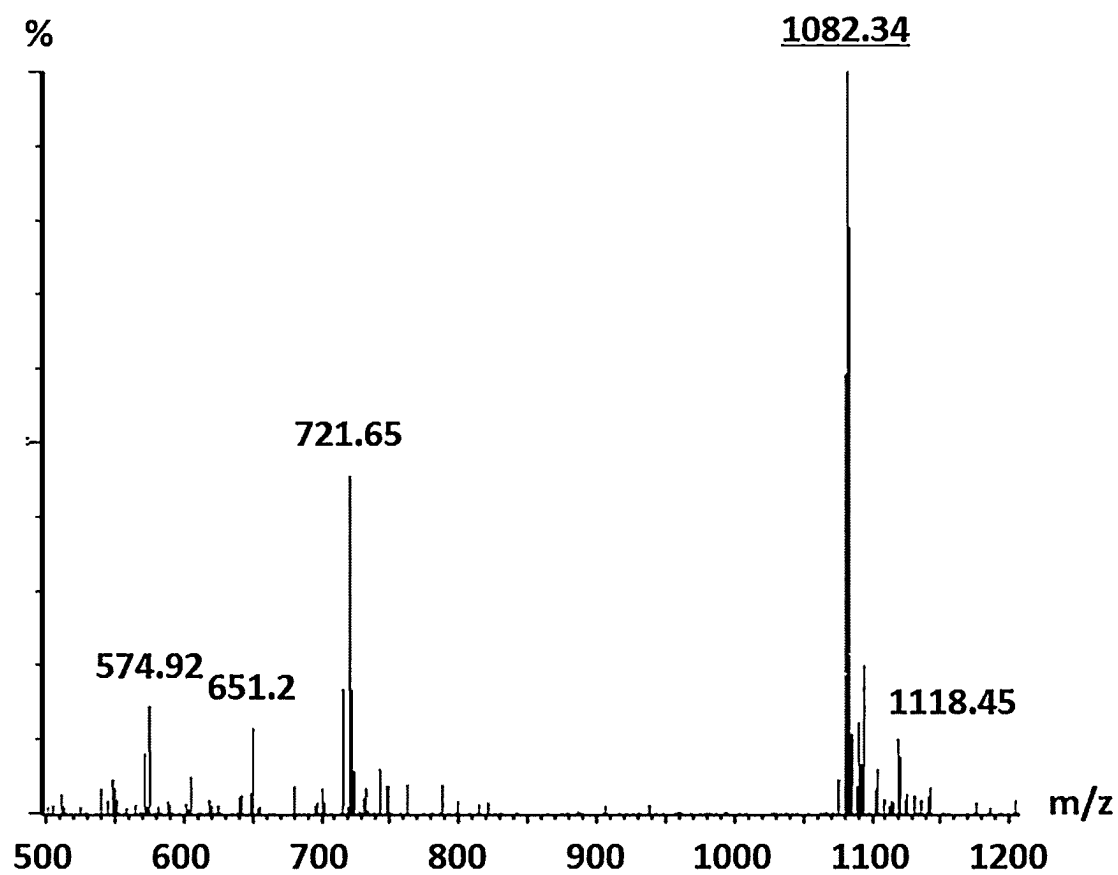


FIG. 38

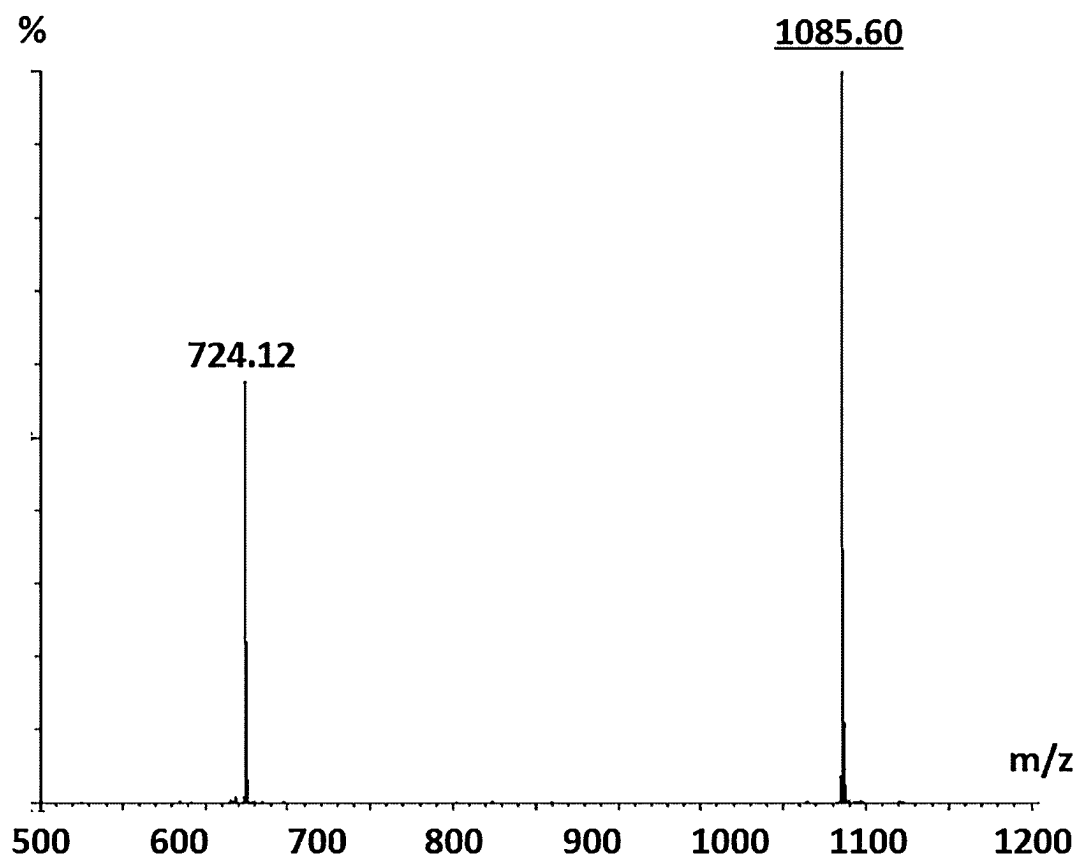


FIG. 39

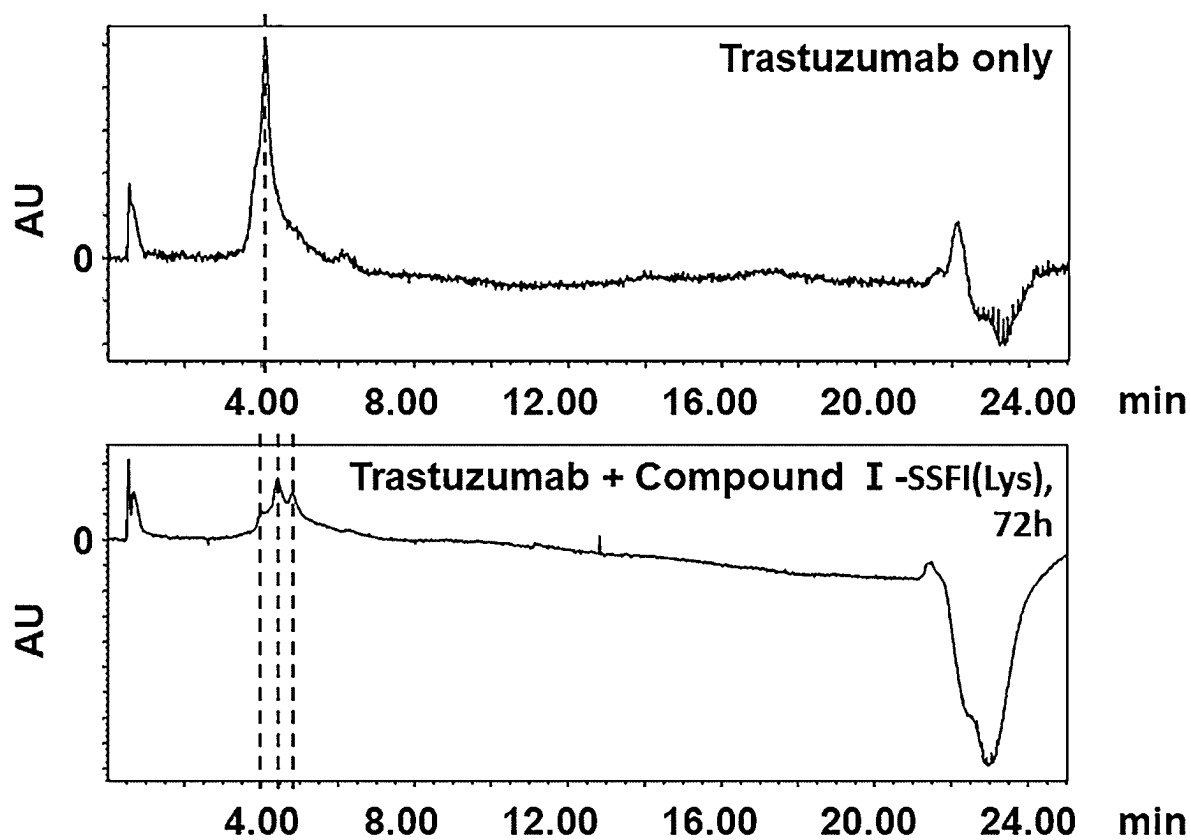


FIG. 40

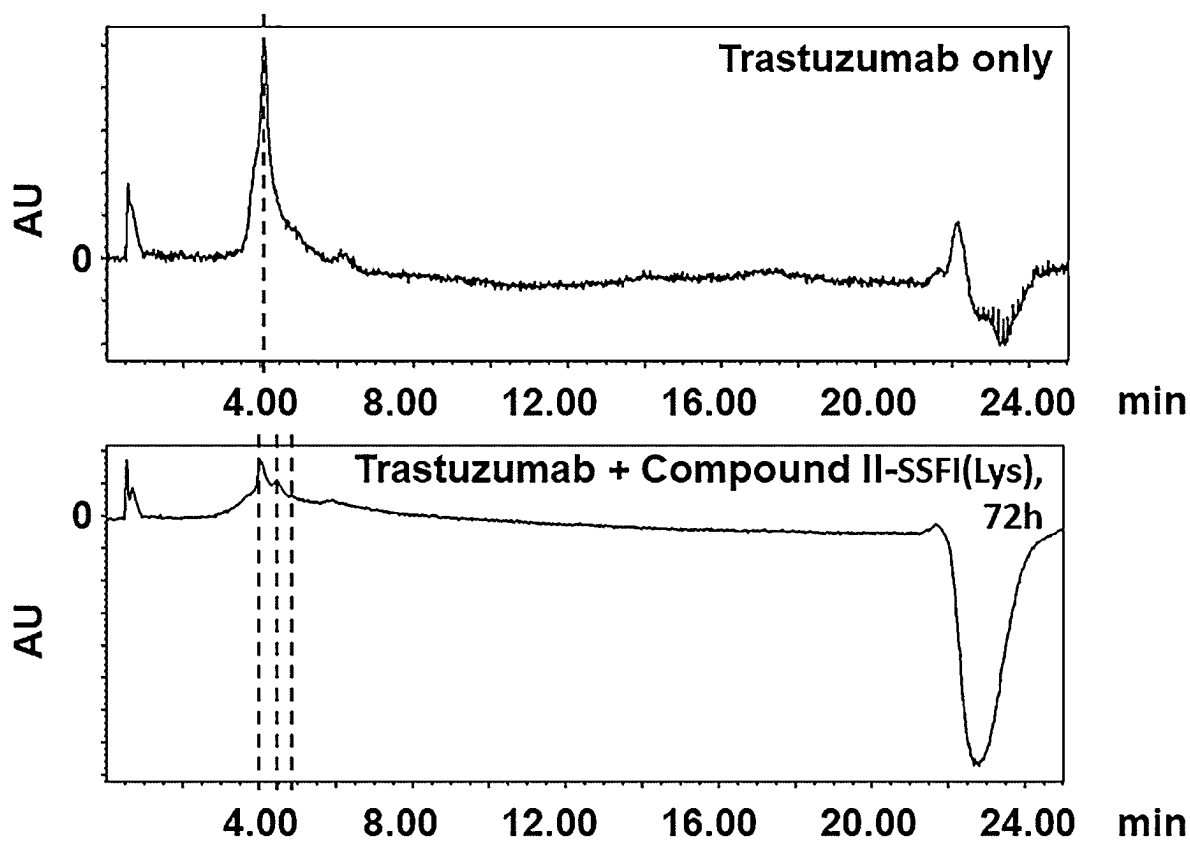
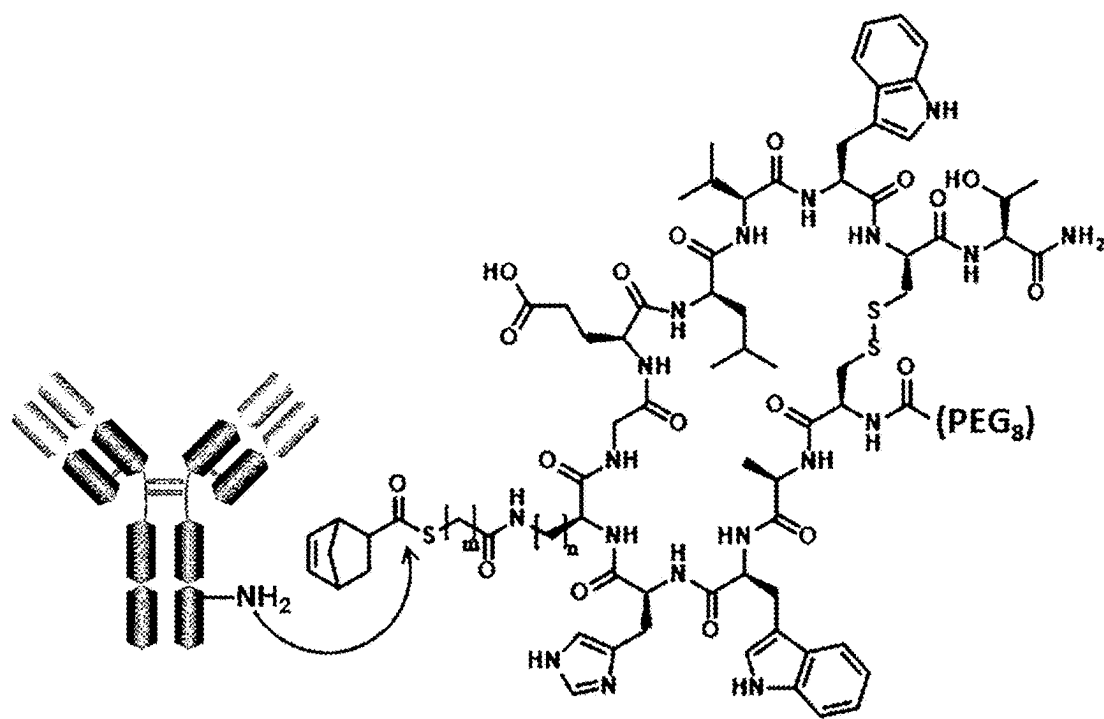


FIG. 41



(chemical structure of formula 16 pegylated)

FIG. 42

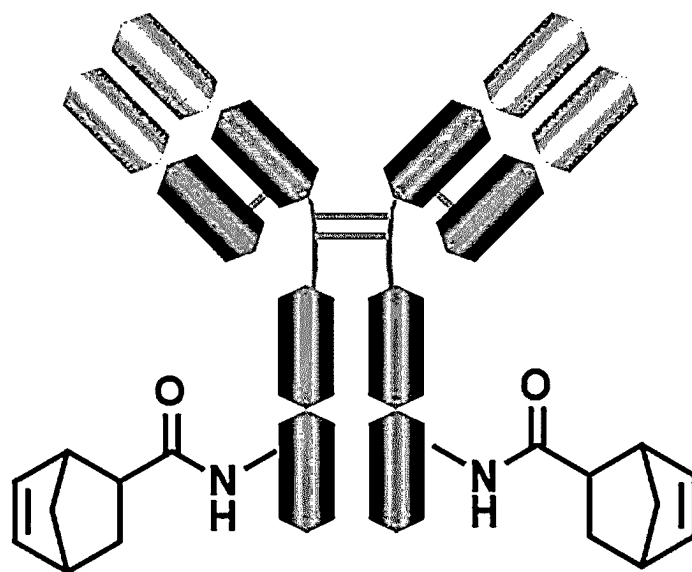


FIG. 43

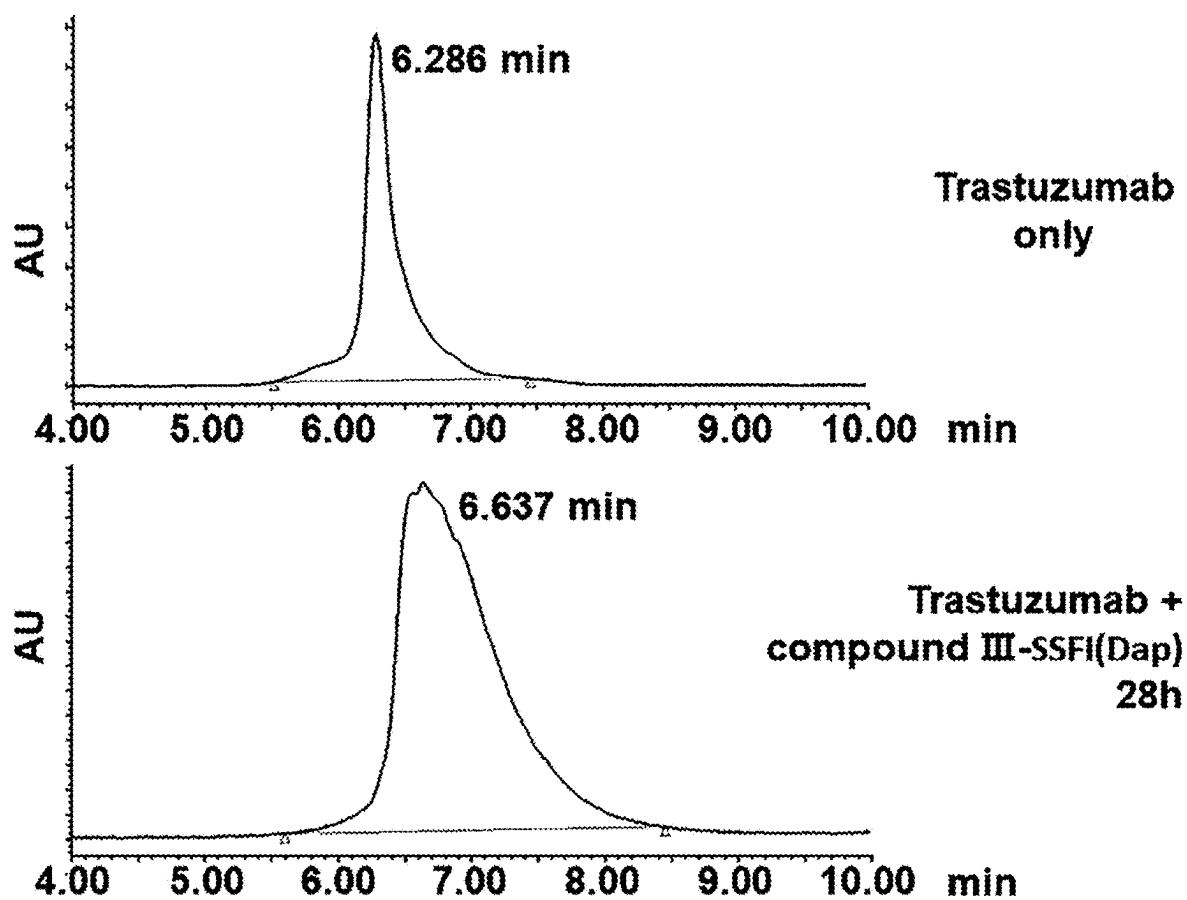


FIG. 44

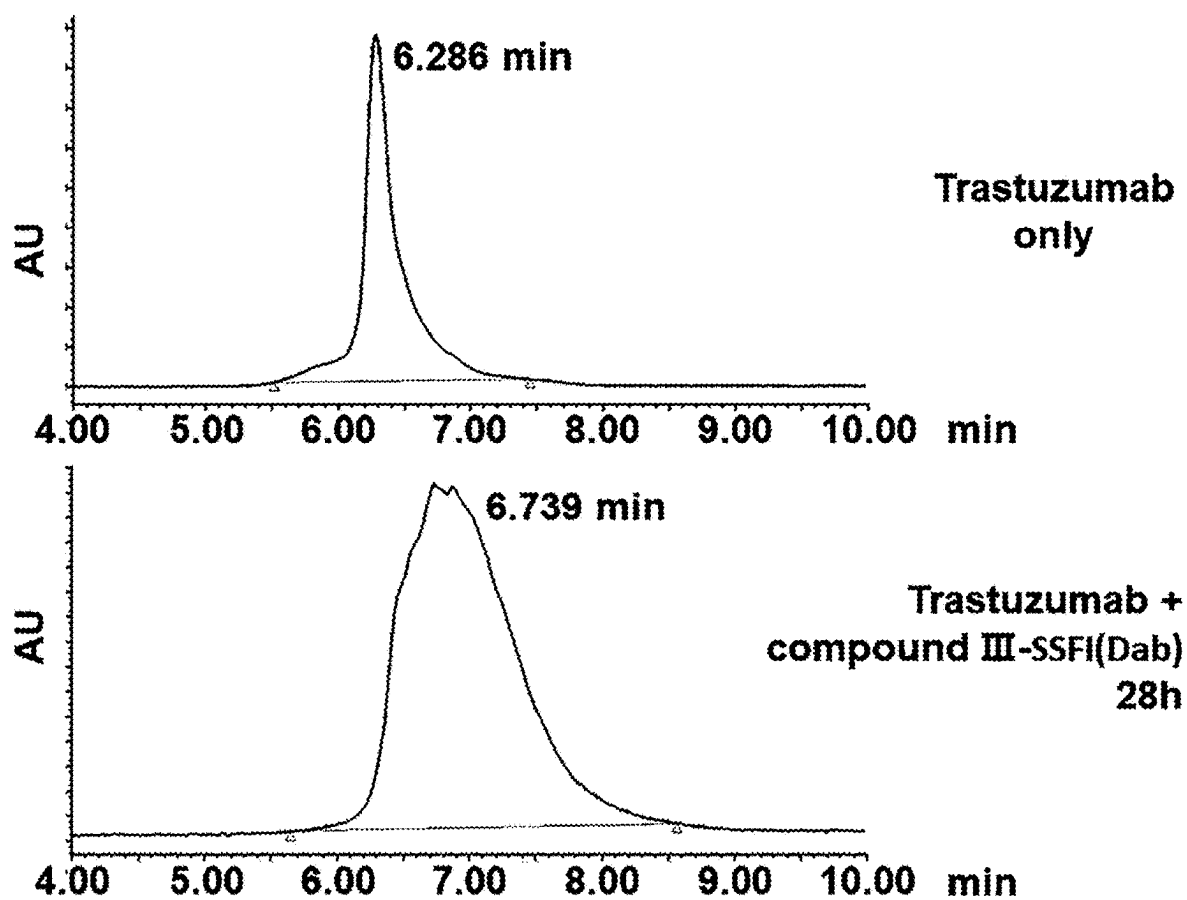


FIG. 45

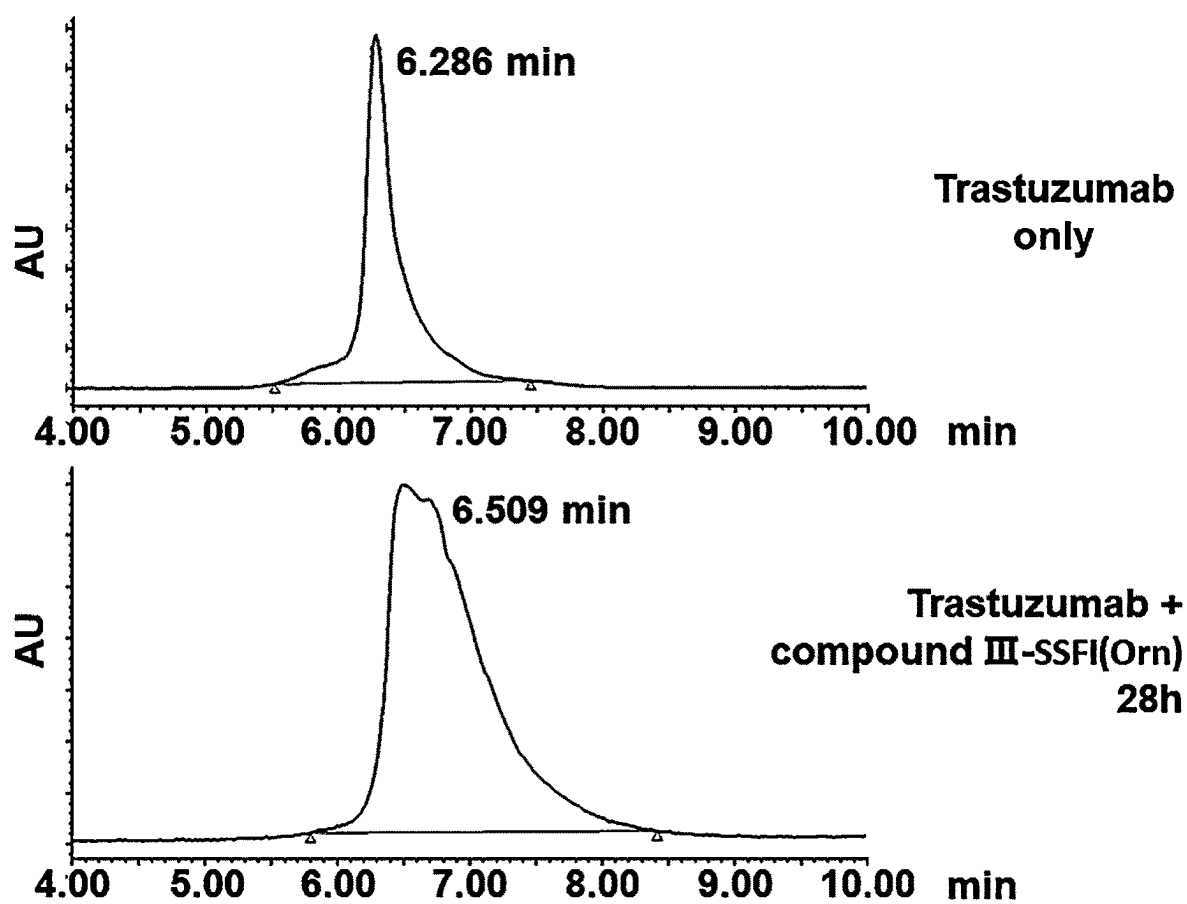


FIG. 46

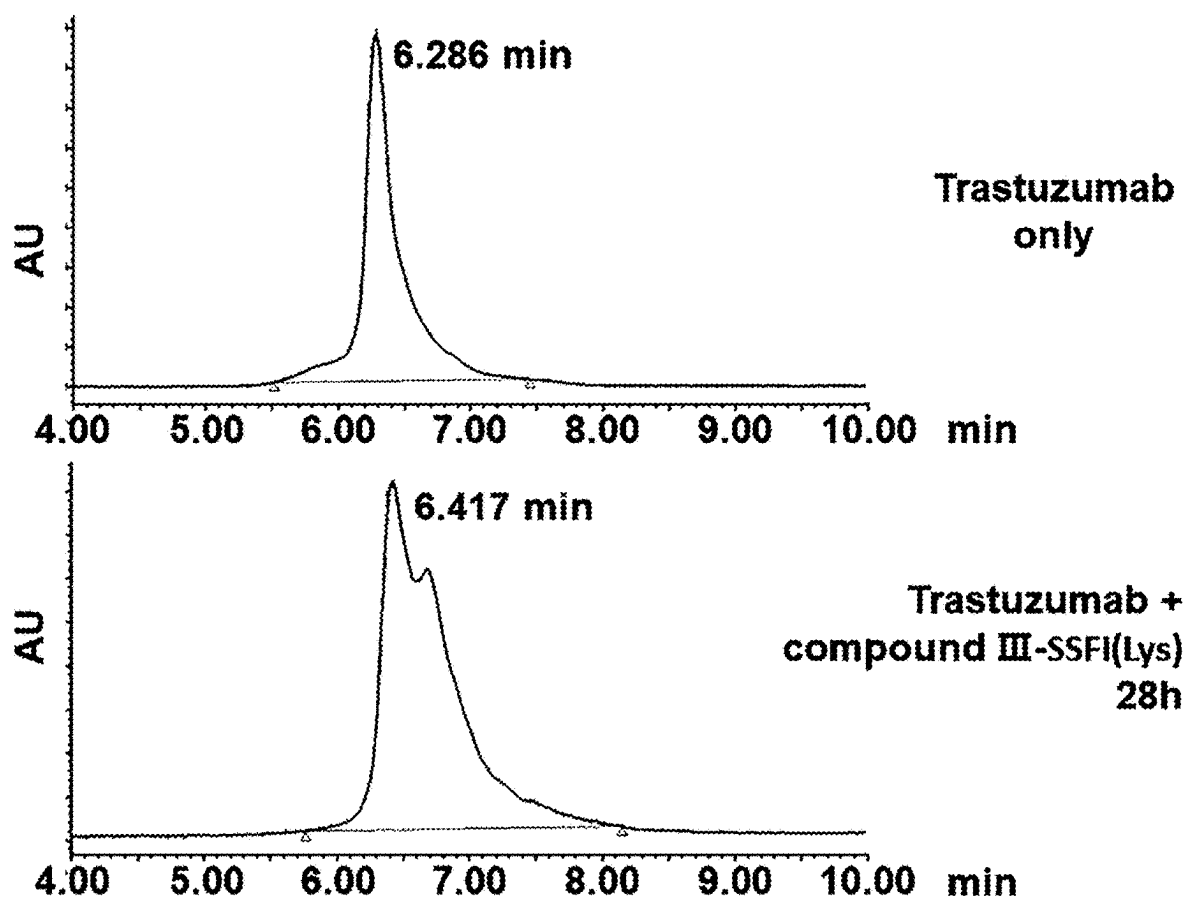


FIG. 47

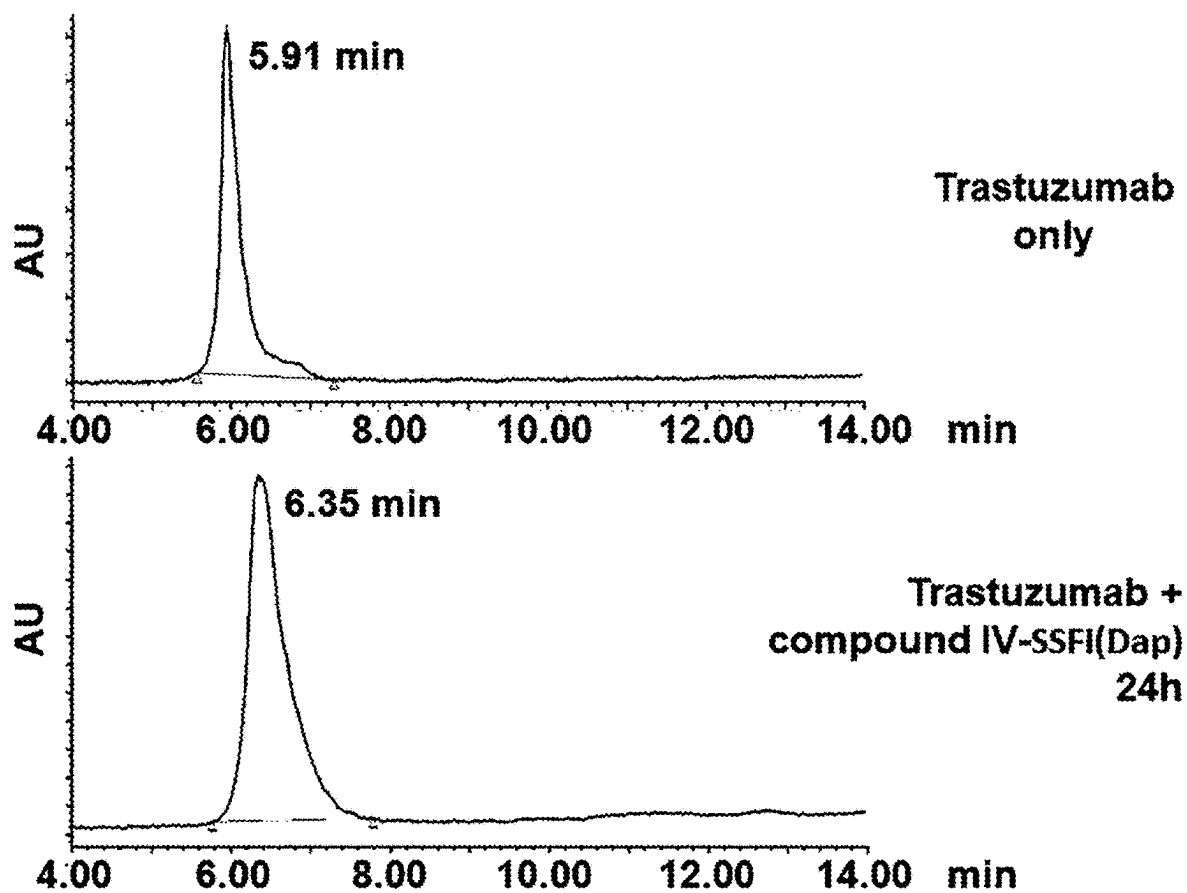


FIG. 48

Trastuzumab only (Fc/2)

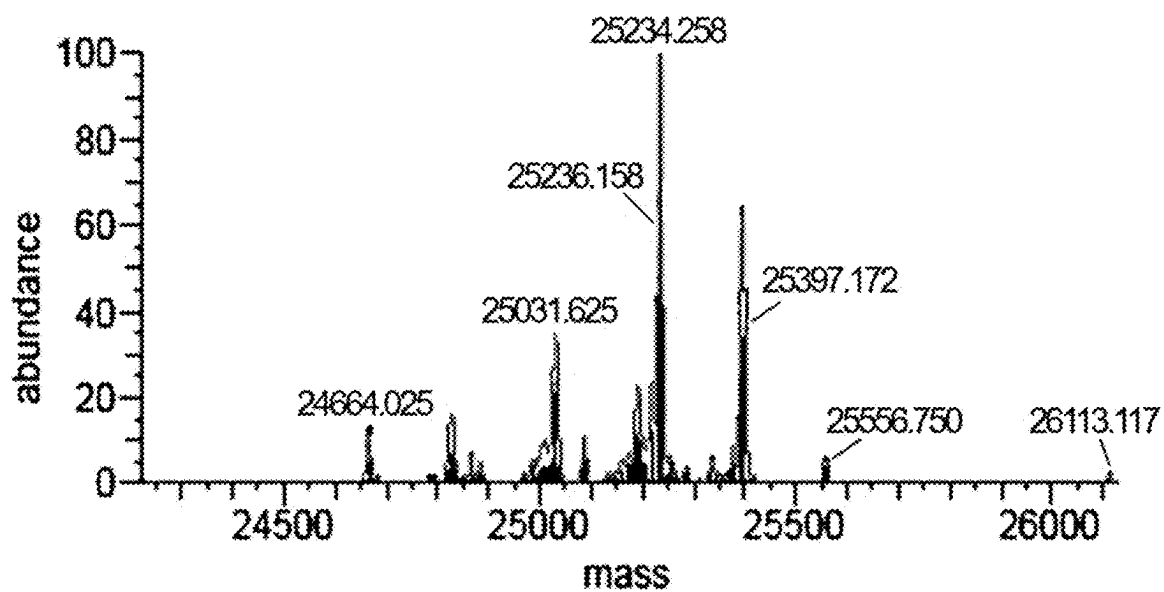


FIG. 49

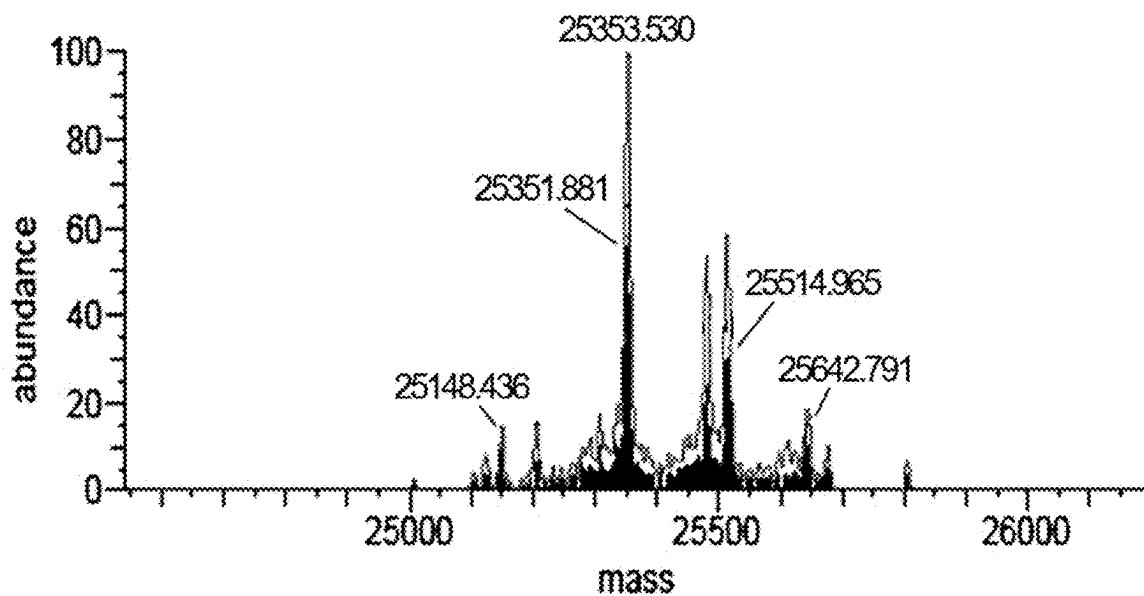
Trastuzumab-norbornene conjugate (Fc/2)

FIG. 50

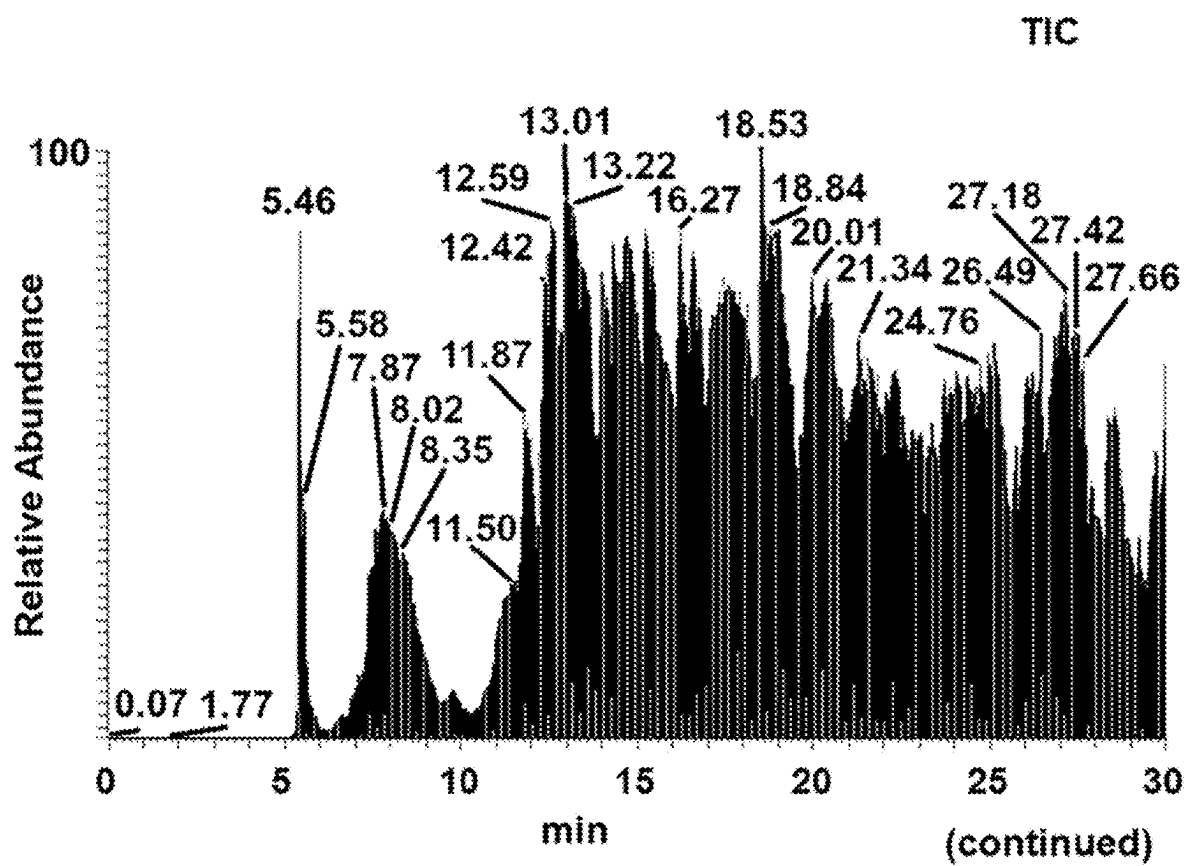


FIG. 51

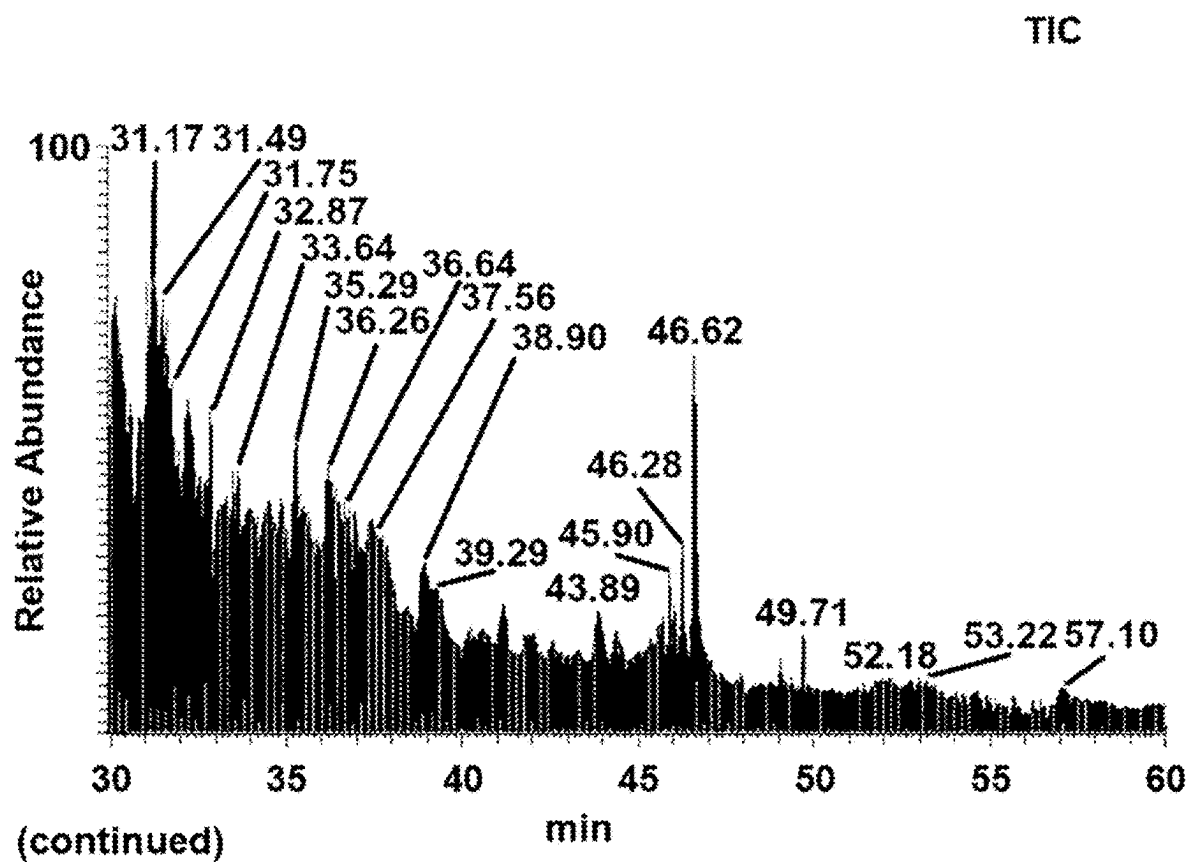


FIG. 52

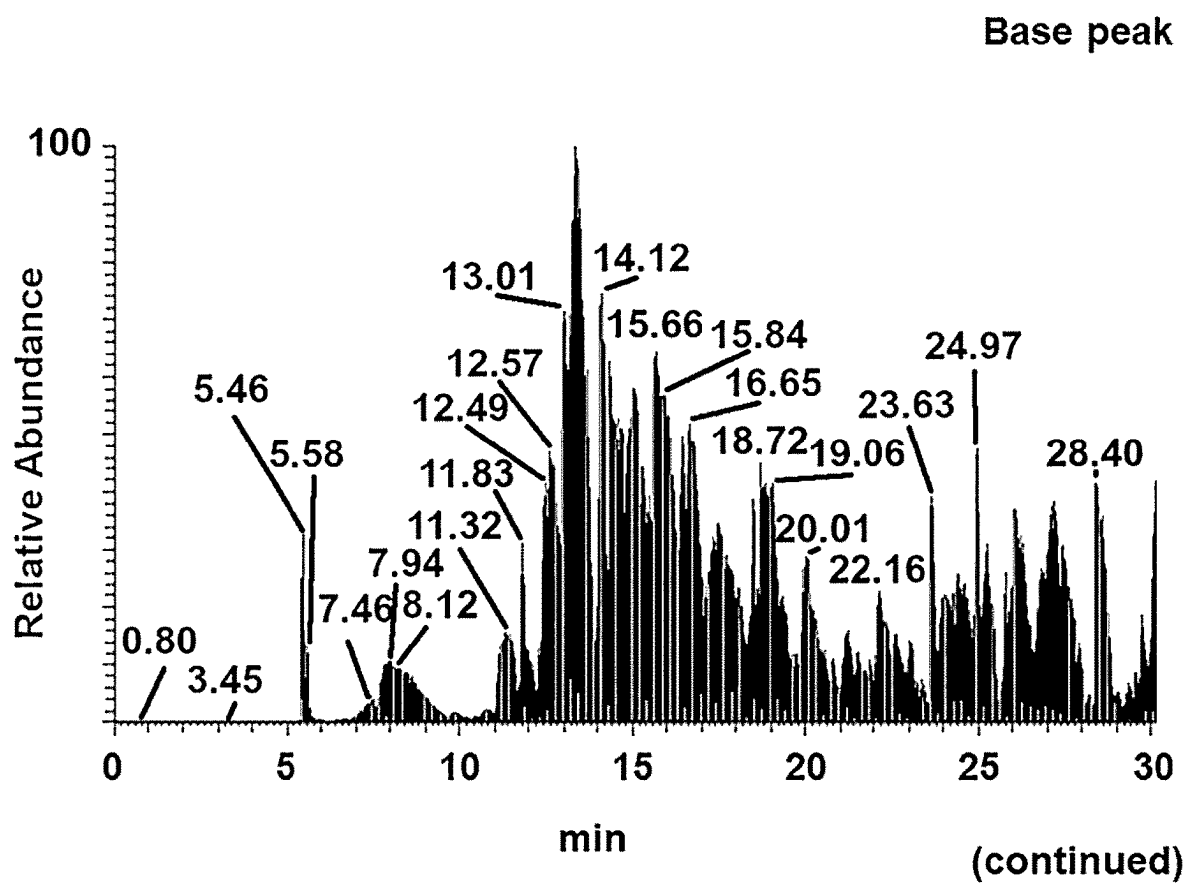


FIG. 53

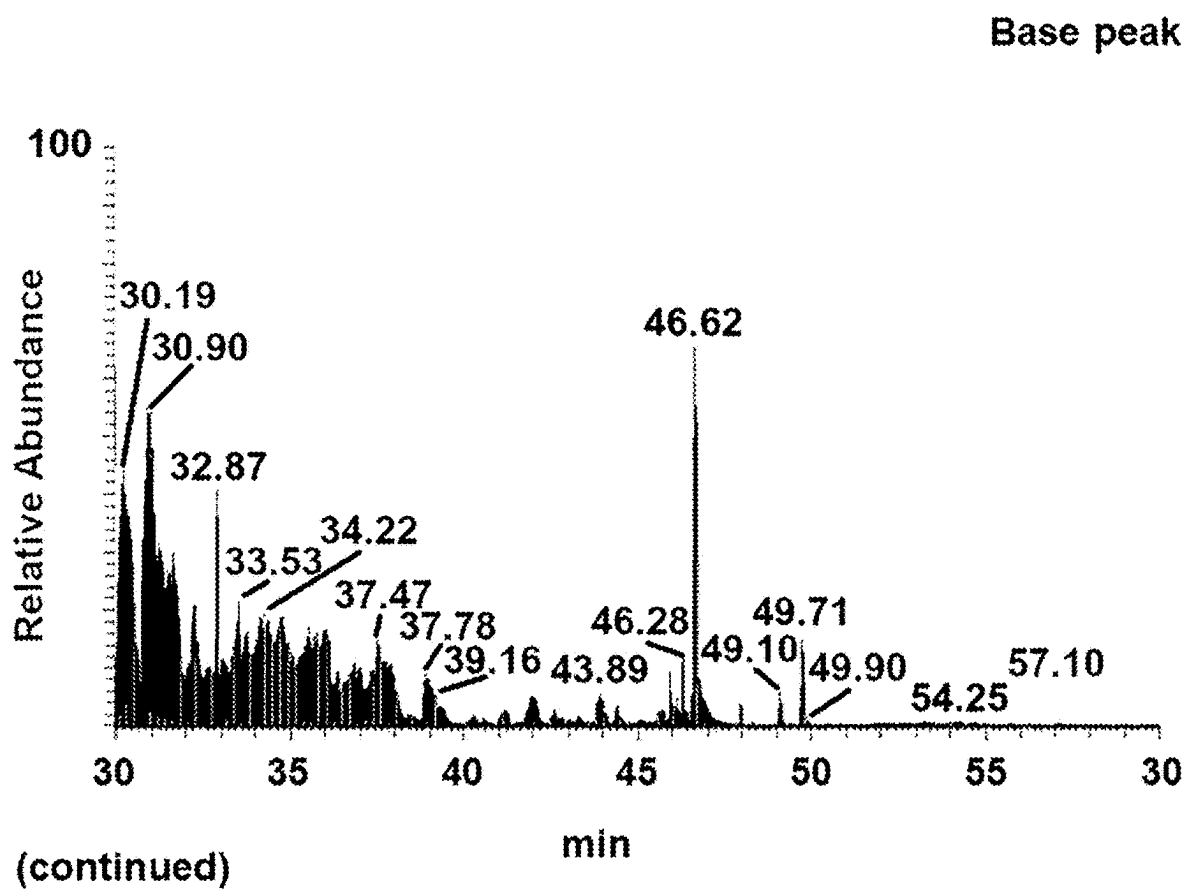


FIG. 54

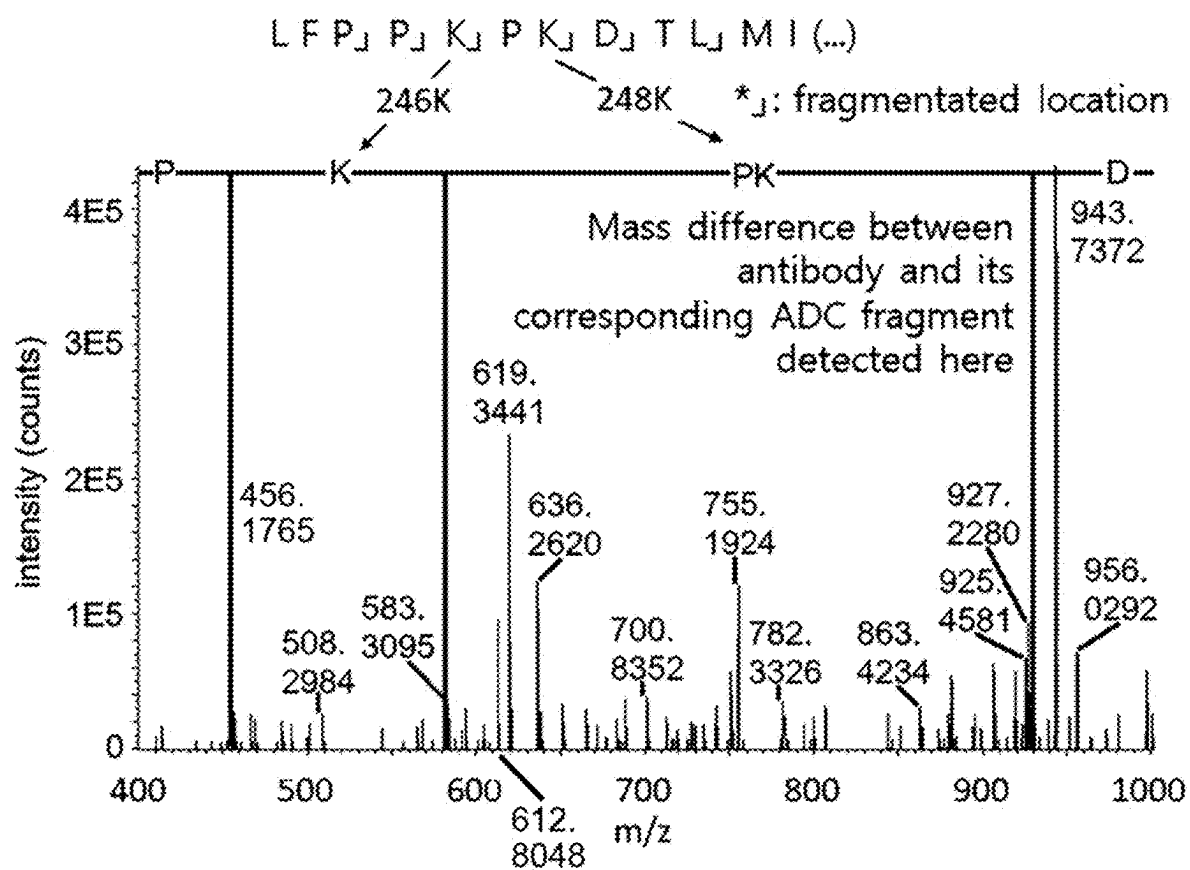


FIG. 55

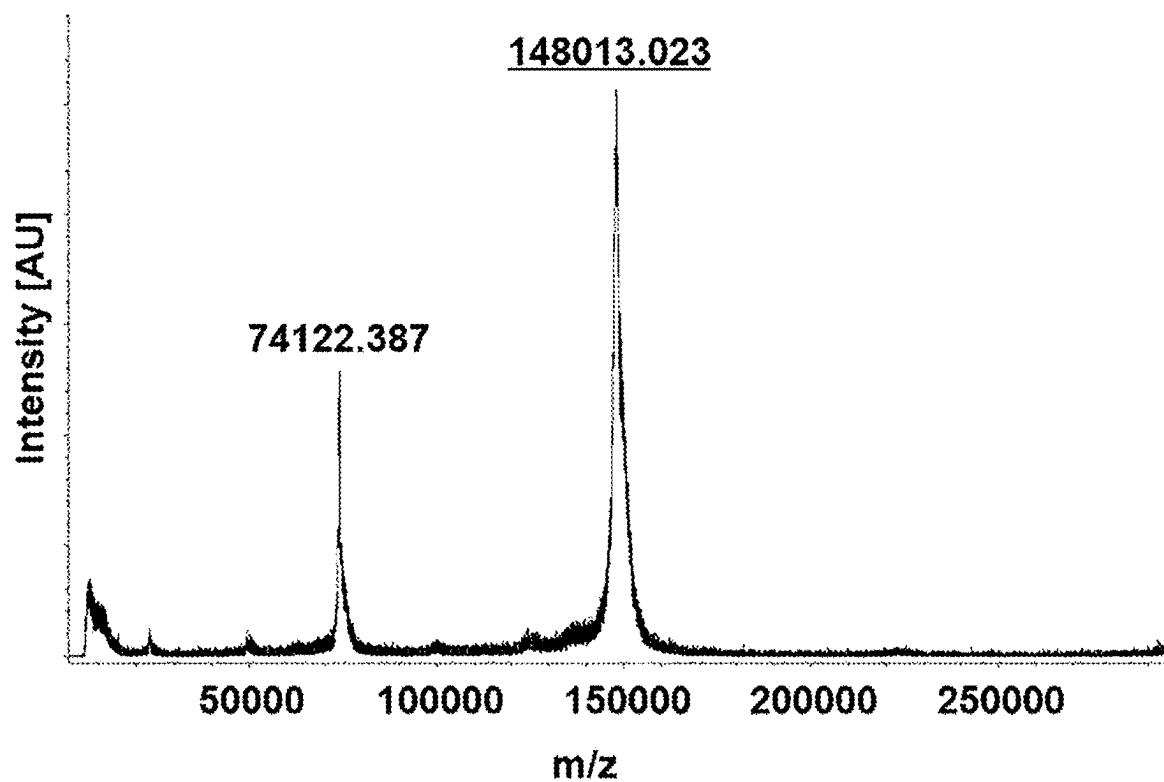


FIG. 56

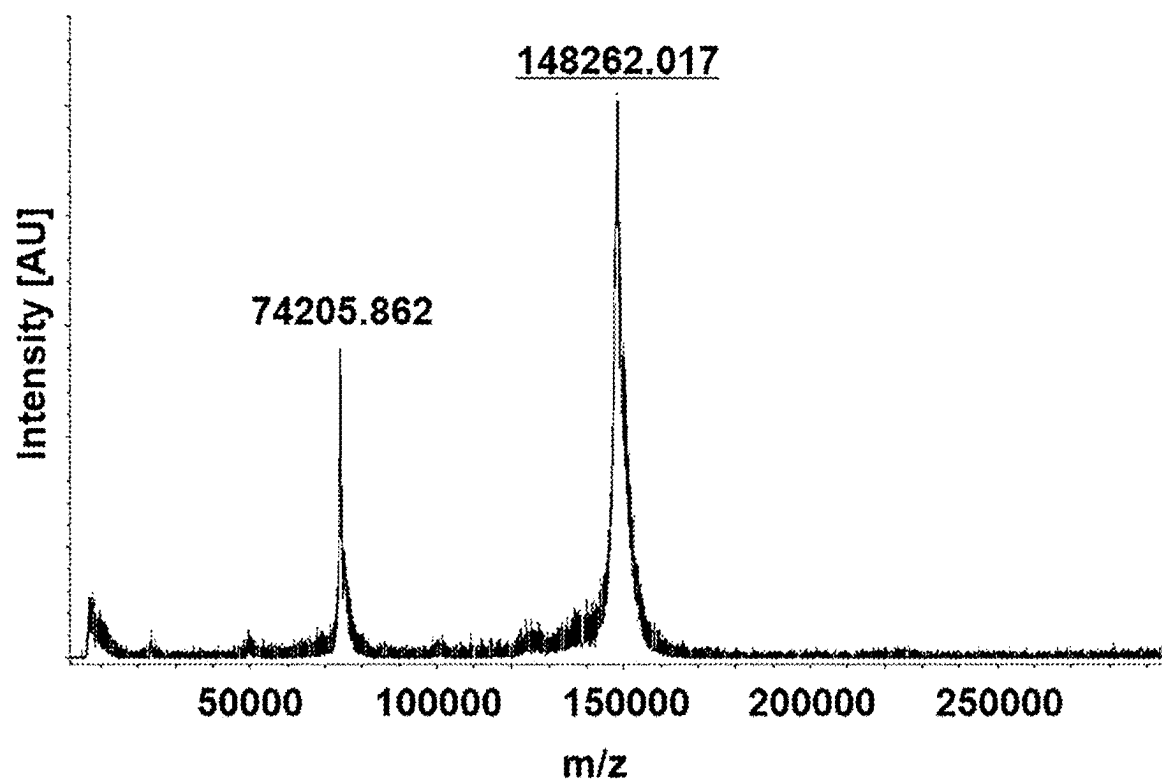


FIG. 57

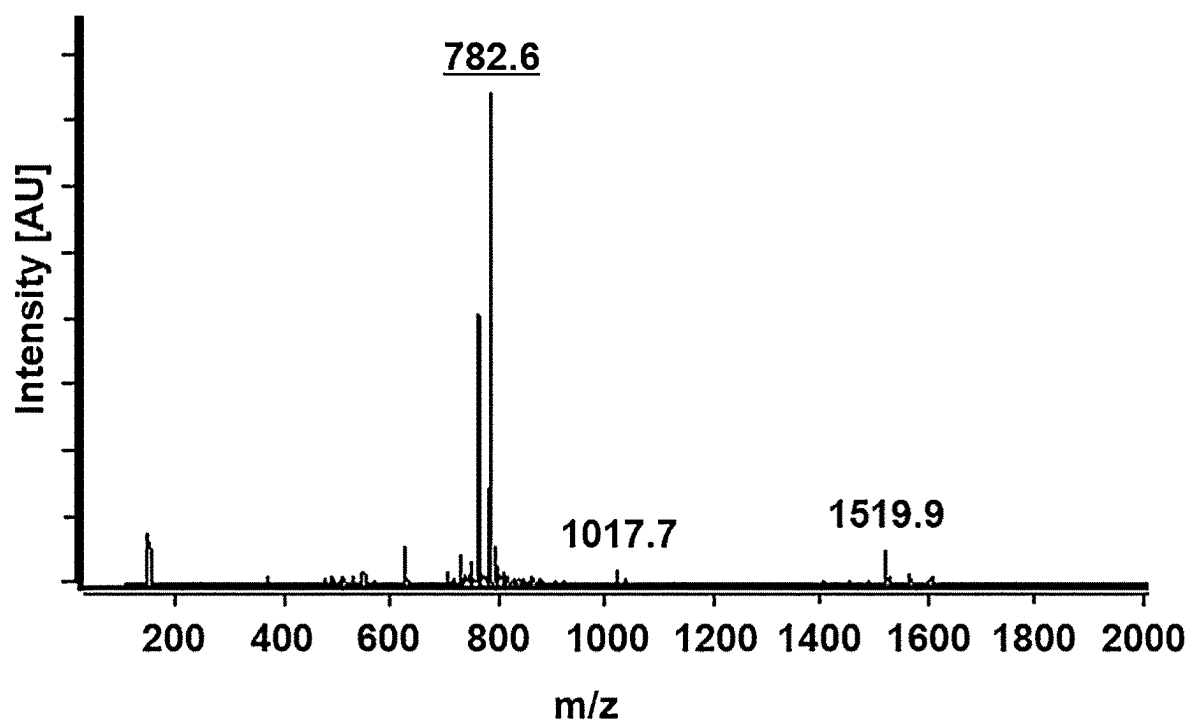


FIG. 58

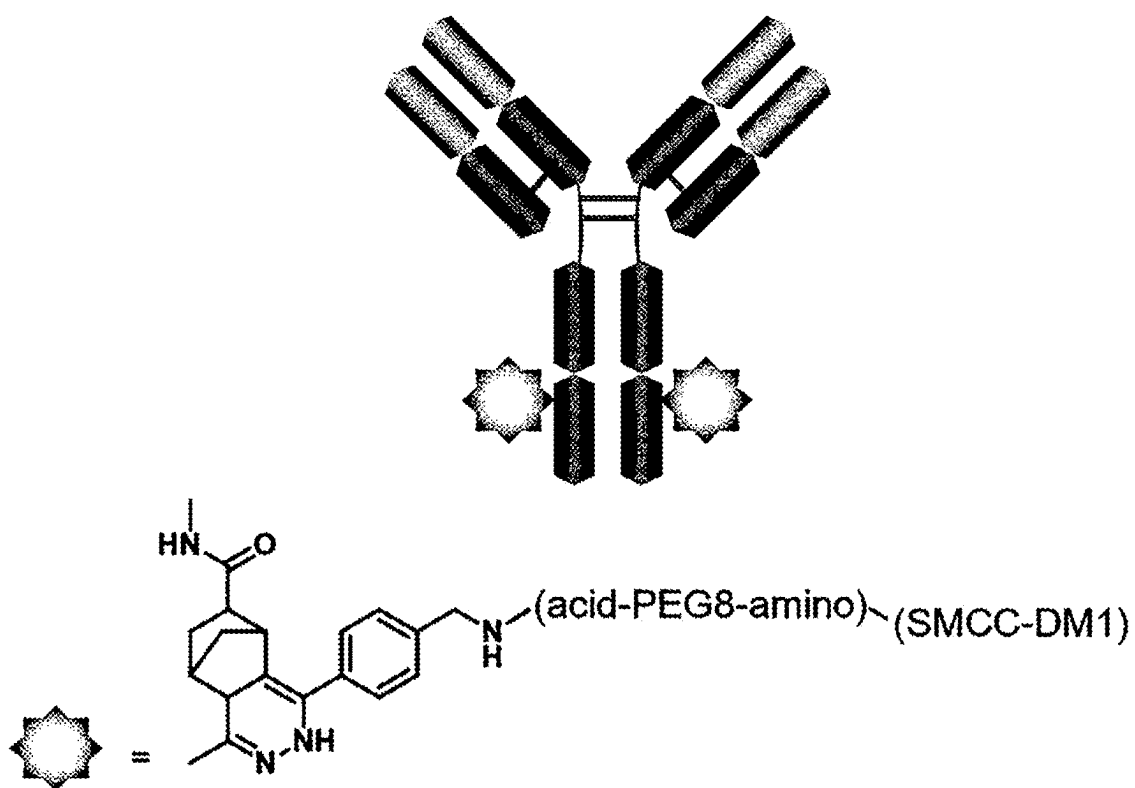


FIG. 59

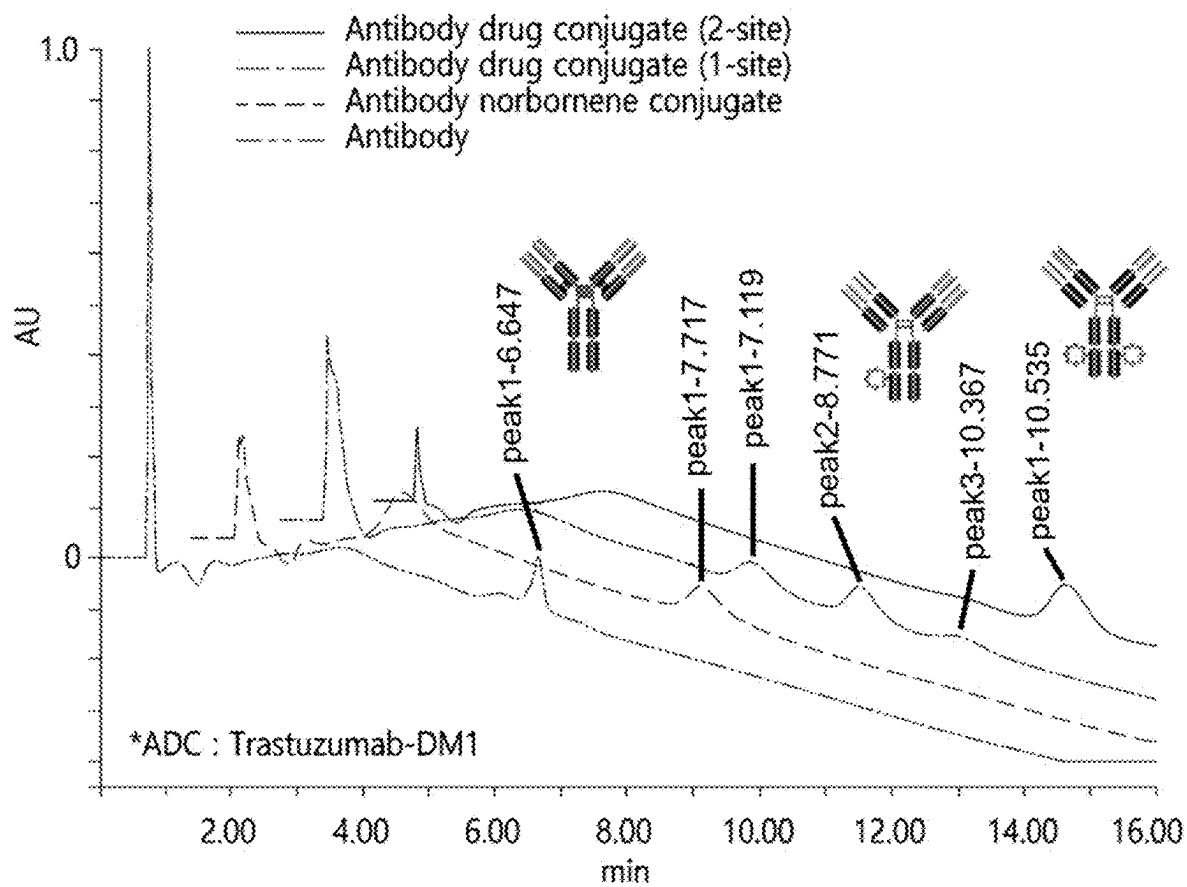


FIG. 60

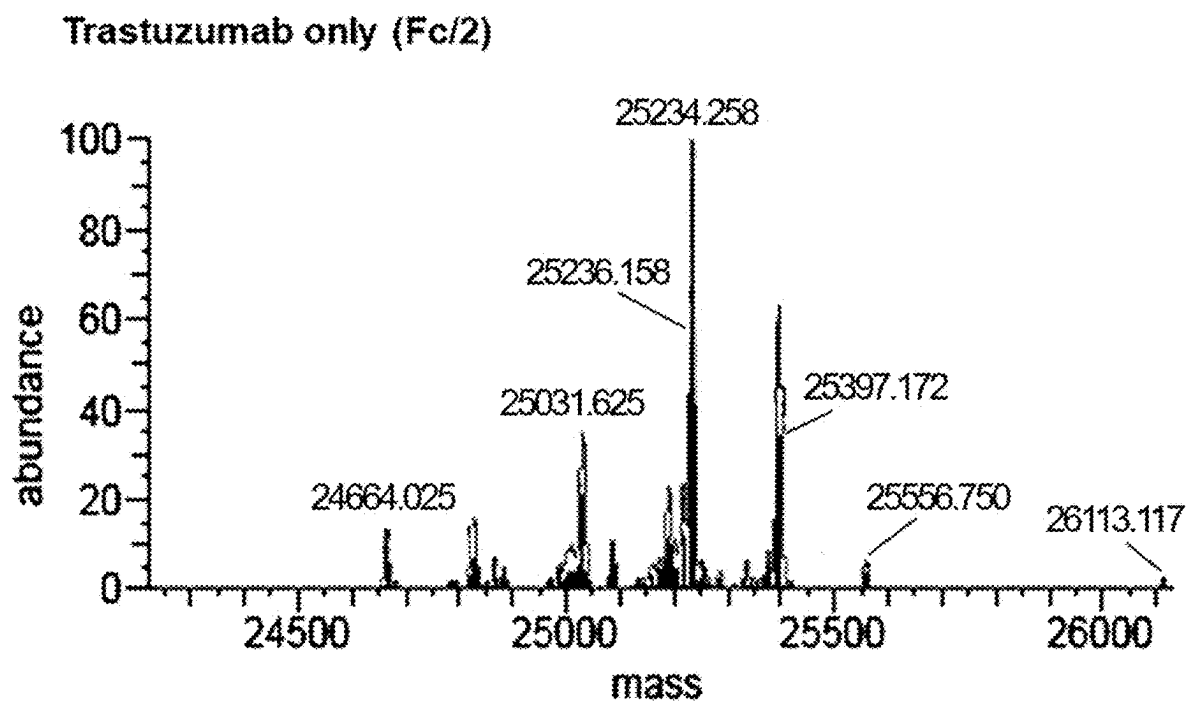
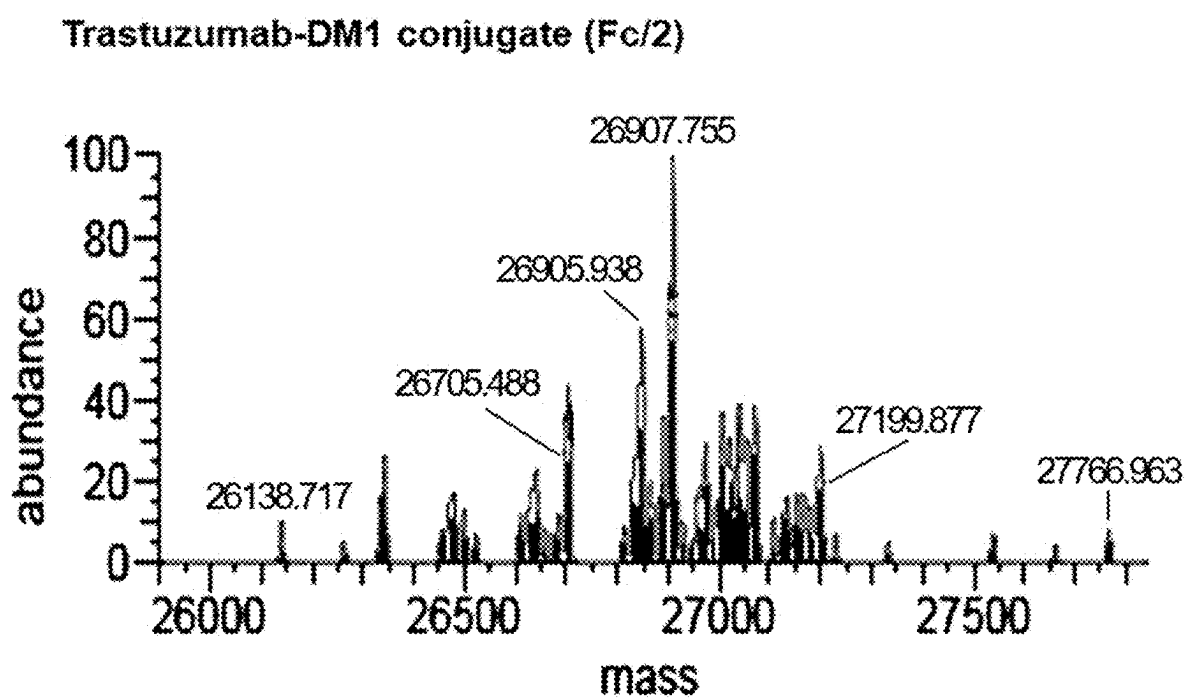


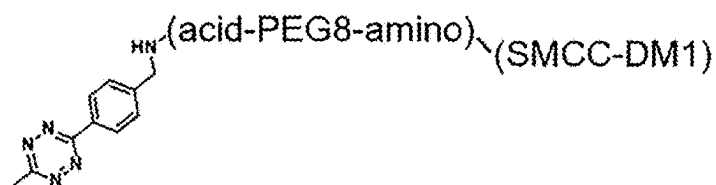
FIG. 61



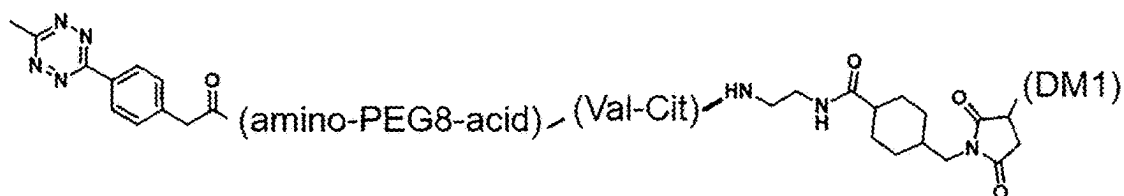
* Exact mass of DM1: 1688.80

FIG. 62

1. NC-DM1 : non-cleavable DM1 (tetrazine-PEG8-DM1)



2. VC-DM1 : cleavable DM1 (tetrazine-PEG8-valine-citrulline-DM1)



3. VC-MMAE : cleavable DM1 (tetrazine-PEG8-valine-citrulline-PAB-MMAE)

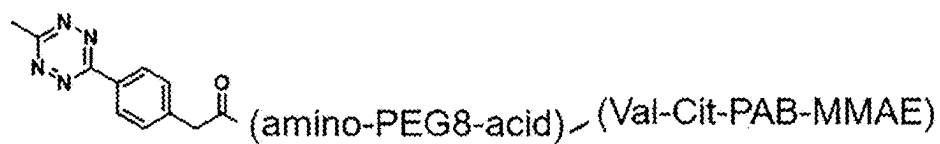


FIG. 63

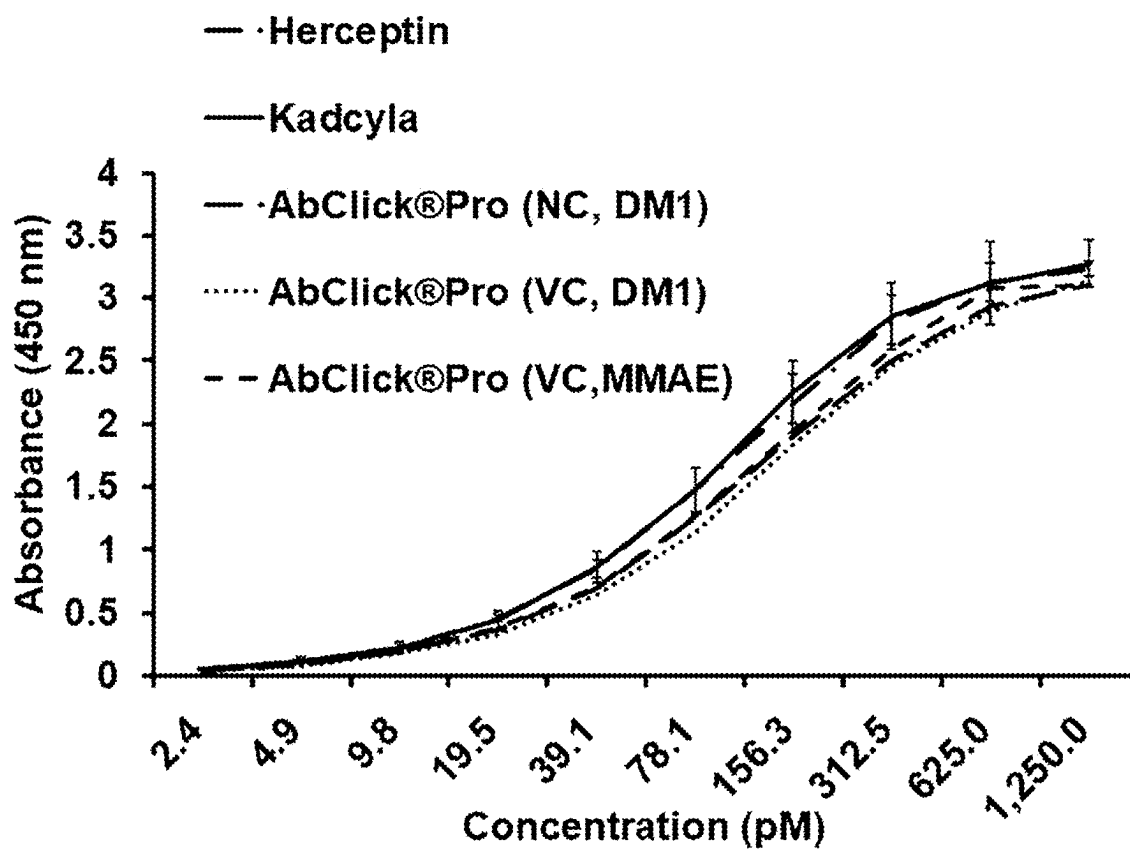


FIG. 64

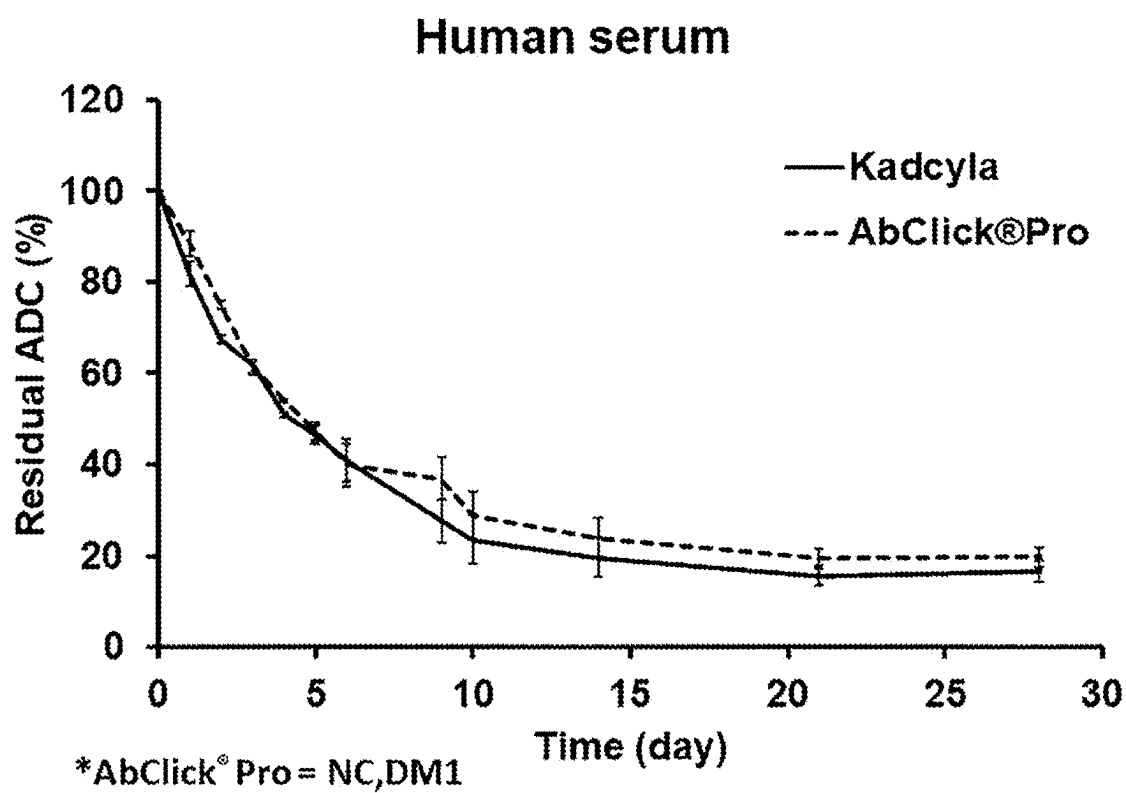


FIG. 65

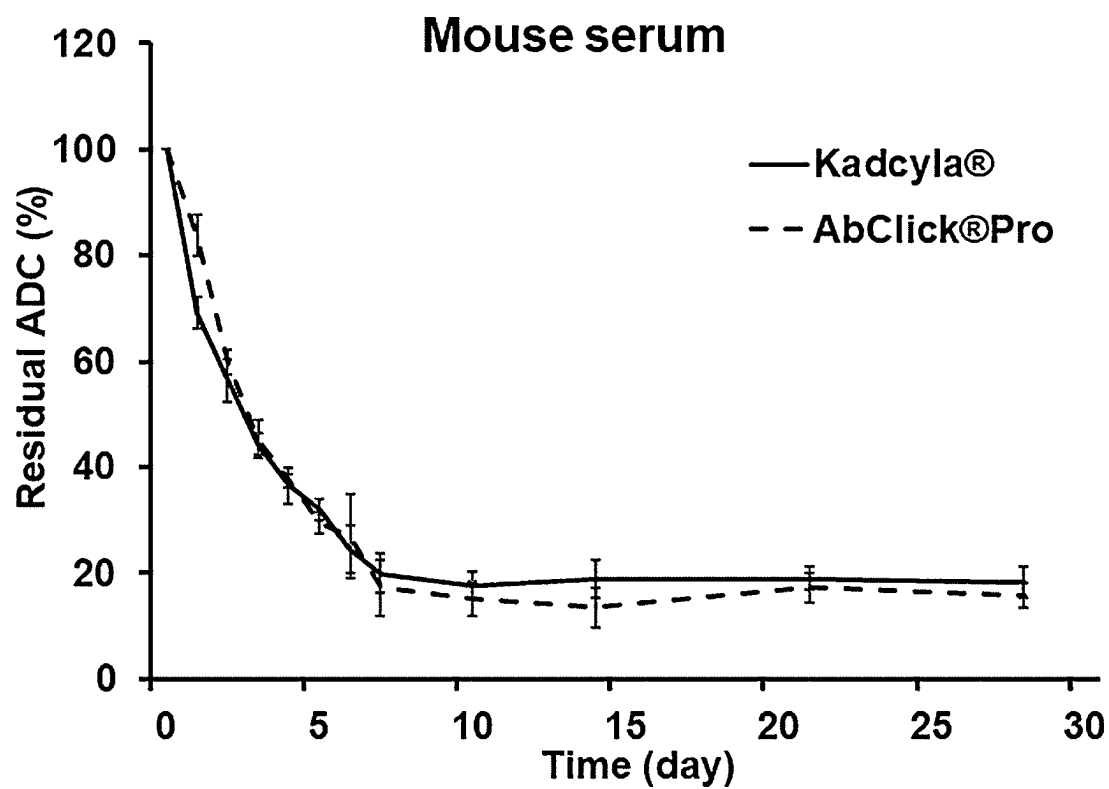


FIG. 66

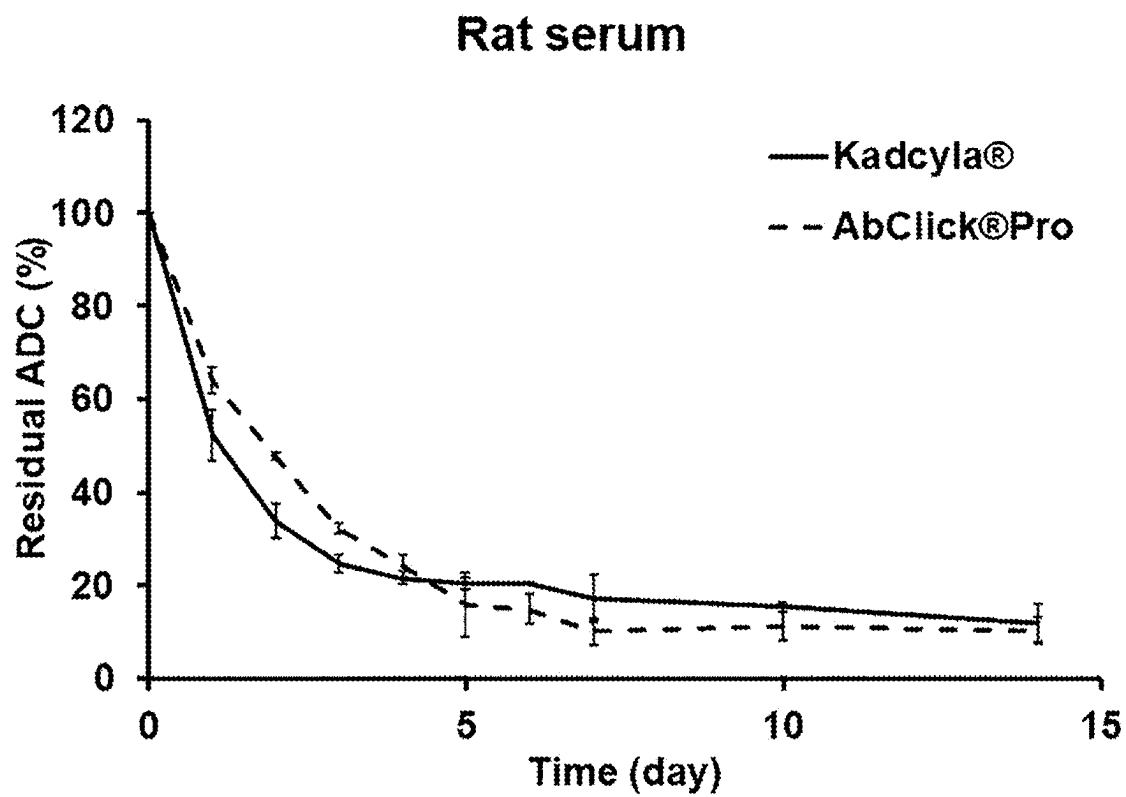


FIG. 67

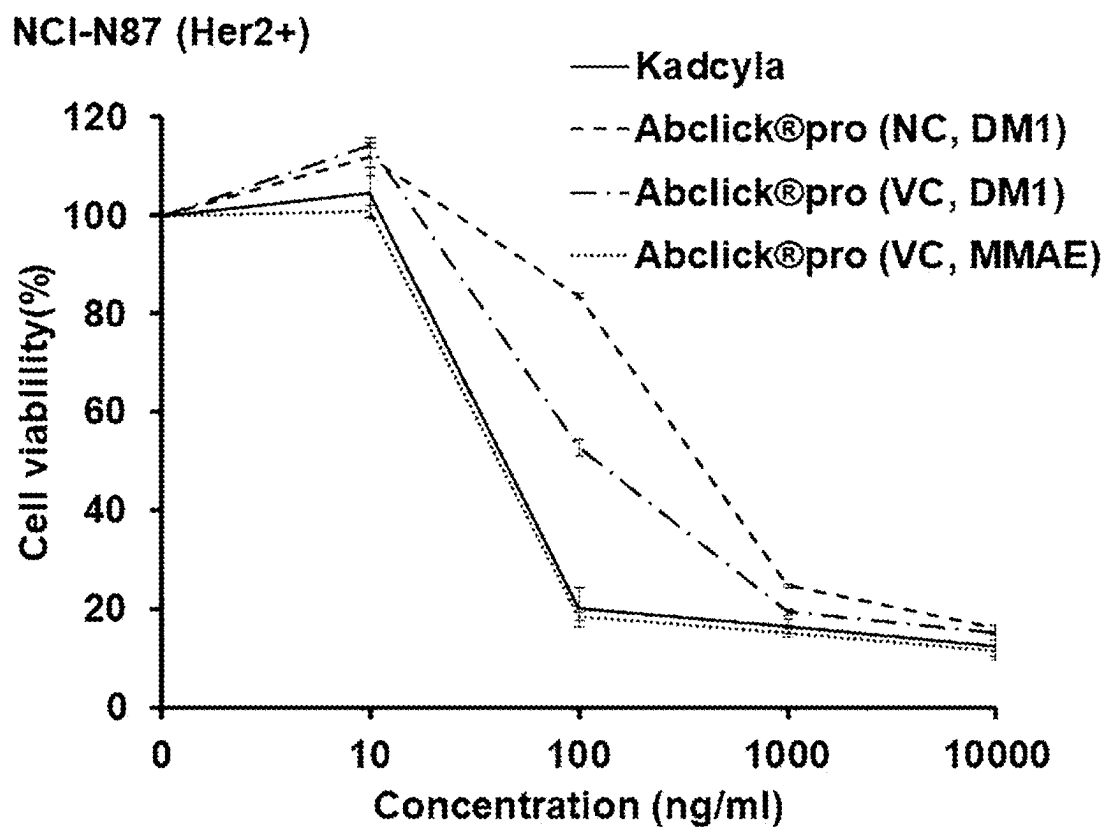


FIG. 68

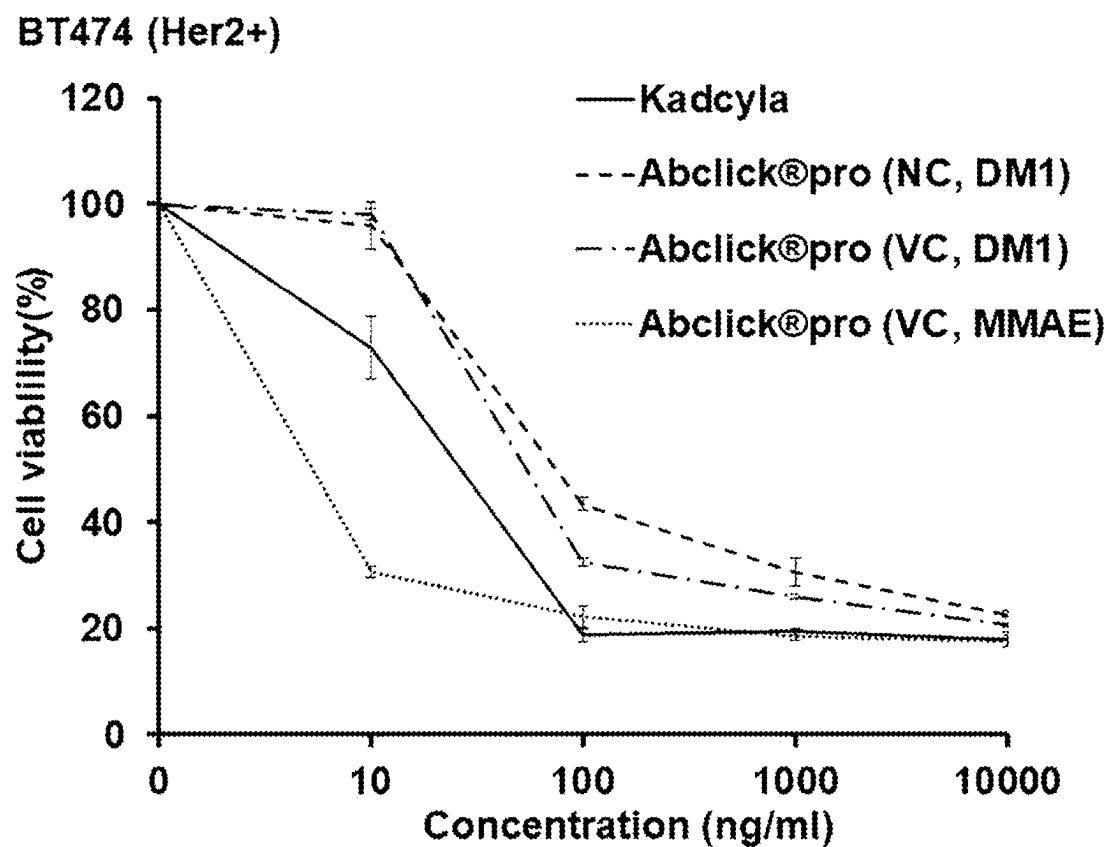


FIG. 69

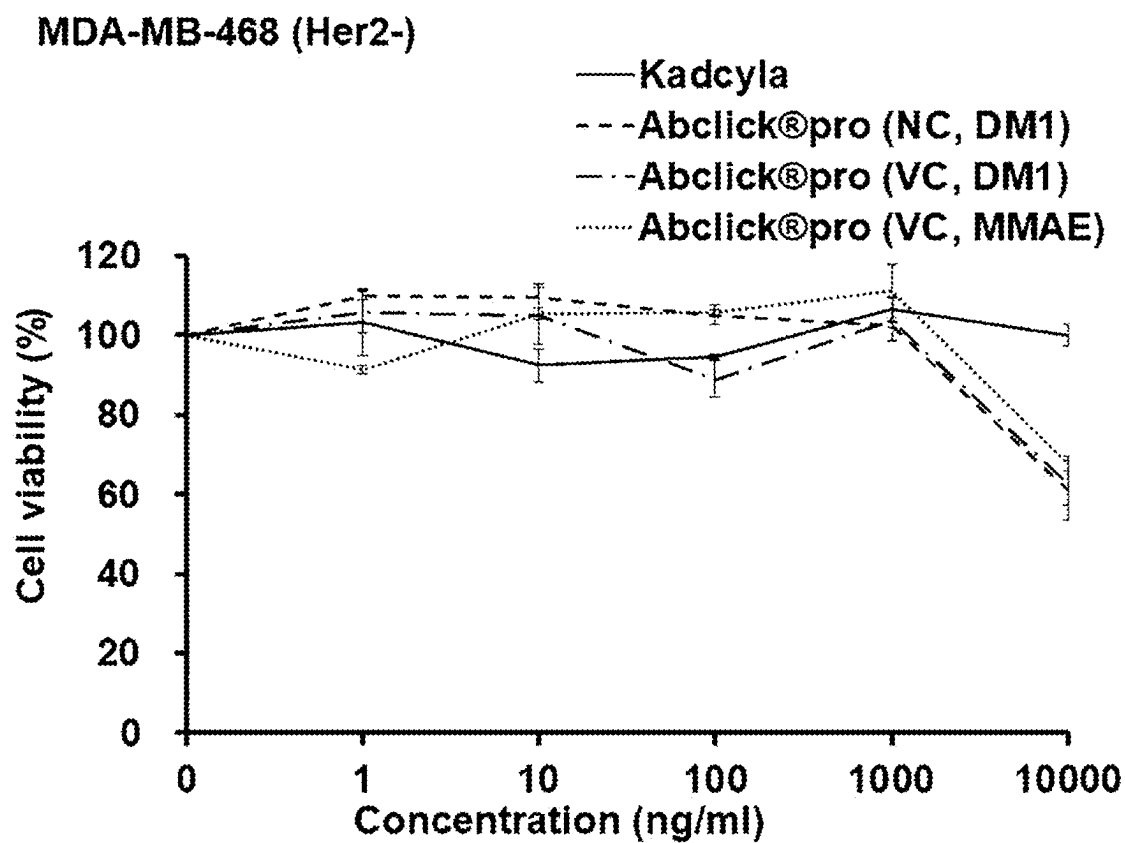


FIG. 70

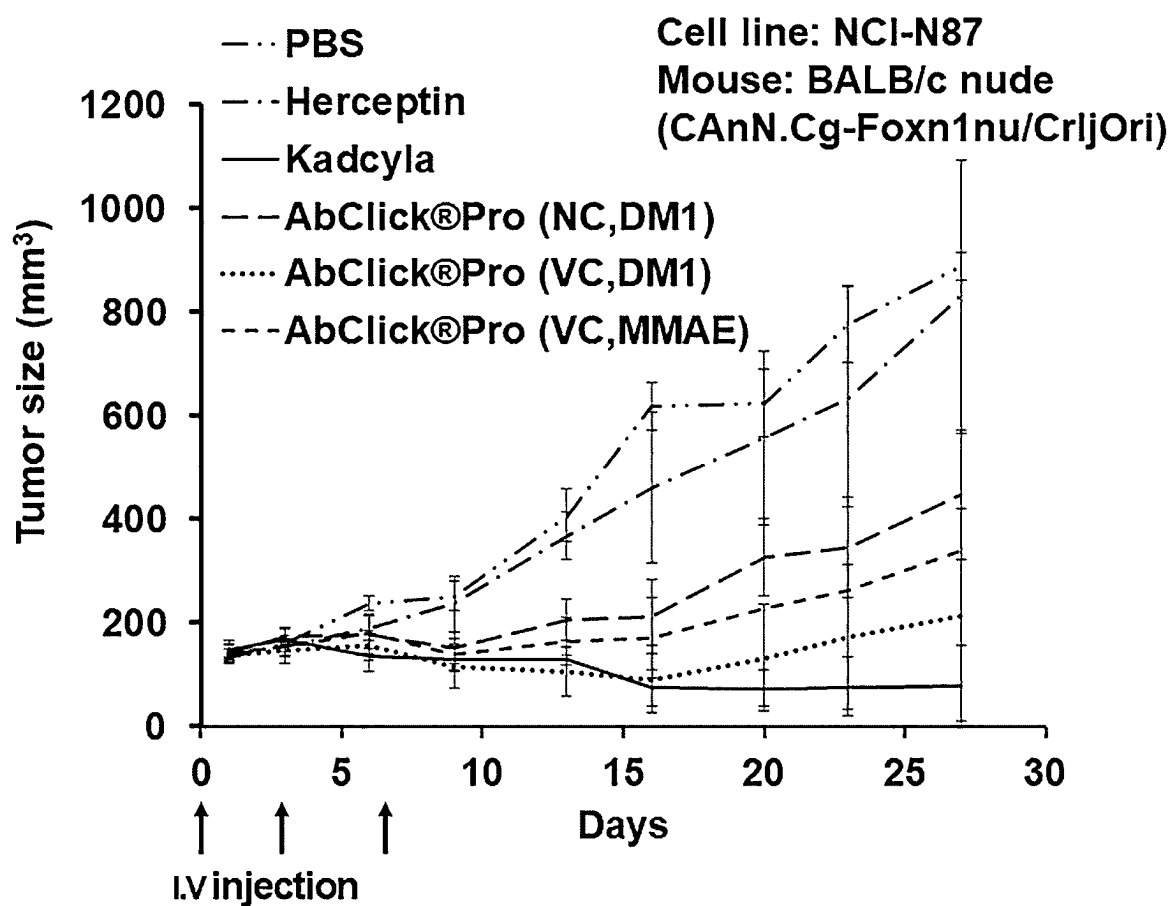


FIG. 71

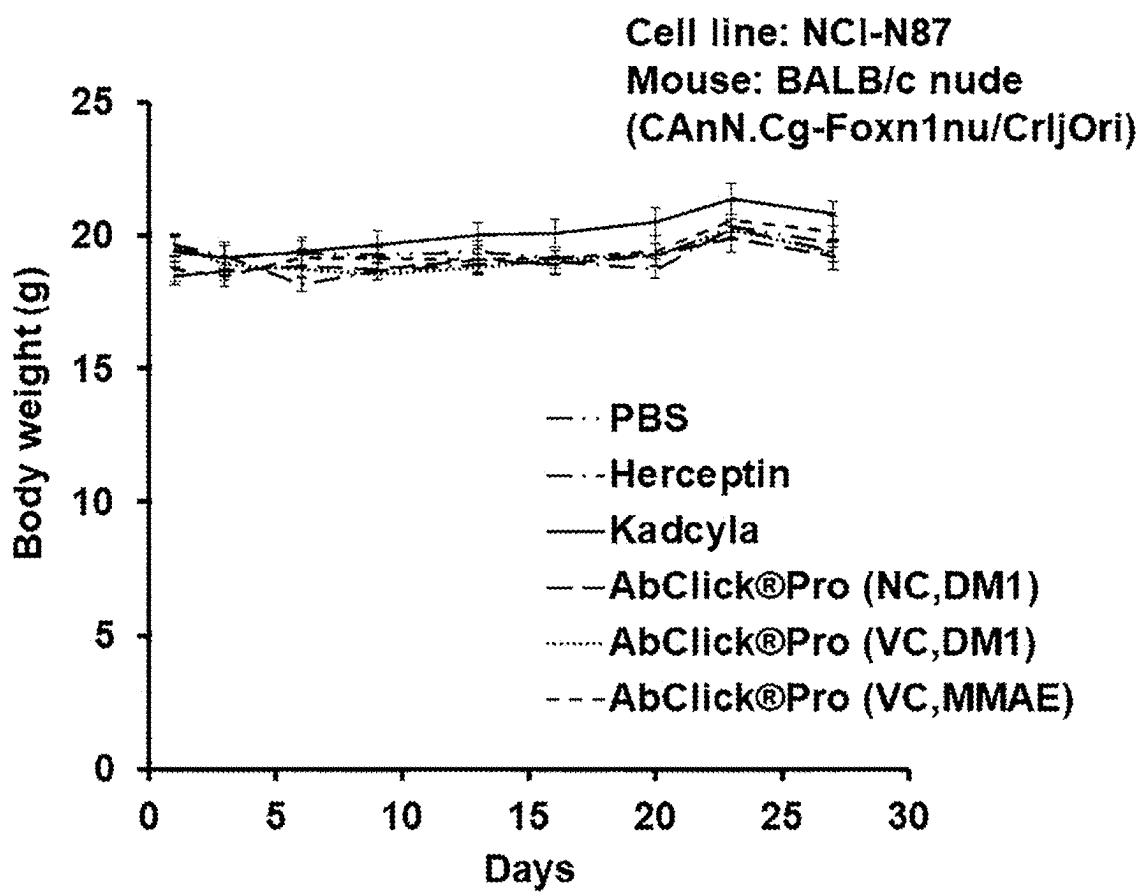
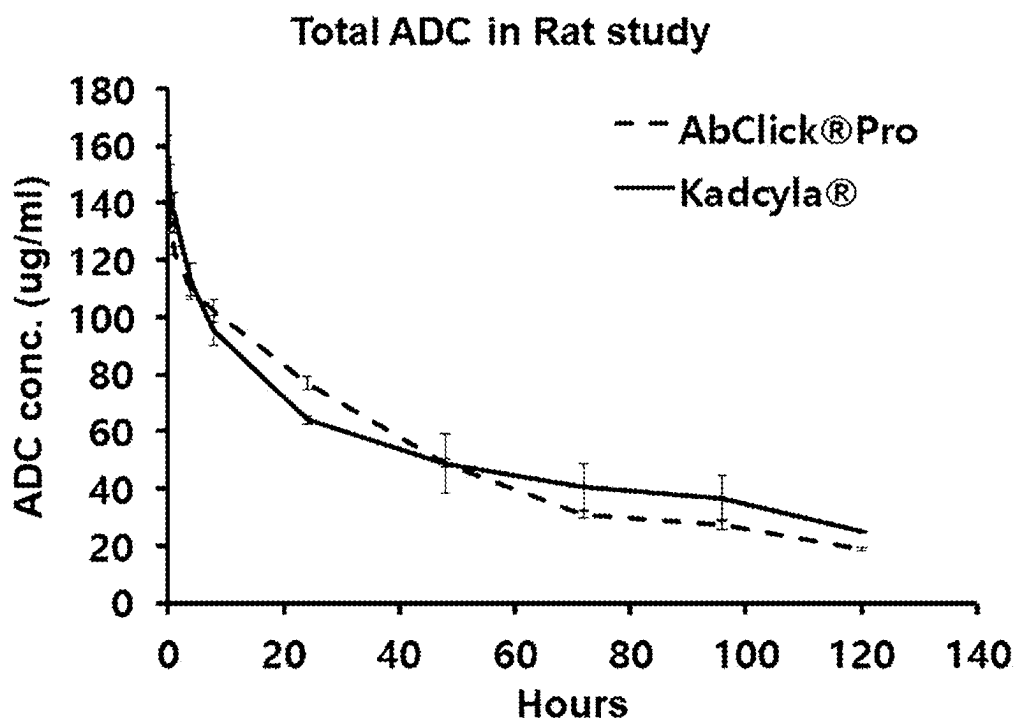


FIG. 72



*AbClick® Pro = NC,DM1

Dose = 5 mg/kg of ADC (n = 3)

FIG. 73

1

SITE-SPECIFIC ANTIBODY CONJUGATION AND ANTIBODY-DRUG CONJUGATE AS SPECIFIC EMBODIMENT THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a continuation application of U.S. patent application Ser. No. 16/980,503, filed on 14 Sep. 2020, which is a national phase application of PCT Application No. PCT/KR2020/003282, filed on Mar. 9, 2020, which claims priority to U.S. Provisional Patent Application No. 62/815,557, filed on Mar. 8, 2019. The entire disclosure of the applications identified in this paragraph is incorporated herein by reference.

FIELD

The present invention relates to technology for selectively labeling a certain amino acid residue of an antibody with substances (low-molecular-weight compounds, synthetic polymers, biopolymers (i.e., peptides, carbohydrates, proteins, and the like)) or linking a molecule (hereinafter referred to as a "cargo") to be delivered to a certain target cell or tissue to the antibody. Also, the present invention relates to technology for labeling a certain site of an antibody with a desired number of substances or linking a desired number of cargos to certain amino acid residues of the antibody. In addition, the present invention encompasses an antibody complex prepared by the method or a method using a fragment complex of the antibody.

BACKGROUND

Antibodies are biomolecules that have a function of recognizing certain molecules, and have been used for various industrial applications. For example, an antibody may be used to detect or screen for a certain substance and check a route through which the certain substance moves in the body or cells. Also, the antibody may be used for therapeutic purposes by inducing an immune response against the certain substance.

To expand the functions of such an antibody, there has been attempts to improve the antibody's ability. Typically, there has been an attempt to label an antibody with foreign substances in order to supplement or expand the antibody's functions. Typically, when an antibody is labeled with a fluorescent substance, the antibody may be used for a fluorescence assay, or when an antibody is labeled with an agent for treating a certain disease, the antibody may be used to maximize a therapeutic effect of the antibody. These attempts and technology are generally referred to as "antibody labeling." The present invention relates to a novel method of labeling an antibody.

In the beginning, the antibody labeling was achieved by randomly attaching a foreign substance to an antibody. However, this method has a lot of problems. The antibodies thus prepared have a problem in that they have poor homogeneity. These antibodies have a problem in that their effects remain uneven because there are a difference in the number of substances attached to each of the antibodies and a difference in binding sites for the antibodies. This problem is a great barrier to the development of antibody-drug conjugate (ADC) technology that requires high safety and reproducibility.

Also, the method has a problem in that an antibody's ability to recognize may be significantly degraded. An

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antibody consists of an Fab domain including an antigen-binding domain that recognizes an antigen, and an Fc domain involved in the crystallization of the antibody. Random labeling resulted in a highly degraded antibody's ability to recognize by allowing a foreign substance to bind to an antigen-binding domain of an antibody or a site adjacent to the antigen-binding domain.

Therefore, there has been a demand for technology for uniformly labeling an antibody in a site-specific manner in the related art. Although some techniques were developed, e.g., genetically modulating or modifying an antibody and the like, most of them are ineffective in technical and economic aspects. Accordingly, the present invention is designed to solve the problems, and thus is directed to technology capable of specifically linking a certain substance (or a "moiety") to a certain site of an antibody without any additional modulation of the antibody and delivering a certain substance (a drug or a labeled substance) into cells or tissues using the antibody as well.

SUMMARY

Technical Problem

The present invention provides technology for specifically transferring a chemical functional group to a certain site of an antibody. In one specific embodiment, the present invention provides technology for specifically transferring a chemical functional group to lysine 246 in the antibody. In another specific embodiment, the present invention provides technology for specifically transferring a chemical functional group to lysine 248 in the antibody. In still another specific embodiment, the present invention provides technology for specifically transferring a chemical functional group to lysine 246 and 248 in the antibody.

The present invention provides technology capable of linking a desired number of chemical functional groups to an antibody. In one specific embodiment, the present invention provides an antibody or a fragment thereof, which has two certain moieties bound thereto. In another specific embodiment, the present invention provides an antibody or a fragment thereof, which has four certain moieties bound thereto.

The present invention provides technology for specifically linking a cargo moiety to a certain site of an antibody. In one specific embodiment, the present invention provides technology for specifically linking a cargo moiety to lysine 246 in the antibody. In another specific embodiment, the present invention provides technology for specifically linking a cargo moiety to lysine 248 in the antibody. In still another specific embodiment, the present invention provides technology for specifically linking a cargo moiety to lysine 246 and 248 in the antibody.

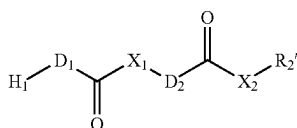
The present invention provides technology for linking a desired number of cargo moieties to an antibody. In one specific embodiment, the present invention provides an antibody or a fragment thereof, which has two cargo moieties bound thereto. In another specific embodiment, the present invention provides an antibody or a fragment thereof, which has four cargo moieties bound thereto.

The present invention provides a method of using the aforementioned antibody or antibody fragment. In one specific embodiment, the present invention provides a method of treating a certain disease using an antibody-drug complex.

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Technical Solution

The present application provides A compound of Formula 2:



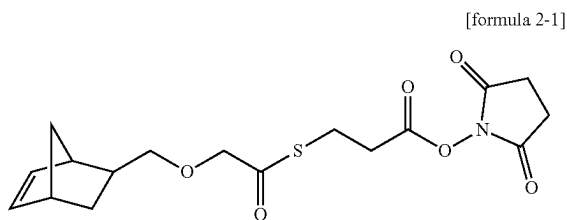
[formula 2]

wherein, H₁ is a first click chemistry functional group, D₁ is selected from a covalent bond, C₁₋₄alkylene, C₂₋₄alkenylene, C₂₋₄alkynylene, and C₃₋₈-cycloalkylene, X₁ is S, D₂ is selected from C₁₋₇alkylene, C₂₋₇alkenylene, C₂₋₇alkynylene, and C₃₋₈-cycloalkylene, X₂ is O, R₂' is N-succinimide, p-nitrophenyl, or pentafluorophenyl.

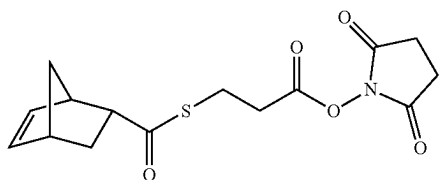
Also, the present application provides the compound wherein H₁ is selected from terminal alkyne, azide, strained alkyne, diene, dienophile, alkene, thiol, and tetrazine. Furthermore, the present application provides the compound wherein H₁ is selected from norbornene, tetrazine, azide and dibenzocyclooctyne-amine.

Additionally, the present application provides the compound wherein R₂' is N-succinimide.

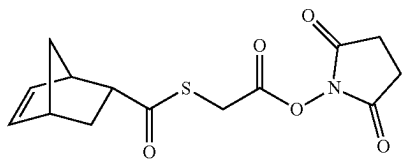
Additionally, the present application provides the compound wherein the formula 2 is formula 2-1, 2-2, or 2-3:



[formula 2-1]



[formula 2-2]

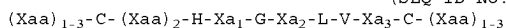


[formula 2-3]

the present application provides a peptide of formula 4-2:

[formula 4-2]

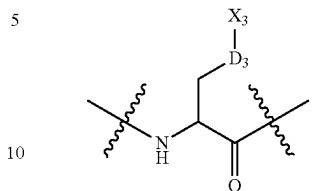
(SEQ ID NO: 6)



wherein, each Xaa is independently any amino acid residue that is not a cysteine residue, C is a cysteine residue, H is a histidine residue, G is a glycine residue, Xa₂ is a glutamic acid residue or an asparagine residue, L is a leucine residue, V is a valine residue, Xa₃ is selected from a

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tryptophan residue, a naphthylalanine residue, and a phenylalanine residue, and Xa₁ is



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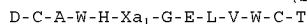
wherein D₃ is a covalent bond or C₁₋₃alkylene and X₃ is NH₂, wherein the peptide consists of 13 to 17 amino acid residues, wherein the peptide exhibits the activity of binding to human immunoglobulin G (IgG), and wherein the cysteine residue that is between two to four amino acids from the N-terminus of formula 4-2 and the cysteine residue that is between two to four amino acids from the C-terminus of formula 4-2 are optionally linked.

Also, the present application provides the peptide wherein D₃ is a covalent bond, methylene, or ethylene

Additionally, the present application provides the peptide wherein the formula 4-2 is formula 4-6:

[formula 4-6]

(SEQ ID NO: 10)

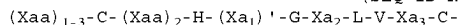


wherein, D is an aspartic acid residue, A is an alanine residue, E is a glutamic acid residue, W is a tryptophan residue, and T is a threonine residue.

The present application provides a peptide-compound conjugate of formula 6-2:

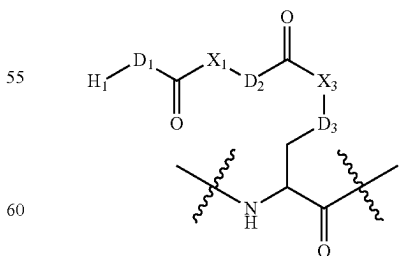
[formula 6-2]

(SEQ ID NO: 15)



wherein, each Xaa is independently any amino acid residue that is not a cysteine residue, C is a cysteine residue, H is a histidine residue, G is a glycine residue, Xa₂ is a glutamic acid residue or an asparagine residue, L is a leucine residue, V is a valine residue, Xa₃ is selected from a tryptophan residue, a naphthylalanine residue, and a phenylalanine residue, and

(Xa₁)' is



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wherein H₁ is a first click chemistry functional group, D₁ is selected from a covalent bond, C₁₋₄alkylene, C₂₋₄alkenylene, C₂₋₄alkynylene, and C₃₋₈-cycloalkylene, X₁ is S, D₂ is selected from C₁₋₇alkylene, C₂₋₇alkenylene,

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C_{2-7} alkynylene, and C_{3-8} -cycloalkynylene, D_3 is a covalent bond, or C_{1-3} alkylene, and X_3 is NH, wherein the peptide consists of 13 to 17 amino acid residues, wherein the peptide exhibits the activity of binding to human immunoglobulin G (IgG), and wherein the cysteine residue that is between two to four amino acids from the N-terminus of formula 6-2 and the cysteine residue that is between two to four amino acids from the C-terminus of formula 6-2 are optionally linked.

Also, the present application provides the peptide-compound conjugate wherein a distance from a beta carbon of the $(Xa_1)'$ to a first carbonyl carbon of the $(Xa_1)'$ is less than approx. 11.668 Å.

Additionally, the present application provides the peptide-compound conjugate wherein D_2 is C_y alkylene, C_y alkenylene, or C_y alkynylene, D_3 is C_x alkylene, wherein y is an integer greater than or equal to 1, and $1 \leq x+y \leq 5$.

Additionally, the present application provides the peptide-compound conjugate wherein a distance from a beta carbon of the $(Xa_1)'$ to a first carbonyl carbon of the $(Xa_1)'$ is greater than approx. 16.208 Å.

Additionally, the present application provides the peptide-compound conjugate wherein D_2 is C_y alkylene, C_y alkenylene, or C_y alkynylene, and D_3 is C_x alkylene, wherein y is an integer greater than or equal to 1, and $9 \leq x+y \leq 12$. Furthermore, the present application provides the peptide-compound conjugate wherein D_2 is C_y alkenylene, or C_y alkynylene.

Additionally, the present application provides the peptide-compound conjugate wherein a distance from a beta carbon of the $(Xa_1)'$ to a first carbonyl carbon of the $(Xa_1)'$ is approx. 11.668 Å to approx. 16.208 Å.

Additionally, the present application provides the peptide-compound conjugate wherein D_2 is C_y alkylene, C_y alkenylene, or C_y alkynylene, and D_3 is C_x alkylene, wherein y is an integer greater than or equal to 1, and $6 \leq x+y \leq 8$.

Additionally, the present application provides the peptide-compound conjugate wherein the formula 6-2 is formula 6-3:

[formula 6-3]

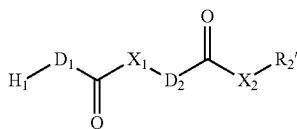
(SEQ ID NO: 16)

D-C-A-W-H-(Xa_1)'-G-E-L-V-W-C-T

wherein, D is an aspartic acid residue, A is an alanine residue, E is a glutamic acid residue, W is a tryptophan residue, T is a threonine residue.

Additionally, the present application provides the peptide-compound conjugate wherein D_1 is a covalent bond and D_2 is methylene.

The present application provides a method for preparing an agent for transferring a first click chemical functional group to an antibodycomprising reacting a compound of formula 2:



wherein, H_1 is a first click chemistry functional group, D_1 is selected from a covalent bond, C_{1-4} alkylene, C_{2-4} alkenylene, C_{2-4} alkynylene, and C_{3-8} -cycloalkynylene, X_1 is S, D_2 is selected from C_{1-7} alkylene, C_{2-7} alkenylene,

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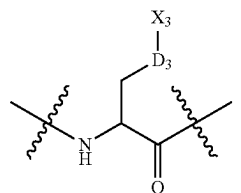
C_{2-7} alkynylene, and C_{3-8} -cycloalkynylene, X_2 is O, R_2' is N-succinimide, p-nitrophenyl, or pentafluorophenyl, with a peptide of formula 4-2:

[formula 4-2]

(SEQ ID NO: 6)

 $(Xaa)_1-C-(Xaa)_2-H-Xa^1-G-Xa_2-L-V-Xa_3-C-(Xaa)_1-3$

wherein, each Xaa is independently any amino acid residue that is not a cysteine residue, C is a cysteine residue, H is a histidine residue, G is a glycine residue, Xa_2 is a glutamic acid residue or an asparagine residue, L is a leucine residue, V is a valine residue, Xa_3 is selected from a tryptophan residue, a naphthylalanine residue, and a phenylalanine residue, and

 Xa_1 is

wherein D_3 is a covalent bond or C_{1-3} alkylene and X_3 is NH_2 , wherein the peptide consists of 13 to 17 amino acid residues, wherein the peptide exhibits the activity of binding to human immunoglobulin G (IgG), and wherein the cysteine residue that is between two to four amino acids from the N-terminus of formula 4-2 and the cysteine residue that is between two to four amino acids from the C-terminus of formula 4-2 are optionally linked.

Also, the present application provides the method for preparing an agent for transferring a first click chemical functional group to an antibodywherein D_2 is C_y alkylene, C_y alkenylene, or C_y alkynylene, and D_3 is C_x alkylene, wherein y is an integer greater than or equal to 1, and $1 \leq x+y \leq 5$, characterized in that the agent for transferring a first click chemical functional group to an antibodyprepared by the method can react with an antibody to deliver the first click chemistry functional group specifically to a 248 a lysine residue of an Fc domain of the antibody.

Additionally, the present application provides the method for preparing an agent for transferring a first click chemical functional group to an antibodywherein D_2 is C_y alkylene, C_y alkenylene, or C_y alkynylene, and D_3 is C_x alkylene, wherein y is an integer greater than or equal to 1, and $1 \leq x+y \leq 5$, characterized in that the agent for transferring a first click chemical functional group to an antibodyprepared by the method can react with an antibody to deliver the first click chemistry functional group specifically to a 248 a lysine residue of an Fc domain of the antibody.

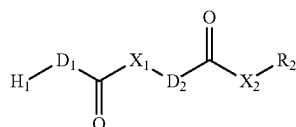
Additionally, the present application provides the method for preparing an agent for transferring a first click chemical functional group to an antibodywherein D_2 is C_y alkylene, C_y alkenylene, or C_y alkynylene, and D_3 is C_x alkylene, wherein y is an integer greater than or equal to 1, and $9 \leq x+y \leq 12$, characterized in that the agent for transferring a first click chemical functional group to an antibodyprepared by the method can react with an antibody to deliver the first click chemistry functional group specifically to a 246 a lysine residue of an Fc domain of the antibody.

Additionally, the present application provides the method for preparing an agent for transferring a first click chemical

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functional group to an antibody wherein D_2 is C_y alkylene, C_y alkenylene, or C_y alkynylene, and D_3 is C_x alkylene, wherein y is an integer greater than or equal to 1, and $6 \leq x+y \leq 8$, characterized in that the agent for transferring a first click chemical functional group to an antibody prepared by the method can react with an antibody to deliver the first click chemistry functional group selectively to a 246 lysine residue or a 248 lysine residue of an Fc domain of the antibody

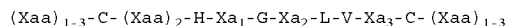
The present application provides a kit for preparing a first click chemistry functional group transferring to an antibody comprising a compound of formula 2:



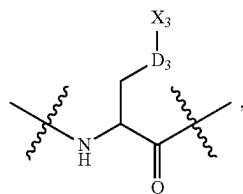
wherein, H_1 is a first click chemistry functional group, D_1 is selected from a covalent bond, C_{1-4} alkylene, C_{2-4} alkenylene, C_{2-4} alkynylene, and C_{3-8} cycloalkynylene, X_1 is S, D_2 is selected from C_{1-7} alkylene, C_{2-7} alkenylene, C_{2-7} alkynylene, and C_{3-8} cycloalkynylene, X_2 is O, R_2' is N-succinimide, p-nitrophenyl, or pentafluorophenyl; and a peptide of formula 4-2:

[formula 4-2]

(SEQ ID NO: 6)



wherein, each Xaa is independently any amino acid residue that is not a cysteine residue, C is a cysteine residue, H is a histidine residue, G is a glycine residue, Xa_2 is a glutamic acid residue or an asparagine residue, L is a leucine residue, V is a valine residue, Xa_3 is selected from a tryptophan residue, a naphthylalanine residue, and a phenylalanine residue, and

 Xa_1 is

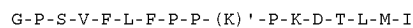
wherein D_3 is a covalent bond or C_{1-3} alkylene and X_3 is NH_2 , wherein the peptide consists of 13 to 17 amino acid residues, wherein the peptide exhibits the activity of binding to human immunoglobulin G (IgG), and wherein the cysteine residue that is between two to four amino acids from the N-terminus of formula 4-2 and the cysteine residue that is between two to four amino acids from the C-terminus of formula 4-2 are optionally linked.

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The present application provides an antibody or a fragment thereof comprising one or more amino acid sequence selected from formula 8-1, formula 8-2, and formula 8-3:

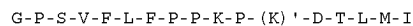
[formula 8-1]

(SEQ ID NO: 20)



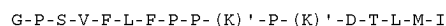
[formula 8-2]

(SEQ ID NO: 21)

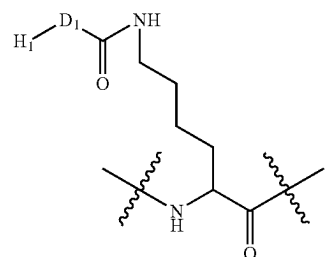


[formula 8-3]

(SEQ ID NO: 22)



wherein, G is a glycine residue, P is a proline residue, S is a serine residue, V is a valine residue, F is a phenylalanine residue, L is a leucine residue, K is a lysine residue, D is an aspartic acid residue, T is a threonine residue, M is a methionine residue, I is an isoleucine residue, and

 $(K)'$ is

wherein H_1 is a first click chemistry functional group, D_1 is selected from a covalent bond, C_{1-4} alkylene, C_{2-4} alkenylene, C_{2-4} alkynylene, and C_{3-8} cycloalkynylene.

Also, the present application provides the antibody or a fragment thereof wherein D_1 is a covalent bond.

Additionally, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 8-1 and not comprising the amino acid sequence of formula 8-2 and 8-3. Furthermore, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 8-1 in both two Fc domains.

Additionally, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 8-2 and not comprising the amino acid sequence of formula 8-1 and 8-3. Furthermore, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 8-2 in both two Fc domains

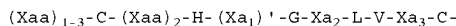
Additionally, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 8-3 and not comprising the amino acid sequence of formula 8-1 and 8-2. Furthermore, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 8-3 in both two Fc domains.

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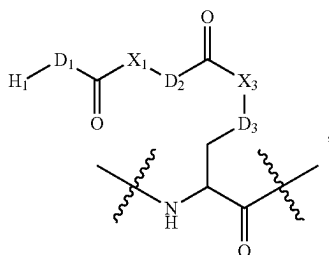
The present application provides a method for preparing an antibody or a fragment thereof comprising first click chemistry functional group comprising reacting a peptide-compound conjugate of formula 6-2:

[formula 6-2]

(SEQ ID NO: 15)



wherein, each Xaa is independently any amino acid residue that is not a cysteine residue, C is a cysteine residue, H is a histidine residue, G is a glycine residue, Xa₂ is a glutamic acid residue or an asparagine residue, L is a leucine residue, V is a valine residue, Xa₃ is selected from a tryptophan residue, a naphthylalanine residue, and a phenylalanine residue, and

(Xa₁)' is

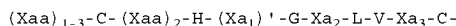
wherein H₁ is a first click chemistry functional group, D₁ is selected from a covalent bond, C₁₋₄alkylene, C₂₋₄alkenylene, C₂₋₄alkynylene, and C₃₋₈-cycloalkynylene, X₁ is S, D₂ is selected from C₁₋₇alkylene, C₂₋₇alkenylene, C₂₋₇alkynylene, and C₃₋₈-cycloalkynylene, D₃ is a covalent bond or C₁₋₃alkylene and X₃ is NH, wherein the peptide consists of 13 to 17 amino acid residues, wherein the peptide exhibits the activity of binding to human immunoglobulin G (IgG), and wherein the cysteine residue that is between two to four amino acids from the N-terminus of formula 6-2 and the cysteine residue that is between two to four amino acids from the C-terminus of formula 6-2 are optionally linked, with an antibody or a fragment thereof.

The present application provides a kit for preparing an antibody or a fragment thereof comprising first click chemistry functional group comprising

a peptide-compound conjugate of formula 6-2:

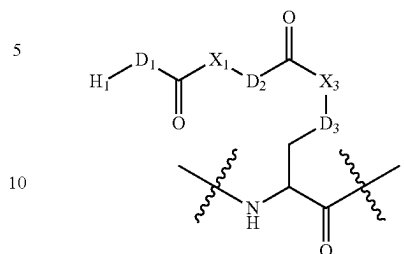
[formula 6-2]

(SEQ ID NO: 15)



wherein, each Xaa is independently any amino acid residue that is not a cysteine residue, C is a cysteine residue, H is a histidine residue, G is a glycine residue, Xa₂ is a glutamic acid residue or an asparagine residue, L is a leucine residue, V is a valine residue, W is a tryptophan residue, and

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(Xa₁)' is

wherein H₁ is a first click chemistry functional group, D₁ is selected from a covalent bond, C₁₋₄alkylene, C₂₋₄alkenylene, C₂₋₄alkynylene, and C₃₋₈-cycloalkynylene, X₁ is S, D₂ is selected from C₁₋₇alkylene, C₂₋₇alkenylene, C₂₋₇alkynylene, and C₃₋₈-cycloalkynylene, D₃ is a covalent bond or C₁₋₃alkylene and X₃ is NH, wherein the peptide consists of 13 to 17 amino acid residues, wherein the peptide exhibits the activity of binding to human immunoglobulin G (IgG), and wherein the cysteine residue that is between two to four amino acids from the N-terminus of formula 6-2 and the cysteine residue that is between two to four amino acids from the C-terminus of formula 6-2 are optionally linked; and an antibody or a fragment thereof.

The present application provides a compound of formula 9:



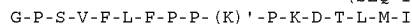
[formula 9]

wherein, C_m is a cargo moiety, H₂ is a second click chemistry functional group.

The present application provides a method for preparing an antibody-drug conjugate comprising reacting an antibody or a fragment thereof comprising one or more amino acid sequence selected from formula 8-1, formula 8-2, and formula 8-3:

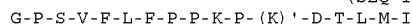
[formula 8-1]

(SEQ ID NO: 20)



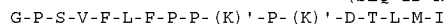
[formula 8-2]

(SEQ ID NO: 21)



[formula 8-3]

(SEQ ID NO: 22)

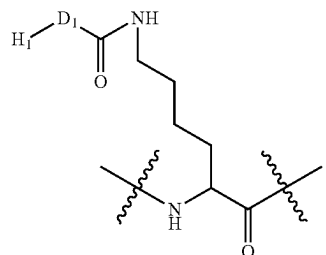


wherein, G is a glycine residue, P is a proline residue, S is a serine residue, V is a valine residue, F is a phenylalanine residue, L is a leucine residue, K is a lysine residue, D is an aspartic acid residue, T is a threonine residue, M is a methionine residue, I is an isoleucine residue, and

(K)' is

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wherein H_1 is a first click chemistry functional group, D_1 is selected from a covalent bond, C_{1-4} alkylene, C_{2-4} alkenylene, C_{2-4} alkynylene, and C_{3-8} cycloalkynylene, with a compound of formula 9:



wherein, C_m is a cargo moiety, H_2 is a second click chemistry functional group which is complementary with the first click chemistry functional group.

The present application provides a kit for preparing an antibody-drug conjugate comprising an antibody or a fragment thereof comprising one or more amino acid sequence selected from formula 8-1, formula 8-2, and formula 8-3:

[formula 8-1] (SEQ ID NO: 20)

G-P-S-V-F-L-F-P-P- (K)' -P-K-D-T-L-M-I

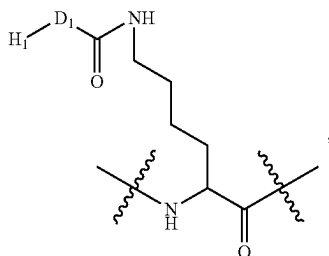
[formula 8-2] (SEQ ID NO: 21)

G-P-S-V-F-L-F-P-P-K-P- (K)' -D-T-L-M-I

[formula 8-3] (SEQ ID NO: 22)

G-P-S-V-F-L-F-P-P- (K)' -P- (K)' -D-T-L-M-I

wherein, G is a glycine residue, P is a proline residue, S is a serine residue, V is a valine residue, F is a phenylalanine residue, L is a leucine residue, K is a lysine residue, D is an aspartic acid residue, T is a threonine residue, M is a methionine residue, I is an isoleucine residue, and (K)' is



wherein H_1 is a first click chemistry functional group, D_1 is selected from a covalent bond, C_{1-4} alkylene, C_{2-4} alkenylene, C_{2-4} alkynylene, and C_{3-8} cycloalkynylene; and a compound of formula 9:



wherein, C_m is a cargo moiety, H_2 is a second click chemistry functional group which is complementary with the first click chemistry functional group.

The present application provides an antibody or a fragment thereof comprising one or more amino acid sequence selected from formula 10-1, formula 10-2, and formula 10-3:

[formula 10-1] (SEQ ID NO: 23)

G-P-S-V-F-L-F-P-P- (K)" -P-K-D-T-L-M-I

[formula 10-2] (SEQ ID NO: 24)

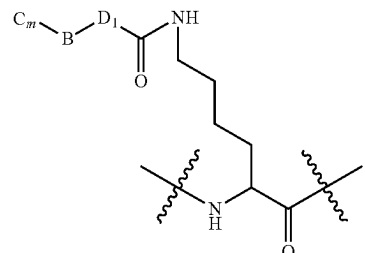
G-P-S-V-F-L-F-P-P-K-P- (K)" -D-T-L-M-I

[formula 10-3] (SEQ ID NO: 25)

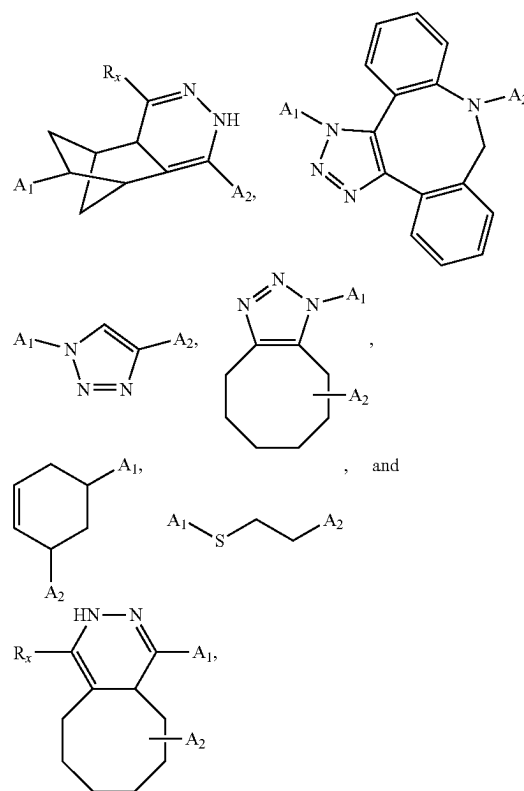
G-P-S-V-F-L-F-P-P- (K)" -P- (K)" -D-T-L-M-I

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wherein, G is a glycine residue, P is a proline residue, S is a serine residue, V is a valine residue, F is a phenylalanine residue, L is a leucine residue, K is a lysine residue, D is an aspartic acid residue, T is a threonine residue, M is a methionine residue, I is an isoleucine residue, and (K)" is



wherein C_m is a cargo moiety, B is selected from



wherein A_1 and A_2 is connected to the cargo moiety or D_1 and they are not both connected to same, R_x is selected from H, halogen, and C_{1-3} alkyl, D_1 is selected from a covalent bond, C_{1-4} alkylene, C_{2-4} alkenylene, C_{2-4} alkynylene, and C_{3-8} cycloalkynylene.

Also, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 10-1 and not comprising the amino acid sequence of formula 10-2 and 10-3. Furthermore, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 10-1 in both two Fc domains.

Additionally, the present application provides the antibody or a fragment thereof comprising the amino acid

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sequence of formula 10-2 and not comprising the amino acid sequence of formula 10-1 and 10-3. Furthermore, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 10-2 in both two Fc domains.

Additionally, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 10-3 and not comprising the amino acid sequence of formula 10-1 and 10-2. Furthermore, the present application provides the antibody or a fragment thereof comprising the amino acid sequence of formula 10-3 in both two Fc domains.

Additionally, the present application provides the antibody or a fragment thereof wherein the cargo moiety comprises a drug moiety. Further, the present application provides the antibody or a fragment thereof wherein the cargo moiety comprises more than one drug moiety. Or further, the present application provides the antibody or a fragment thereof wherein the drug moiety is an anticancer agent. Furthermore, the present application provides the antibody or a fragment thereof wherein the anticancer agent is at least one selected from DM1, DM3, DM4, abrin, ricin A, *pseudomonas* exotoxin, cholera toxin, diphtheria toxin, tumor necrosis factor, α -interferon, β -interferon, nerve growth factor, platelet derived growth factor, tissue plasminogen activator, a cytokine, an apoptotic agent, an anti-angiogenic agent, a lymphokine, taxane, a DNA-alkylating agent, anthracycline, tubulysin analogs, duocarmycin analogs, auristatin E, auristatin F, maytansinoid, a cytotoxic agent comprising a reactive polyethylene glycol moiety, taxon, cytochalasin B, gramicidin D₁, ethidium bromide, emetine, mitomycin, etoposide, teniposide, vincristine, vinblastine, t. colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoid, procaine, tetracaine, lidocaine, propranolol, puromycin, methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine, mechlorethamine, thiotepachlorambucil, meiphalan, carmustine, lomustine, cyclophosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, cisplatin, dactinomycin, bleomycin, anthramycin, calicheamicin, Gemcitabine, bendamustine, bortezomib, carboplatin, cabazitaxel, dasatinib, docetaxel, epirubicin, erlotinib, everolimus, gemcitabine, gefitinib, idarubicin, imatinib, hydroxyurea, lapatinib, leuporelin, melphalan, nedaplatin, nilotinib, oxaliplatin, pazopanib, pemetrexed, picoplatin, romidepsin, satraplatin, sorafenib, vemurafenib, sunitinib, teniposide, triplatin, and vinorelbine.

The present application provides a pharmaceutical composition for treating cancer comprising aforementioned antibody-drug conjugate

Also, the present application provides the pharmaceutical composition for treating cancer wherein the cancer is selected from bladder cancer, bone cancer, brain tumor, breast cancer, heart cancer, cervical cancer, colorectal cancer, rectal cancer, esophageal cancer, fibrosarcoma, gastric cancer, gastrointestinal cancer, head and neck cancer, Kaposi sarcoma, renal cancer, leukemia, liver cancer, lung cancer, lymphoma, melanoma, myeloma, ovarian cancer, pancreatic cancer, penile cancer, prostate cancer, testicular germ cell cancer, thymoma and thymic carcinoma.

Additionally, the present application provides the pharmaceutical composition for treating cancer wherein the cancer is breast cancer.

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The present application provides a method for treating cancer comprising administering a pharmaceutical composition comprising aforementioned antibody-drug conjugate to a subject.

Also, the present application provides the method for treating cancer wherein the cancer is selected from bladder cancer, bone cancer, brain tumor, breast cancer, heart cancer, cervical cancer, colorectal cancer, rectal cancer, esophageal cancer, fibrosarcoma, gastric cancer, gastrointestinal cancer, head and neck cancer, Kaposi sarcoma, renal cancer, leukemia, liver cancer, lung cancer, lymphoma, melanoma, myeloma, ovarian cancer, pancreatic cancer, penile cancer, prostate cancer, testicular germ cell cancer, thymoma and thymic carcinoma.

Furthermore, the present application provides the method for treating cancer wherein the cancer is breast cancer.

Advantageous Effects

An antibody product according to the present invention can have a certain number of chemical functional groups or cargo moieties labeled at a certain site thereof. Therefore, the present invention can provide an antibody product having high uniformity. Also, the present invention can provide an antibody product whose antibody functions are not degraded. That is, the present invention can provide an antibody product whose antibody binding affinity and half-life are not degraded. The present invention is of great significance as being the first technology allowing site-specific labeling of an antibody without any complicated processes.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows a partial sequence of an Fc domain of SEQ ID NO: 1 and numbers in the sequence numbered according to the EU numbering system.

FIG. 2 shows positions of lysine residues in an Fc domain including lysine 246 and 248.

FIG. 3 and FIG. 4 show the topology between SSFI and the Fc domain.

FIG. 5 shows Xa₁ of the SSFI and lysine 246 and 248 in the Fc domain.

FIG. 6 shows a distance between an amine group of the lysine 246 and a beta carbon of the Xa₁ of the SSFI.

FIG. 7 shows a distance between an amine group of the lysine 248 and a beta carbon of the Xa₁ of the SSFI.

FIG. 8 shows a structure of R₁'-L₂-SSFI and a distance (L_c) between a beta carbon and a first carbonyl carbon of (Xa₁)'.

FIG. 9 shows the topology between the R₁'-L₂-SSFI and the Fc domain so that a side chain of (Xa₁)' has a direction (a dotted arrow) parallel with the x axis on the drawing.

FIG. 10 shows the conditions used to allow the first carbonyl carbon to react well with lysine 248.

FIG. 11 shows the conditions used to allow the first carbonyl carbon to react well with lysine 246.

FIG. 12 shows the conditions used to allow the first carbonyl carbon to selectively react with lysine 246 or 248.

FIG. 13 shows an FcRn binding site and lysine 246 and 248 in the Fc domain.

FIG. 14 shows the analysis of a binding structure between SSFI and the Fc domain in comparison with a binding site between Fc and of FcRn.

FIG. 15 shows a method of synthesizing Compound I.

FIG. 16 shows the results of confirming a structure of Compound I by means of mass spectrometry.

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FIG. 17 shows a method of synthesizing Compound II.
 FIG. 18 shows the results of confirming a structure of Compound II by means of mass spectrometry.
 FIG. 19 shows a method of synthesizing Compound III.
 FIG. 20 shows the results of confirming a structure of Compound III by means of mass spectrometry.
 FIG. 21 and FIG. 22 show a method of synthesizing Compound IV.
 FIG. 23 shows the results of confirming a structure of Compound IV by means of mass spectrometry.
 FIG. 24 shows the results of confirming a structure of SSFI (6Lys) by means of mass spectrometry.
 FIG. 25 shows the results of confirming a structure of SSFI (6Orn) by means of mass spectrometry.
 FIG. 26 shows the results of confirming a structure of SSFI (6Dab) by means of mass spectrometry.
 FIG. 27 shows the results of confirming a structure of SSFI (6Dap) by means of mass spectrometry.
 FIG. 28 shows the results of confirming a structure of DD2 by means of mass spectrometry.
 FIG. 29 shows the results of confirming a structure of DD3 by means of mass spectrometry.
 FIG. 30 shows the results of confirming a structure of DD4 by means of mass spectrometry.
 FIG. 31 shows the results of confirming a structure of DD5 by means of mass spectrometry.
 FIG. 32 shows the results of confirming a structure of DD6 by means of mass spectrometry.
 FIG. 33 shows the results of confirming a structure of Compound I-SSFI (6Lys) by means of mass spectrometry.
 FIG. 34 shows the results of confirming a structure of Compound II-SSFI (6Lys) by means of mass spectrometry.
 FIG. 35 shows the results of confirming a structure of Compound III-SSFI (6Lys) by means of mass spectrometry.
 FIG. 36 shows the results of confirming a structure of Compound III-SSFI (6Orn) by means of mass spectrometry.
 FIG. 37 shows the results of confirming a structure of Compound III-SSFI (6Dab) by means of mass spectrometry.
 FIG. 38 shows the results of confirming a structure of Compound III-SSFI (6Dap) by means of mass spectrometry.
 FIG. 39 shows the results of confirming a structure of Compound IV-SSFI (6Dap) by means of mass spectrometry.
 FIG. 40 shows the results of observing a binding reaction using trastuzumab and Compound I-SSFI (6Lys) by means of HIC-HPLC.
 FIG. 41 shows the results of observing a binding reaction using trastuzumab and Compound II-SSFI (6Lys) by means of HIC-HPLC.
 FIG. 42 shows a reaction of an antibody with Compound III-SSFI (6Dap, Dab, Orn, or Lys).
 FIG. 43 shows a structure of Ab (246/248)-Norbornene as the final product.
 FIG. 44 shows the results of observing a binding reaction using trastuzumab and Compound III-SSFI (6Dap) by means of HIC-HPLC.
 FIG. 45 shows the results of observing a binding reaction using trastuzumab and Compound III-SSFI (6Dab) by means of HIC-HPLC.
 FIG. 46 shows the results of observing a binding reaction using trastuzumab and Compound III-SSFI (6Orn) by means of HIC-HPLC.
 FIG. 47 shows the results of observing a binding reaction using trastuzumab and Compound III-SSFI (6Lys) by means of HIC-HPLC.
 FIG. 48 shows the results of observing a binding reaction using trastuzumab and Compound IV-SSFI (6Dap) by means of HIC-HPLC.

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FIG. 49 and FIG. 50 show an increase of molecular weight spectra by antibody-norbornene binding.
 FIG. 51, FIG. 52, FIG. 53 and FIG. 54 show the MS/MS chromatogram results of trastuzumab and antibody-norbornene complexes.
 FIG. 55 shows the sequence matching results by means of the MS/MS spectrum.
 FIG. 56 shows the mass spectrum of trastuzumab measured to confirm a trastuzumab-azide structure.
 FIG. 57 shows the mass spectrum of a trastuzumab-azide complex measured to confirm the trastuzumab-azide structure.
 FIG. 58 shows the results of confirming a structure of tetrazine-DM1 by means of mass spectrometry.
 FIG. 59 shows a structure of a Compound III-SSFI (6Dap)-based trastuzumab-DM1 conjugate.
 FIG. 60 shows the results of observing a formation reaction of a trastuzumab-DM1 conjugate by means of HIC-HPLC.
 FIG. 61 and FIG. 62 show an increase of molecular weight spectra by norbornene-tetrazine-DM1 binding.
 FIG. 63 shows structures of three designs of payloads bound to an antibody labeled with norbornene.
 FIG. 64 shows the results of analyzing antigen binding affinity of an antibody-drug conjugate.
 FIG. 65, FIG. 66 and FIG. 67 show the results of analyzing serum stability of the antibody-drug conjugate.
 FIG. 68, FIG. 69 and FIG. 70 show the results of evaluating a medicinal effect of the antibody-drug conjugate at a cellular level.
 FIG. 71 and FIG. 72 show the results of evaluating a medicinal effect of the antibody-drug conjugate at an animal level.
 FIG. 73 shows the results of a pharmacokinetics test on the antibody-drug conjugate.

DETAILED DESCRIPTION

Definitions

Unless otherwise defined, all technical and scientific terms used in the present invention have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. The following references are provided to give a number of common definitions of the terms used in the present invention to those skilled in the related art: Singleton et al., Dictionary of Microbiology and Molecular Biology (2nd ed. 1994); The Cambridge Dictionary of Science and Technology (Walker ed., 1988); The Glossary of Genetics, 5th Ed., R. Rieger et al. (eds.), Springer Verlag (1991); and Hale & Marham, The Harper Collins Dictionary of Biology (1991). Unless otherwise clearly specified, the following terms used in the present invention shall have the meanings ascribed to them as follows.

In some embodiments, a chemical structure is disclosed together with the corresponding chemical name. When the terms contradict or conflict with each other, the chemical structure takes precedence over the chemical name to understand the meaning of a compound through the chemical structure.

The term "hetero" used in the present invention refers to a compound or a group of compounds including at least one heteroatom. The term "heteroatom" refers to an atom other than a carbon or hydrogen atom, and, for example, includes B, Si, N, P, O, S, and Se. Preferably, the heteroatom includes, among others, polyvalent elements such as N, O, and S, or

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monovalent elements such as F, Cl, Br, and I, but the present invention is not limited thereto.

The term “lower” used in the present invention is used to modify hydrocarbons, for example, alkylenes, and the like, and thus means that the corresponding hydrocarbon has 6 or less carbon atoms. For example, a C₁₋₆ linear or branched alkyl group are referred to by another name such as a “lower alkyl” group.

The term “oxy” used in the present invention refers to a secondary radical (—O—) of an oxygen atom.

The term “alkyl” or “alkane” refers to a linear or branched non-aromatic hydrocarbon that is completely saturated. Unless otherwise defined, a linear or branched alkyl group typically has a 1 to approximately 20 carbon atoms, preferably 1 to approximately 10 carbon atoms. The linear and branched alkyl group includes methyl, ethyl, n-propyl, isopropyl, n-butyl, sec-butyl, tert-butyl, pentyl, hexyl, pentyl, and octyl.

The term “alkenyl” or “alkene” refers to a linear or branched non-aromatic hydrocarbon that contains at least one double bond. Unless otherwise defined, a linear or branched alkenyl group typically has 1 to approximately 20 carbon atoms, preferably 1 to approximately 10 carbon atoms.

The term “alkynyl” or “alkyne” refers to a linear or branched non-aromatic hydrocarbon that contains at least one triple bond. Unless otherwise defined, a linear or branched alkynyl group typically has 1 to approximately 20 carbon atoms, preferably 1 to approximately 10 carbon atoms.

The term “cycloalkane” or “cycloalkyl” group refers to a completed saturated cyclic hydrocarbon. The “cycloalkyl” includes monocyclic and polycyclic rings. Unless otherwise defined, a monocyclic cycloalkyl group generally has 3 to approximately 10 carbon atoms, more generally 3 to 8 carbon atoms. Rings other than the first ring of the polycyclic cycloalkyl may be selected from saturated, unsaturated, and aromatic rings. The cycloalkyl includes a bicyclic molecule that contains 1, 2, or 3 or more atoms shared between two rings. The term “fused cycloalkyl” refers to a polycyclic cycloalkyl in which one ring shares two adjacent atoms with another ring. Rings other than the first ring of the fused polycyclic cycloalkyl may be selected from saturated, unsaturated, and aromatic rings.

The term “cycloalkyne” or “cycloalkynyl” refers to a cyclic hydrocarbon that contains at least one triple bond, which is also referred to as a “strained alkyne.” The “cycloalkynyl” includes monocyclic and polycyclic rings. Unless otherwise defined, a monocyclic cycloalkynyl generally has 3 to approximately 10 carbon atoms, more generally 3 to 8 carbon atoms. Rings other than the first ring of the polycyclic cycloalkynyl may be selected from saturated, unsaturated and aromatic rings. The cycloalkynyl is a bicyclic molecule that contains 1, 2, or 3 or more atoms shared between two rings. The term “fused cycloalkynyl” refers to a polycyclic cycloalkynyl in which one ring shares two adjacent atoms with another ring. Rings other than the first ring of the fused polycyclic cycloalkynyl may be selected from saturated, unsaturated, and aromatic rings.

The term “alkylene” used as a molecule itself or used as part of another molecule refers to a divalent radical derived from an alkane. For example, the group includes —CH₂CH₂—, and —CH₂CH₂CH₂CH₂—, all of which contain 10 or less carbon atoms, but the present invention is not limited thereto. The term “lower alkylene” refers to a short alkylene group that generally has 6 or less carbon atoms.

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Unless stated otherwise, the term “alkylene” is intended to encompass groups represented by the “heteroalkylene” in the present invention.

The term “heteroalkylene” refers to a divalent radical derived from a heteroalkyl, and, for example, includes —CH₂—CH₂—S—CH₂—CH₂—, and —CH₂—S—CH₂—CH₂—NH—CH₂—, but the present invention is not limited thereto. A heteroalkylene group may contain the same or different heteroatoms at each end or all ends of a chain thereof (including an alkyleneoxy, an alkyleneedioxy, an alkyleneamino, an alkylene diamino, an aminooxyalkylene, and the like, but the present invention is not limited thereto). Further, an indication of connection of both ends of the chain is independent of the arrangement of groups of a formula. For example, the formula —C(O)₂R'— refers to both —C(O)₂R'— and —R'(O)₂C—.

The term “alkenylene” used as a molecule itself or used as part of another molecule refers to a divalent radical derived from an alkene. For example, the group includes —CH=CH—, —CH₂CH=CHCH₂—, and —CH=CH—CH=CH—, all of which have 10 or less carbon atoms, but the present invention is not limited thereto. Unless stated otherwise, the term “alkenylene” is intended to encompass heteroalkenylenes in the present invention.

The term “alkynylene” used as a molecule itself or used as part of another molecule refers to a divalent radical derived from an alkyne. For example, the group includes —C≡C—, —CH₂C≡CCH₂—, and —C≡C—C≡C—, all of which have 10 or less carbon atoms, but the present invention is not limited thereto. Unless stated otherwise, the term “alkynylene” is intended to encompass heteroalkynylenes in the present invention.

The term “cycloalkylene” used as a molecule itself or used as part of another molecule refers to a divalent radical derived from a cycloalkene. Unless stated otherwise, the “cycloalkylene” is intended to encompass heterocycloalkylenes in the present invention.

The term “alkylene” used in this specification, examples, and the claims is intended to encompass both “unsubstituted alkylene” and “substituted alkylene.” Among these, the latter refers to an alkylene group that has a substituent replacing a hydrogen atom on one or more carbon atoms of the hydrocarbon. Unless otherwise clearly specified, the substituent may, for example, include a halogen, a hydroxyl group, a carbonyl group (for example, carboxyl, alkoxycarbonyl, formyl, or acyl), a thiocarbonyl group (for example, thioester, thioacetate, or thioformate), an alkoxy group, a phosphoryl group, a phosphate group, a phosphonate group, a phosphinate group, an amino group, an amido group, an amidine group, an imine group, a cyano group, a nitro group, an azido group, a sulfhydryl group, an alkylthio group, a sulfate group, a sulfonate group, a sulfamoyl group, a sulfonamido group, a sulfonyl group, a heterocyclyl group, an aralkyl group, or an aromatic or heteroaromatic group. When properly substituted, it may be understood by those skilled in the related art that a substituted residue on a hydrocarbon chain may itself be substituted. For example, the substituent of the substituted alkylene may include substituted and unsubstituted amino, azido, imino, amido, phosphoryl (including phosphonates and phosphinates), sulfonyl (including sulfates, sulfonamidos, sulfamoyls, and sulfonates), and silyl groups, and may also include ethers, alkylthios, carbonyls (including ketones, aldehydes, carboxylates, and esters), —CF₃, —CN, and equivalents thereof. Exemplary substituted alkyls are as described below. The cycloalkylene may be further substituted with an alkyl, an alkenyl, an alkoxy, an alkylthio, an aminoalkyl, a

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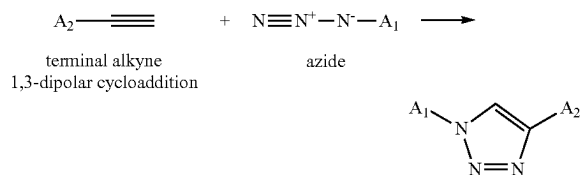
carbonyl-substituted alkyl, $-\text{CF}_3$, $-\text{CN}$, and equivalents thereof. This content is also equally applicable to the alkenylene and the alkynylene.

When used together with a residue such as an alkylene, an alkenylene, or an alkynylene, the term " C_{x-y} " is, for example, intended to encompass residues containing x to y carbon atoms in the chain thereof. For example, the term " C_{x-y} alkylene" refers to a substituted or unsubstituted, linear or branched alkylene group that contains x to y carbon atoms in the chain thereof. For example, it is meant to exemplarily include a haloalkylene group such as difluoromethylene, 2,2,2-trifluoroethylene, and the like. The C_0 alkylene refers to a covalent bond. The terms " C_{2-y} alkenylene" and " C_{2-y} alkynylene" refers to a substituted or unsubstituted unsaturated aliphatic residue to which a definition of length and possible substitution is applied as described in the definition of the alkylene. However, this means that each contains at least one double or triple bond.

The term "click-chemistry reaction" is used as a chemical concept introduced by K. Barry Sharpless of the Scripps Research Institute to explain complementary chemical functional groups and a chemical reaction designed so that two molecules can rapidly and stably form a covalent bond. The click-chemistry reaction does not refer to a certain reaction, but refers to a concept of such a rapid and stable reaction. In any embodiment, the click-chemistry reaction must be modular, wide in scope, give high yields, generate only insignificant by-products, be stereospecific, physiologically stable, driven by a thermodynamic driving force (for example, greater than 84 kJ/mol), and/or have high atom economy. Some reactions are known to satisfy the requirements:

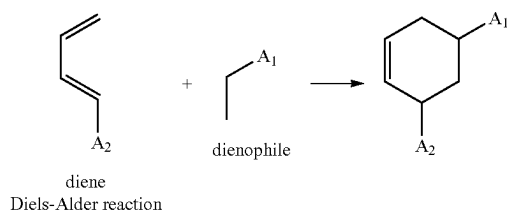
- (1) Huisgen 1,3-dipolar cycloaddition (for example, including a Cu(I)-catalyzed cycloaddition reaction, which is often commonly referred to as a "click reaction"; see Tornøe et al., Journal of Organic Chemistry (2002) 67: 3057-3064): Copper or ruthenium is generally used as a catalyst;

[Schematic diagram of Huisgen 1,3-dipolar cycloaddition]



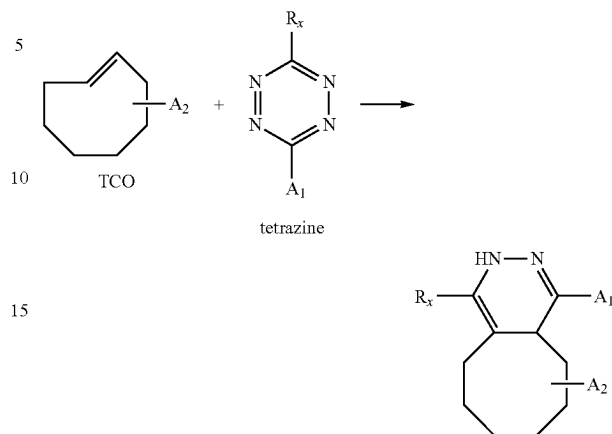
- (2) Diels-Alder reaction, for example cycloaddition (for example, strain-promoted cycloaddition (SPAAC)) including a normal electron-demand Diels-Alder reaction and an inverse electron-demand Diels-Alder reaction, but the present invention is not limited thereto);

[Schematic Diagram of Diels-Alder Reaction]

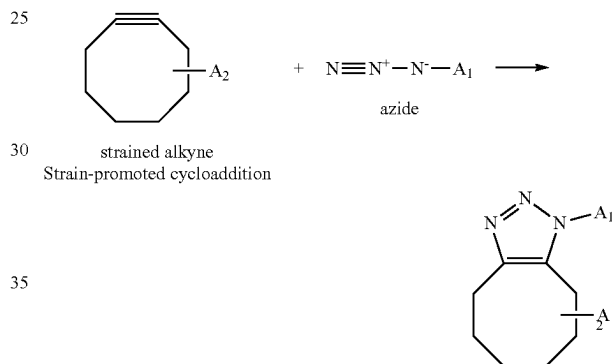


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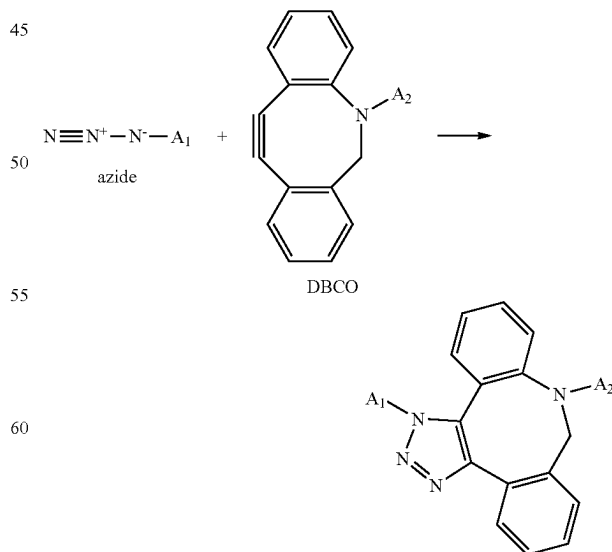
[Example of Diels-Alder Reaction; TCO and Tetrazine]



[Schematic Diagram of Strain-Promoted Cycloaddition]



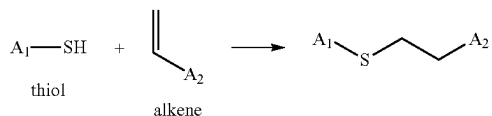
[Example of Strain-Promoted Cycloaddition; Azide and DBCO]



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- (3) Nucleophilic addition to small strained rings such as epoxides and aziridines;
 (4) Nucleophilic addition to activated carbonyl groups;
 (5) Addition to carbon-carbon double bonds or triple bonds.

[Addition of Thiol and Alkene]



The term “click-chemistry functional group” used in the present invention refers to a functional group that participates in a click-chemistry reaction. For example, a strained alkyne (for example, a cyclooctyne) corresponds to a click-chemistry functional group. In general, the click-chemistry reaction requires at least two molecules, each of which contains click-chemistry functional groups complementary to each other. In this way, a pair of click-chemistry functional groups having reactivity with each other is often referred to as “partner click-chemistry functional groups” in the present invention. In strain-promoted cycloaddition of a cyclooctyne with an azide, for example, the azide is a partner click-chemistry functional group for the cyclooctyne and other alkynes. Exemplary click-chemistry functional groups used in the present invention include a terminal alkyne, an azide, a strained alkyne, a diene, a dienophile, a trans-cyclooctene, an alkene, a thiol, and a tetrazine, but the present invention is not limited thereto. Other click-chemistry functional groups are known to those skilled in the related art.

The term “leaving group” used in the present invention has the same concept as well known to those skilled in the related art (Advanced Organic Chemistry: reactions, mechanisms and structure—Fourth Edition of Jerry March, John Wiley and Sons Ed.; 1992, pages 351-357), and refers to a

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chemical functional group linked to any reactant, which moves when the reactant is subjected to a substitution reaction (displacement reaction), for example, a nucleophilic substitution reaction. A good leaving group refers to a leaving group that easily moves during the nucleophilic substitution reaction. Exemplary good leaving groups include a halogen (F, Cl, Br, and I), a tosylate, a mesylate, a triflate, an acetate, a trifluoromethylacetate, a camphor-sulfonate, 2-thioxobenzo[d]thiazol-3(2H)-yl, N-hydrosuccinimide, N-aryloxide, and an aryloxide substituted with one or more electron-withdrawing groups (EWGs), but the present invention is not limited thereto. For example, the aryloxide substituted with one or more electron-withdrawing groups include 2-nitrophenoxide, 4-nitrophenoxide, 2,4-dinitrophenoxide, pentafluorophenoxide, 2-chloro-4-nitrophenoxide, 2,4-dichlorophenoxide, and 2,4,6-chlorophenoxide, and the electron-withdrawing group, for example, includes a halogen (F, Cl, Br, or I), —NO₂, —CN, —C(O)(C₁₋₆ alkyl), —C(O)(aryl), —C(O)O(C₁₋₆ alkyl), —C(O)O(aryl), and the like.

The term “interactome” used in the present invention refers to a protein or a peptide that participates in a protein-protein interaction when there is a protein-protein interaction (PPI) between proteins or between peptides. For example, a chaperone protein and its passenger protein are mutual interactomes. The “protein-protein interaction” means that two or more proteins or peptide molecules interact to come into physical contact with each other with high specificity. In this case, the causative interaction includes an electromagnetic force, a hydrogen bond, and a hydrophobic interaction, but the present invention is not limited thereto.

The term “antibody interactome” used in the present invention refers to an interactome for an antibody including immunoglobulins. Exemplary antibody interactomes are listed in Table 1. Among these, peptides such as Fc-III, and the like are known to have binding activity for an Fc domain of an immunoglobulin. In this case, the peptides are also referred to as “Fc interactomes.”

TABLE 1

List of SSAIs				
Peptide	Binding Constant	Binding Capacity	Elution pH	Remarks
PAM	$K_d = 0.3 \mu\text{M}$	n.a.	3 or 9	Dendrimer comb. library
D-PAM	n.a.	50	3.5	SpA mimic
D-PAM- ϕ	n.a.	10	4	
TKTSRSISIF (SEQ ID NO: 27)	n.a.	n.a.	n.a.	Phage display library
FGRLVSSAIRY (SEQ ID NO: 28)	n.a.	n.a.	n.a.	SpA mimic
Fc-III (DeLano et al.) Fc-III- (Sephacrose)	$K_d = 16 \text{ nM}$	26.6 mg/ μmol	3.5	Phage display cyclic peptide library SpA mimic
FcBP-2	$K_d = 1.8 \text{ nM}$	n.a.	n.a.	Bicyclic peptide
Fc-III-4C	$K_d = 2.5 \text{ nM}$	n.a.	3.5	Bicyclic peptide
EPIHRSTLTALL (SEQ ID NO: 29)	n.a.	320 ug/g	n.a.	Phage display library SpA mimic
APAR (SEQ ID NO: 30)	$K_d = 94 \text{ nM}$	9.1	n.a.	Comb. tetrapeptide library
FcRM	$K_d = 20 \mu\text{M}$	n.a.	2.7	Synthetic cyclic peptide library, Fc γ -receptor mimic

TABLE 1-continued

List of SSAIs				
Peptide	Binding Constant	Binding Capacity	Elution pH	Remarks
HWRGWV (SEQ ID NO: 31)	$K_d = 10 \mu\text{M}$	26.4	4	Comb. library
HYFKFD (SEQ ID NO: 32)	$K_d = 11 \mu\text{M}$	27.0	n.a.	(one-bead-one-peptide)
FRRHL (SEQ ID NO: 33)	$K_d = 26 \mu\text{M}$	33.6	n.a.	SpA mimic
HWCitGWV (SEQ ID NO: 34)	$K_d = 108 \mu\text{M}$	78	n.a.	
D ₂ AAG (SEQ ID NO: 35)	$K_a = 7.9 \times 10^5 \text{M}^{-1}$	36.2	3.6	Comb. library
DAAG (SEQ ID NO: 36)	$K_a = 2.6 \times 10^5 \text{M}^{-1}$	49.6	3.6	SpA mimic
cyclo[(N α -Ac)S(A)-RWHYFK-Lact-E] (SEQ ID NO: 37)	n.a.	n.a.	3.5	Cyclic peptide
cyclo[(N α -Ac)Dap(A)-RWHYFK-Lact-E] (SEQ ID NO: 38)	n.a.	n.a.	3.5	
cyclo[(Link-M-WFRHYK] (SEQ ID NO: 39)	$K_d = 7.6 \mu\text{M}$	19.7	4.0	mRNA display library SpA mimic
NKFRGKYK (SEQ ID NO: 40)	$K_a = 8.6 \times 10^6 \text{M}^{-1}$	DBC-4.9	4.0	Spot peptide array
NARKFYKG (SEQ ID NO: 41)	$K_a = 6.5 \times 10^6 \text{M}^{-1}$	DBC-5.0	4.0	Fcv mimic
FYWHCLDE (1) (SEQ ID NO: 42)	$K_d = 1.5 \mu\text{M}$	104.2	6.0	Biometric design strategy
FYCHWALE (2) (SEQ ID NO: 43)	$K_d = 6.1 \mu\text{M}$	87.6	6.0	Fc-binder (SpA_mimic)
FYCHTIDE (3) (SEQ ID NO: 44)	$K_d = 5.7 \mu\text{M}$	63.7	6.0	Dual affinity ligand
Dual 1/3 (2:1)	$K_d = 0.69 \mu\text{M}$	137.9	6.0	
RRGW (SEQ ID NO: 45)	$K_d = 0.5 \text{nM}$	n.a.	n.a.	Computer design strategy Fc-binder
KHRFNKD (SEQ ID NO: 46)	$K_d = 20 \text{nM}$	n.a.	n.a.	Phage-display library SpA mimic

*Maximal binding capacity (q_m) in mg/mL, DBC: Dynamic binding capacity. The depicted values are towards IgG and may not be directly compared to each other because they were obtained at different conditions or by different methods.
n.a.: Not available.

According to the present invention, the term “antibody”⁴⁰ refers to an immunoglobulin molecule or a fragment thereof. Immunoglobulins are generally well known, and have an ability to specifically bind to a certain antigen. However, because the antibody according to the present invention is a concept that also encompasses a fragment thereof, the antibody does not have to show binding ability to a certain antigen as in case of the Fc fragment. In addition to the naturally occurring immunoglobulins, the antibody is also intended to encompass all recombinant proteins, fusion proteins, chimeric proteins, human immunoglobulins, non-human animal immunoglobulins, and the like, which have the same or similar structures.

According to the present invention, the term “conjugate”⁵⁵ refers to a heterogeneous molecule made when conjugate partners are taken together to form a covalent bond. The covalent bond may be preferably formed by means of a click-chemistry reaction. The term “conjugate partner” refers to each of molecules intended to form a conjugate. In this case, when one wishing to perform conjugation intends to link a certain molecule to any other molecule, the certain molecule may be generally referred to as a “target molecule” or “target protein,” and the any other molecule may be generally referred to as a “cargo molecule” or “cargo moiety” for the sake of convenience.

According to the present invention, the term “carrier moiety” refers to a portion of molecule that constitutes a conjugate, that is, a molecule that functions to enhance serum stability of a molecule linked thereto or extend the half-life of the molecule. Molecules that may be used as the carrier moiety are well known in the related art. Representative examples of the carrier moiety include albumin, gelatin, elastin (including tropoelastin), an elastin-derived polypeptide (α -elastin and elastin-like polypeptides (ELPs)), gliadin, legumin, zein, a soy protein (for example, a soy protein isolate (SPI)), a milk protein, a whey protein, bilirubin, and the like, but the present invention is not limited thereto.

According to the present invention, the term “fluorescent moiety” is intended to encompass dyes or dye reagents used for fluorescence. Molecules that may be used as the dyes and dye reagents are well known in the related art. Representative examples of the fluorescent moiety are as listed in Table 2, but the present invention is not limited thereto (Immuno-tech-Coulter Corp. catalog, “Cytometry Monoclonal Reagent Guide,” 8/95, p.3).

TABLE 2

List of fluorescent moieties				
Fluorochrome	Maximum Absorbance	Excitation at 488 nm	Maximum Emission	Fluorescence
Flourescein	495 nm	Yes	525 nm	Green
isothiocyanate (FITC)				
Phycoerythrin (PE)	488; 565 nm	Yes	575 nm	Orange-red
Energy Coupled Dye [ECD] (PE-Texas-Red)	488; 565 nm	Yes	610-635 nm	Red
Phycoerythrin-Cyanin 5 (PE-Cy5)	488; 565; 652 nm	Yes	670 nm	Deep-red

According to the present invention, the term “drug moiety” refers to a molecule that has a therapeutic effect on any disease. The drug moiety according to the present invention includes those known to a person having ordinary skill in the art as being effective against any disease. Typically, the drug moiety having an anti-cancer effect includes DM1, DM3, DM4, Abrin, Ricin A, a *Pseudomonas* exotoxin, a Cholera toxin, a Diphtheria toxin, a tumor necrosis factor, a interferon, p interferon, a nerve growth factor, a platelet-derived growth factor, a tissue plasminogen activator, a cytokine, an apoptosis-inducing agent, an anti-angiogenic agent, a lymphokine, taxane, a DNA-alkylating agent, anthracyclin, a Tubulysin analogue, a duocarmycin analogue, auristatin E, auristatin F, a maytansinoid, a cytotoxic agent including a reactive polyethylene glycol residue, taxon, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, T. Colchicine, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dihydrotestosterone, a glucocorticoid, procaine, tetracaine, lidocaine, propranolol, puromycin, methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine, mechlorethamine, thiotepe, chlorambucil, Meiphalan, carmustine, lomustine, cyclophosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, cisplatin, dactinomycin, bleomycin, anthramycin, calicheamicin, abiraterone, bendamustine, bortezomib, carboplatin, cabazitaxel, dasatinib, doxorubicin, epirubicin, erlotinib, everolimus, gemcitabine, gefitinib, idarubicin, imatinib, hydroxyurea, lapatinib, leuporelin, melphalan, nedaplatin, nilotinib, oxaliplatin, pazopanib, pemetrexed, picoplatin, romidepsin, satraplatin, sorafenib, vemurafenib, sunitinib, teniposide, triplatin, and vinorelbine, but the present invention is not limited thereto.

According to the present invention, the term “radioactive moiety” refers to a moiety including a radioisotope. Radioisotopic labeling is useful for diagnostic imaging and radiotherapy. Representative radioactive moieties include ^{18}F , ^{11}C , ^{125}I , ^{123}I , ^{124}I , ^{131}I , and $^{99\text{m}}\text{Tc}$, but the present invention is not limited thereto.

The term “pharmaceutically acceptable carrier” may be used in the sense of including an excipient, a diluent, or an adjuvant. The carrier may be, for example, selected from the group consisting of lactose, dextrose, sucrose, sorbitol, mannitol, xylitol, erythritol, maltitol, starch, acacia gum, alginate, gelatin, calcium phosphate, calcium silicate, cellulose, methyl cellulose, polyvinyl pyrrolidone, water, physiological saline, a buffer such as PBS, methylhydroxy benzoate, propylhydroxy benzoate, talc, magnesium stearate, and mineral oil. The carrier may include a filler, an anti-agglomerating agent, a lubricant, a wetting agent, a flavoring agent, an emulsifying agent, a preservative, or a combination thereof.

The term “pharmaceutically acceptable salt” refers to a salt in which biological effectiveness and properties of a proteome and a compound according to the present invention are preserved and that is not undesirable in biological or other aspects. In many cases, the proteome and compound of the present invention may form an acidic and/or basic salt in the presence of a charged group, for example, a charged amino and/or carboxyl group or the likes. A pharmaceutically acceptable acid addition salt may be prepared from an inorganic and organic acid, and a pharmaceutically acceptable base addition salt may be prepared from an inorganic and organic base.

The term “treatment” refers to an approach to obtain beneficial or desirable clinical outcomes. For the object of the present invention, non-limiting examples of the beneficial or desirable clinical outcomes include palliation of symptoms, reduction of a disease range, stabilization (i.e., not worsening) of a disease state, delay of progression or reduction of a progression rate of a disease, (partially or totally) improvement or temporal palliation and mitigation of a disease condition, and whether detectable or not. The treatment denotes all therapeutic treatments, and prophylactic or preventative methods. The treatments include treatments required for disorders to be prevented and already-occurring disorders. “Palliating” a disease means that a range of disease conditions and/or undesirable clinical signs are lowered and/or a time course of progression of the disease is delayed or extended, compared to when the disease is not treated.

The “therapeutically effective amount” (or “effective amount”) refers to a sufficient amount of an active ingredient, for example, an agent according to the present invention, to achieve the treatment when administered to a subject or a patient. Therefore, what constitutes a therapeutically effective amount of a composition according to the present invention may be easily determined by those skilled in the related art. In the context of vision therapy, the “therapeutically effective amount” is an amount that causes an objectively measured change in one or more parameters associated with the treatment of eye diseases or conditions, which include an increase or decrease in expression of one or more genes associated with the eye diseases or conditions, an induction of apoptosis or other cell death pathways, a clinical improvement in symptoms, a decrease in abnormal neovascularization or inflammation, and the like. Of course, the therapeutically effective amount may vary depending on a certain subject and condition to be treated, the weight and age of a subject, the severity of a disease condition, a certain compound to be selected, a subsequent dosing schedule, the administration time adjustment, a mode of administration, and the like, all of which may be easily determined by those skilled in the related art. In the context of combination therapy, it should be understood that what constitutes a

therapeutically effective amount of a certain active ingredient may be different from what constitutes a therapeutically effective amount of an active ingredient that is administered for monotherapy (that is, a treatment regimen using one chemical entity as the active ingredient).

The “subject” or “patient” refers to an animal in need of treatment which may be achieved by the molecule of the present invention. The animal to be treated according to the present invention includes a vertebrate. Particularly preferred examples of the animal include mammals, for example, bovine, canine, equine, feline, ovine, porcine, and primate animals (including human and non-human primates).

The term “about” or “approximately” refers to an amount, level, value, number, frequency, percentage, dimension, size, quantity, weight, or length that varies by 30, 25, 20, 15, 10, 9, 8, 7, 6, 5, 4, 3, 2, or 1% with respect to the reference amount, level, value, number, frequency, percentage, dimension, size, quantity, weight, or length.

1. Antibody

It should be understood that the description of the present invention is intended to provide a description of the structure, academic system, and physiological action of an antibody provided to aid in understanding the present invention, and the antibody falling within the scope of rights of the present invention is not intended to limit the description of the present invention.

An antibody consists of two heavy chains and two light chains as known in the art. When an antibody is simply divided in a functional aspect, the antibody is divided into a fragment antigen-binding variable region (Fab) including a light chain and a fragment crystallizable region composed as a portion of a heavy chain. The Fab includes a paratope that binds to an antigen, and refers to a region that allows an antibody to have specific binding activity for an antigen as known in the art. As the Fc domain is a ligand for an Fc receptor (FcR) in cells, it plays an important role in inducing an immune response. Also, the Fc domain plays an important role in extending the half-life of an antibody by binding to a neonatal Fc receptor so that the antibody can be repeatedly internalized into the cells.

From these facts, it is possible to deduce some desirable directivity of labeling an antibody: (1) First, it is desirable to label an antibody at a position spaced apart from a paratope of the antibody. When the labeling is performed on the paratope or at a position adjacent to the paratope, a binding affinity of the antibody for an antigen may be significantly lowered. (2) Second, it is desirable to label an antibody at a position spaced apart from a recognition site of FcR including FcRn. When the labeling is performed on the recognition site of the receptor or at a position adjacent to the recognition site, an immune response-inducing function of the antibody may be reduced, or the half-life of the antibody may be shortened. Information on the binding activity motif of the Fc domain may be found in DeLano, W. L. (2000): Convergent Solutions to Binding at a Protein-Protein Interface; Science, 287(5456), 1279-1283., and W. Lance Martin et al. (2001), Molecular Cell, Vol. 7, 867-877, April, 2001.

In the present invention, when reference to an amino acid sequence of the Fc domain of the antibody is made, numbers in the sequence are numbered according to the EU numbering system unless stated otherwise. The EU numbering system has been widely used as a sequencing system for the Fc domain after research on the sequence of IgG, as described in Edelman G M, et al., The covalent structure of

an entire gamma-G immunoglobulin molecule; Proc. Natl. Acad. Sci. USA., 1969 May; 63(1): 78-85.

1.1. Search for Desirable Labeling Site

A labeling site of an antibody may be designed in consideration of the criteria for the labeling site, that is, (1) a site spaced apart from a paratope; and (2) a site spaced apart from a recognition site of FcR including FcRn. Examples of the amino acids used in a bioconjugation reaction typically include lysine, cysteine, and tyrosine. Lysine 246 (Lys₂₄₆) and lysine 248 (Lys₂₄₈) present in an Fc domain of the antibody are residues that satisfy all the requirements, and thus are both desirable labeling sites. A sequence of the Fc domain including the Lys₂₄₆ and Lys₂₄₈ residues is GPSVFLFPPKPKDTLMI, and the sequence and numbers in the sequence numbered according to the EU numbering system are shown in FIG. 1 (FIG. 1, SEQ ID NO: 1).

In specific embodiments, the antibody according to the present invention may include a sequence of SEQ ID NO: 1 or derivatives thereof. Also, the antibody according to the present invention may include a derivative of SEQ ID NO: 1 in which lysine 246 is substituted. In addition, the antibody according to the present invention may include a derivative of SEQ ID NO: 1 in which lysine 248 is substituted. Furthermore, the antibody according to the present invention may include a derivative of SEQ ID NO: 1 in which lysine 246 and 248 are substituted.

The sequence of SEQ ID NO: 1 or derivatives thereof include sequences mutated in an acceptable range. In specific embodiments, the mutated sequences may have a homology that is greater than or equal to approximately 90%, approximately 85%, approximately 80%, approximately 75%, or approximately 70% with respect to the sequence of SEQ ID NO: 1 or derivatives thereof. In specific embodiment specified below, it should be understood that the derivatives of SEQ ID NO: 1 represented by Formulas 7-1 to 7-3, 8-1 to 8-3, and 10-1 to 10-3 also include sequences mutated in an acceptable range.

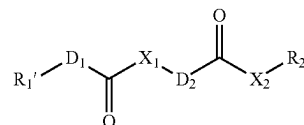
FIG. 2 shows positions of lysine residues in an Fc domain including lysine 246 and 248.

2. Linker (R₁'-L₁)

The present invention discloses a novel compound capable of being used to label an antibody. For the sake of convenience, such a compound is herein referred to as a linker, which is indicated by the symbol “R₁'-L₁.”

The present invention provides a compound having a structure of the following Formula 1:

[Formula 1]



wherein R₁' is a first chemical functional group,

D₁ is any alkylene, alkenylene, or alkynylene,

X₁ is an element that is more electronegative than carbon,

D₂ is any alkylene, alkenylene, or alkynylene,

X₂ is an element that is more electronegative than carbon, and

R₂' is a second chemical functional group.

In Formula 1, a carbonyl group connected to D₁ is referred to as a first carbonyl group. Also, a carbonyl group connected to D₂ is referred to as a second carbonyl group.

In specific embodiments, R₁' may include a click-chemistry functional group. Also, the click-chemistry functional

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group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R_1' may include two or more click-chemistry functional groups.

In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-CH_2OCH_2-$. In addition, D_1 may be a covalent bond. When D_1 is a covalent bond, R_1' and the carbon of the first carbonyl group are directly connected to each other. Hereinafter, when describing a structure of a compound, all the alkylene, the alkenylene, the alkynylene, and the cycloalkylene are intended to include a heteroalkylene, a heteroalkynylene, a heterocycloalkylene, respectively, the contents of which are also specified in the section "Definitions."

In specific embodiments, X_1 may be NR_1 , S, or O, wherein R_1 may be H, a halogen, or a substituted or unsubstituted C_{1-3} alkylene. Also, X_1 may be S. X_1 may attract electrons from the carbon of the first carbonyl group to activate the first carbonyl group.

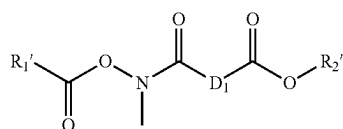
In specific embodiments, D_2 may include any one selected from a C_{1-7} alkylene, a C_{2-7} alkenylene, a C_{2-7} alkynylene, and a C_{3-8} cycloalkylene. Also, D_2 may be a C_{1-2} alkylene. Furthermore, D_2 may be methylene.

In specific embodiments, X_2 may be NR_1 , S, or O, wherein R_1 may be H, a halogen, or a substituted or unsubstituted C_{1-3} alkylene. Also, X_2 may be O. X_2 may attract electrons from the carbon of the second carbonyl group to activate the second carbonyl group.

In specific embodiments, R_2' may be a halogen, N-succinimide, p-nitrophenyl, or pentafluorophenyl.

In specific embodiments, R_2' and X_2 taken together may form a leaving group. For example, X_2 may be O, and R_2' may be N-succinimide, p-nitrophenyl, or pentafluorophenyl. Also, R_2' may be N-succinimide. When a good leaving group is connected to the second carbonyl group, reactivity of the second carbonyl group may be enhanced. For example, an N-hydroxysuccinimide ester (an NHS ester) is known to show very high reactivity.

The present invention provides a compound having a structure of the following Formula 1-2:



[Formula 1-2]

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wherein R_1' is a first chemical functional group,

D_1 is any alkylene, alkenylene, or alkynylene, and

R_2' is a second chemical functional group.

In specific embodiments, R_1' may include a click-chemistry functional group. Also, the click-chemistry functional group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R_1' may include two or more click-chemistry functional groups.

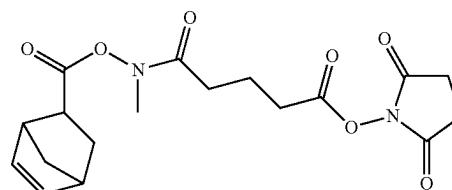
In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a C_{1-7} alkylene, a C_{2-7} alkenylene, a C_{2-7} alkynylene, and a C_{3-3} cycloalkylene. Also, D_1 may be a C_{1-2} alkylene. Furthermore, D_1 may be methylene.

In specific embodiments, R_2' and O taken together may form a leaving group. In this case, R_2' may be N-succinimide, p-nitrophenyl, or pentafluorophenyl. Also, R_2' may be N-succinimide.

In specific embodiments, the compound represented by Formula 1-2 may have a structure of the following Formula 1-3:

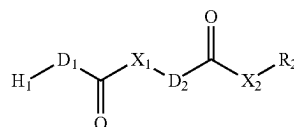
[Formula 1-3]



2.1. Linker Including First Click-Chemistry Functional Group (H_1-L_1)

The present invention provides a compound having a structure of the following Formula 2:

[Formula 2]



wherein H_1 is a first click-chemistry functional group,

D_1 is any alkylene, alkenylene, or alkynylene,

X_1 is an element that is more electronegative than carbon,

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D₂ is any alkylene, alkenylene, or alkynylene,
X₂ is an element that is more electronegative than carbon,
and

R₂' is a second chemical functional group. The linker having the structure of Formula 2 is herein referred to as a "linker including a first click-chemistry functional group," which is indicated by the symbol "H₁-L₁."

In Formula 2, a carbonyl group connected to D₁ is referred to as a first carbonyl group. Also, a carbonyl group connected to D₂ is referred to as a second carbonyl group.

In specific embodiments, H₁ may include any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, H₁ may be an azide, or a strained alkyne. Further, H₁ may be an azide, or dibenzocyclooctyne-amine. Additionally, H₁ may be a diene, or a dienophile. Further, H₁ may be a tetrazine, or a norbornene. Alternatively, H₁ may be a tetrazine, or a trans-cyclooctene.

In specific embodiments, D₁ may include any one selected from a covalent bond, a C₁₋₄ alkylene, a C₂₋₄ alkenylene, a C₂₋₄ alkynylene, and a C₃₋₈ cycloalkylene. Also, D₁ may be —CH₂OCH₂—. In addition, D₁ may be a covalent bond.

In specific embodiments, X₁ may be NR₁, S, or O, wherein R₁ may be H, a halogen, or a substituted or unsubstituted C₁₋₃ alkylene. Also, X₁ may be S.

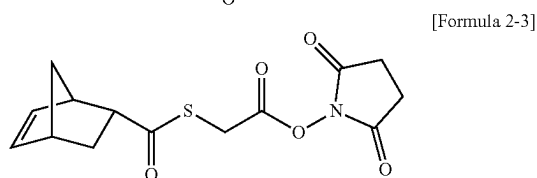
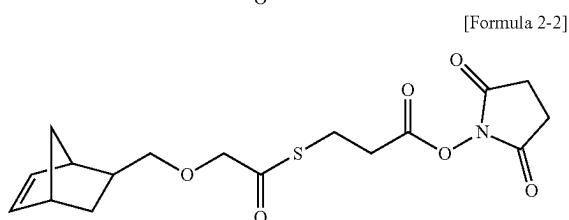
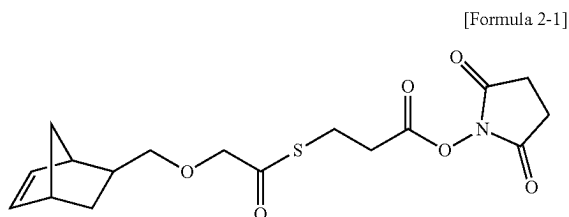
In specific embodiments, D₂ may include any one selected from a C₁₋₇ alkylene, a C₂₋₇ alkenylene, a C₂₋₇ alkynylene, and a C₃₋₃ cycloalkylene. Also, D₂ may be a C₁₋₂ alkylene. Furthermore, D₂ may be methylene.

In specific embodiments, X₂ may be NR₁, S, or O, wherein R₁ may be H, a halogen, or a substituted or unsubstituted C₁₋₃ alkylene. Also, X₂ may be O.

In specific embodiments, R₂' may be a halogen, N-succinimide, p-nitrophenyl, or pentafluorophenyl.

In specific embodiments, R₂' and X₂ taken together may form a leaving group. For example, X₂ may be O, and R₂' may be N-succinimide, p-nitrophenyl, or pentafluorophenyl. Also, R₂' may be N-succinimide.

In specific embodiments, the compound represented by Formula 2 may have any one structure selected from the following Formulas 2-1 to 2-3:

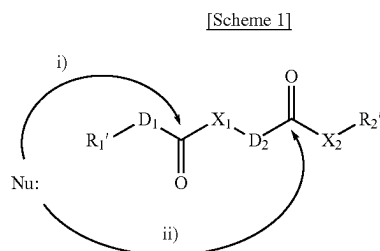


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2.2. The Position at which a Substitution Reaction Occurs May be Specifically Determined Due to the Difference in Reactivity Between the First Carbonyl Group and the Second Carbonyl Group

When the linkers of Formulas 1 and 2, and sub-examples thereof are designed, sites for substitution reaction may be specified based on the design of X₁, X₂, and R₂'.

The linker disclosed in the present invention functions to transfer R₁' to a target molecule by means of a substitution reaction occurring between the activated first and/or second carbonyl groups. Such a substitution reaction is schematically shown as in the following Scheme 1.



In Scheme 1, the target molecule is designated "Nu:" for the sake of convenience. The Nu: that serves as a nucleophile because it has an unshared electron pair causes a nucleophilic acyl substitution reaction with the first carbonyl group and/or second carbonyl group to form a bond with a linker. In the nucleophilic acyl substitution reaction, the reactivity of the carbonyl group may be determined by the basicity of the leaving group. Therefore, the reactivity of the carbonyl group is known to increase in the order of the carboxylate, the amide, the carboxylic acid, the ester, the thioester, and the acyl phosphate. Also, the carbonyl group such as an NHS ester, or the like is known to have high reactivity because the carbonyl group forms a very stable leaving group.

In specific embodiments, X₁ and X₂ may be elements that are more electronegative than carbon. Also, X₁ and X₂ may be NR₁, S, or O, wherein R₁ may be H, a halogen, or a substituted or unsubstituted C₁₋₃ alkylene. Because X₁ and X₂ taken together with the residue(s) to which they are attached may form a leaving group, the carbonyl group may be activated.

In this case, the activation of the carbonyl group may optionally allow Nu: to i) preferentially react with the first carbonyl group or ii) preferentially react with the second carbonyl group. This tendency of reaction may be determined depending on a difference in reactivity between the first carbonyl group and the second carbonyl group. For example, when the basicity of a leaving group including X₁, is lower than that of a leaving group including X₂, the first carbonyl group may react first. In another embodiment, when the basicity of the leaving group including X₂ is lower than that of the leaving group including X₁, the second carbonyl group may react first.

In preferred embodiments, the reactivity of the second carbonyl group is preferably higher than that of the first carbonyl group in the linker according to the present invention. According to the present invention, a selective reaction was achieved by allowing X₂-R₂' connected to the second carbonyl group to form a good leaving group, and designing the first carbonyl group to be an amide, a thioester, an ester, and the like, which show mild reactivity. For example, in the case of the linker of Formula 1-3, the second carbonyl group

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may be an NHS ester, thereby allowing it to react faster than the first carbonyl group (a thioester).

The prior art disclosed in Publication Nos. US 2018/0141976 A1 and WO 2018/199337 A1 is similar to the present invention in terms of the form of an agent for transferring a first chemical functional group to an antibody (see section 4 below), but is different from the present invention in that the cross-linker includes two NHS esters. The two carbonyl groups of the cross-linker have the same reactivity, and it is difficult to prepare a desired agent for transferring a chemical functional group with high yield. Also, the cross-linker has a high probability of reacting with two SSFIs due to high reactivity of the NHS esters. According to the present invention, such problems have been solved by designing the first carbonyl group to be a thioester, and the like, which have mild reactivity.

3. Site-Specific Antibody Interactome (SSAI)

According to the present invention, there is disclosed a novel peptide for bringing a molecule to be labeled into close contact with a certain site of an antibody. For the sake of convenience, such a peptide is herein referred to as a site-specific antibody interactome, which is indicated by the symbol "SSAI."

The SSAI provided according to the present invention may have binding activity for a certain site of an antibody.

In specific embodiments, the SSAI may have binding activity for an Fc domain of the antibody. In this case, the SSAI may preferably have binding activity for a site spaced apart from a paratope of the antibody.

In specific embodiments, the SSAI may have binding activity for the Fc domain of the antibody. In this case, the SSAI may preferably have binding activity for an FcRn binding site of the antibody or a site spaced apart from residues of the antibody which affects the FcRn binding site of the antibody.

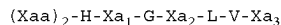
3.1: Site-Specific Fc Interactome (SSFI)

In the SSAI, a peptide having specific binding activity for the Fc domain is herein referred to as a "site-specific Fc interactome", which is indicated by the symbol "SSFI."

In specific embodiments, the SSFI may include an amino acid sequence represented by the following Formula 3:

[Formula 3]

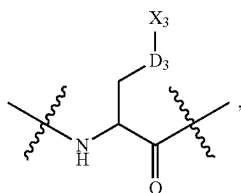
(SEQ ID NO: 3)



wherein each Xaa is independently any amino acid except cysteine,

H is histidine, G is glycine, Xa₂ is glutamic acid or asparagine, L is leucine, V is valine, Xa₃ is selected from tryptophan, naphthylalanine, and phenylalanine, and

Xa₁ is



wherein D₃ is a covalent bond or a C₁₋₃ alkylene, and X₃ is NH₂, OH, or SH. As an analogue of the sequence AWHLGELVW (SEQ ID NO: 2), which is a sequence found to have binding activity for the Fc domain in the articles

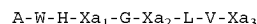
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"Dias, R. L. A., et al (2006), Protein Ligand Design: From Phage Display to Synthetic Protein Epitope Mimetics in Human Antibody Fc-Binding Peptidomimetics; Journal of the American Chemical Society, 128(8), 2726-2732; and DeLano, W. L., et al., Convergent solutions to binding at a protein-protein interface; Science 2000, 287, 1279-1283," it has binding activity for the Fc domain. The Fc binding activity motif according to the present invention has characteristics as follows: 1) First, key residues having Fc binding activity are specified. 2) The motif is also designed to allow a nucleophilic substitution reaction with a linker by changing the 4th leucine of SEQ ID NO: 2 into Xa₁ having a free electron pair. (X)_n means that it consists of n Xs, and (X)_{n-m} means that it consists of n or more and m or less Xs. Hereinafter, the carbon connected to D₃ is referred to as "beta carbon (β-carbon)."

Also, the amino acid sequence of Formula 3 may have a structure represented by the following Formula 3-1:

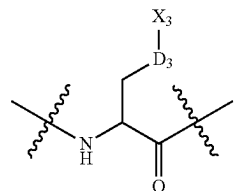
[Formula 3-1]

(SEQ ID NO: 4)



wherein A is alanine, H is histidine, G is glycine, Xa₂ is glutamic acid or asparagine, L is leucine, V is valine, Xa_α is selected from tryptophan, naphthylalanine, and phenylalanine, and

Xa₁ is



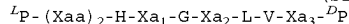
wherein D₃ is a covalent bond or a C₁₋₃ alkylene, and X₃ is NH₂, OH, or SH. In Formulas 3 and 3-1, X₃ may be NH₂. Alternatively, Xa₂ may be glutamic acid. Alternatively, Xa₃ may be tryptophan.

When the peptide including each of amino acid sequences of Formula 3 and sub-examples thereof is in a cyclic peptide form in which internal residues are connected to each other, the peptide is known to have better binding activity. The present invention provides a cyclic peptide including the amino acid sequence of Formula 3.

The present invention provides a cyclic peptide having a structure of the following Formula 4-1:

[Formula 4-1]

(SEQ ID NO: 5)

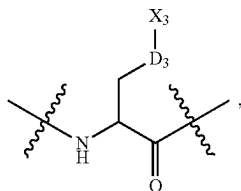


wherein N-terminal ^LP and ^DP form a D-proline-L-proline template,

each Xaa is independently any amino acid except cysteine, H is histidine, G is glycine, Xa₂ is glutamic acid or asparagine, L is leucine, V is valine, Xa₃ is selected from tryptophan, naphthylalanine, and phenylalanine, and

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Xa₁ is



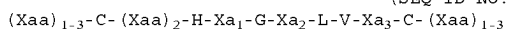
wherein D₃ is a covalent bond or a C₁₋₃ alkylene, and X₃ is NH₂, OH, or SH.

In specific embodiments, X₃ may be NH₂. In specific embodiments, (X)₂ may be AW. In specific embodiments, Xa₂ may be glutamic acid. In specific embodiments, Xa₃ may be tryptophan.

The present invention provides a cyclic peptide having a structure of the following Formula 4-2:

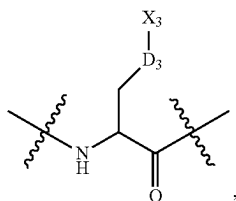
[Formula 4-2]

(SEQ ID NO: 6)



wherein each Xaa is independently any amino acid except cysteine,

C is cysteine, H is histidine, G is glycine, Xa₂ is glutamic acid or asparagine, L is leucine, V is valine, Xa₃ is selected from tryptophan, naphthylalanine, and phenylalanine, and Xa₁ is



wherein D₃ is a covalent bond or a C₁₋₃ alkylene, and X₃ is NH₂, OH, or SH.

In specific embodiments, the peptide may consist of 13 or more and 17 or less amino acid residues.

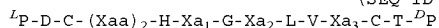
In specific embodiments, cysteine located 2 to 4 amino acids from the N-terminus and cysteine located 2 to 4 amino acids from the C-terminus may be optionally connected to each other.

In specific embodiments, X₃ may be NH₂. In specific embodiments, (X)₂ may be AW. In specific embodiments, Xa₂ may be glutamic acid. In specific embodiments, Xa_a may be tryptophan.

In specific embodiments, one of the residues constituting the N-terminal (X)₁₋₃ and one of the residues constituting the C-terminal (X)₁₋₃ may be bound to each other. In one exemplary embodiment, the peptide of Formula 4-2 may have a structure of the following Formula 4-3:

[Formula 4-3]

(SEQ ID NO: 7)



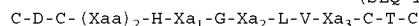
wherein N-terminal ^LP and ^DP may form a D-proline-L-proline template.

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In another exemplary embodiment, the peptide of Formula 4-2 may have a structure of the following Formula 4-4:

[Formula 4-4]

(SEQ ID NO: 8)

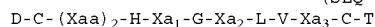


wherein the N-terminal cysteine and the C-terminal cysteine may be connected to each other.

The present invention provides a cyclic peptide having a structure of the following Formula 4-5:

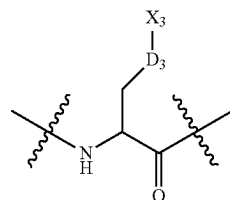
[Formula 4-5]

(SEQ ID NO: 9)



wherein D is aspartic acid, T is threonine, each Xaa is independently any amino acid except cysteine,

C is cysteine, H is histidine, G is glycine, Xa₂ is glutamic acid or asparagine, L is leucine, V is valine, Xa_a is selected from tryptophan, naphthylalanine, and phenylalanine, and Xa₁ is



wherein D₃ is a covalent bond or a C₁₋₃ alkylene, and X₃ is NH₂, OH, or SH.

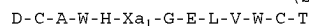
In specific embodiments, the peptide may consist of 13 or more and 17 or less amino acid residues.

In specific embodiments, cysteine located 2 amino acids from the N-terminus and cysteine located 2 amino acids from the C-terminus may be optionally connected to each other. In specific embodiments, X₃ may be NH₂. In specific embodiments, (X)₂ may be AW. In specific embodiments, Xa₂ may be glutamic acid. In specific embodiments, Xa_a may be tryptophan.

In specific embodiments, the Formula 4-5 may be identical to the following Formula 4-6:

[Formula 4-6]

(SEQ ID NO: 10)



wherein A is alanine, and E is glutamic acid.

The peptides having the structures of Formulas 4-1 to 4-6 may have binding activity for an antibody. Also, the peptides may have binding activity for immunoglobulin G (IgG). In addition, the peptides may have binding activity for an Fc domain of the antibody.

In specific embodiments, the N-terminus of the SSFI according to the present invention may be succinylated. In specific embodiments, the SSFI according to the present invention may include a polar amino acid residue at the N-terminal (X)₁₋₃ thereof. In other specific embodiments, the SSFI according to the present invention may include a polar amino acid residue at the C-terminal (X)₁₋₃ thereof. In this case, the polar amino acid residue includes an acidic

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amino acid and a basic amino acid. Also, the polar amino acid residue may include glutamic acid or aspartic acid.

3.2. Site-specific Fc interactome according to the present invention may be arranged with an Fc domain of an antibody with a specific topology.

In the present content, the compound of Formula 4-6 is provided as one example to aid in understanding the present invention, but the scope of the present invention is not limited thereto. It will be noted that the following description also applies to the compounds of Formulas 3, 3-1, and 4-1 to 4-6, and the compound of Formula 4-6 is merely provided as one example for the sake of convenience.

As previously described above, the SSFI according to the present invention has binding activity for the Fc domain of the antibody. In this case, the SSFI may be arranged with the Fc domain with a specific topology due to the interaction between amino acid residues. The representative interaction between the SSFI sequence according to the present invention and the Fc domain includes (1) a salt linkage of the SSFI with histidine 433 in the Fc domain, (2) a hydrogen bond of the SSFI with asparagine 434, (3) a salt linkage of the SSFI with glutamic acid 380, (4) a salt linkage of the SSFI with arginine 255, and the like. These interactions and specific topologies thus formed may be determined from the research results already known in the art (see DeLano, W. L., et al., Convergent solutions to binding at a protein-protein interface; Science 2000, 287, 1279-1283).

When the SSFI according to the present invention is designed, it is important for the SSFI to form a stable topology with the Fc domain. This is because an agent for transferring a first chemical functional group to the antibody according to the present invention, and a labeling process using the same are designed based on the topology between the SSFI and the Fc domain found in research (see the following sections 5.2, 5.3, and 5.4). When the interaction between the SSFI and the Fc domain becomes unstable during the design of SSFI to disturb a topology between their molecules, it is unfavorable as the interaction may give a negative effect on the labeling process.

One embodiment of the design principle will be described with reference to an exemplary compound. The SSFI represented by Formula 4-6 has a structure as follows:

[Formula 4-6]

(SEQ ID NO: 10)

D-C-A-W-H-Xa₁-G-E-L-V-W-C-T

The results of simulating the topology between SSFI set forth in SEQ ID NO: 10 and the Fc domain based on the data in the articles, and the like are shown in FIGS. 3 and 4 (see DeLano, W. L., et al., Convergent solutions to binding at a protein-protein interface; Science 2000, 287, 1279-1283). In this case, a histidine residue at position 5 in the SSFI forms a salt linkage with glutamic acid 380 in the Fc domain, indicating that this salt linkage has a significant effect on the topology between the SSFI and the Fc domain (see a dotted line in FIG. 4). Therefore, it is desirable that the histidine residue and its position are not changed during the design of the SSFI (see 'H' next to the residue Xa₁ in Formulas 3, 3-1, and 4-1 to 4-6). In addition, it was confirmed that glutamic acid 8 is a residue that shows electronegativity, and thus forms a salt linkage with arginine 255 in the Fc domain, which shows electropositivity, thereby exerting a significant effect on the topology between the SSFI and the Fc domain (see a dotted line in FIG. 4). Therefore, the corresponding residue is preferably an acidic amino acid that may correspond to glutamic acid, and may be replaced with asparagine

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(see the residue Xa₂ in Formulas 3, 3-1, and 4-1 to 4-6). When the amino acid residues are replaced with other amino acid residues or are substituted with any functional groups, this may have an effect on the intermolecular interaction, thereby exerting an effect on the topology between the SSFI and the Fc domain.

In addition, glycine at position 7 in the sequence of SEQ ID NO: 10 is a small amino acid that is required to form a bent structure of the SSFI. Therefore, it is desirable that the glycine residue and its position are not changed during the design of the SSFI (see 'G' between the residues Xa₁ and Xa₂ in Formulas 3, 3-1, and 4-1 to 4-6).

FIG. 5 shows a topology between lysine residues of the Fc domain and the SSFI. Based on the topology, it can be seen that the lysine residues of the Fc domain located closest to the residue Xa₁ are lysine 246 and 248 (FIG. 5).

A distance between an amine group of lysine 246 and the beta carbon of Xa₁ was measured. As a result, it was confirmed that the minimum distance is measured to be approximately 11.668 Å (hereinafter referred to as "D_{246,min}"), and the maximum distance is measured to be approximately 20.765 Å (hereinafter referred to as "D_{246,max}") as the bonds constituting a lysine branch rotate (FIG. 6).

A distance between an amine group of lysine 248 and the beta carbon of Xa₁ was measured. As a result, it was confirmed that the minimum distance is measured to be approximately 6.723 Å (hereinafter referred to as "D_{248,min}"), and the maximum distance is measured to be approximately 16.208 Å (hereinafter referred to as "D_{248,max}") as the bonds constituting a lysine branch rotate (FIG. 7).

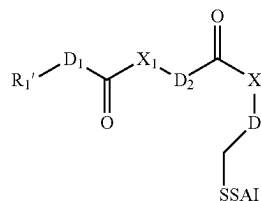
As will be described in the following section 5.3, the distance relationship may be an important consideration in the design of the linker and the SSFI, as well as the agent for transferring a first chemical functional group to an antibody.

4. Agent for transferring first chemical functional group to antibody; Conjugate of R₁'-L₁ and SSAI (R₁'-L₂-SSAI)

According to the present invention, there is disclosed an agent for transferring a first chemical functional group to an antibody. Such a compound is herein indicated by the symbol "R₁'-L₂-SSAI." The compound is also referred to as a conjugate of R₁'-L₁ and SSAI (an R₁'-L₂-SSAI conjugate) depending on the structure thereof.

The present invention provides R₁'-L₂-SSAI having a structure of the following Formula 5:

[Formula 5]



wherein R₁' is a first chemical functional group, D₁ is any alkylene, alkenylene, or alkynylene, X₁ is an element that is more electronegative than carbon, D₂ is any alkylene, alkenylene, or alkynylene, D₃ is a covalent bond or a C₁₋₃ alkylene, X₃ is NH, O, or S, and SSAI is a site-specific antibody interactome.

In specific embodiments, R₁' may include a click-chemistry functional group. Also, the click-chemistry functional group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a

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thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R_1' may include two or more click-chemistry functional groups.

In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-\text{CH}_2\text{OCH}_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, X_1 may be NR_1 , S, or O, wherein R_1 may be H, a halogen, or a substituted or unsubstituted C_{1-3} alkylene. Also, X_1 may be S.

In specific embodiments, D_2 may include any one selected from a C_{1-7} alkylene, a C_{2-7} alkenylene, a C_{2-7} alkynylene, and a C_{3-8} cycloalkylene. Also, D_2 may be a C_{1-2} alkylene. Furthermore, D_2 may be methylene.

In specific embodiments, X_3 may be NH.

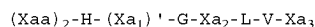
In specific embodiments, the SSAI may be a peptide sequence having binding activity for Fab. In other specific embodiments, the SSAI may be a peptide sequence having binding activity for the Fc domain.

When the SSAI in Formula 5 is SSFI, this is indicated by the symbol " $R_1'-L_2$ -SSFI." The $R_1'-L_2$ -SSFI according to the present invention is produced by means of a nucleophilic substitution reaction of X_{a1} of the SSFI according to the present invention with a second carbonyl group of $R_1'-L_1$ (see the following section 4-2 and Scheme 2). Therefore, the $R_1'-L_2$ -SSFI according to the present invention includes those in which X_{a1} of the SSFI in Formulas 3, 3-1, and 4-1 to 4-6 is substituted with $(X_{a1})'$, but the present invention is not limited to the following exemplary embodiments thereof.

The present invention provides an $R_1'-L_2$ -SSFI including an amino acid sequence of the following Formula 5-1:

[Formula 5-1]

(SEQ ID NO: 11)

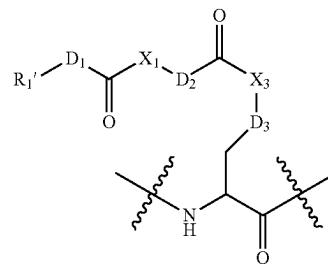


wherein each Xaa is independently any amino acid except cysteine,

H is histidine, G is glycine, X_{a2} is glutamic acid or asparagine, L is leucine, V is valine, X_{a3} is selected from tryptophan, naphthylalanine, and phenylalanine, and

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$(X_{a1})'$ is



wherein R_1' is a first chemical functional group, D_1 is any alkylene, alkenylene, or alkynylene, X_1 is an element that is more electronegative than carbon, D_2 is any alkylene, alkenylene, or alkynylene, D_3 is a covalent bond or a C_{1-3} alkylene, and X_3 is NH, O, or S.

In specific embodiments, R_1' may include a click-chemistry functional group. Also, the click-chemistry functional group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R_1' may include two or more click-chemistry functional groups.

In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, or a drug moiety. Also, R_1' may include a VC linker. In addition, R_1' may include a radioactive moiety. Also, R_1' may include a drug moiety. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-\text{CH}_2\text{OCH}_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, X_1 may be NR_1 , S, or O, wherein R_1 may be H, a halogen, or a substituted or unsubstituted C_{1-3} alkylene.

In specific embodiments, D_2 may include any one selected from a C_{1-7} alkylene, a C_{2-7} alkenylene, a C_{2-7} alkynylene, and a C_{3-8} cycloalkylene. Also, D_2 may be a C_{1-2} alkylene. Furthermore, D_2 may be methylene.

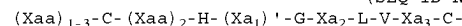
In specific embodiments, X_3 may be NH.

In specific embodiments, $(X)_2$ may be AW. In specific embodiments, X_{a2} may be glutamic acid. In specific embodiments, X_{a3} may be tryptophan.

The present invention provides an $R_1'-L_2$ -SSFI having a structure of the following Formula 5-2:

[Formula 5-2]

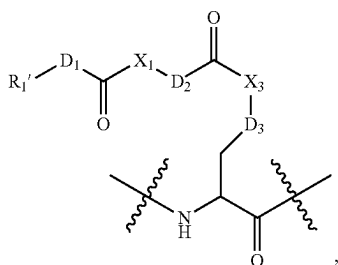
(SEQ ID NO: 12)



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wherein each Xaa is independently any amino acid except cysteine,

C is cysteine, H is histidine, G is glycine, Xa₂ is glutamic acid or asparagine, L is leucine, V is valine, Xa₃ is selected from tryptophan, naphthylalanine, and phenylalanine, and (Xa₁)' is



wherein R₁' is a first chemical functional group,

D₁ is any alkylene, alkenylene, or alkynylene,

X₁ is an element that is more electronegative than carbon,

D₂ is any alkylene, alkenylene, or alkynylene,

D₃ is a covalent bond or a C₁₋₃ alkylene, and

X₃ is NH, O, or S.

In specific embodiments, R₁' may include a click-chemistry functional group. Also, the click-chemistry functional group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R₁' may include two or more click-chemistry functional groups.

In other specific embodiments, R₁' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R₁' may include a drug moiety. In addition, R₁' may include a VC linker. In other specific embodiments, R₁' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D₁ may include any one selected from a covalent bond, a C₁₋₄ alkylene, a C₂₋₄ alkenylene, a C₂₋₄ alkynylene, and a C₃₋₈ cycloalkylene. Also, D₁ may be —CH₂OCH₂—. In addition, D₁ may be a covalent bond.

In specific embodiments, X₁ may be NR₁, S, or O, wherein R₁ may be H, a halogen, or a substituted or unsubstituted C₁₋₃ alkylene.

In specific embodiments, D₂ may include any one selected from a C₁₋₇ alkylene, a C₂₋₇ alkenylene, a C₂₋₇ alkynylene, and a C₃₋₈ cycloalkylene. Also, D₂ may be a C₁₋₂ alkylene. Furthermore, D₂ may be methylene.

In specific embodiments, Formula 5-2 may consist of 13 or more and 17 or less amino acid residues (including (Xa₁)').

In specific embodiments, cysteine located 2 to 4 amino acids from the N-terminus and cysteine located 2 to 4 amino acids from the C-terminus may be optionally connected to each other.

In specific embodiments, X₃ may be NH. In specific embodiments, (X)₂ may be AW. In specific embodiments, Xa₂ may be glutamic acid. In specific embodiments, Xa₃ may be tryptophan.

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In specific embodiments, one of the residues constituting the N-terminal (X)₁₋₃ and one of the residues constituting the C-terminal (X)₁₋₃ may be bound to each other.

In specific embodiments, Formula 5-2 may be identical to Formula 5-3:

[Formula 5-3]

(SEQ ID NO: 13)

D-C-A-W-H-(Xa₁)'-G-E-L-V-W-C-T

wherein A is alanine, and E is glutamic acid.

The R₁'-L₂-SSAI or R₁'-L₂-SSFI having the structures of Formulas 5 and 5-1 to 5-3 may have binding activity for an antibody. Also, the R₁'-L₂-SSAI or R₁'-L₂-SSFI may have binding activity for immunoglobulin G (IgG). In addition, the R₁'-L₂-SSAI or R₁'-L₂-SSFI may have binding activity for an Fc domain of the antibody.

4.1. Agent for transferring first click-chemistry functional group to antibody; Conjugate of H₁-L₁ and SSAI (H₁-L₂-SSAI)

According to the present invention, there is disclosed an agent for transferring a first click-chemistry functional group to an antibody. Such a compound is herein indicated by the symbol "H₁-L₂-SSAI." The compound is also referred to as a conjugate of H₁-L₁ and SSAI (an H₁-L₂-SSAI conjugate) depending on the structure thereof. In this case, when the SSAI is SSFI, this is indicated by the symbol 'H₁-L₂-SSFI.'

The H₁-L₂-SSAI or H₁-L₂-SSFI according to the present invention includes those in which R₁' contains a click-chemistry functional group in the section '4. Agent for transferring first chemical functional group to antibody,' but the present invention is not limited to the following exemplary embodiments thereof.

The present invention provides an H₁-L₂-SSFI including an amino acid sequence of the following Formula 6-1:

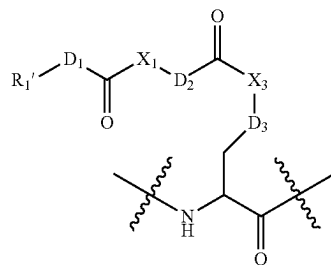
[Formula 6-1]

(SEQ ID NO: 14)

(Xaa)₂-H-(Xa₁)'-G-Xa₂-L-V-Xa₃

wherein each Xaa is independently any amino acid except cysteine,

H is histidine, G is glycine, Xa₂ is glutamic acid or asparagine, L is leucine, V is valine, Xa₃ is selected from tryptophan, naphthylalanine, and phenylalanine, and (Xa₁)' is



wherein H₁ is a first click-chemistry functional group,

D₁ is any alkylene, alkenylene, or alkynylene,

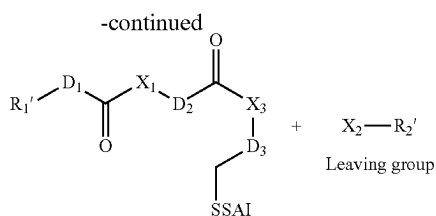
X₁ is an element that is more electronegative than carbon,

D₂ is any alkylene, alkenylene, or alkynylene,

D₃ is a covalent bond or a C₁₋₃ alkylene, and

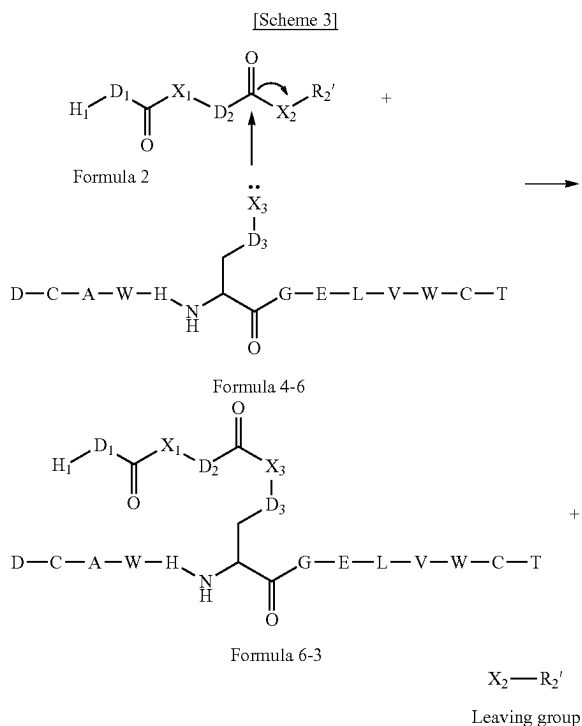
X₃ is NH, O, or S.

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The R_1' - L_2 -SSAI according to the present invention may be prepared by allowing a site-specific antibody interactome (SSAI) to react with the linker (R_1' - L_1) according to the present invention. The SSAI according to the present invention is designed to include a nucleophile X_3 . The X_3 may attack an activated carbonyl group included in the linker to cause a nucleophilic substitution reaction. In this case, as the X_3 attacks the second carbonyl carbon, the R_1' - L_2 -SSAI according to the present invention is prepared.

As a more specific example, a method of preparing a compound of Formula 6-3 will be described. In specific embodiments, the compound of Formula 6-3 may be prepared through a reaction of the following Scheme 3.



The SSAI including the amino acid sequence of Formula 3 or 3-1 according to the present invention and having each of the structures of Formulas 4-1 to 4-6 includes an X_{a1} residue including a nucleophile X_3 . As the X_3 attacks the second carbonyl group included in the linker, the R_1' - L_2 -SSAI according to the present invention is prepared.

In specific embodiments, the basicity of the leaving group including X_2 may be lower than that of the leaving group including X_1 , in the linker according to the present invention. In specific embodiments, the linker according to the present invention may allow X_2 - R_2' connected to the second

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carbonyl group to form a good leaving group. Also, X_2 may be O, and R_2' may be N-succinimide, p-nitrophenyl, or pentafluorophenyl. In this case, the reactivity of the second carbonyl group is higher than that of the first carbonyl group. Therefore, X_3 of the SSAI may specifically attack the second carbonyl group of the linker.

According to the present invention, there is disclosed a method of preparing an agent for transferring a first chemical functional group to an antibody.

The present invention provides a method of preparing an R_1' - L_2 -SSAI, which comprises:

reacting a linker according to the present invention with a site-specific antibody interactome according to the present invention.

In specific embodiments, the linker may be any one selected from Formulas 1, 2, and 2-1 to 2-3.

In specific embodiments, the site-specific antibody interactome may include or have any one structure selected from Formulas 3, 3-1, and 4-1 to 4-6.

Also, the linker may have the structure of Formula 2, and the site-specific antibody interactome may have any one structure selected from Formulas 4-2 to 4-6. Furthermore, the site-specific antibody interactome may have the structure of Formula 4-6.

According to the present invention, there is disclosed a method of preparing an agent for transferring a first click-chemistry functional group to an antibody.

The present invention provides a method of preparing an H_1 - L_2 -SSAI, which comprises:

reacting a linker according to the present invention with a site-specific antibody interactome according to the present invention.

In specific embodiments, the linker may be any one selected from Formulas 2, and 2-1 to 2-3.

In specific embodiments, the site-specific antibody interactome may include or have any one structure selected from Formulas 3, 3-1, and 4-1 to 4-6.

Also, the linker may have the structure of Formula 2, and the site-specific antibody interactome may have any one structure selected from Formulas 4-2 to 4-6. Furthermore, the site-specific antibody interactome may have the structure of Formula 4-6.

According to the present invention, there is disclosed a kit for preparing an agent for transferring a first chemical functional group to an antibody.

The present invention provides a kit for preparing an agent for transferring a first chemical functional group to an antibody, which comprises the linker according to the present invention and the site-specific antibody interactome according to the present invention.

In specific embodiments, the linker may be any one selected from Formulas 1, 2, and 2-1 to 2-3.

In specific embodiments, the site-specific antibody interactome may include or have any one structure selected from Formulas 3, 3-1, and 4-1 to 4-6.

Also, the linker may have the structure of Formula 2, and the site-specific antibody interactome may have any one structure selected from Formulas 4-2 to 4-6. Furthermore, the site-specific antibody interactome may have the structure of Formula 4-6.

According to the present invention, there is disclosed a kit for preparing an agent for transferring a first click-chemistry functional group to an antibody.

The present invention provides a kit for preparing an agent for transferring a first click-chemistry functional group to an antibody, which comprises a linker containing the first

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click-chemistry functional group according to the present invention, and a site-specific antibody interactome.

In specific embodiments, the linker may be any one selected from Formulas 2, and 2-1 to 2-3.

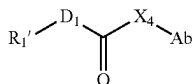
In specific embodiments, the site-specific antibody interactome may include or have any one structure selected from Formulas 3, 3-1, and 4-1 to 4-6.

Also, the linker may have the structure of Formula 2, and the site-specific antibody interactome may have any one structure selected from Formulas 4-2 to 4-6. Furthermore, the site-specific antibody interactome may have the structure of Formula 4-6.

5. Antibody Containing First Chemical Functional Group (R_1' -Ab)

According to the present invention, there is disclosed an antibody containing a first chemical functional group. Such a compound is herein indicated by the symbol " R_1' -Ab."

The present invention provides an R_1' -Ab represented by Formula 7:



[Formula 7]

wherein R_1' is a first chemical functional group, D_1 is any alkylene, alkenylene, or alkynylene, X_4 is NH, O, or S, and Ab is an antibody.

In specific embodiments, R_1' may be a click-chemistry functional group. Also, R_1' may include any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, R_1' may be an azide, or a strained alkyne. Further, R_1' may be an azide, or dibenzocyclooctyne-amine. Additionally, R_1' may be a diene, or a dienophile. Further, R_1' may be a tetrazine, or a norbornene. Alternatively, R_1' may be a tetrazine, or a trans-cyclooctene.

In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-CH_2OCH_2-$. In addition, D_1 may be a covalent bond.

In one specific embodiment, X_4 may be NH.

In specific embodiments, Ab may be a human antibody. In other specific embodiments, Ab may be a non-human animal antibody. In specific embodiments, Ab may be immunoglobulin G (IgG). In specific embodiments, Ab may be a whole antibody. In other specific embodiments, Ab may be a fragment of the antibody.

In specific embodiments, X_4 and Ab may be connected via an Fab domain of Ab. In other specific embodiments, X_4 and Ab may be connected via an Fc domain of Ab. Also, X_4 and Ab may be connected via lysine 246 or 248 in the Fc domain of Ab. Furthermore, X_4 and Ab may be connected via lysine 246 in the Fc domain of Ab. Further, X_4 and Ab may be connected via lysine 248 in the Fc domain of Ab. Alternatively, X_4 and Ab may be connected via lysine 246 and 248 in the Fc domain of Ab. In specific embodiments, X_4 and Ab may be connected via only one of two Fc domains of Ab. In

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other specific embodiments, X_4 and Ab may be connected via both of the two Fc domains of Ab.

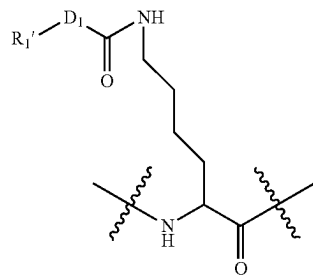
The present invention provides an antibody or a fragment thereof, which includes an amino acid sequence of the following Formula 7-1:

[Formula 7-1]

(SEQ ID NO: 17)

G-P-S-V-F-L-F-P-P-(K)-P-K-D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and (K)' is



wherein R_1' is a first chemical functional group, and D_1 is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has R_1' connected via lysine 246 in an Fc domain thereof, or a site corresponding to the lysine 246.

In specific embodiments, R_1' may include a click-chemistry functional group. Also, the click-chemistry functional group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R_1' may include two or more click-chemistry functional groups.

In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-CH_2OCH_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody.

In specific embodiments, the antibody may include the amino acid sequence of Formula 7-1 in only one of the two

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Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 7-1 in both of the two Fc domains thereof.

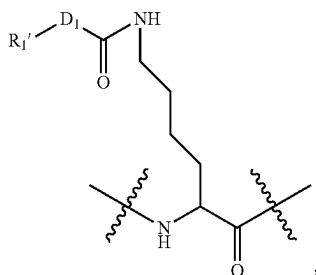
The present invention provides an antibody or a fragment thereof, which includes an amino acid sequence of the following Formula 7-2:

[Formula 7-2]

(SEQ ID NO: 18) 10

G-P-S-V-F-L-F-P-P-K-P- (K)' -D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and (K)' is



wherein R_1' is a first chemical functional group, and D_1 is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has R_1' connected via lysine 248 in an Fc domain thereof, or a site corresponding to the lysine 248.

In specific embodiments, R_1' may include a click-chemistry functional group. Also, the click-chemistry functional group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R_1' may include two or more click-chemistry functional groups.

In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-CH_2OCH_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody.

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In specific embodiments, the antibody may include the amino acid sequence of Formula 7-2 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 7-2 in both of the two Fc domains thereof.

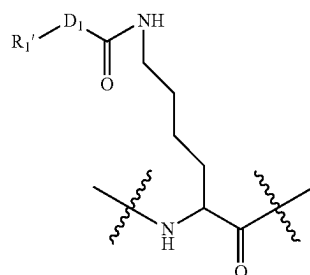
The present invention provides an antibody or a fragment thereof, which includes an amino acid sequence of the following Formula 7-3:

[Formula 7-3]

(SEQ ID NO: 19)

G-P-S-V-F-L-F-P-P- (K)' -P- (K)' -D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and (K)' is



wherein R_1' is a first chemical functional group, and D_1 is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has R_1' connected via lysine 246 and 248 in an Fc domain thereof, or sites corresponding to the lysine 246 and 248.

In specific embodiments, R_1' may include a click-chemistry functional group. Also, the click-chemistry functional group may include one or more selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, the click-chemistry functional group may be selected from an azide, or a strained alkyne. Further, the click-chemistry functional group may be selected from an azide, or dibenzocyclooctyne-amine. Additionally, the click-chemistry functional group may be selected from a diene, or a dienophile. Further, the click-chemistry functional group may be selected from a tetrazine, or a norbornene. Alternatively, the click-chemistry functional group may be selected from a tetrazine, or a trans-cyclooctene. In addition, R_1' may include two or more click-chemistry functional groups.

In other specific embodiments, R_1' may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-CH_2OCH_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody.

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In specific embodiments, the antibody may include the amino acid sequence of Formula 7-3 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 7-3 in both of the two Fc domains thereof.

The present invention provides an antibody or a fragment thereof, which includes one or more amino acid sequences selected from Formulas 7-1, 7-2, and 7-3. In this case, the contents of the sequences of Formulas 7-1 to 7-3 are as described above.

In specific embodiments, D_1 may be a covalent bond.

In specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 7-1 and may not include the amino acid sequences of Formulas 7-2 and 7-3. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 7-1 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 7-1 in both of the two Fc domains thereof.

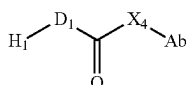
In other specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 7-2 and may not include the amino acid sequences of Formulas 7-1 and 7-3. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 7-2 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 7-2 in both of the two Fc domains thereof.

In specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 7-3 and may not include the amino acid sequences of Formulas 7-1 and 7-2. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 7-3 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 7-3 in both of the two Fc domains thereof.

5.1. Antibody containing first click-chemistry functional group

According to the present invention, there is disclosed an antibody containing a first click-chemistry functional group. Such a compound is herein indicated by the symbol " H_1 -Ab."

The present invention provides an H_1 -Ab represented by Formula 8:



wherein H_1 is a first click-chemistry functional group, D_1 is any alkylene, alkenylene, or alkynylene, X_4 is NH, O, or S, and Ab is an antibody.

In specific embodiments, H_1 may include any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, H_1 may be an azide, or a strained alkyne. Further, H_1 may be an azide, or dibenzocyclooctyne-amine. Additionally, H_1 may be a diene, or a dienophile. Further, H_1 may be a tetrazine, or a norbornene. Alternatively, H_1 may be a tetrazine, or a trans-cyclooctene.

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In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-\text{CH}_2\text{OCH}_2-$. In addition, D_1 may be a covalent bond.

In one specific embodiment, X_4 may be NH.

In specific embodiments, Ab may be a human antibody. In other specific embodiments, Ab may be a non-human animal antibody. In specific embodiments, Ab may be immunoglobulin G (IgG). In specific embodiments, Ab may be a whole antibody. In other specific embodiments, Ab may be a fragment of the antibody. In specific embodiments, Ab may be a wild-type antibody. In other specific embodiments, Ab may be a manipulated antibody.

In specific embodiments, X_4 and Ab may be connected via an Fab domain of Ab. In other specific embodiments, X_4 and Ab may be connected via an Fc domain of Ab. Also, X_4 and Ab may be connected via lysine 246 or 248 in the Fc domain of Ab. Furthermore, X_4 and Ab may be connected via lysine 246 in the Fc domain of Ab. Further, X_4 and Ab may be connected via lysine 248 in the Fc domain of Ab. Alternatively, X_4 and Ab may be connected via lysine 246 and 248 in the Fc domain of Ab. In specific embodiments, X_4 and Ab may be connected via only one of two Fc domains of Ab. In other specific embodiments, X_4 and Ab may be connected via both of the two Fc domains of Ab.

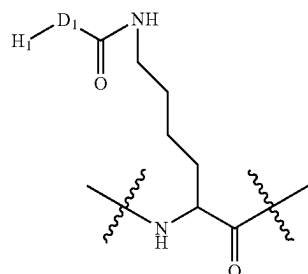
The present invention provides an antibody or a fragment thereof, which includes an amino acid sequence of the following Formula 8-1:

[Formula 8-1]

(SEQ ID NO: 20)

G-P-S-V-F-L-F-P-P-(K)-P-K-D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and (K)' is



wherein H_1 is a first click-chemistry functional group, and D_1 is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has H_1 connected via lysine 246 in an Fc domain thereof, or a site corresponding to the lysine 246.

In specific embodiments, H_1 may include any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, H_1 may be an azide, or a strained alkyne. Further, H_1 may be an azide, or dibenzocyclooctyne-amine. Additionally, H_1 may be a diene, or a dienophile. Further, H_1 may be a tetrazine, or a norbornene. Alternatively, H_1 may be a tetrazine, or a trans-cyclooctene.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a

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C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-\text{CH}_2\text{OCH}_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody. In specific embodiments, the antibody may be a wild-type antibody. In other specific embodiments, the antibody may be a manipulated antibody.

In specific embodiments, the antibody may include the amino acid sequence of Formula 8-1 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 8-1 in both of the two Fc domains thereof.

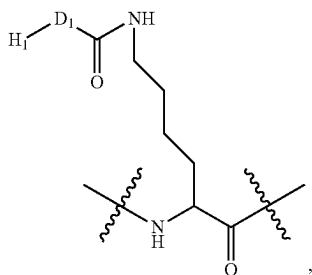
The present invention provides an antibody or a fragment thereof, which includes an amino acid sequence of the following Formula 8-2:

[Formula 8-2]

(SEQ ID NO: 21)

G-P-S-V-F-L-F-P-P-K-(K)'-D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and (K)' is



wherein H_1 is a first click-chemistry functional group, and D_1 is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has H_1 connected via lysine 248 in an Fc domain thereof, or a site corresponding to the lysine 248.

In specific embodiments, H_1 may include any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, H_1 may be an azide, or a strained alkyne. Further, H_1 may be an azide, or dibenzocyclooctyne-amine. Additionally, H_1 may be a diene, or a dienophile. Further, H_1 may be a tetrazine, or a norbornene. Alternatively, H_1 may be a tetrazine, or a trans-cyclooctene.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-\text{CH}_2\text{OCH}_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific

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embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody. In specific embodiments, the antibody may be a wild-type antibody. In other specific embodiments, the antibody may be a manipulated antibody.

In specific embodiments, the antibody may include the amino acid sequence of Formula 8-2 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 8-2 in both of the two Fc domains thereof.

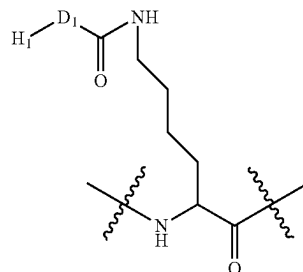
The present invention provides an antibody or a fragment thereof, which includes an amino acid sequence of the following Formula 8-3:

[Formula 8-3]

(SEQ ID NO: 22)

G-P-S-V-F-L-F-P-P-(K)'-P-(K)'-D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and (K)' is



wherein H_1 is a first click-chemistry functional group, and D_1 is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has H_1 connected via lysine 246 and 248 in an Fc domain thereof, or sites corresponding to the lysine 246 and 248.

In specific embodiments, H_1 may include any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, H_1 may be an azide, or a strained alkyne. Further, H_1 may be an azide, or dibenzocyclooctyne-amine. Additionally, H_1 may be a diene, or a dienophile. Further, H_1 may be a tetrazine, or a norbornene. Alternatively, H_1 may be a tetrazine, or a trans-cyclooctene.

In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-\text{CH}_2\text{OCH}_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody. In specific embodiments, the antibody may be a wild-type antibody. In other specific embodiments, the antibody may be a manipulated antibody.

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In specific embodiments, the antibody may include the amino acid sequence of Formula 8-3 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 8-3 in both of the two Fc domains thereof.

The present invention provides an antibody or a fragment thereof, which includes one or more amino acid sequences selected from Formulas 8-1, 8-2, and 8-3. In this case, the contents of the sequences of Formulas 8-1 to 8-3 are as described above.

In specific embodiments, D_1 may be a covalent bond.

In specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 8-1 and may not include the amino acid sequences of Formulas 8-2 and 8-3. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 8-1 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 8-1 in both of the two Fc domains thereof.

In other specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 8-2 and may not include the amino acid sequences of Formulas 8-1 and 8-3. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 8-2 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 8-2 in both of the two Fc domains thereof.

In specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 8-3 and may not include the amino acid sequences of Formulas 8-1 and 8-2. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 8-3 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 8-3 in both of the two Fc domains thereof.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody. In specific embodiments, the antibody may be a wild-type antibody. In other specific embodiments, the antibody may be a manipulated antibody.

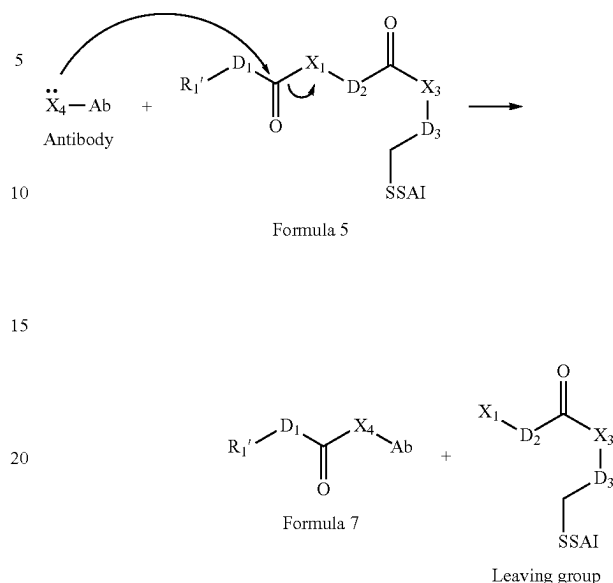
5.2. Method of Preparing Antibody Containing First Chemical Functional Group

According to the present invention, there is disclosed a method of preparing the R_1' -Ab, and H_1 -Ab (hereinafter generally referred to as " R_1' -Ab"). It will be noted that a description of the following preparation methods is provided to aid in understanding the present invention.

For example, a method of preparing a compound of Formula 7 will be described. In specific embodiments, the compound of Formula 7 may be prepared through a reaction of the following Scheme 4.

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[Scheme 4]

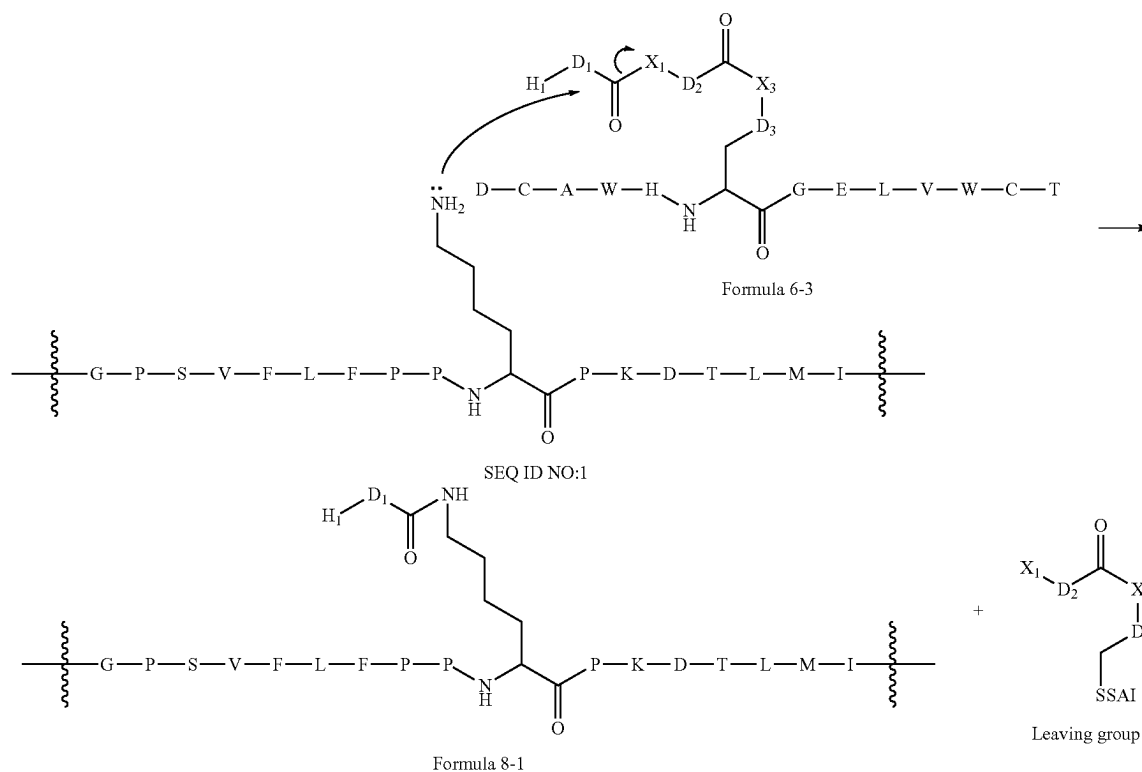


Because a first carbonyl group of the linker according to the present invention has mild reactivity, cross-linking may be realized when the first carbonyl group of the linker has a close positional relationship with an amine group of an antibody. First, the R_1' -L₂-SSAI represented by Formula 5 is brought into close contact with a certain site of the antibody to create an environment in which a reaction may occur. Because the R_1' -L₂-SSAI according to the present invention has an activated carbonyl group (i.e., a carbonyl group connected to X_1) corresponding to the first carbonyl group of the linker, the R_1' -L₂-SSAI of the present invention may trigger a nucleophilic substitution reaction. In this case, the compound of Formula 7 may be prepared since an atom (X_4) having a free electron pair present in the antibody serves as a nucleophile to attack the carbonyl group. In this case, by design of the linker, a site-specific antibody interactome (SSAI) leaves while being included in a leaving group and the SSAI is removed from the final product. These characteristics have a positive effect on physical properties of an antibody product, as shown in the section 5.8 below.

In specific embodiments, the X_4 may be NH_2 . Also, the X_4 may be NH_2 of a lysine residue. In other specific embodiments, the X_4 may be SH. Also, the X_4 may be SH of a cysteine residue. In other specific embodiments, the X_4 may be OH.

As a more specific example, a method of preparing an antibody or a fragment thereof, which includes the amino acid sequence of Formula 8-3, will be described. In specific embodiments, the compound including the amino acid sequence of Formula 8-3 may be prepared through a reaction of the following Scheme 5.

[Scheme 5]



The compound having a structure of Formula 6-3 (a structure in which two cysteine residues are optionally connected) is directed towards the Fc domain of the antibody because the compound has an SSFI sequence (see section 3.2). In this case, the antibody including an amino acid sequence of SEQ ID NO: 1 includes lysine residues in an Fc domain thereof, and such a lysine residue may serve as a nucleophile. The exemplary embodiment shows a case where lysine 246 in the Fc domain or a residue corresponding to the lysine 246 (hereinafter referred to as "lysine 246") serves as a nucleophile. In this case, an amine group of the lysine 246 attacks a first carbonyl group of Formula 6-3 to produce a compound of Formula 8-1. In this way, H₁ or R₁ may be transferred to the lysine residue of the Fc domain of the antibody. In this case, to which certain lysine residue the chemical functional group is transferred may depend on the design of the linker, the SSAI, and the R₁'-L₂-SSAI, as described in the sections 5.3 and 5.4 below.

According to the present invention, there is disclosed a method of preparing an antibody containing a first chemical functional group.

The present invention provides a method of preparing an R₁'-Ab, which includes:

reacting an agent for transferring a first chemical functional group to an antibody according to the present invention with an antibody or a fragment thereof.

In specific embodiments, the agent for transferring a first chemical functional group to an antibody may be any one selected from Formulas 5, 5-1 to 5-3, and 6-1 to 6-3.

In specific embodiments, the antibody or fragment thereof may be a human antibody. In other specific embodiments, the antibody or fragment thereof may be a non-human

animal antibody. In specific embodiments, the antibody or fragment thereof may be immunoglobulin G (IgG). In specific embodiments, the antibody or fragment thereof may be a whole antibody. In other specific embodiments, the antibody or fragment thereof may be a fragment of the antibody. In specific embodiments, the antibody or fragment thereof may be a wild-type antibody. In other specific embodiments, the antibody or fragment thereof may be a manipulated antibody.

In specific embodiments, the present invention provides a method of preparing an antibody having a first chemical functional group transferred to a specific lysine residue of an Fc domain thereof. In this case, those of aspects as described in the section 5.4 below may be used as the agent for transferring a first chemical functional group to an antibody.

According to the present invention, there is disclosed a method of preparing an antibody containing a first click-chemistry functional group.

The present invention provides a method of preparing an H₁-Ab, which includes:

reacting an agent for transferring a first click-chemistry functional group to an antibody according to the present invention with an antibody or a fragment thereof.

In specific embodiments, the agent for transferring a first click-chemistry functional group to an antibody may be any one selected from Formulas 6-1 to 6-3. Also, the agent for transferring a first click-chemistry functional group to an antibody may have the structure of Formula 6-3.

In specific embodiments, the antibody or fragment thereof may be a human antibody. In other specific embodiments, the antibody or fragment thereof may be a non-human animal antibody. In specific embodiments, the antibody or

fragment thereof may be immunoglobulin G (IgG). In specific embodiments, the antibody or fragment thereof may be a whole antibody. In other specific embodiments, the antibody or fragment thereof may be a fragment of the antibody. In specific embodiments, the antibody or fragment thereof may be a wild-type antibody. In other specific embodiments, the antibody or fragment thereof may be a manipulated antibody.

In specific embodiments, the present invention provides a method of preparing an antibody having a first click-chemistry functional group transferred to a specific lysine residue of an Fc domain thereof. In this case, those of aspects as described in the section 5.4 below may be used as the agent for transferring a first click-chemistry functional group to an antibody.

According to the present invention, there is disclosed a kit for preparing an antibody or a fragment thereof containing a first chemical functional group.

The present invention provides a kit for preparing an antibody or a fragment thereof containing a first chemical functional group, which includes an agent for transferring a first chemical functional group to an antibody according to the present invention, and an antibody or a fragment thereof.

In specific embodiments, the agent for transferring a first chemical functional group to an antibody may be any one selected from Formulas 5, 5-1 to 5-3, and 6-1 to 6-3.

In specific embodiments, the antibody or fragment thereof may be a human antibody. In other specific embodiments, the antibody or fragment thereof may be a non-human animal antibody. In specific embodiments, the antibody or fragment thereof may be immunoglobulin G (IgG). In specific embodiments, the antibody or fragment thereof may be a whole antibody. In other specific embodiments, the antibody or fragment thereof may be a fragment of the antibody. In specific embodiments, the antibody or fragment thereof may be a wild-type antibody. In other specific embodiments, the antibody or fragment thereof may be a manipulated antibody.

Also, the present invention provides a kit for preparing an antibody or a fragment thereof containing a first chemical functional group, which includes:

- a linker ($R_1'-L_1$) according to the present invention;
- a site-specific antibody interactome according to the present invention; and
- an antibody or a fragment thereof.

In specific embodiments, the linker may be any one selected from Formulas 1, 2, and 2-1 to 2-3.

In specific embodiments, the site-specific antibody interactome may be any one selected from Formulas 3, 3-1, and 4-1 to 4-6.

In specific embodiments, the antibody or fragment thereof may be a human antibody. In other specific embodiments, the antibody or fragment thereof may be a non-human animal antibody. In specific embodiments, the antibody or fragment thereof may be immunoglobulin G (IgG). In specific embodiments, the antibody or fragment thereof may be a whole antibody. In other specific embodiments, the antibody or fragment thereof may be a fragment of the antibody. In specific embodiments, the antibody or fragment thereof may be a wild-type antibody. In other specific embodiments, the antibody or fragment thereof may be a manipulated antibody.

The present invention provides a kit for preparing an antibody having a first chemical functional group transferred to a specific lysine residue of an Fc domain thereof. In this case, those of aspects as described in the section 5.4 below may be used as the agent for transferring a first chemical

functional group to an antibody, the linker, and the site-specific antibody interactome.

According to the present invention, there is disclosed a kit for preparing an antibody or a fragment thereof containing a first click-chemistry functional group.

The present invention provides a kit for preparing an antibody or a fragment thereof containing a first click-chemistry functional group, which includes an agent for transferring a first click-chemistry functional group to an antibody according to the present invention, and an antibody or a fragment thereof.

In specific embodiments, the agent for transferring a first click-chemistry functional group to an antibody may be any one selected from Formulas 6-1 to 6-3. Also, the agent for transferring a first click-chemistry functional group to an antibody may have the structure of Formula 6-3.

In specific embodiments, the antibody or fragment thereof may be a human antibody. In other specific embodiments, the antibody or fragment thereof may be a non-human animal antibody. In specific embodiments, the antibody or fragment thereof may be immunoglobulin G (IgG). In specific embodiments, the antibody or fragment thereof may be a whole antibody. In other specific embodiments, the antibody or fragment thereof may be a fragment of the antibody. In specific embodiments, the antibody or fragment thereof may be a wild-type antibody. In other specific embodiments, the antibody or fragment thereof may be a manipulated antibody.

Also, the present invention provides a kit for preparing an antibody or a fragment thereof containing a first click-chemistry functional group, which includes:

- a linker (H_1-L_1) according to the present invention;
- a site-specific antibody interactome according to the present invention; and
- an antibody or a fragment thereof.

In specific embodiments, the linker may be any one selected from Formulas 2, and 2-1 to 2-3.

In specific embodiments, the site-specific antibody interactome may be any one selected from Formulas 3, 3-1, and 4-1 to 4-6.

In specific embodiments, the antibody or fragment thereof may be a human antibody. In other specific embodiments, the antibody or fragment thereof may be a non-human animal antibody. In specific embodiments, the antibody or fragment thereof may be immunoglobulin G (IgG). In specific embodiments, the antibody or fragment thereof may be a whole antibody. In other specific embodiments, the antibody or fragment thereof may be a fragment of the antibody. In specific embodiments, the antibody or fragment thereof may be a wild-type antibody. In other specific embodiments, the antibody or fragment thereof may be a manipulated antibody.

The present invention provides a kit for preparing an antibody having a first click-chemistry functional group transferred to a specific lysine residue of an Fc domain thereof. In this case, those of aspects as described in the section 5.4 below may be used as the agent for transferring a first click-chemistry functional group to an antibody, the linker, and the site-specific antibody interactome.

5.3. Function of $(Xa_1)'$ and Design Principle of Location of $(Xa_1)'$ on $R_1'-L_2$ -SSFI

In the present content, the compound of Formula 6-3 is provided as one example to aid in understanding the present invention, but the scope of the present invention is not limited thereto. It will be noted that the following description also applies to the compounds of Formulas 5, 5-1 to 5-3,

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and 6-1 to 6-3, and the compound of Formula 6-3 is merely provided as one example for the sake of convenience.

As discussed in the section 5.2, (Xa₁)' of the R₁'-L₂-SSFI functions to transfer R₁' to an antibody by means of a nucleophilic substitution reaction. According to the present invention, it is assumed that the conditions for facilitating a nucleophilic substitution reaction satisfy the following requirements: (1) (Xa₁)' is adjacent to a lysine residue of an Fc domain, and (2) a side chain to which R₁' is bound is directed towards the lysine residue. It was expected that the yield and uniformity of a process would drop as the position of (Xa₁)' and the direction of the side chain become farther from the lysine residue of the Fc domain. Also, (3) it will be preferred that a substitution position of (Xa₁)' does not have a great influence on the interaction between the SSFI and the Fc domain (see section 3.2).

As indirectly seen in FIGS. 3 and 5, it was confirmed that the positions in the SSFI that satisfy the requirement of (1) in relation to lysine 246 and 248 in the Fc domain are positions 5, 6, 7, and 8 based on the following Formula 6-3.

D-C-A-W-H—(Xa₁)'-G-E-L-V-W-C-T (SEQ ID NO:

16)

[Formula 6-3]

In this case, as histidine corresponding to position 5 forms a salt linkage with glutamic acid 380 in the Fc domain, replacement of the histidine may affect the interaction between the SSFI and the Fc domain. Therefore, this does not satisfy the requirement of (3). Also, because the direction of this side chain is not close to the lysine 246 and 248, this does not satisfy the requirement of (2). Because glycine corresponding to position 7 helps to form a bent structure of the SSFI, it is not desirable to replace the glycine residue with a large (Xa₁)' residue. As glutamic acid corresponding to position 8 forms a salt linkage with arginine 255 in the Fc domain, replacement of the glutamic acid may affect the interaction between the SSFI and the Fc domain. Therefore, this does not satisfy the requirement of (3) (see section 3.2). It was judged that position 6 is most suitable for the position of (Xa₁)' because position 6 satisfies all the requirements of (1), (2), and (3). Therefore, the R₁'-L₂-SSFI according to the present invention has been completed based on these facts.

5.4. The position of R₁' transferred to an antibody may vary depending on the length of D₂ in R₁'-L₁ and D₃ in SSFI.

The present content is intended to explain a preferred design principle of the R₁'-L₂-SSFI according to the present invention. According to the design of the R₁'-L₂-SSFI, it is possible to specifically transfer R₁' to a specific lysine residue of the Fc domain. Also, the present content is intended to explain a design principle of the R₁'-L₁ and SSFI to prepare a preferred R₁'-L₂-SSFI.

A person of ordinary skill in the art who reads the section 5.2 as described above may recognize that a nucleophilic substitution reaction may occur when the first carbonyl carbon of (Xa₁)' of the R₁'-L₂-SSFI is located adjacent to an amine group of lysine of the Fc domain. Based on the description of the section 3.2 as described above, a person of ordinary skill in the art may also recognize that the R₁'-L₂-SSFI is arranged with the Fc domain with a specific topology, wherein amine groups of lysine 246 and 248 in the Fc domain are spaced apart a certain distance from the beta carbon of (Xa₁)' (FIG. 5). By using this combination to design the R₁'-L₂-SSFI, it was expected that it is possible to specifically label a desired lysine residue when (1) a distance (hereinafter generally referred to as "L_c") between the beta carbon of (Xa₁)' and the first carbonyl carbon is the same as

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or similar to (2) a distance between the beta carbon of (Xa₁)' and the amine groups of lysine 246 and 248 in the Fc domain in the specific topology.

The structure of the R₁'-L₂-SSFI according to the present invention and the L_c are shown in FIG. 8. As shown in FIG. 8, D₃, X₃, carbon atoms, D₂, and X₁, are located between the beta carbon of (Xa₁)' and the first carbonyl carbon. Among these, D₃ and X₃ are associated with the design of SSFI, and D₂ and X₁, are associated with the design of L₁-R₁'.

For the sake of convenience, the invention was embodied on the assumption that D₃ is a C_x alkylene, X₃ is N, D₂ is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and X₁, is S, wherein y is an integer greater than or equal to 1. In this case, it was assumed that the alkylene, the alkenylene, and the alkynylene have the same length. The structure of the linker shown in FIG. 8 was modeled using the software Discovery Studio to calculate L_c values. As a result, the L_c values determined with respect to the x+y value are listed in Table 3. In the calculation process, the chain was modeled when it had the longest length.

TABLE 3

Lc values with respect to x + y value	
x + y	Lc (Å)
1 (x = 0, y = 1)	6.616
2	7.784
3	9.102
4	10.299
5	11.599
6	12.816
7	14.100
8	15.334
9	16.601
10	17.848
11	19.099
12	20.357
13	21.592
14	22.857

Hereinafter, the accompanying drawings provided to show a topology between the R₁'-L₂-SSFI and a lysine residue of the Fc domain are provided based on FIG. 9. FIG. 9 shows a topology between an R₁'-L₂-SSFI and an Fc domain so that a direction (a dotted arrow) of a side chain of (Xa₁)' is parallel with the x axis in the drawing. As shown in FIG. 9, it can be seen that the side chain of (Xa₁)' is directed towards the amine groups of the lysine 246 and 248 (see section 5.3). Also, it can be seen from the drawing that the side chain of (Xa₁)' may specifically react with the lysine 246 or 248 depending on the length of L_c. FIGS. 10 to 12 are diagrams viewing a diagram of FIG. 9 in a direction (a thick solid line arrow) parallel with the y-axis in the drawing along the exemplary length of L_c.

The distances (D_{246,min}, D_{246,max}, D_{248,min}, and D_{248,max}) between the beta carbon of (Xa₁)' and the amine groups of lysine 246 and 248 in the Fc domain are described in the section 3.2. The beta carbon of (Xa₁)' is closer to the lysine 248 in the Fc domain than the lysine residue at position 246. Therefore, it was expected that the first carbonyl carbon would react well with the lysine 248 when the L_c value is shorter than D_{246,min} (FIG. 10), the first carbonyl carbon would react well with the lysine 246 when the L_c value is longer than D_{248,max} (FIG. 11), and the first carbonyl carbon would selectively react with the lysine 246 and 248 when the L_c value is longer than or equal to D_{246,min} and shorter than or equal to D_{248,max} (FIG. 12).

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According to the present invention, there is disclosed an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 248 in an Fc domain of an antibody (see FIG. 11).

The present invention provides an agent for transferring a first chemical functional group to an antibody, characterized in that a distance (L_c) between the beta carbon of $(Xa_1)'$ and the first carbonyl carbon is shorter than $D_{246,min}$ (approximately 11.668 Å). In this case, L_c may have a value of approximately 6.5 Å, approximately 7 Å, approximately 8 Å, approximately 9 Å, approximately 10 Å, approximately 11 Å, or approximately 11.5 Å.

In specific embodiments, when the R_1' - L_2 -SSFI has any one structure selected from Formulas 5, 5-1 to 5-3, and 6-1 to 6-3, D_3 is a C_x alkylene, X_3 is N, D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, X_1 is S, wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $1 \leq x+y \leq 5$. Also, the sum of x and y may be 1, 2, 3, 4, or 5. For example, x may be 0, and y may be $1 \leq y \leq 5$. In another exemplary embodiment, x may be 1, and y may be $1 \leq y \leq 4$. In still another exemplary embodiment, x may be 2, and y may be $1 \leq y \leq 3$. In yet another exemplary embodiment, x may be 3, and y may be $1 \leq y \leq 2$. The corresponding numerical range is determined based on the value listed in Table 3.

According to the present invention, there is disclosed an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 246 in an Fc domain of an antibody.

According to one aspect of the present invention, the present invention provides an agent for transferring a first chemical functional group to an antibody, characterized in that a distance (L_c) between the beta carbon of $(Xa_1)'$ and the first carbonyl carbon is longer than $D_{248,max}$ (approximately 16.208 Å). For example, L_c may have a value of approximately 16.5 Å, approximately 17 Å, approximately 18 Å, approximately 19 Å, approximately 20 Å, or approximately 20.5 Å.

In specific embodiments, when the R_1' - L_2 -SSFI has any one structure selected from Formulas 5, 5-1 to 5-3, and 6-1 to 6-3, D_3 is a C_x alkylene, X_3 is N, D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, X_1 is S, wherein y may be an integer greater than or equal to 1, and the sum of x and y may be greater than or equal to 9. Also, the sum of x and y may be 9, 10, 11, or 12. For example, x may be 0, and y may be $9 \leq y \leq 12$. In another exemplary embodiment, x may be 1, and y may be $8 \leq y \leq 11$. In still another exemplary embodiment, x may be 2, and y may be $7 \leq y \leq 10$. In yet another exemplary embodiment, x may be 3, and y may be $6 \leq y \leq 9$. Optionally, D_2 may be an alkynylene. When D_2 is an alkynylene, the side chain of $(Xa_1)'$ may become stereoscopically rigid, thereby preventing bending of the side chain.

According to the present invention, there is disclosed an R_1' - L_2 -SSFI for selectively transferring a first chemical functional group to lysine 246 or 248 in an Fc domain of an antibody.

According to one aspect of the present invention, the present invention provides an agent for transferring a first chemical functional group to an antibody, characterized in that a distance (L_c) between the beta carbon of $(Xa_1)'$ and the first carbonyl carbon has a value longer than or equal to $D_{246,min}$ (approximately 11.668 Å) and shorter than or equal to $D_{248,max}$ (approximately 16.208 Å). In this case, L_c may have a value of approximately 11.668 Å, approximately 12 Å, approximately 13 Å, approximately 14 Å, approximately 15 Å, approximately 15.5 Å, approximately 16 Å, or approximately 16.208 Å.

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In specific embodiments, when the R_1' - L_2 -SSFI has any one structure selected from Formulas 5, 5-1 to 5-3, and 6-1 to 6-3, D_3 is a C_x alkylene, X_3 is N, D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, X_1 is S, wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $6 \leq x+y \leq 8$. In this case, the sum of x and y may be 6, 7, or 8. For example, x may be 0, and y may be $6 \leq y \leq 8$. In another exemplary embodiment, x may be 1, and y may be $5 \leq y \leq 7$. In still another exemplary embodiment, x may be 2, and y may be $4 \leq y \leq 6$. In yet another exemplary embodiment, x may be 3, and y may be $3 \leq y \leq 5$.

According to the present invention, there is disclosed a method of preparing an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 248 in an Fc domain of an antibody.

As one example of the method of preparing an agent for transferring first chemical functional group to an antibody as described in the section 4.2, the present invention provides a method of preparing an R_1' - L_2 -SSAI, which is characterized by including:

reacting a linker according to the present invention with a site-specific antibody interactome according to the present invention,

wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $1 \leq x+y \leq 5$. Also, the R_1' - L_2 -SSAI prepared by the method may react with an antibody to specifically transfer a first chemical functional group to lysine 248 in an Fc domain of the antibody.

According to the present invention, there is disclosed a kit for preparing an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 248 in an Fc domain of an antibody.

As one example of the method of preparing an agent for transferring first chemical functional group to an antibody as described in the section 4.2, the present invention provides a kit for preparing an R_1' - L_2 -SSAI, which is characterized by including:

a linker according to the present invention; and

a site-specific antibody interactome according to the present invention,

wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $1 \leq x+y \leq 5$.

According to the present invention, there is disclosed a method of preparing an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 246 in an Fc domain of an antibody.

As one example of the method of preparing an agent for transferring first chemical functional group to an antibody as described in the section 4.2, the present invention provides a method of preparing an R_1' - L_2 -SSAI, which is characterized by including:

reacting a linker according to the present invention with a site-specific antibody interactome according to the present invention,

wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

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wherein y is an integer greater than or equal to 1, and the sum of x and y is $9 \leq x+y \leq 12$. Also, the R_1' - L_2 -SSAI prepared by the method may react with an antibody to specifically transfer a first chemical functional group to lysine 246 in an Fc domain of the antibody.

According to the present invention, there is disclosed a kit for preparing an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 246 in an Fc domain of an antibody.

As one example of the method of preparing an agent for transferring first chemical functional group to an antibody as described in the section 4.2, the present invention provides a kit for preparing an R_1' - L_2 -SSAI, which is characterized by including:

a linker according to the present invention; and
a site-specific antibody interactome according to the present invention,

wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $9 \leq x+y \leq 12$.

According to the present invention, there is disclosed a method of preparing an R_1' - L_2 -SSFI for selectively transferring a first chemical functional group to lysine 246 or 248 in an Fc domain of an antibody.

As one example of the method of preparing an agent for transferring first chemical functional group to an antibody as described in the section 4.2, the present invention provides a method of preparing an R_1' - L_2 -SSAI, which is characterized by including:

reacting a linker according to the present invention with a site-specific antibody interactome according to the present invention,

wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $6 \leq x+y \leq 8$. Also, the R_1' - L_2 -SSAI prepared by the method may react with an antibody to selectively transfer a first chemical functional group to lysine 246 or 248 in an Fc domain of the antibody.

According to the present invention, there is disclosed a kit for preparing an R_1' - L_2 -SSFI for selectively transferring a first chemical functional group to lysine 246 or 248 in an Fc domain of an antibody.

As one example of the method of preparing an agent for transferring first chemical functional group to an antibody as described in the section 4.2, the present invention provides a kit for preparing an R_1' - L_2 -SSAI, which is characterized by including:

a linker according to the present invention; and
a site-specific antibody interactome according to the present invention,

wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $6 \leq x+y \leq 8$.

According to the present invention, there is disclosed a method of preparing an antibody having a first chemical functional group specifically transferred to lysine 248 in an Fc domain thereof.

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As one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a method of preparing an R_1' -Ab, which is characterized by including:

reacting an agent for transferring a first chemical functional group to an antibody according to the present invention with an antibody or a fragment thereof,

wherein, in the agent for transferring a first chemical functional group to an antibody, a distance (L_c) between the beta carbon of $(X_{A1})'$ and the first carbonyl carbon is shorter than approximately 11.668 Å.

In specific embodiments, in the agent for transferring a first chemical functional group to an antibody, D_3 is a C_x alkylene, and D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene,

wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $1 \leq x+y \leq 5$.

According to the present invention, there is disclosed a kit for preparing an antibody having a first chemical functional group specifically transferred to lysine 248 in an Fc domain thereof.

As one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a kit for preparing an R_1' -Ab, which is characterized by including:

an agent for transferring a first chemical functional group to an antibody according to the present invention; and
an antibody or a fragment thereof,

wherein, in the agent for transferring a first chemical functional group to an antibody, a distance (L_c) between the beta carbon of $(X_{A1})'$ and the first carbonyl carbon is shorter than approximately 11.668 Å.

In specific embodiments, in the agent for transferring a first chemical functional group to an antibody, D_3 is a C_x alkylene, and D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene,

wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $1 \leq x+y \leq 5$.

Optionally, as one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a kit for preparing an R_1' -Ab, which is characterized by including:

a linker according to the present invention;
a site-specific antibody interactome according to the present invention; and
an antibody or a fragment thereof,

wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $1 \leq x+y \leq 5$.

According to the present invention, there is disclosed a method of preparing an antibody having a first chemical functional group specifically transferred to lysine 246 in an Fc domain thereof.

As one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a method of preparing an R_1' -Ab, which is characterized by including:

allowing an antibody or a fragment thereof to react with an agent for transferring a first chemical functional group to an antibody according to the present invention,

wherein, in the agent for transferring a first chemical functional group to an antibody, a distance (L_c)

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between the beta carbon of $(Xa_1)'$ and the first carbonyl carbon is longer than approximately 16.208 Å.

In specific embodiments, in the agent for transferring a first chemical functional group to an antibody, D_3 is a C_x alkylene, and D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene,

wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $9 \leq x+y \leq 12$.

According to the present invention, there is disclosed a kit for preparing an antibody having a first chemical functional group specifically transferred to lysine 246 in an Fc domain thereof.

As one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a kit for preparing an R_1' -Ab, which is characterized by including:

an agent for transferring a first chemical functional group to an antibody according to the present invention; and an antibody or a fragment thereof,

wherein, in the agent for transferring a first chemical functional group to an antibody, a distance (L_c) between the beta carbon of $(Xa_1)'$ and the first carbonyl carbon is longer than approximately 16.208 Å.

In specific embodiments, in the agent for transferring a first chemical functional group to an antibody, D_3 is a C_x alkylene, and D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene,

wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $9 \leq x+y \leq 12$.

Optionally, as one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a kit for preparing an R_1' -Ab, which is characterized by including:

a linker according to the present invention; a site-specific antibody interactome according to the present invention; and

an antibody or a fragment thereof, wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $9 \leq x+y \leq 12$.

According to the present invention, there is disclosed a method of preparing an antibody having a first chemical functional group selectively transferred to lysine 246 or 248 in an Fc domain thereof.

As one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a method of preparing an R_1' -Ab, which is characterized by including:

reacting an agent for transferring a first chemical functional group to an antibody according to the present invention with an antibody or a fragment thereof,

wherein, in the agent for transferring a first chemical functional group to an antibody, a distance (L_c) between the beta carbon of $(Xa_1)'$ and the first carbonyl carbon is longer than or equal to approximately 11.668 Å and shorter than or equal to approximately 16.208 Å.

In specific embodiments, in the agent for transferring a first chemical functional group to an antibody, D_3 is a C_x alkylene, and D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene,

wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $6 \leq x+y \leq 8$.

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According to the present invention, there is disclosed a kit for preparing an antibody having a first chemical functional group selectively transferred to lysine 246 or 248 in an Fc domain thereof.

As one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a kit for preparing an R_1' -Ab, which is characterized by including:

an agent for transferring a first chemical functional group to an antibody according to the present invention; and an antibody or a fragment thereof,

wherein, in the agent for transferring a first chemical functional group to an antibody, a distance (L_c) between the beta carbon of $(Xa_1)'$ and the first carbonyl carbon is longer than or equal to approximately 11.668 Å and shorter than or equal to approximately 16.208 Å.

In specific embodiments, in the agent for transferring a first chemical functional group to an antibody, D_3 is a C_x alkylene, and D_2 is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene,

wherein y may be an integer greater than or equal to 1, and the sum of x and y may be $6 \leq x+y \leq 8$.

Optionally, as one example of the method of preparing an antibody containing a first chemical functional group as described in the section 5.2, the present invention provides a kit for preparing an R_1' -Ab, which is characterized by including:

a linker according to the present invention; a site-specific antibody interactome according to the present invention; and

an antibody or a fragment thereof, wherein D_2 in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and

D_3 in the site-specific antibody interactome is a C_x alkylene,

wherein y is an integer greater than or equal to 1, and the sum of x and y is $6 \leq x+y \leq 8$.

According to the present invention, there is disclosed a method of preparing an antibody having a first chemical functional group transferred to both lysine 246 and 248 in an Fc domain thereof.

The present invention provides a method of preparing an R_1' -Ab, which includes:

reacting a first agent for transferring a first chemical functional group to an antibody with an antibody or a fragment thereof; and

reacting a second agent for transferring a first chemical functional group to an antibody with the antibody or fragment thereof.

In specific embodiments, the first agent for transferring a first chemical functional group to an antibody may be an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 248 in the Fc domain as described above, and the second agent for transferring a first chemical functional group to an antibody may be an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 246 in the Fc domain as described above.

In specific embodiments, the first agent for transferring a first chemical functional group to an antibody may be an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 246 in the Fc domain as described above, and the second agent for transferring a first chemical functional group to an antibody may be an R_1' - L_2 -SSFI for specifically transferring a first chemical functional group to lysine 248 in the Fc domain as described above.

In specific embodiments, the reacting of the first agent for transferring a first chemical functional group to an antibody

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with the antibody or fragment thereof; and the reacting of the second agent for transferring a first chemical functional group to an antibody with the antibody or fragment thereof may be sequentially performed.

According to the present invention, there is disclosed a kit for preparing an antibody having a first chemical functional group transferred to both lysine 246 and 248 in an Fc domain thereof.

The present invention provides a kit for preparing an R₁'-Ab, which includes:

- a first agent for transferring a first chemical functional group to an antibody;
- a second agent for transferring a first chemical functional group to an antibody; and
- an antibody or a fragment thereof.

In specific embodiments, the first agent for transferring a first chemical functional group to an antibody may be an R₁'-L₂-SSFI for specifically transferring a first chemical functional group to lysine 248 in the Fc domain as described above, and the second agent for transferring a first chemical functional group to an antibody may be an R₁'-L₂-SSFI for specifically transferring a first chemical functional group to lysine 246 in the Fc domain as described above.

In specific embodiments, the first agent for transferring a first chemical functional group to an antibody may be an R₁'-L₂-SSFI for specifically transferring a first chemical functional group to lysine 246 in the Fc domain as described above, and the second agent for transferring a first chemical functional group to an antibody may be an R₁'-L₂-SSFI for specifically transferring a first chemical functional group to lysine 248 in the Fc domain as described above.

5.5. It is Possible to Transfer First Chemical Functional Group More Site-Specifically by Modulating the Reactivity of the First Carbonyl Group.

As described above in the section 5.2, an antibody containing a first chemical functional group may be prepared by a nucleophile of the antibody attacking a first carbonyl group of the R₁'-L₂-SSFI. The first carbonyl group is characterized by having milder reactivity than the second carbonyl group of the linker (see section 2.2). The reactivity of the first carbonyl group may be modulated so that a certain amine group of the antibody can react with the first carbonyl group only when the certain amine group is in the vicinity of the first carbonyl group. In this way, the present invention allows the first carbonyl group not to react well with any amine group through the vicinity conditions for reaction, and enables high positional specificity of the reaction.

The prior art disclosed in Publication Nos. US 2018/0141976 A1 and WO 2018/199337 A1 aims to site-selectively modulate lysine 246 or 248 in an Fc domain using an analogue of Fc-III. Because the prior art uses a disuccinimidyl cross-linker, the first carbonyl group and the second carbonyl group are equally highly reactive (see section 2.2). Therefore, because the first carbonyl group has high reactivity and has no vicinity conditions for reaction, any lysine residue is highly likely to be labeled.

5.6. The antibody containing a first chemical functional group prepared according to the present invention has high uniformity and yield.

The technical problem and the technical solution to specifically transfer (1) "a desired number" of first chemical functional groups to (2) "certain sites of an antibody" have been described in detail with reference to the sections 5.3 and 5.4. As can be seen from Experimental Examples below, it can be seen that the chemical functional group of the antibody containing a first chemical functional group prepared by means of the technical solution is transferred to a

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certain lysine residue in the Fc domain thereof with high specificity and yield (see Experimental Example 3.3). Therefore, the present invention has been completed based on these facts.

As shown in the background art, the conjugate product having high uniformity due to antibody labeling has advantages in that (1) the functions of the antibody conjugate are uniformly guaranteed, (2) the antibody conjugate is safe due to its predictable effects, and (3) a drop in function of an antibody may be avoided because it is possible to label an antibody while avoiding a functional region of the antibody.

The prior art disclosed in Publication Nos. US 2018/0141976 A1 and WO 2018/199337 A1 aims to site-selectively modulate lysine 246 or 248 in an Fc domain using an analogue of Fc-III. However, according to the prior art, the uniformity and yield of the antibody may be inevitably low due to two reasons. According to the prior art, first, because a distance (L_c) between the beta carbon of (Xa₁)' and the first carbonyl carbon is approximately 15 Å, it is possible to selectively label lysine at either position 246 or 248, but it is impossible to specifically select one of the lysine 246 and 248 to be labeled. Second, a first carbonyl group of a cross-linker according to the prior art has no vicinity conditions for reaction due to high reactivity (see section 5.5). As a result, the first carbonyl group is highly likely to react with any lysine residue. Therefore, the prior art has a drawback in that the uniformity and yield of an antibody-labeled product are inferior to those of the present invention.

5.7. No functions of the antibody are lowered because the antibody containing a first chemical functional group according to the present invention has no FcRn binding site blocked therein.

The design principle for the antibody labeling site being (1) spaced apart from the paratope; and (2) spaced apart from the recognition site of FcR including FcRn has been described above in the section 1.1. Because the lysine 246 and 248 are included in the Fc domain, the lysine 246 and 248 are spaced apart from the paratope, but the corresponding site may overlap the recognition site of FcRn, which may be problematic.

It is known that FcRn has various functions, and particularly plays an important role in extending the half-life of the antibody by participating in IgG recycling. When the antibody is used in vivo, that is, when the antibody is used as a therapeutic agent, a contrast medium, and the like, the interaction between FcRn and the antibody may not occur smoothly, thereby making it impossible for the antibody to function normally due to the short half-life of the antibody.

5.7.1. Lysine 246 and Lysine 248 are Spaced Apart from the FcRn Binding Site of the Antibody

The spacing between the lysine residues in the Fc domain and the FcRn binding site of Fc was an important consideration to choose the lysine residue in the Fc domain. FIG. 13 shows a binding structure between an Fc domain and FcRn and positions of the FcRn binding site and lysine 246 and 248 in the Fc domain. Papers and computer modeling were employed to show a binding structure between the Fc domain and FcRn (Ying T, Ju T W, Wang Y, Prabakaran P, Dimitrov D S., Interactions of IgG1 CH2 and CH3 Domains with FcRn; Front Immunol. 2014; 5:146., Monnet C, Jorieux S, Urbain R, et al., Selection of IgG Variants with Increased FcRn Binding Using Random and Directed Mutagenesis: Impact on Effector Functions. Front Immunol. 2015; 6:39.) (see the left image of FIG. 13). The FcRn binding site and the lysine 246 and 248 disclosed in the same article data are indicated in the Fc domain (see the right image of FIG. 13).

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Referring to the two images, it can be seen that side chains of the lysine 246 and 248 are directed in a direction opposite the FcRn binding site.

From these facts, it was contemplated that, when an antibody labeling site is chosen, the antibody labeling site does not affect the interaction between the FcRn and Fc domain. In effect, it was confirmed that the half-life of the prepared antibody is not reduced so that the antibody labeling site does not influence binding of FcRn as intended (see Experimental Example 5).

5.8. A Process of Preparing the Antibody Containing a First Chemical Functional Group According to the Present Invention is Characterized by SSAI Leaving,

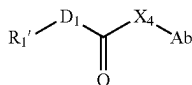
As seen from Schemes 4 and 5 in the section 5.2, a process of preparing the R_1' -Ab according to the present invention is characterized in that SSAI leaves the final product R_1' -Ab because the SSAI is included in a leaving group of a nucleophilic substitution reaction. The binding structure between the Fc domain and the SSFI according to the present invention (see section 3.2) was analyzed in comparison with the binding site between Fc and FcRn. As a result, it was confirmed that a portion of the FcRn binding site is covered with the SSFI (FIG. 14). In this regard, it can be expected that, when the SSFI does not leave during the process of preparing the R_1' -Ab, the SSFI has a negative influence on the physical properties (e.g., half-life, and the like) of the antibody.

The prior art disclosed in Publication No. US 2018/0141976 A1 aims to site-selectively modulate lysine 246 or 248 in an Fc domain using an analogue of Fc-III. However, the corresponding prior art is different from the present invention in that SSFI is included in the final antibody-labeled product because the SSFI does not leave during the preparation process.

The prior art disclosed in Publication No. WO 2018/199337 A1 aims to site-selectively modulate lysine 246 or 248 in an Fc domain using an analogue of Fc-III, wherein SSFI is not included in the final antibody-labeled product. However, the corresponding prior art further includes cleaving a "cleavable linker that is a divalent group" in order to remove the SSFI during the preparation process, but the present invention has an advantage in that the SSFI leaves during a conjugation reaction without any additional processes. Also, the cleaving of the cleavable linker that is a divalent group according to the corresponding invention has a drawback in that a disulfide bond forming a structure of the antibody may be cleaved. Because the present invention uses a nucleophilic substitution reaction which occurs easily without any special conditions, this problem may be dramatically improved.

5.9. No side reactions other than a conjugate formation reaction occurs because the antibody containing a first click-chemistry functional group according to the present invention does not contain a highly bioreactive chemical functional group.

The antibody prepared by the method of preparing an R_1' -Ab according to the present invention has a structure represented by Formula 7:



[Formula 7]

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When R_1' is transferred to lysine 246 or 248, X_4 may be NH, and D_1 may be any alkylene, alkenylene, or alkynylene. In this case, $-NH-(CO)-$ connected to the antibody is an amide bond that is generally stable in vivo. Also, D_1 generally has a structure which is not highly bioreactive. Therefore, it is expected that the R_1' -Ab prepared according to the present invention has high safety because a highly bioreactive structure is not added to molecules other than the labeled molecule (R_1').

In particular, when the R_1' -Ab is an H_1 -Ab having a click-chemistry residue transferred thereto, a highly bioreactive structure is not added to the R_1' -Ab. Therefore, the yield of the click-chemistry reaction may be improved because no secondary reactions other than the click-chemistry reaction occur.

The prior art disclosed Publication No. WO 2018/199337 A1 aims to site-selectively modulate lysine 246 or 248 in an Fc domain using an analogue of Fc-III and use a biorthogonal functional group. However, the corresponding prior art is different from the present invention in that the antibody product according to the corresponding prior art is allowed to have an additional chemical functional group such as thiol, hydroxy, carboxylic acid, phosphoric acid, amine, and the like during the cleaving of the "cleavable linker that is a divalent group." Such an additional chemical functional group is a bioreactive functional group, and thus may affect the safety of the antibody product.

6. Payload (C_m-H_2)

According to the present invention, there is disclosed a payload. Such a compound is herein indicated by the symbol " C_m-H_2 ."

The present invention provides a C_m-H_2 represented by Formula 9:



wherein C_m is a cargo moiety,

H_2 is a second click-chemistry functional group.

In specific embodiments, C_m may include a carrier moiety, a fluorescent moiety, a drug moiety, or a radioactive moiety. Also, R_1' may include a drug moiety. In addition, R_1' may include a VC linker. In other specific embodiments, R_1' may include an antibody or an analogue thereof, which includes a paratope.

In specific embodiments, when C_m includes a drug moiety, the drug moiety may be an anti-cancer drug. Also, the anti-cancer drug may include one or more selected from DM1, DM3, DM4, Abrin, Ricin A, a *Pseudomonas* exotoxin, a Cholera toxin, a Diphtheria toxin, a tumor necrosis factor, a interferon, p interferon, a nerve growth factor, a platelet-derived growth factor, a tissue plasminogen activator, a cytokine, an apoptosis-inducing agent, an anti-angiogenic agent, a lymphokine, taxane, a DNA-alkylating agent, anthracyclin, a Tubulysin analogue, a duocarmycin analogue, auristatin E, auristatin F, a maytansinoid, a cytotoxic agent including a reactive polyethylene glycol residue, taxon, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, T. Colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dihydrotestosterone, a glucocorticoid, procaine, tetracaine, lidocaine, propranolol, puromycin, methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine, mechlorethamine, thiotepe, chlorambucil, Meiphalan, carmustine, lomustine, cyclophosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, cisplatin, dactinomycin, bleomycin, anthramycin, calicheamicin, abiraterone, bendamustine, bortezomib, carbo-

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platin, cabazitaxel, dasatinib, doxorubicin, epirubicin, erlotinib, everolimus, gemcitabine, gefitinib, idarubicin, imatinib, hydroxyurea, lapatinib, leuprorelin, melphalan, nedaplatin, nilotinib, oxaliplatin, pazopanib, pemetrexed, picoplatin, romidepsin, satraplatin, sorafenib, vemurafenib, sunitinib, teniposide, triplatin, and vinorelbine.

In specific embodiments, C_m may include a plurality of carrier moieties, fluorescent moieties, drug moieties, or radioactive moieties. Also, C_m may include two or more drug moieties. In addition, C_m may include a VC linker.

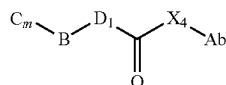
In specific embodiments, H_2 may include any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine. Furthermore, H_2 may be an azide, or a strained alkyne. Further, H_2 may be an azide, or dibenzocyclooctyne-amine. Additionally, H_2 may be a diene, or a dienophile. Further, H_2 may be a tetrazine, or a norbornene. Alternatively, H_2 may be a tetrazine, or a trans-cyclooctene.

In specific embodiments, when H_2 is to react with the H_1 -Ab according to the present invention, H_2 may be complementary to the first click-chemistry functional group (H_1).

7. Antibody-Payload Conjugate (C_m -Ab)

According to the present invention, there is disclosed a conjugate of an antibody and a payload (i.e., an antibody-payload conjugate). Such a compound is herein indicated by the symbol " C_m -Ab."

The present invention provides a C_m -Ab represented by Formula 10:



[Formula 10]

wherein C_m is a cargo moiety,

B is a structure formed by a click-chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group,

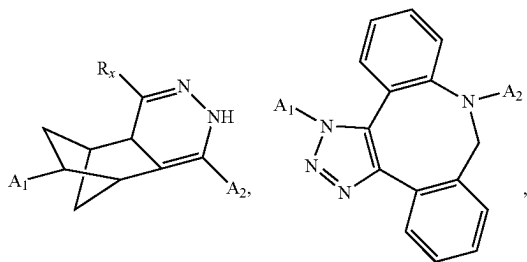
D_1 is any alkylene, alkenylene, or alkynylene,

X_4 is NH, O, or S, and

Ab is an antibody.

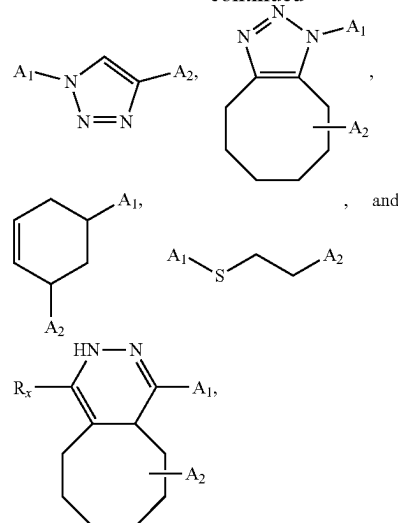
The contents of the cargo moiety apply to the contents disclosed in the section '6. Payload.'

In specific embodiments, B may be a structure formed by a click-chemistry reaction between any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine and its partner "click-chemistry functional group." Also, B may be any one selected from

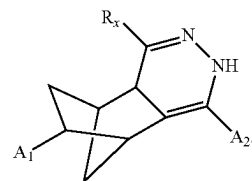


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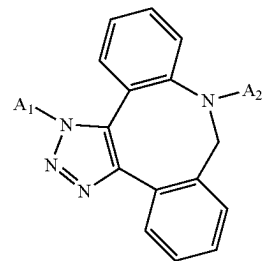
-continued



wherein both A_1 and A_2 are connected to the cargo moiety or D_1 so that the A_1 and A_2 cannot be connected to the same moiety, and R_x may be selected from H, a halogen, and a C_{1-3} alkyl. Furthermore, B may be



Additionally, B may be



In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-CH_2OCH_2-$. In addition, D_1 may be a covalent bond.

In one specific embodiment, X_4 may be NH.

In specific embodiments, Ab may be a human antibody. In other specific embodiments, Ab may be a non-human animal antibody. In specific embodiments, Ab may be immunoglobulin G (IgG). In specific embodiments, Ab may be a whole antibody. In other specific embodiments, Ab may be a fragment of the antibody.

In specific embodiments, X_4 and Ab may be connected via an Fab domain of Ab. In other specific embodiments, X_4 and Ab may be connected via an Fc domain of Ab. Also, X_4 and Ab may be connected via lysine 246 or 248 in the Fc domain of Ab. Furthermore, X_4 and Ab may be connected via lysine

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246 in the Fc domain of Ab. Further, X₄ and Ab may be connected via lysine 248 in the Fc domain of Ab. Alternatively, X₄ and Ab may be connected via lysine 246 and 248 in the Fc domain of Ab. In specific embodiments, X₄ and Ab may be connected via only one of two Fc domains of Ab. In other specific embodiments, X₄ and Ab may be connected via both of the two Fc domains of Ab.

The present invention provides an antibody or a fragment thereof including an amino acid sequence of the following 10
Formula 10-1:

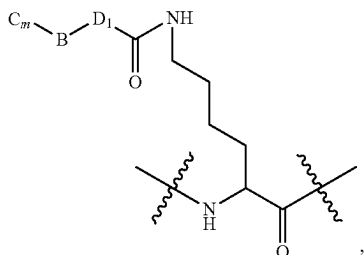
[Formula 10-1]

(SEQ ID NO: 23) 15

G-P-S-V-F-L-F-P-P- (K) " -P-K-D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and

$(K)''$ is



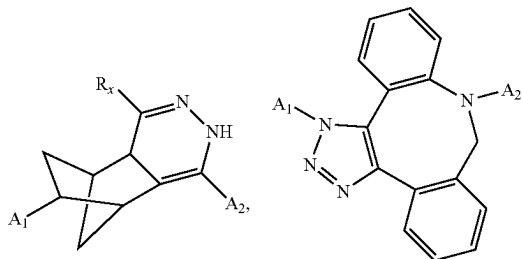
wherein C_m is a cargo moiety,

B is a structure formed by a click-chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group, and

D₁ is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has a cargo moiety conjugated to lysine 246 in an Fc domain thereof, or a site corresponding to the lysine 246.

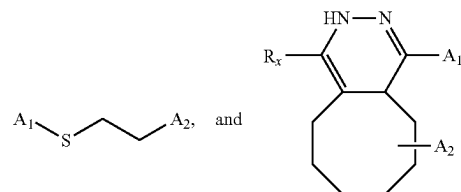
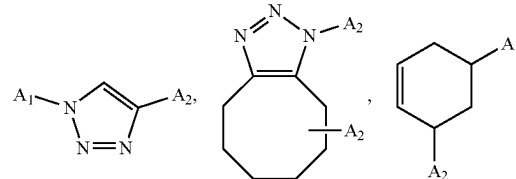
The contents of the cargo moiety apply to the contents disclosed in the section '6. Payload.'

In specific embodiments, B may be a structure formed by a click-chemistry reaction between any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine and its partner “click-chemistry functional group.” Also, B may be any one selected from

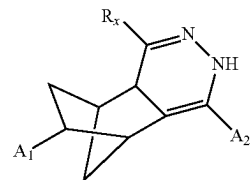


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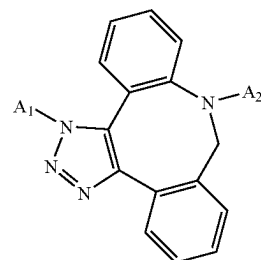
-continued



and wherein both A₁ and A₂ are connected to the cargo moiety or D₁ so that the A₁ and A₂ cannot be connected to the same moiety, and R_x may be selected from H, a halogen, and a C₁₋₃ alkyl. Furthermore, B may be



Additionally, B may be



In specific embodiments, D₁ may include any one selected from a covalent bond, a C₁₋₄ alkylene, a C₂₋₄ alkenylene, a C₂₋₄ alkynylene, and a C₃₋₈ cycloalkylene. Also, D₁ may be —CH₂OCH₂—. In addition, D₁ may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody.

In specific embodiments, the antibody may include the amino acid sequence of Formula 10-1 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 10-1 in both of the two Fc domains thereof.

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The present invention provides an antibody or a fragment thereof including an amino acid sequence of the following Formula 10-2:

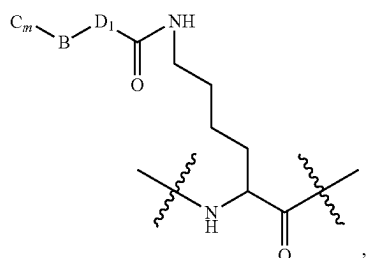
[Formula 10-2]

(SEQ ID NO: 24)

G-P-S-V-F-L-F-P-P-K-P-(K)-D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and

(K)" is



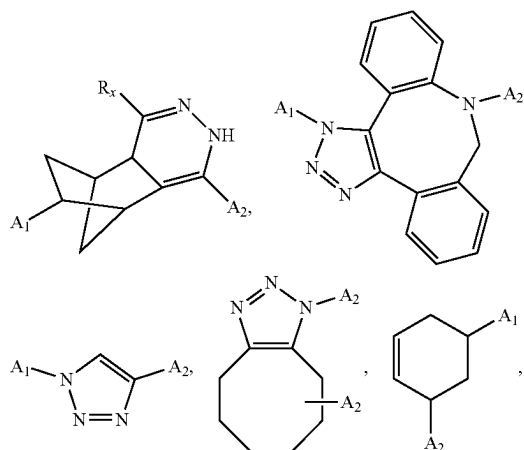
wherein C_m is a cargo moiety,

B is a structure formed by a click-chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group, and

D_1 is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has a cargo moiety conjugated to lysine 246 in an Fc domain thereof, or a site corresponding to the lysine 246.

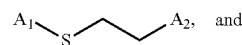
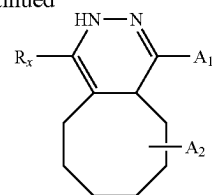
The contents of the cargo moiety apply to the contents disclosed in the section '6. Payload.'

In specific embodiments, B may be a structure formed by a click-chemistry reaction between any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine and its partner "click-chemistry functional group." Also, B may be any one selected from

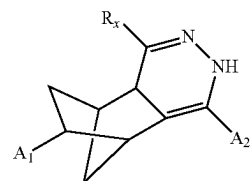


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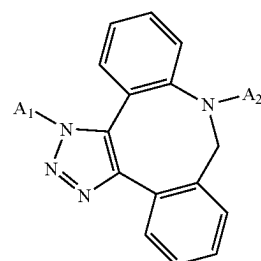
-continued



wherein both A_1 and A_2 are connected to the cargo moiety or D_1 so that the A_1 and A_2 cannot be connected to the same moiety, and R_x may be selected from H, a halogen, and a C_{1-3} alkyl. Furthermore, B may be



Additionally, B may be



In specific embodiments, D_1 may include any one selected from a covalent bond, a C_{1-4} alkylene, a C_{2-4} alkenylene, a C_{2-4} alkynylene, and a C_{3-8} cycloalkylene. Also, D_1 may be $-CH_2OCH_2-$. In addition, D_1 may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody.

In specific embodiments, the antibody may include the amino acid sequence of Formula 10-2 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 10-2 in both of the two Fc domains thereof.

The present invention provides an antibody or a fragment thereof including an amino acid sequence of the following Formula 10-3:

[Formula 10-3]

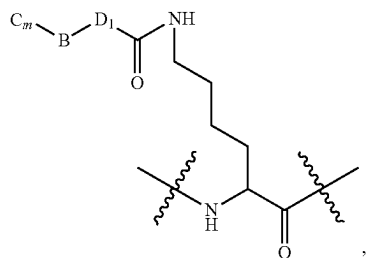
(SEQ ID NO: 25)

G-P-S-V-F-L-F-P-P-(K)-P-(K)-D-T-L-M-I

wherein G is glycine, P is proline, S is serine, V is valine, F is phenylalanine, L is leucine, K is lysine, D is aspartic acid, T is threonine, M is methionine, I is isoleucine, and

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(K)" is each independently



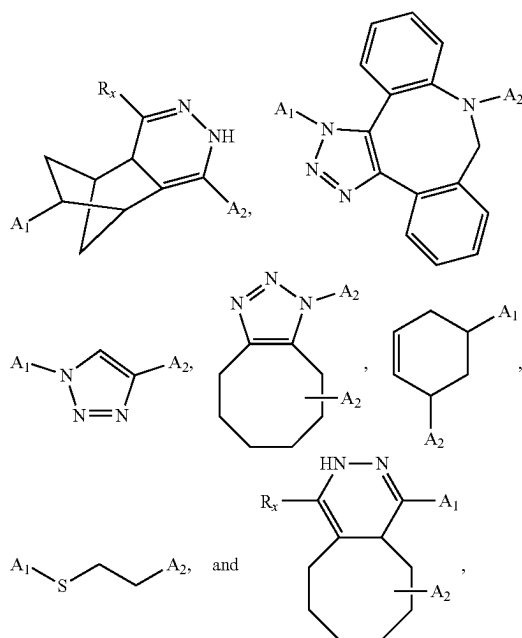
wherein C_m is a cargo moiety,

B is a structure formed by a click-chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group, and

D₁ is any alkylene, alkenylene, or alkynylene. The antibody or fragment thereof has a cargo moiety conjugated to lysine 246 in an Fc domain thereof, or a site corresponding to the lysine 246.

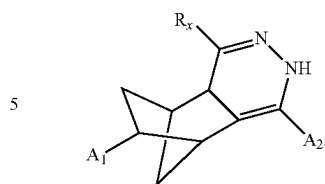
The contents of the cargo moiety apply to the contents disclosed in the section “6. Payload.”

In specific embodiments, B may be a structure formed by a click-chemistry reaction between any one selected from an alkyne, an azide, a strained alkyne, a diene, a dienophile, an alkene, a thiol, and a tetrazine and its partner “click-chemistry functional group.” Also, B may be any one selected from

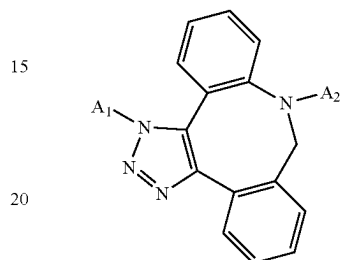


wherein both A₁ and A₂ are connected to the cargo moiety or D₁ so that the A₁ and A₂ cannot be connected to the same moiety, and R_x may be selected from H, a halogen, and a C₁₋₃ alkyl. Furthermore, B may be

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¹⁰ Additionally, B may be



In specific embodiments, D₁ may include any one selected from a covalent bond, a C₁₋₄ alkylene, a C₂₋₄ alkenylene, a C₂₋₄ alkynylene, and a C₃₋₈ cycloalkylene. Also, D₁ may be —CH₂OCH₂—. In addition, D₁ may be a covalent bond.

In specific embodiments, the antibody may be a human antibody. In other specific embodiments, the antibody may be a non-human animal antibody. In specific embodiments, the antibody may be immunoglobulin G (IgG). In specific embodiments, the antibody may be a whole antibody. In other specific embodiments, the antibody may be a fragment of the antibody.

In specific embodiments, the antibody may include the amino acid sequence of Formula 10-3 in only one of the two Fc domains thereof. In other specific embodiments, the antibody may include the amino acid sequence of Formula 10-3 in both of the two Fc domains thereof.

The present invention provides an antibody or a fragment thereof including one or more amino acid sequences selected from Formulas 10-1, 10-2, and 10-3. In this case, the contents of Formulas 10-1 to 10-3 are as described above.

In specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 10-1 and may not include the amino acid sequences of Formulas 10-2 and 10-3. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 10-1 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 10-1 in both of the two Fc domains thereof.

In other specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 10-2 and may not include the amino acid sequences of Formulas 10-1 and 10-3. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 10-2 in only one of the two Fc domains thereof. In addition, the antibody may include the amino acid sequence of Formula 10-2 in both of the two Fc domains thereof.

In specific embodiments, the antibody or fragment thereof may include only the amino acid sequence of Formula 10-3 and may not include the amino acid sequences of Formulas 10-1 and 10-2. Also, the antibody or fragment thereof may include the amino acid sequence of Formula 10-3 in only one of the two Fc domains thereof. In addition, the antibody

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may include the amino acid sequence of Formula 10-3 in both of the two Fc domains thereof.

7.1. Antibody-Drug Conjugate (ADC)

According to the present invention, there is disclosed a novel antibody-drug conjugate (ADC). The ADC according to the present invention means that a payload in the antibody-payload conjugate includes a drug moiety.

The present invention provides a C_m -Ab according to the present invention in which a cargo moiety includes a drug moiety.

In specific embodiments, the cargo moiety may include two or more drug moieties.

In specific embodiments, the drug moiety may be an anti-cancer drug. Also, the anti-cancer drug may include one or more selected from DM1, DM3, DM4, Abrin, Ricin A, a *Pseudomonas* exotoxin, a Cholera toxin, a Diphtheria toxin, a tumor necrosis factor, a interferon, β interferon, a nerve growth factor, a platelet-derived growth factor, a tissue plasminogen activator, a cytokine, an apoptosis-inducing agent, an anti-angiogenic agent, a lymphokine, taxane, a DNA-alkylating agent, anthracycline, a Tubulysin analogue, a duocarmycin analogue, auristatin E, auristatin F, a maytansinoid, a cytotoxic agent including a reactive polyethylene glycol residue, taxon, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, T. Colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dihydrotestosterone, a glucocorticoid, procaine, tetracaine, lidocaine, propranolol, puromycin, methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine, mechlorethamine, thiotepa, chlorambucil, Meiphalan, carmustine, lomustine, cyclophosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, cisplatin, dactinomycin, bleomycin, anthramycin, calicheamicin, abiraterone, bendamustine, bortezomib, carboplatin, cabazitaxel, dasatinib, doxetaxel, epirubicin, erlotinib, everolimus, gemcitabine, gefitinib, idarubicin, imatinib, hydroxyurea, lapatinib, leuporelin, melphalan, nedaplatin, nilotinib, oxaliplatin, pazopanib, pemetrexed, picoplatin, romidepsin, satraplatin, sorafenib, vemurafenib, sunitinib, teniposide, triplatin, and vinorelbine.

In specific embodiments, the cargo moiety may include a VC linker.

7.2. Method of Preparing Antibody-Payload Conjugate

According to the present invention, there is disclosed a method of preparing an antibody-payload conjugate.

The antibody-payload conjugate of the present invention may be prepared by allowing a C_m -H₂ to react with an H₁-Ab. In this case, the contents of the H₁-Ab apply to the contents disclosed in the section 5.1, and the contents disclosed in the section 6 apply for the C_m -H₂. When the first click-chemistry functional group of H₁-Ab and the second click-chemistry functional group of C_m -H₂ are complementary to each other, that is, when the first click-chemistry functional group of H₁-Ab and the second click-chemistry functional group of C_m -H₂ function as partner click-chemistry functional groups, a click-chemistry reaction may occur to prepare an antibody-payload conjugate according to the present invention. The contents of the click-chemistry reaction were sufficiently described above in the section "Definition." The click-chemistry reaction is a biorthogonal reaction, and has an advantage in that the reaction occurs at a very rapid reaction rate and is used to form a strong binding structure. Therefore, the click-chemistry reaction has the same advantages as in the following sections 7.3 and 7.4.

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The present invention provides a method of preparing an antibody-payload conjugate, characterized in that the method includes reacting an antibody containing a first click-chemistry functional group according to the present invention with a payload according to the present invention, wherein H₂ of the payload is a second click-chemistry functional group complementary to the first click-chemistry functional group.

In this case, the antibody provided in the section 5.1 applies to the antibody containing the first click-chemistry functional group, and the payload provided in the section 6 applies to the payload. Because all the antibodies containing a first click-chemistry functional group in the lysine 246 and/or the lysine 248 are disclosed in the section 5.1, it is desirable that the antibody-payload conjugates in which a cargo moiety is conjugated to the lysine 246 and/or the lysine 248 are disclosed to a fully reproducible extent (see Formulas 10 and 10-1 to 10-3 in the section 7).

The present invention provides a kit for preparing an antibody-payload conjugate, characterized in that the kit includes an antibody containing a first click-chemistry functional group according to the present invention; and a payload according to the present invention, wherein H₂ of the payload is a second click-chemistry functional group complementary to the first click-chemistry functional group.

In this case, the antibody provided in the section 5.1 applies to the antibody containing the first click-chemistry functional group, and the payload provided in the section 6 applies to the payload.

Optionally, the present invention provides a kit for preparing an antibody-payload conjugate, which includes:

- an antibody;
- an agent for transferring a first click-chemistry functional group to an antibody according to the present invention;
- a payload according to the present invention.

In specific embodiments, the agent for transferring a first click-chemistry functional group to an antibody may be an H₁-L₂-SSFI for specifically transferring a first click-chemistry functional group to lysine 246 or 248 in an Fc domain of an antibody according to the present invention. In other specific embodiments, the agent for transferring a first click-chemistry functional group to an antibody may be an H₁-L₂-SSFI for specifically transferring a first click-chemistry functional group to lysine 246 in an Fc domain of an antibody according to the present invention. In still another specific embodiment, the agent for transferring a first click-chemistry functional group to an antibody may be an H₁-L₂-SSFI for specifically transferring a first click-chemistry functional group to lysine 248 in an Fc domain of an antibody according to the present invention.

Optionally, the present invention provides a kit for preparing an antibody-payload conjugate, which includes:

- an antibody;
- a linker according to the present invention;
- a site-specific antibody interactome according to the present invention; and
- a payload according to the present invention.

In specific embodiments, D₂ in the linker is a C_y alkylene, a C_y alkenylene, or a C_y alkynylene, and D₃ in the site-specific antibody interactome is a C_x alkylene, wherein y may be an integer greater than or equal to 1. Also, the sum of x and y may be 1 ≤ x + y ≤ 5. In addition, the sum of x and y may be 6 ≤ x + y ≤ 8. Further, the sum of x and y may be 9 ≤ x + y ≤ 12.

7.3. The Antibody-Payload Conjugate According to the Present Invention Uses a Biorthogonal Reaction to Form a Stable Bond.

The antibody-payload conjugate according to the present invention is formed through a click-chemistry reaction. The click-chemistry reaction is a biorthogonal reaction that does not influence biochemical phenomena naturally occurring in vivo. Also, a bond formed by the biorthogonal reaction is not recognized by in vivo lyases. Therefore, the antibody-payload conjugate according to the present invention has an advantage in that the antibody and the cargo moiety form a very stable bond in vivo.

7.4. The Antibody-Payload Conjugate Prepared According to the Present Invention has High Uniformity and Yield.

As described above in the section 5.6, the antibody containing a first click-chemistry functional group according to the present invention has high uniformity and yield. Therefore, the antibody-payload conjugate prepared from the antibody according to the present invention has an advantage in that it also has high uniformity and yield. The uniformity of C_m -Ab has to be high in order that the antibody conjugate has uniform performance.

In the case of the antibody conjugates prepared by the prior art disclosed in Publication Nos. US 2018/0141976 A1 and WO 2018/199337 A1, since the antibody conjugates have poor yield and uniformity, as described above in the section 5.6, there may be a negative effect on the uniformity of performance.

7.5. No Functions of the Antibody are Lowered Because the Antibody-Payload Conjugate According to the Present Invention has No FcRn Binding Site Blocked Therein.

The C_m -Ab according to present invention has the same advantages as described in the sections 5.7 and 5.8. In particular, pharmacokinetic (PK) characteristics of the ADC used in vivo may be greatly improved because the ADC has an increased half-life.

An FcRn binding site of the ADC prepared by the prior art disclosed in Publication No. US 2018/0141976 A1 is blocked because SSFI is included in the final antibody-labeled product. Therefore, the ADC according to the present invention has superior PK characteristics, compared to the ADC according to the corresponding prior art.

7.6. The Antibody-Drug Conjugate According to the Present Invention is Stable Because the Antibody-Drug Conjugate does not Contain a Highly Bioreactive Chemical Functional Group.

The C_m -Ab according to the present invention has the same advantages as described in the section 5.9. In particular, pharmacodynamic (PD) characteristics of the ADC used in vivo may be greatly improved because the ADC enables a smooth antibody-drug action.

In the case of the ADC prepared by the prior art disclosed in Publication No. WO 2018/199337 A1, an additional chemical functional group such as thiol, hydroxy, a carboxylic acid, phosphoric acid, an amine, and the like are included in the final antibody-labeled product. Therefore, the ADC according to the present invention has superior PD characteristics, compared to the ADC according to the corresponding prior art.

8. Composition

According to the present invention, there is disclosed a composition including an antibody. In this case, the antibody may be an antibody containing a first chemical functional group according to the present invention. Alternatively, the antibody may be an antibody-payload conjugate according to the present invention. Also, the antibody may be an antibody-drug conjugate according to the present invention.

The composition according to the present invention may be used for various applications depending on the function of an antibody included in the composition. For example, when a first chemical functional group or a cargo moiety includes a radioactive moiety, the corresponding composition may be used as a radioactive contrast medium, and the like. Alternatively, when the first chemical functional group or the cargo moiety includes a fluorescent moiety, the corresponding composition may be used as a label used in an enzyme-linked immunosorbent assay (ELISA), and the like. Alternatively, when the first chemical functional group or the cargo moiety includes a drug moiety, the corresponding composition may be used as a pharmaceutical composition. In this case, components of the composition generally used in the related art fall within the scope of the present invention. Also, compositional ratios of the corresponding components generally used in the art also fall within the scope of the present invention.

The present invention provides a composition including an antibody containing a first chemical functional group according to the present invention. In specific embodiments, the antibody containing the first chemical functional group may be at least one selected from Formulas 7, 7-1 to 7-3, 8, and 8-1 to 8-3.

Also, the present invention provides a composition including an antibody containing a first click-chemistry functional group according to the present invention. In specific embodiments, the antibody containing the first click-chemistry functional group may be at least one selected from Formulas 8, and 8-1 to 8-3.

In addition, the present invention provides a composition including an antibody-payload conjugate according to the present invention. In this case, the contents described in the section 7 apply to the contents of the antibody-payload conjugate.

Optionally, the present invention provides a composition including an antibody-drug conjugate according to the present invention. In this case, the contents described in the section 7.1 apply to the contents of the antibody-drug conjugate.

Pharmaceutical Composition

The following content relates to the where the composition is a pharmaceutical composition used for diagnostic, prophylactic and/or therapeutic purposes. Only within the content "pharmaceutical composition" here, all the R_1 -Ab, H_1 -Ab, C_m -Ab, and ADC according to the present invention are used interchangeably with the term "antibody or fragment thereof."

The present invention provides a pharmaceutical composition including an antibody or a fragment thereof according to the present invention. Also, the pharmaceutical composition may be a composition for treating cancer. Furthermore, the cancer may include any one selected from bladder cancer, bone cancer, brain cancer, breast cancer, heart cancer, cervical cancer, colon cancer, rectal cancer, esophageal cancer, fibrosarcoma, gastric cancer, stomach cancer, head and neck cancer, Kaposi's sarcoma, renal cancer, leukemia, liver cancer, lung cancer, lymphoma, melanoma, myeloma, ovarian cancer, pancreatic cancer, penile cancer, prostate cancer, testicular germ cell cancer, thymoma, and thymic carcinoma. Further, the cancer may be breast cancer.

The present invention provides a therapeutic method which includes administering a pharmaceutical composition including an antibody or a fragment thereof according to the present invention into a subject. Also, the therapeutic method may be a method of treating cancer. Furthermore, the cancer may include any one selected from bladder

cancer, bone cancer, brain cancer, breast cancer, heart cancer, cervical cancer, colon cancer, rectal cancer, esophageal cancer, fibrosarcoma, gastric cancer, stomach cancer, head and neck cancer, Kaposi's sarcoma, renal cancer, leukemia, liver cancer, lung cancer, lymphoma, melanoma, myeloma, ovarian cancer, pancreatic cancer, penile cancer, prostate cancer, testicular germ cell cancer, thymoma, and thymic carcinoma. Further, the cancer may be breast cancer.

To prepare a pharmaceutical or sterilized composition including an antibody or a fragment thereof, the antibody or fragment thereof according to the present invention may be mixed with a pharmaceutically acceptable carrier or excipient. The composition may further include one or more other therapeutic agents which are suitable for treating or preventing cancer (e.g., breast cancer, colorectal cancer, lung cancer, multiple myeloma, ovarian cancer, liver cancer, gastric cancer, pancreatic cancer, acute myelogenous leukemia, chronic myelogenous leukemia, osteosarcoma, squamous cell carcinoma, peripheral nerve sheath tumors, schwannoma, head and neck cancer, bladder cancer, esophageal cancer, Barrett's esophageal cancer, glioblastoma, clear cell sarcoma of soft tissue, malignant mesothelioma, neurofibromatosis, kidney cancer, melanoma, prostate cancer, benign prostate hypertrophy (BPH), gynecomastia, rhabdomyosarcoma, and endometriosis).

A formulation of therapeutic and diagnostic agents may be prepared by mixing with a physiologically acceptable carrier, an excipient, or a stabilizing agent, for example, in the form of a freeze-dried powder, slurry, an aqueous solution, a lotion, or a suspension (see, for example, Hardman et al., Goodman and Gilman's The Pharmacological Basis of Therapeutics, McGraw-Hill, New York, N.Y., 2001; Gennaro, Remington: The Science and Practice of Pharmacy, Lippincott, Williams, and Wilkins, New York, N.Y., 2000; Avis, et al. (eds.), Pharmaceutical Dosage Forms: Parenteral Medications, Marcel Dekker, N Y, 1993; Lieberman, et al. (eds.), Pharmaceutical Dosage Forms: Tablets, Marcel Dekker, N Y, 1990; Lieberman, et al. (eds.) Pharmaceutical Dosage Forms: Disperse Systems, Marcel Dekker, N Y, 1990; Weiner and Kotkoskie, Excipient Toxicity and Safety, Marcel Dekker, Inc., New York, N.Y., 2000).

In specific embodiments, the clinical service form (CSF) of the antibody-drug conjugate according to the present invention is a lyophilisate present in a vial containing ADC, sodium succinate, and polysorbate 20. The lyophilisate may be reconstituted with water for injection, and the solution includes ADC, sodium succinate, sucrose and polysorbate 20 at approximately pH 5.0. For sequential intravenous administration, the resulting solution will be usually further diluted into a carrier solution.

The choice of a therapeutic dosing regimen depends on various factors including a rate of serum or tissue replacement of a substance, a level of symptoms, the immunogenicity of the substance, and the accessibility of target cells in a biological matrix. In specific embodiments, the dosing regimens maximize an amount of a therapeutic agent to be delivered to a patient so as to satisfy acceptable levels of side effects. Therefore, an amount of a biological agent to be administered depends in part on the certain substance and the severity of the condition being treated. Guidance is available for selection of the appropriate doses of antibodies, cytokines, and small molecules (see, for example, Wawrzynczak, Antibody Therapy, Bios Scientific Pub. Ltd, Oxfordshire, U K, 1996; Kresina (ed.), Monoclonal Antibodies, Cytokines and Arthritis, Marcel Dekker, New York, N.Y., 1991; Bach (ed.), Monoclonal Antibodies and Peptide Therapy in Autoimmune Diseases, Marcel Dekker, New

York, N.Y., 1993; Baert et al., New Engl. J. Med. 348:601-608, 2003; Milgrom et al., New Engl. J. Med. 341:1966-1973, 1999; Slamon et al., New Engl. J. Med. 344:783-792, 2001; Beniaminovitz et al., New Engl. J. Med. 342:613-619, 2000; Ghosh et al., New Engl. J. Med. 348:24-32, 2003; Lipsky et al., New Engl. J. Med. 343:1594-1602, 2000).

The appropriate dose is, for example, determined by a clinical practitioner using parameters or factors known in the related art or suspected to affect treatment, or using parameters or factors expected to affect treatment. In general, the dose begins with an amount somewhat smaller than the optimal dose and then increases in small increments until the desirable or optimal effects are achieved relative to any negative side effects. Important diagnostic measures, for example, include symptoms of inflammation or levels of inflammatory cytokines produced.

An actual dose level of the active ingredient in the pharmaceutical composition according to the present invention may vary in order to achieve an effective amount of the active ingredient to realize desired therapeutic responses in a certain patient, a composition, and a mode of administration without causing any toxicity in patients. The chosen dose level may be determined depending on various pharmacokinetic factors including the activities of a certain composition of the present invention used, or esters, salts or amides thereof, a route of administration, an administration time, a secretion rate of a certain compound used, a duration of treatment, other drugs, compounds and/or substances used in combination with the certain compound used, the age, sex, weight, condition, overall health, and previous medical history of a patient to be treated, and other factors known in the medical field.

The composition including an antibody or a fragment thereof according to the present invention may be given by continuous infusion, or given in a single dose, for example, daily, weekly, 1 to 7 times per week, bi-weekly, once every three weeks, once every four weeks, once every five weeks, once every six weeks, once every seven weeks, or once every eight weeks. The dose may be given intravenously, subcutaneously, topically, orally, intranasally, intrarectally, intramuscularly, intracerebrally, or by inhalation. A certain dosing protocol involves the maximum dose or administration frequency that avoids significant unwanted side effects.

For the antibody or fragment thereof according to the present invention, the dose to be administered to a patient may be in a range of 0.0001 mg/kg to 100 mg/kg (patient weight). The dose may be in a range of 0.0001 mg/kg to 20 mg/kg, 0.0001 mg/kg to 10 mg/kg, 0.0001 mg/kg to 5 mg/kg, 0.0001 to 2 mg/kg, 0.0001 to 1 mg/kg, 0.0001 mg/kg to 0.75 mg/kg, 0.0001 mg/kg to 0.5 mg/kg, 0.0001 mg/kg to 0.25 mg/kg, 0.0001 to 0.15 mg/kg, 0.0001 to 0.10 mg/kg, 0.001 to 0.5 mg/kg, 0.01 to 0.25 mg/kg, or 0.01 to 0.10 mg/kg (patient weight). The dose of the antibody or fragment thereof according to the present invention may be calculated by multiplying the weight (kilogram (kg)) of a patient by the dose (mg/kg) to be administered.

The antibody or fragment thereof according to the present invention may be repeatedly administered, and the administration may be performed at intervals of at least a day, two days, three days, five days, 10 days, 15 days, 30 days, 45 days, 2 months, 75 days, 3 months, or at least 6 months. In specific embodiments, the antibody or fragment thereof according to the present invention may be repeatedly administered every three weeks.

An effective amount to be administered to a certain patient may vary depending on factors such as the condition to be treated, the overall health of a patient, the mode, route and

dosage of administration, and the severity of side effects (see, for example, Maynard et al., *A Handbook of SOPs for Good Clinical Practice*, Interpharm Press, Boca Raton, Fla., 1996; Dent, *Good Laboratory and Good Clinical Practice*, Urch Publ., London, U K, 2001).

The route of administration may be, for example, by topical or skin application, by intravenous, intraperitoneal, intracerebral, intramuscular, intraocular, intra-arterial, intramedullary, intralesional injection or infusion, or by a sustained release system or an implant (see, for example, Sidman et al., *Biopolymers* 22:547-556, 1983; Langer et al., *J. Biomed. Mater. Res.* 15:167-277, 1981; Langer, *Chem. Tech.* 12:98-105, 1982; Epstein et al., *Proc. Natl. Acad. Sci. USA* 82:3688-3692, 1985; Hwang et al., *Proc. Natl. Acad. Sci. USA* 77:4030-4034, 1980; U.S. Pat. Nos. 6,350,466 and 6,316,024). When necessary, the composition may also include a solubilizing agent and a topical anesthetic (for example, lidocaine) for relieving pain at a site of injection. For example, the composition may also be formulated using an inhaler or a nebulizer, or may be used for pulmonary administration by formulation using an aerosolizing agent. See, for example, U.S. Pat. Nos. 6,019,968, 5,985,320, 5,985,309, 5,934,272, 5,874,064, 5,855,913, 5,290,540, and 4,880,078; and PCT Publication Nos. WO 92/19244, WO 97/32572, WO 97/44013, WO 98/31346, and WO 99/66903, the entire contents of which are hereby incorporated by reference.

Also, the composition of the present invention may be administered through one or more routes of administration using one or more various methods known in the related art. As recognized by a person of ordinary skill in the art, the route and/or mode of administration will depend on the desired results. The route of administration selected for the antibody or fragment thereof according to the present invention includes intravenous, intramuscular, intradermal, intraperitoneal, subcutaneous, intraspinal, or other parenteral routes of administration, for example, routes of administration by injection or infusion. Parenteral administration generally represents a mode of administration by injection other than the enteral and topical administration, and includes intravenous, intramuscular, intra-arterial, intrathecal, intracapsular, intraorbital, intracardial, intradermal, intraperitoneal, transtracheal, subcutaneous, subepidermal, intra-articular, subcapsular, subarachnoid, intraspinal, epidural, and intrasternal injections and infusions, but the present invention is not limited thereto. Alternatively, the composition of the present invention may be administered through a non-parenteral route of administration, for example, a topical, intraepidermal or intramucosal route of administration, and may be, for example, administered intranasally, orally, vaginally, intrarectally, sublingually, or topically. In one exemplary embodiment, the antibody or fragment thereof according to the present invention may be administered by infusion. In another exemplary embodiment, the antibody or fragment thereof according to the present invention may be subcutaneously administered.

When the antibody or fragment thereof according to the present invention is administered using a controlled or delayed release system, a pump may be used to achieve the controlled or delayed release (see Langer, *supra*; Sefton, *CRC Crit. Ref Biomed. Eng.* 14:20, 1987; Buchwald et al., *Surgery* 88:507, 1980; Saudek et al., *N. Engl. J. Med.* 321:574, 1989). A polymeric material may be used to achieve the controlled or delayed release in the therapy of the present invention (see, for example, *Medical Applications of Controlled Release*, Langer and Wise (eds.), CRC Pres., Boca Raton, Fla., 1974; *Controlled Drug Bioavail-*

ability, Drug Product Design and Performance, Smolen and Ball (eds.), Wiley, New York, 1984; Ranger and Peppas, *J. Macromol. Sci. Rev. Macromol. Chem.* 23:61, 1983; also see Levy et al., *Science* 228:190, 1985; During et al., *Ann. Neurol.* 25:351, 1989; Howard et al., *J. Neurosurg.* 7 1:105, 1989; U.S. Pat. Nos. 5,679,377; 5,916,597; 5,912,015; 5,989,463; 5,128,326; PCT Publication No. WO 99/15154; and PCT Publication No. WO 99/20253). Examples of polymers used for a sustained release formulation include poly(2-hydroxyethyl methacrylate), poly(methyl methacrylate), poly(acrylic acid), poly(ethylene-co-vinyl acetate), poly(methacrylic acid), polyglycolide (PLG), a polyanhydride, poly(N-vinyl pyrrolidone), poly(vinyl alcohol), polyacrylamide, poly(ethylene glycol), polylactide (PLA), poly(lactide-co-glycolide) (PLGA), and a polyorthoester, but the present invention is not limited thereto. In one exemplary embodiment, the polymer used for a sustained release formulation is inert, has no filterable impurities, and is stable upon storage, sterilization, and biodegradability. The controlled or delayed release system may be located adjacent to a prophylactic or therapeutic target, and thus requires a portion of the systemic dose (see, for example, Goodson, in *Medical Applications of Controlled Release*, *supra*, vol. 2, pp. 115-138, 1984).

The controlled release system is discussed in the review of the article "Langer, *Science* 249:1527-1533, 1990." Any prior art known to a person of ordinary skill in the art may be used to produce a sustained release formulation including one or more antibodies or a fragments thereof according to the present invention. See, for example, U.S. Pat. No. 4,526,938, PCT Publication No. WO 91/05548, PCT Publication No. WO 96/20698, Ning et al., *Radiotherapy & Oncology* 39:179-189, 1996; Song et al., *PDA Journal of Pharmaceutical Science & Technology* 50:372-397, 1995; Cleek et al., *Pro. Int'l. Symp. Control. Rel. Bioact. Mater.* 24:853-854, 1997; and Lam et al., *Proc. Int'l. Symp. Control Rel. Bioact. Mater.* 24:759-760, 1997, the entire contents of which are hereby incorporated by reference.

When the antibody or fragment thereof according to the present invention is topically administered, the antibody or fragment thereof may be formulated into forms of an ointment, a cream, a transdermal patch, a lotion, a gel, a shampoo, a spray, an aerosol, a solution, or an emulsion, or other forms well known to a person of ordinary skill in the art. See, for example, Remington's *Pharmaceutical Sciences and Introduction to Pharmaceutical Dosage Forms*, 19th ed., Mack Pub. Co., Easton, Pa. (1995). A non-sprayable topical dosage form includes a carrier or one or more excipients suitable for topical application. In some cases, viscous, semisolid, or solid forms having a higher kinematic viscosity than water may be generally used as the topical dosage form. Suitable preparations include a solution, a suspension, an emulsion, a cream, an ointment, a powder, a liniment, a salve, and the like, all of which are sterilized, or mixed with an adjuvant (for example, a preservative, a stabilizing agent, a wetting agent, a buffer, or a salt) influencing various characteristics, for example, such as osmotic pressure, when necessary, but the present invention is not limited thereto. In some cases, other suitable topical dosage forms include a sprayable aerosol formulation filled in a mixture of compressed volatile substances (for example, a gaseous propellant such as freon) or a squeeze bottle after the active ingredient is combined with a solid or liquid inert carrier. When necessary, a moisturizing agent or a humectant may be also added to the pharmaceutical composition and the dosage form. Examples of such an additional component are known in the related art.

When the composition including an antibody or a fragment thereof is intranasally administered, the composition may be formulated into forms of an aerosol, a spray, mist, or drops. In particular, a prophylactic or therapeutic agent for use in the present invention may be conveniently delivered in an aerosol spray dosage form from a pressurized pack or an atomizer using a suitable propellant (for example, dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide, or other suitable gases). In the case of the pressurized aerosol, the dosage unit may be determined by providing a valve to deliver a metered amount. A capsule and cartridge (for example, consisting of gelatin) for use in an inhaler or an insufflator may be formulated to include a powder mixture of compounds, and a suitable powder base, for example, lactose or starch.

Methods for co-administration or treatment with a second therapeutic agent, for example, a cytokine, a steroid, a chemotherapeutic agent, an antibiotic, or radiation are known in the related art (see, for example, Hardman et al., (eds.) (2001) Goodman and Gilman's The Pharmacological Basis of Therapeutics, 10th ed., McGraw-Hill, New York, N.Y.; Poole and Peterson (eds.) (2001) Pharmacotherapeutics for Advanced Practice: A Practical Approach, Lippincott, Williams & Wilkins, Phila., Pa.; Chabner and Longo (eds.) (2001) Cancer Chemotherapy and Biotherapy, Lippincott, Williams & Wilkins, Phila., Pa.). An effective amount of the therapeutic agent may reduce symptoms by at least 10%; at least 20%; at least approximately 30%; at least 40%, or at least 50%.

An additional therapy (for example, a prophylactic or therapeutic agent) that may be administered in combination with the antibody or fragment thereof according to the present invention may be administered together with the antibody or fragment thereof according to the present invention at intervals of less than 5 minutes, intervals of less than 30 minutes, intervals of an hour, intervals of approximately an hour, intervals of approximately 1 to approximately 2 hours, intervals of approximately 2 hours to approximately 3 hours, intervals of approximately 3 hours to approximately 4 hours, intervals of approximately 4 hours to approximately 5 hours, intervals of approximately 5 hours to approximately 6 hours, intervals of approximately 6 hours to approximately 7 hours, intervals of approximately 7 hours to approximately 8 hours, intervals of approximately 8 hours to approximately 9 hours, intervals of approximately 9 hours to approximately 10 hours, intervals of approximately 10 hours to approximately 11 hour, intervals of approximately 11 hour to approximately 12 hours, intervals of approximately 12 hours to 18 hours, intervals of 18 hours to 24 hours, intervals of 24 hours to 36 hours, intervals of 36 hours to 48 hours, intervals of 48 hours to 52 hours, intervals of 52 hours to 60 hours, intervals of 60 hours to 72 hours, intervals of 72 hours to 84 hours, intervals of 84 hours to 96 hours, or intervals of 96 hours to 120 hours. Two or more therapeutic agents may be administered during the same visit by a patient.

In specific embodiments, the antibody or fragment thereof according to the present invention may be formulated to insure a proper distribution in vivo. For example, the blood-brain barrier (BBB) excludes a number of highly hydrophilic compounds. To ensure therapeutic compounds of the invention cross the BBB (when necessary), they can be, for example, formulated in liposomes. For a method of preparing liposomes, for example, see U.S. Pat. Nos. 4,522,811; 5,374,548; and 5,399,331. The liposomes may include one or more moieties that are selectively transported into specific cells or organs, thereby enhancing targeted drug delivery (see, for example, Ranade, (1989) J. Clin. Pharmacol.

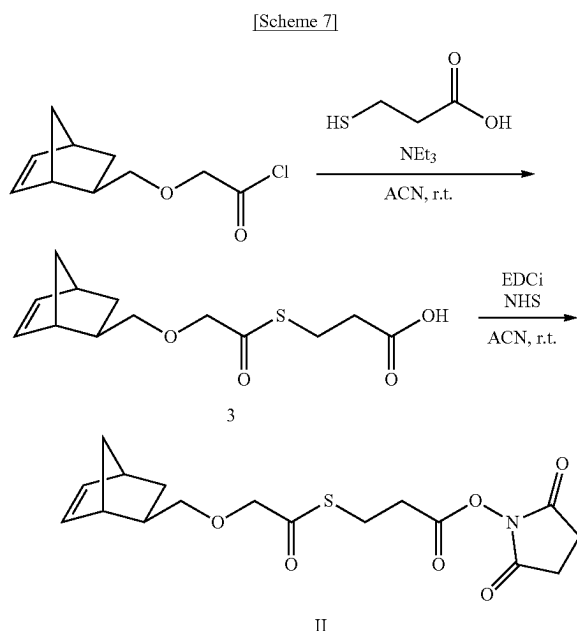
29:685). Exemplary targeting moieties include folate or biotin (see, for example, U.S. Pat. No. 5,416,016 (to Low et al.)); mannosides (see Umezawa et al., (1988) Biochem. Biophys. Res. Commun. 153:1038); antibodies (see Bloeman et al., (1995) FEBS Lett. 357:140; Owais et al., (1995) Antimicrob. Agents Chemother. 39:180); a surfactant protein A receptor (see Briscoe et al., (1995) Am. J. Physiol. 1233:134); p 120 (see Schreier et al., (1994) J. Biol. Chem. 269:9090), and the like. Also, see K. Keinänen; M. L. Laukkanen (1994) FEBS Lett. 346:123; J. J. Killian; I. J. Fidler (1994) Immunomethods 4:273.

The present invention provides a protocol for administering a pharmaceutical composition, which includes the antibody or fragment thereof according to the present invention, to a subject in need thereof alone or in combination with other therapies. The therapeutic agent (for example, a prophylactic or therapeutic agent) for combination therapy of the present invention may be simultaneously or sequentially administered to the subject. The therapeutic agent (for example, a prophylactic or therapeutic agent) for combination therapy of the present invention may also be administered periodically. A periodic therapy includes administering a first therapeutic agent (for example, a first prophylactic or therapeutic agent) for a predetermined period of time, administering a second therapeutic agent (for example, a second prophylactic or therapeutic agent) for a predetermined period of time, and then sequentially repeating the steps of administration, that is, a predetermined cycle for reducing the occurrence of resistance to one of the therapeutic agents (for example, agonists), and/or preventing or reducing side effects of one of the therapeutic agents (for example, agonists), and/or improving efficacy of the therapeutic agents.

The therapeutic agent (for example, a prophylactic or therapeutic agent) for combination therapy of the present invention may be simultaneously administered to a subject.

The term "simultaneously" means that therapies (for example, prophylactic or therapeutic agents) need not be administered at exactly the same time without limitation, but are rather administered to a subject sequentially with the pharmaceutical composition including an antibody or a fragment thereof according to the present invention, and are administered at time intervals that may serve to provide higher benefits compared to when the antibody of the present invention is administered at a different time from other therapy(ies). For example, the respective therapeutic agents may be sequentially administered to a subject at the same time or different points of time in any order; but should be administered at sufficiently short time intervals to provide a desired therapeutic or prophylactic effect when they are not administered at the same time. The respective therapeutic agents may be administered to a subject in any adequate form through any suitable route of administration. In various exemplary embodiments, the therapeutic agent (for example, a prophylactic or therapeutic agent) may be administered to a subject at intervals of less than 15 minutes, intervals of less than 30 minutes, intervals of less than an hour, intervals of approximately an hour, intervals of approximately 1 to approximately 2 hours, intervals of approximately 2 hours to approximately 3 hours, intervals of approximately 3 hours to approximately 4 hours, intervals of approximately 4 hours to approximately 5 hours, intervals of approximately 5 hours to approximately 6 hours, intervals of approximately 6 hours to approximately 7 hours, intervals of approximately 7 hours to approximately 8 hours, intervals of approximately 8 hours to approximately 9 hours, intervals of approximately 9 hours to approximately 10 hours, intervals

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1.1.2. Compound II (SO₂ Linker: NHS & Norbornene)

Synthesis of Compound II (Scheme 7, FIG. 17)

Synthesis of Compound 3

0.55 g (2.33 mmol, 1.0 eq.) of Compound 1 was dissolved in 5 mL of acetonitrile (ACN), and 0.2 mL (2.33 mmol, 1.0 eq.) of 3-mercaptopropionic acid and 0.49 mL (3.49 mmol, 1.5 eq.) of triethylamine were then added dropwise while stirring at room temperature. The reaction solution was stirred for 11 hours, and then concentrated under reduced pressure. Subsequently, water was added to the reaction solution, and the reaction solution was extracted three times with ethyl acetate (EA). An organic layer was dried over magnesium sulfate, concentrated under reduced pressure, and then purified by silica gel column chromatography (DCM:MeOH=10:1) to obtain 0.56 g (yield: 89%) of a target compound.

Synthesis of Compound II

0.56 g (2.07 mmol, 1.0 eq.) of Compound 3 was dissolved in 10 mL of acetonitrile (ACN), and 0.81 g (4.21 mmol, 2.0 eq.) of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI) and 0.65 g (5.61 mmol, 2.7 eq.) of N-hydroxysuccinimide (NHS) were added while stirring at room temperature. The reaction solution was stirred for 12 hours, and then concentrated under reduced pressure. Subsequently, water was added to the reaction solution, and the reaction solution was extracted three times with EA. An organic layer was dried over magnesium sulfate, and concentrated under reduced pressure. Then, the residue was purified by silica gel column chromatography (EA:Hex=2:1) to obtain 0.63 g (yield: 83%) of a target compound.

Confirmation of Structure of Compound II

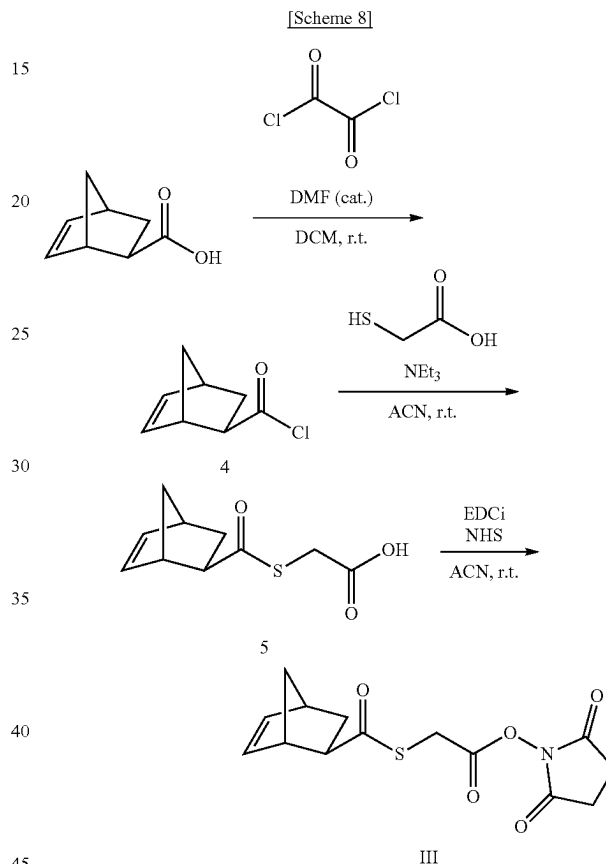
¹H NMR (500 MHz, CDCl₃) δ 6.16-6.14 (m, 2H), 4.45 (s, 1H), 4.21-4.08 (m, 1H), 3.69-3.58 (m, 1H), 3.51 (dt, J=14.5,

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8.9 Hz, 1H), 3.22 (t, J=7.1 Hz, 2H), 2.96 (t, J=7.1 Hz, 2H), 2.79-2.83 (m, 6H), 1.81-1.67 (m, 1H), 1.37-1.21 (m, 4H), 1.13-1.18 (m, 1H).

LRMS (ESI): m/z 385.1 [M+NH₄⁺]

The structure of Compound II was confirmed by mass spectrometry. The results are shown in FIG. 18.

1.1.3. Compound III (SO₃ Linker: NHS & Norbornene)

Synthesis of Compound III (Scheme 8, FIG. 19)

Synthesis of Compound 4

0.5 g (3.62 mmol, 1.0 eq.) of exo-5-norbornenecarboxylic acid was dissolved in 20 mL of DCM, and 0.028 mL (0.37 mmol, 0.94 eq.) of DMF and 0.63 mL (7.24 mmol, 2.0 eq.) of oxalyl chloride were then added dropwise while stirring at room temperature. The reaction solution was stirred for 3 hours, and then concentrated under reduced pressure to obtain 0.47 g (yield: 83%) of a target compound.

Synthesis of Compound 5

0.47 g (3.0 mmol, 1.0 eq.) of Compound 4 was dissolved in 15 mL of acetonitrile (ACN), and 0.21 mL (3.0 mmol, 1.0 eq.) of thioglycolic acid and 0.63 mL (4.5 mmol, 1.5 eq.) of triethylamine were then added dropwise while stirring at room temperature. The reaction solution was stirred for 18 hours, and then concentrated under reduced pressure. Sub-

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sequently, water was added to the reaction solution, and the reaction solution was extracted three times with ethyl acetate (EA). An organic layer was dried over magnesium sulfate, concentrated under reduced pressure, and then purified by column chromatography (DCM:MeOH=10:1) to obtain 0.3 g (yield: 47%) of a target compound.

^1H NMR (500 MHz, CDCl_3) δ 9.82 (brs, 1H), 6.24-6.07 (m, 2H), 3.76 (s, 2H), 3.12 (s, 1H), 2.97 (s, 1H), 2.54 (dd, $J=9.0$, 4.7 Hz, 1H), 2.04-1.87 (m, 1H), 1.57 (d, $J=8.5$ Hz, 1H), 1.48-1.35 (m, 2H).

Synthesis of Compound III

0.26 g (1.22 mmol, 1.0 eq.) of Compound 5 was dissolved in 15 mL of acetonitrile (ACN), and 0.35 g (1.83 mmol, 1.5 eq.) of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI) and 0.28 g (2.44 mmol, 2.0 eq.) of N-hydroxysuccinimide (NHS) were added while stirring at room temperature. The reaction solution was stirred for 3 hours, and then concentrated under reduced pressure. Subsequently, water was added to the reaction solution, and the reaction solution was extracted three times with EA. An organic layer was recovered, dried over magnesium sulfate, and then concentrated under reduced pressure. Then, the residue was purified on a silica gel column using chromatography (EA:Hex=2:1) to obtain 0.32 g (yield: 85%) of a target compound.

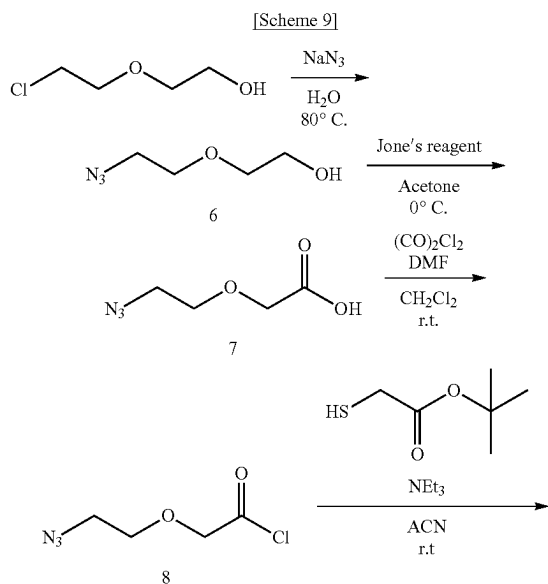
Confirmation of structure of Compound III

^1H NMR (500 MHz, CDCl_3) δ 6.21 (dd, $J=5.5$, 3.0 Hz, 1H), 6.15 (dd, $J=5.5$, 3.1 Hz, 1H), 4.01 (s, 2H), 3.15 (s, 1H), 2.98 (s, 1H), 2.86 (s, 4H), 2.54 (dd, $J=9.2$, 4.6 Hz, 1H), 2.02 (dt, $J=11.9$, 4.0 Hz, 1H), 1.59 (d, $J=8.6$ Hz, 1H), 1.46-1.36 (m, 2H).

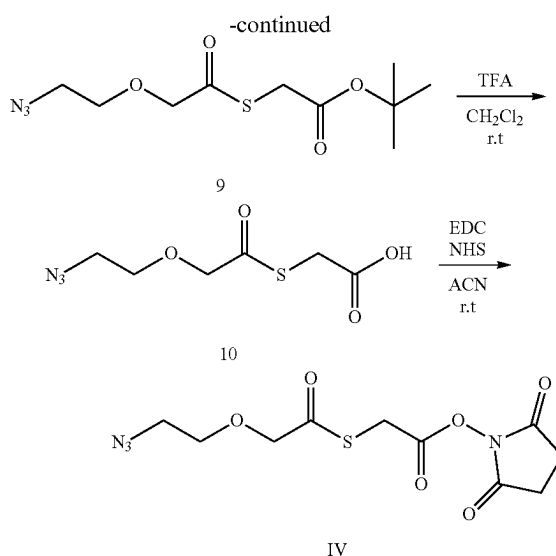
LCMS (ESI): m/z 332.16 $[\text{M}+\text{Na}^+]$

The structure of Compound III was confirmed by mass spectrometry. The results are shown in FIG. 20.

1.1.4. Compound IV (SO4 Linker: NHS & Azide)



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Synthesis of Compound IV (Scheme 9, FIGS. 21 and 22)

Synthesis of Compound 6

2-(2-Chloroethoxy)ethanol (2 mL, 18.94 mmol) was dissolved in distilled water (12 mL), and NaN_3 (3.08 g, 47.35 mmol, 2.5 eq.) was added thereto. The resulting mixture was stirred at 80° C. for 16 hours. The reaction mixture was cooled to room temperature, and poured into 5% NaOH (aq.) (20 mL), and then stirred for approximately 10 minutes. The reaction mixture was extracted three times with diethyl ether, and an organic layer was dried over magnesium sulfate, and filtered. The filtrate was concentrated under reduced pressure to obtain 2.47 g (yield: 99%) of a target compound.

Synthesis of Compound 7

2.47 g (18.84 mmol) of Compound 6 was dissolved in acetone (50 mL), and a 1 M Jone's reagent (75.36 mL, 75.36 mmol, 4 eq.) was then slowly added at 0° C. The reaction mixture was stirred at 0° C. for 3 hours, warmed to room temperature, and then stirred for approximately 10 minutes. The reaction mixture was extracted three times with ethyl acetate (EA), and an organic layer was dried over magnesium sulfate, and filtered. The filtrate was concentrated under reduced pressure to obtain 2.69 g (yield: 98%) of a target compound.

Synthesis of Compound 8

2.69 g (18.57 mmol, 1.0 eq.) of Compound 7 was dissolved in 50 mL of DCM, and 0.1 mL (1.29 mmol, 0.07 eq.) of DMF and 2.43 mL (27.86 mmol, 1.5 eq.) of oxalyl chloride were then added dropwise while stirring at room temperature. The reaction solution was stirred for 3 hours, and then concentrated under reduced pressure to obtain 2.42 g (yield: 80%) of a target compound.

Synthesis of Compound 9

0.88 g (5.40 mmol, 1.0 eq.) of Compound 8 was dissolved in 30 mL of DCM, and 0.8 g (5.40 mmol, 1.0 eq.) of

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tert-butyl 2-mercaptoacetate and 1.41 mL (8.10 mmol, 1.5 eq.) of N,N-diisopropylethylamine were then added dropwise while stirring at room temperature. The reaction mixture was stirred for 2 hours, and then concentrated under reduced pressure. Subsequently, water was added to the reaction solution, and the reaction solution was extracted three times with dichloromethane (DCM). An organic layer was dried over magnesium sulfate, concentrated under reduced pressure, and then purified by column chromatography (Hex:EA=5:1) to obtain 0.73 g (yield: 49%) of a target compound.

Synthesis of Compound 10

0.73 g (2.66 mmol, 1 eq.) of Compound 9 was dissolved in 10 mL of DCM, and 10 mL (129.8 mmol, 48 eq.) of trifluoroacetic acid was then added dropwise while stirring at room temperature. The reaction solution was stirred for 8 hours, and then concentrated under reduced pressure to obtain 0.40 g (yield: 70%) of a target compound.

Synthesis of Compound IV

1.97 g (9.0 mmol, 1.0 eq.) of Compound 10 was dissolved in 25 mL of acetonitrile (ACN), and 2.58 g (13.5 mmol, 1.5

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eq.) of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI) and 2.07 g (18.0 mmol, 2.0 eq.) of N-hydroxysuccinimide (NHS) were added while stirring at room temperature. The reaction solution was stirred for 3 hours, and then concentrated under reduced pressure. Subsequently, water was added to the reaction solution, and the reaction solution was extracted three times with EA. An organic layer was recovered, dried over magnesium sulfate, and then concentrated under reduced pressure. Then, the residue was purified on a silica gel column using chromatography (EA:Hex=2:1) to obtain 1.03 g (yield: 36%) of a target compound.

Confirmation of Structure of Compound IV

^1H NMR (500 MHz, CDCl_3) δ 4.30 (s, 2H), 4.00 (s, 2H), 3.87-3.75 (m, 2H), 3.55-3.47 (m, 2H), 2.86 (s, 4H).

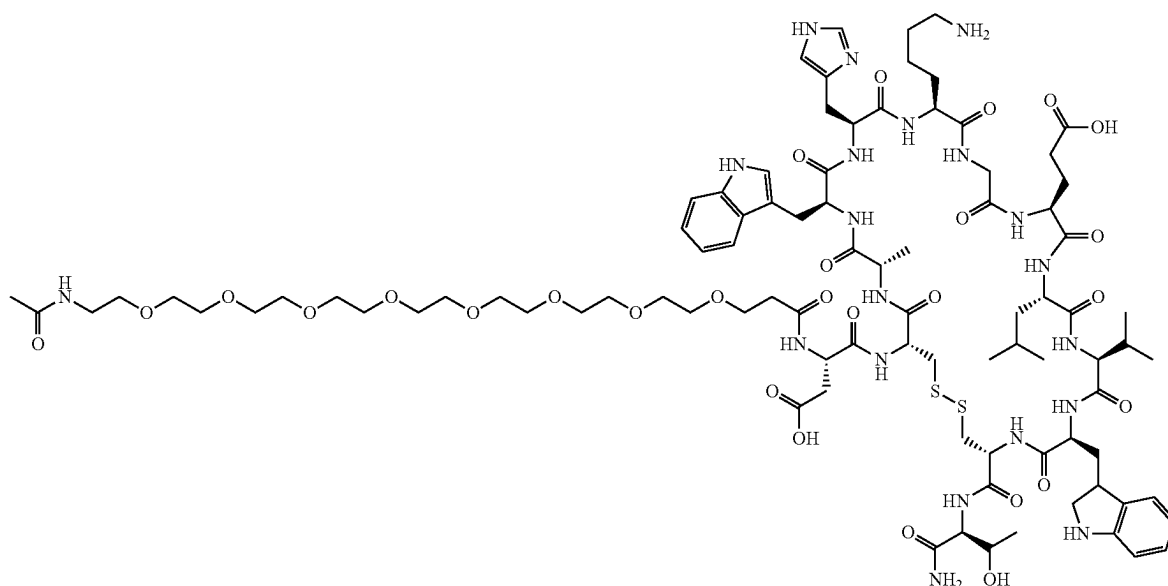
LRMS (ESI): m/z 334.0 $[\text{M}+\text{NH}_4^+]$

The structure of Compound IV was confirmed by mass spectrometry. The results are shown in FIG. 23.

1.2. Synthesis and Confirmation of Structure of Site-Specific Fc Interactome (SSFI)

1.2.1. Synthesis and Confirmation of Structure of SSFI (where Xa_1 is Lysine)

Synthesis of SSFI (6Lys)



FcBP (6Lys)

List and order of introduction of Fmoc amino acids used Fmoc-L-Thr(tBu)-OH, Fmoc-Cys(Trt)-OH, Fmoc-L-Trp(Boc)-OH, Fmoc-L-Val-OH, Fmoc-L-Leu-OH, Fmoc-L-Glu(OtBu)-OH, Fmoc-Gly-OH, Fmoc-Lys(Boc)-OH, Fmoc-L-His(Trt)-OH, Fmoc-L-Trp(Boc)-OH, Fmoc-Ala-OH, Fmoc-Cys(Trt)-OH, Fmoc-Asp(tBu)-OH.

Preparation Method

(a) Introduction of Amino Acids

Amounts of reagents used in the following process were based on 0.25 mmole. 0.5 g (0.48 mmole/g) of a clear amide resin (Peptide International Inc., USA) was put into a synthesis reactor, and 1 mmole of each Fmoc-amino acid block was weighed to prepare a peptide amino acid sequence from the C-terminus to the N-terminus in the above order.

A reaction for activating an Fmoc-amino acid to attach the activated residue to a clear amide resin was performed sequentially from the C-terminal amino acid.

Removal of Fmoc was performed in 20% piperidine-containing DMF, and activation and introduction of the residue was performed by mixing amino acids prepared to correspond to the sequence with 2 mL of a 0.5 M HOBt-containing DMF solution, 2 mL of a 0.5 M HBTU-containing DMF solution, and 174 μ L of DIPEA for 5 minutes, and mixing the resulting mixture for 2 hours in a reactor containing the resin.

Confirmation of an introduction reaction was carried out using a Kaiser test method. When the introduction reaction was confirmed to be incomplete, the introduction reaction was repeated once more, or capping was performed using a 20% Ac₂O-containing DMF solution. In each of the introduction reaction and the Fmoc removal, the resin was thoroughly washed with DMF and DCM before moving to the next step. This process was repeatedly performed until the targeted peptide sequence was completed.

(b) Introduction of H-PEG8-OH

To introduce H-PEG₈-OH to the N-terminus of the sequence after the introduction of the amino acids was completed, 1 mL of a 0.5 M Fmoc-N-amido-dPEG8-acid-in-DMF solution, 1 mL of a 0.5 M HBTU-containing DMF solution, 1 mL of a 0.5 M HOBt-containing DMF solution, and 87 μ L of DIPEA were mixed for 5 minutes, and the resulting mixture was mixed for 2 hours in a reactor containing a reactive resin.

Progression of the reaction was monitored by the Kaiser test method. When the unreacted amine was found to remain, the reaction time was extended for another 1 to 3 hours, or the reaction solution was discarded, and the aforementioned reaction process was repeated again. Removal of the N-terminal Fmoc protective group was performed using 20% piperidine-containing DMF, and the resin to which the peptide was attached was then dried and weighed.

(c) 250 mg of the resin to which the peptide was attached as prepared in the step (b) was stirred with 2 mL of a mixed solution of TFA, TIS, water and EDT (94:1.0:2.5:2.5) at room temperature for 120 minutes to cleave the peptide from the resin. The cleaved mixture was filtered, and the filtrate was concentrated to about half its volume using nitrogen gas. Thereafter, ether was poured into the mixture to precipitate the peptide. The precipitated peptide was washed three times with ether, and dried under nitrogen gas. The dried precipitate was dissolved in 0.1% TFA-30% ACN-containing water, stirred for 6 hours, and then concentrated.

The concentrate was dissolved at a concentration of 0.1 mg/mL in a 5%-DMSO-20%-ACN-containing 0.01 M ammonium acetate buffer (pH 6.5) solution, and then stirred for 3 days while being exposed to air. Progression of a disulfide bond-forming reaction was monitored by HPLC. When the reaction was found not to progress any more, the reaction solution was freeze-dried to obtain a peptide precipitate.

(d) Purification

The peptide precipitate obtained by freeze-drying in the step (c) was separated under the prep-LC primary purification conditions listed in the following Table 4, further purified under the prep-LC secondary purification conditions listed in the following Table 5, and then freeze-dried. The resulting peptide was confirmed to have a purity of 90% or more by analytical HPLC, and the molecular weight of the synthesized peptide was confirmed using an LC/MS mass spectrometer.

H-PEG8-Asp-Cys*-Ala-Trp-His-Lys-Gly-Glu-Leu-Val-Trp-Cys*-Thr-NH₂ (Cys*: disulfide bonding sites)

TABLE 4

Prep-LC primary purification condition			
Prep-LC primary purification condition			
Equipment name	Waters 2525 pump, Waters 2487 UV detector		
Column	Waters, XBridge Prep C18 5 μ m OBD 19 $\times$ 250 mm		
Mobile phase A/B	0.1% TFA-20% ACN/0.1% TFA in 80% ACN		
Gradient	0 min $\rightarrow$ 30 min:B 0% $\rightarrow$ B 100%		
Flow rate	17 mL/min	Detection wavelength	UV 280 nm

TABLE 5

Prep-LC secondary purification condition			
Prep-LC secondary purification condition			
Equipment name	Waters 2525 pump, Waters 2487 UV detector		
Column	Waters, XBridge Prep C18 5 μ m OBD 19 $\times$ 250 mm		
Mobile phase A/B	0.1% TFA-30% ACN/0.1% TFA in 70% ACN		
Gradient	0 min $\rightarrow$ 30 min:B 0% $\rightarrow$ B 100%		
Flow rate	15 mL/min	Detection wavelength	UV 280 nm

Confirmation of Structure of SSFI (6Lys)

Synthesis of SSFI (6Lys) was confirmed by molecular weight measurement by matrix-assisted laser desorption/ionization (MALDI) mass spectrometry.

Measuring equipment: Ultraflex extreme (Bruker)

Measuring matrix: CHCA (α -Cyano-4-hydroxycinnamic acid) & DHB (2,5-Dihydroxybenzoic acid)

Calculated molecular weight: 2010.29 g/mol

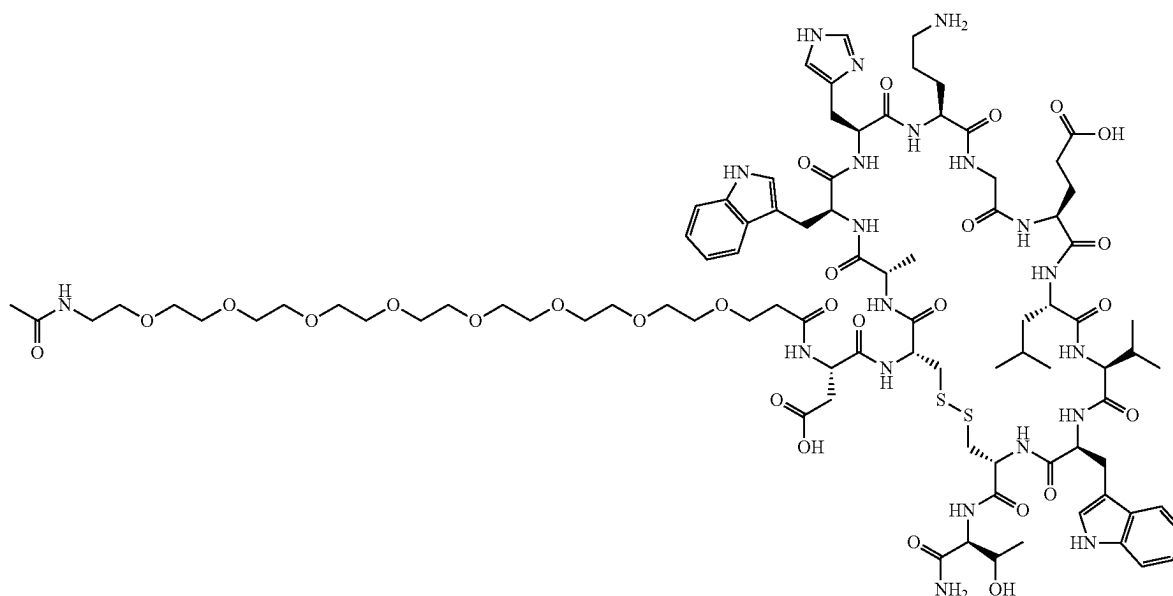
Measured molecular weight (M+H)⁺: 2011.89 g/mol

The mass spectrometry results of SSFI (6Lys) are shown in FIG. 24.

1.2.2. Synthesis and Confirmation of Structure of SSFI (where Xa₁ is Ornithine)

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FcBP (6Ornithine)

Synthesis of SSFI (6Orn)

List and order of introduction of Fmoc amino acids used
Fmoc-L-Thr(tBu)-OH, Fmoc-Cys(Trt)-OH, Fmoc-L-Trp
(Boc)-OH, Fmoc-L-Val-OH, Fmoc-L-Leu-OH, Fmoc-
L-Glu(OtBu)-OH, Fmoc-Gly-OH, Fmoc-Orn(Boc)-
OH, Fmoc-L-His(Trt)-OH, Fmoc-L-Trp(Boc)-OH
Fmoc-Ala-OH, Fmoc-Cys(Trt)-OH, Fmoc-Asp(tBu)-
OH.

Preparation Method

(a) Introduction of Amino Acids

Amounts of reagents used in the following process were
based on 0.25 mmole. 0.5 g (0.48 mmole/g) of a clear amide resin (Peptide International Inc., USA) was put into a
synthesis reactor, and 1 mmole of each Fmoc-amino acid
block was weighed to prepare a peptide amino acid sequence
from the C-terminus to the N-terminus in the above.

A reaction for activating an Fmoc-amino acid to attach the
activated residue to a clear amide resin was performed
sequentially from the C-terminal amino acid.

Removal of Fmoc was performed in 20% piperidine-
containing DMF, and activation and introduction of the
residue was performed by mixing amino acids prepared to
correspond to the sequence with 2 mL of a 0.5 M HOBt-
containing DMF solution, 2 mL of a 0.5 M HBTU-contain-
ing DMF solution, and 174 μ L of DIPEA for 5 minutes, and
mixing the resulting mixture for 2 hours in a reactor con-
taining the resin.

Confirmation of an introduction reaction was carried out
using a Kaiser test method. When the introduction reaction
was confirmed to be incomplete, the introduction reaction
was repeated once more, or capping was performed using a
20% Ac₂O-containing DMF solution. In each of the intro-
duction reaction and the Fmoc removal, the resin was
thoroughly washed with DMF and DCM before moving to
the next step. This process was repeatedly performed until
the targeted peptide sequence was completed.

(b) Introduction of H-PEG8-OH

To introduce H-PEG₈-OH to the N-terminus of the
sequence after the introduction of the amino acids was

completed, 1 mL of a 0.5 M Fmoc-N-amido-dPEG8-acid-
in-DMF solution, 1 mL of a 0.5 M HBTU-containing DMF
solution, 1 mL of a 0.5 M HOBt-containing DMF solution,
and 87 μ L of DIPEA were mixed for 5 minutes, and the
resulting mixture was mixed for 2 hours in a reactor con-
taining a reactive resin.

Progression of the reaction was monitored by the Kaiser
test method. When the unreacted amine was found to
remain, the reaction time was extended for another 1 to 3
hours, or the reaction solution was discarded, and the
aforementioned reaction process was repeated again.
Removal of the N-terminal Fmoc protective group was
performed using 20% piperidine-containing DMF, and the
resin to which the peptide was attached was then dried and
weighed.

(c) 250 mg of the resin to which the peptide was attached
as prepared in the step (b) was stirred with 2 mL of a mixed
solution of TFA, TIS, water and EDT (94:1.0:2.5:2.5) at
room temperature for 120 minutes to cleave the peptide from
the resin. The cleaved mixture was filtered, and the filtrate
was concentrated to about half its volume using nitrogen
gas. Thereafter, ether was poured into the mixture to pre-
cipitate the peptide. The precipitated peptide was washed
three times with ether, and dried under nitrogen gas. The
dried precipitate was dissolved in 0.1% TFA-30% ACN-
containing water, stirred for 6 hours, and then concentrated.

The concentrate was dissolved at a concentration of 0.1
mg/mL in a 5%-DMSO-20%-ACN-containing 0.01 M
ammonium acetate buffer (pH 6.5) solution, and then stirred
for 3 days while being exposed to air. Progression of a
disulfide bond-forming reaction was monitored by HPLC.
When the reaction was found not to progress any more, the
reaction solution was freeze-dried to obtain a peptide pre-
cipitate.

(d) Purification

The peptide precipitate obtained by freeze-drying in the
step (c) was separated under the prep-LC primary purifica-
tion conditions listed in the following Table 6, further
purified under the prep-LC secondary purification condi-

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tions listed in the following Table 7, and then freeze-dried. The resulting peptide was confirmed to have a purity of 90% or more by analytical HPLC, and the molecular weight of the synthesized peptide was confirmed using an LC/MS mass spectrometer.

H-PEG8-Asp-Cys*-Ala-Trp-His-Orn-Gly-Glu-Leu-Val-Trp-Cys*-Thr-NH₂(Cys*: disulfide bonding sites)

TABLE 6

Prep-LC primary purification condition	
Prep-LC primary purification condition	
Equipment name	Waters 2525 pump, Waters 2487 UV detector
Column	Waters, XBridge Prep C18 5 μ m OBD 19 $\times$ 250 mm
Mobile phase A/B	0.1% TFA-20% ACN/0.1% TFA in 80% ACN
Gradient	0 min $\rightarrow$ 30 min:B 0% $\rightarrow$ B 100%
Flow rate	17 mL/min Detection wavelength UV 280 nm

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TABLE 7

Prep-LC secondary purification condition	
Prep-LC secondary purification condition	
Equipment name	Waters 2525 pump, Waters 2487 UV detector
Column	Waters, XBridge Prep C18 5 μ m OBD 19 $\times$ 250 mm
Mobile phase A/B	0.1% TFA-30% ACN/0.1% TFA in 70% ACN
Gradient	0 min $\rightarrow$ 30 min:B 0% $\rightarrow$ B 100%
Flow rate	15 mL/min Detection wavelength UV 280 nm

Confirmation of Structure of SSFI (6Orn)

Synthesis of SSFI (6Ornithine) was Confirmed by Molecular Weight Measurement by Matrix-Assisted Laser Desorption/Ionization (MALDI) Mass Spectrometry.

Measuring equipment: Ultraflex extreme (Bruker)

Measuring matrix: CHCA (α -Cyano-4-hydroxycinnamic acid) & DHB (2,5-Dihydroxybenzoic acid)

Calculated molecular weight: 1996.26 g/mol

Measured molecular weight (M+2H)²⁺: 999.13 g/mol

The mass spectrometry results of SSFI (6Orn) are shown in FIG. 25.

1.2.3. Synthesis and confirmation of structure of SSFI (where Xa₁ is diaminobutyric acid (Dab))



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Synthesis of SSFI (6Dab)

List and order of introduction of Fmoc amino acids used
Fmoc-L-Thr(tBu)-OH, Fmoc-Cys(Trt)-OH, Fmoc-L-Trp
(Boc)-OH, Fmoc-L-Val-OH, Fmoc-L-Leu-OH, Fmoc-
L-Glu(OtBu)-OH, Fmoc-Gly-OH, Fmoc-Dab(Boc)-
OH, Fmoc-L-His(Trt)-OH, Fmoc-L-Trp(Boc)-OH
Fmoc-Ala-OH, Fmoc-Cys(Trt)-OH, Fmoc-Asp(tBu)-
OH.

Preparation Method

(a) Introduction of Amino Acids

Amounts of reagents used in the following process were
based on 0.25 mmole. 0.5 g (0.48 mmole/g) of a clear amide
resin (Peptide International Inc., USA) was put into a
synthesis reactor, and 1 mmole of each Fmoc-amino acid
block was weighed to prepare a peptide amino acid sequence
from the C-terminus to the N-terminus in the above order.

A reaction for activating an Fmoc-amino acid to attach the
activated residue to a clear amide resin was performed
sequentially from the C-terminal amino acid.

Removal of Fmoc was performed in 20% piperidine-
containing DMF, and activation and introduction of the
residue was performed by mixing amino acids prepared to
correspond to the sequence with 2 mL of a 0.5 M HOBt-
containing DMF solution, 2 mL of a 0.5 M HBTU-contain-
ing DMF solution, and 174 μ L of DIPEA for 5 minutes, and
mixing the resulting mixture for 2 hours in a reactor con-
taining the resin.

Confirmation of an introduction reaction was carried out
using a Kaiser test method. When the introduction reaction
was confirmed to be incomplete, the introduction reaction
was repeated once more, or capping was performed using a
20% Ac₂O-containing DMF solution. In each of the intro-
duction reaction and the Fmoc removal, the resin was
thoroughly washed with DMF and DCM before moving to
the next step. This process was repeatedly performed until
the targeted peptide sequence was completed.

(b) Introduction of H-PEG8-OH

To introduce H-PEG₈-OH to the N-terminus of the
sequence after the introduction of the amino acids was
completed, 1 mL of a 0.5 M Fmoc-N-amido-dPEG8-acid-
in-DMF solution, 1 mL of a 0.5 M HBTU-containing DMF
solution, 1 mL of a 0.5 M HOBt-containing DMF solution,
and 87 μ L of DIPEA were mixed for 5 minutes, and the
resulting mixture was mixed for 2 hours in a reactor con-
taining a reactive resin.

Progression of the reaction was monitored by the Kaiser
test method. When the unreacted amine was found to
remain, the reaction time was extended for another 1 to 3
hours, or the reaction solution was discarded, and the
aforementioned reaction process was repeated again.
Removal of the N-terminal Fmoc protective group was
performed using 20% piperidine-containing DMF, and the
resin to which the peptide was attached was then dried and
weighed.

(c) 250 mg of the resin to which the peptide was attached
as prepared in the step (b) was stirred with 2 mL of a mixed
solution of TFA, TIS, water and EDT (94:1.0:2.5:2.5) at
room temperature for 120 minutes to cleave the peptide from
the resin. The cleaved mixture was filtered, and the filtrate
was concentrated to about half its volume using nitrogen
gas. Thereafter, ether was poured into the mixture to pre-
cipitate the peptide. The precipitated peptide was washed
three times with ether, and dried under nitrogen gas. The

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dried precipitate was dissolved in 0.1% TFA-30% ACN-
containing water, stirred for 6 hours, and then concentrated.

The concentrate was dissolved at a concentration of 0.1
mg/mL in a 5%-DMSO-20%-ACN-containing 0.01 M
ammonium acetate buffer (pH 6.5) solution, and then stirred
for 3 days while being exposed to air. Progression of a
disulfide bond-forming reaction was monitored by HPLC.
When the reaction was found not to progress any more, the
reaction solution was freeze-dried to obtain a peptide pre-
cipitate.

(d) Purification

The peptide precipitate obtained by freeze-drying in the
step (c) was separated under the prep-LC primary purifica-
tion conditions listed in the following Table 8, further
purified under the prep-LC secondary purification condi-
tions listed in the following Table 9, and then freeze-dried.
The resulting peptide was confirmed to have a purity of 90%
or more by analytical HPLC, and the molecular weight of
the synthesized peptide was confirmed using an LC/MS
mass spectrometer.

H-PEG8-Asp-Cys*-Ala-Trp-His-Dab-Gly-Glu-Leu-Val-

Trp-Cys*-Thr-NH₂

(Cys*: disulfide bonding sites)

TABLE 8

Prep-LC primary purification condition	
Prep-LC primary purification condition	
Equipment name	Waters 2525 pump, Waters 2487 UV detector
Column	Waters, XBridge Prep C18 5 μ m OBD 19 $\times$ 250 mm
Mobile phase A/B	0.1% TFA-20% ACN/0.1% TFA in 80% ACN
Gradient	0 min $\rightarrow$ 30 min:B 0% $\rightarrow$ B 100%
Flow rate	17 ml/min
Detection wavelength	UV 280 nm

TABLE 9

Prep-LC secondary purification condition	
Prep-LC secondary purification condition	
Equipment name	Waters 2525 pump, Waters 2487 UV detector
Column	Waters, XBridge Prep C18 5 μ m OBD 19 $\times$ 250 mm
Mobile phase A/B	0.1% TFA-30% ACN/0.1% TFA in 70% ACN
Gradient	0 min $\rightarrow$ 30 min:B 0% $\rightarrow$ B 100%
Flow rate	15 mL/min
Detection wavelength	UV 280 nm

Confirmation of structure of SSFI (6Dab)

Synthesis of SSFI (6Dab) was confirmed by molecular
weight measurement by matrix-assisted laser desorption/
ionization (MALDI) mass spectrometry.

Measuring equipment: Ultraflex extreme (Bruker)

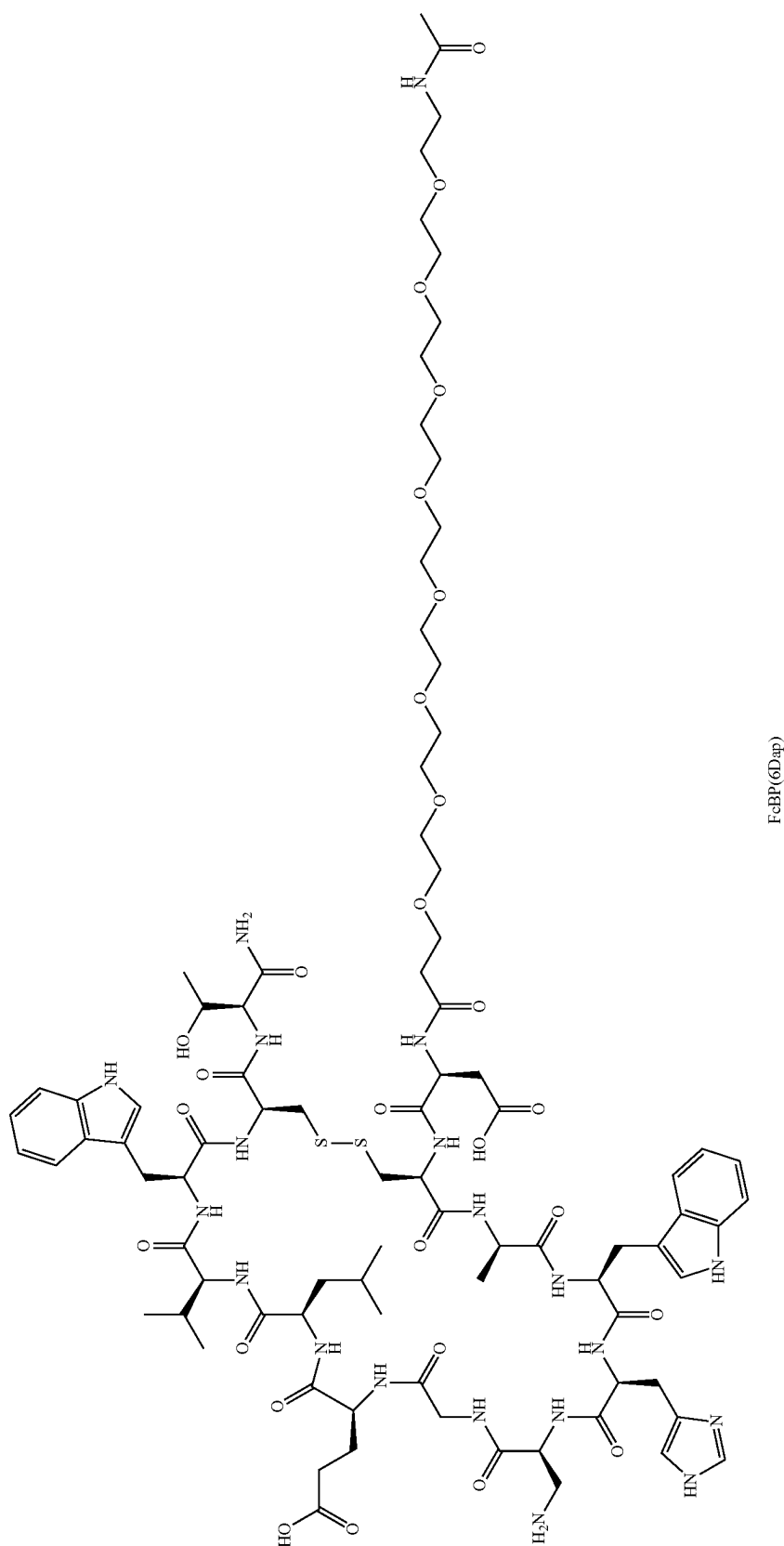
Measuring matrix: CHCA (α -Cyano-4-hydroxycinnamic
acid) & DHB (2,5-Dihydroxybenzoic acid)

Calculated molecular weight: 1982.24 g/mol

Measured molecular weight (M+2H)²⁺: 992.33 g/mol

The mass spectrometry results of SSFI (6Dab) are shown
in FIG. 26.

1.2.4. Synthesis and Confirmation of Structure of SSFI
(where Xa₁ is Diaminopropionic Acid (Dap))



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Synthesis of SSFI (6Dap)

Sequence: Nor-PEG8-DCAWHA(beta amino alanine, Dap)GELVWCT-CONH₂

List and order of introduction of Fmoc amino acids used Fmoc-L-Thr(tBu)-OH, Fmoc-Cys(Trt)-OH, Fmoc-L-Trp (Boc)-OH, Fmoc-L-Val-OH, Fmoc-L-Leu-OH, Fmoc-L-Glu(OtBu)-OH, Fmoc-Gly-OH, Fmoc-Dap(Boc)-OH, Fmoc-L-His(Trt)-OH, Fmoc-L-Trp(Boc)-OH, Fmoc-Ala-OH, Fmoc-Cys(Trt)-OH, Fmoc-Asp(tBu)-OH.

Preparation Method

(a) Introduction of Amino Acids

Amounts of reagents used in the following process were based on 0.25 mmole. 0.5 g (0.48 mmole/g) of a clear amide resin (Peptide International Inc., USA) was put into a synthesis reactor, and 1 mmole of each Fmoc-amino acid block was weighed to prepare a peptide amino acid sequence from the C-terminus to the N-terminus in the above order.

A reaction for activating an Fmoc-amino acid to attach the activated residue to a clear amide resin was performed sequentially from the C-terminal amino acid.

Removal of Fmoc was performed in 20% piperidine-containing DMF, and activation and introduction of the residue was performed by mixing amino acids prepared to correspond to the sequence with 2 mL of a 0.5 M HOBt-containing DMF solution, 2 mL of a 0.5 M HBTU-containing DMF solution, and 174 µL of DIPEA for 5 minutes, and mixing the resulting mixture for 2 hours in a reactor containing the resin.

Confirmation of an introduction reaction was carried out using a Kaiser test method. When the introduction reaction was confirmed to be incomplete, the introduction reaction was repeated once more, or capping was performed using a 20% Ac₂O-containing DMF solution. In each of the introduction reaction and the Fmoc removal, the resin was thoroughly washed with DMF and DCM before moving to the next step. This process was repeatedly performed until the targeted peptide sequence was completed.

(b) Introduction of H-PEG8-OH

To introduce H-PEG₈-OH to the N-terminus of the sequence after the introduction of the amino acids was completed, 1 mL of a 0.5 M Fmoc-N-amido-dPEG8-acid-in-DMF solution, 1 mL of a 0.5 M HBTU-containing DMF solution, 1 mL of a 0.5 M HOBt-containing DMF solution, and 87 µL of DIPEA were mixed for 5 minutes, and the resulting mixture was mixed for 2 hours in a reactor containing a reactive resin.

Progression of the reaction was monitored by the Kaiser test method. When the unreacted amine was found to remain, the reaction time was extended for another 1 to 3 hours, or the reaction solution was discarded, and the aforementioned reaction process was repeated again. Removal of the N-terminal Fmoc protective group was performed using 20% piperidine-containing DMF, and the resin to which the peptide was attached was then dried and weighed.

(c) 250 mg of the resin to which the peptide was attached as prepared in the step (b) was stirred with 2 mL of a mixed solution of TFA, TIS, water and EDT (94:1.0:2.5:2.5) at room temperature for 120 minutes to cleave the peptide from the resin. The cleaved mixture was filtered, and the filtrate was concentrated to about half its volume using nitrogen gas. Thereafter, ether was poured into the mixture to precipitate the peptide. The precipitated peptide was washed three times with ether, and dried under nitrogen gas. The dried precipitate was dissolved in 0.1% TFA-30% ACN-containing water, stirred for 6 hours, and then concentrated.

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The concentrate was dissolved at a concentration of 0.1 mg/mL in a 5%-DMSO-20%-ACN-containing 0.01 M ammonium acetate buffer (pH 6.5) solution, and then stirred for 3 days while being exposed to air. Progression of a disulfide bond-forming reaction was monitored by HPLC. When the reaction was found not to progress any more, the reaction solution was freeze-dried to obtain a peptide precipitate.

(d) Purification

The peptide precipitate obtained by freeze-drying in the step (c) was separated under the prep-LC primary purification conditions listed in the following Table 10, further purified under the prep-LC secondary purification conditions listed in the following Table 11, and then freeze-dried. The resulting peptide was confirmed to have a purity of 90% or more by analytical HPLC, and the molecular weight of the synthesized peptide was confirmed using an LC/MS mass spectrometer.

H-PEG8-Asp-Cys*-Ala-Trp-His-Dap-Gly-Glu-Leu-Val-

Trp-Cys*-Thr-NH₂

(Cys*: disulfide bonding sites)

TABLE 10

Prep-LC primary purification condition	
Prep-LC primary purification condition	
Equipment name	Waters 2525 pump, Waters 2487 UV detector
Column	Waters, XBridge Prep C18 5 µm OBD 19 × 250 mm
Mobile phase A/B	0.1% TFA-20% ACN/0.1% TFA in 80% ACN
Gradient	0 min → 30 min:B 0% → B 100%
Flow rate	17 mL/min Detection wavelength UV 280 nm

TABLE 12

Prep-LC secondary purification condition	
Prep-LC secondary purification condition	
Equipment name	Waters 2525 pump, Waters 2487 UV detector
Column	Waters, XBridge Prep C18 5 µm OBD 19 × 250 mm
Mobile phase A/B	0.1% TFA-30% ACN/0.1% TFA in 70% ACN
Gradient	0 min → 30 min:B 0% → B 100%
Flow rate	15 mL/min Detection wavelength UV 280 nm

Confirmation of structure of SSFI (6Dap)

Measuring equipment: Quattro Premier Xe (Waters)

Calculated molecular weight: 1968.21 g/mol

Measured molecular weight (M+2H)²⁺: 984.71 g/mol

The mass spectrometry results of SSFI (6Dap) are shown in FIG. 27.

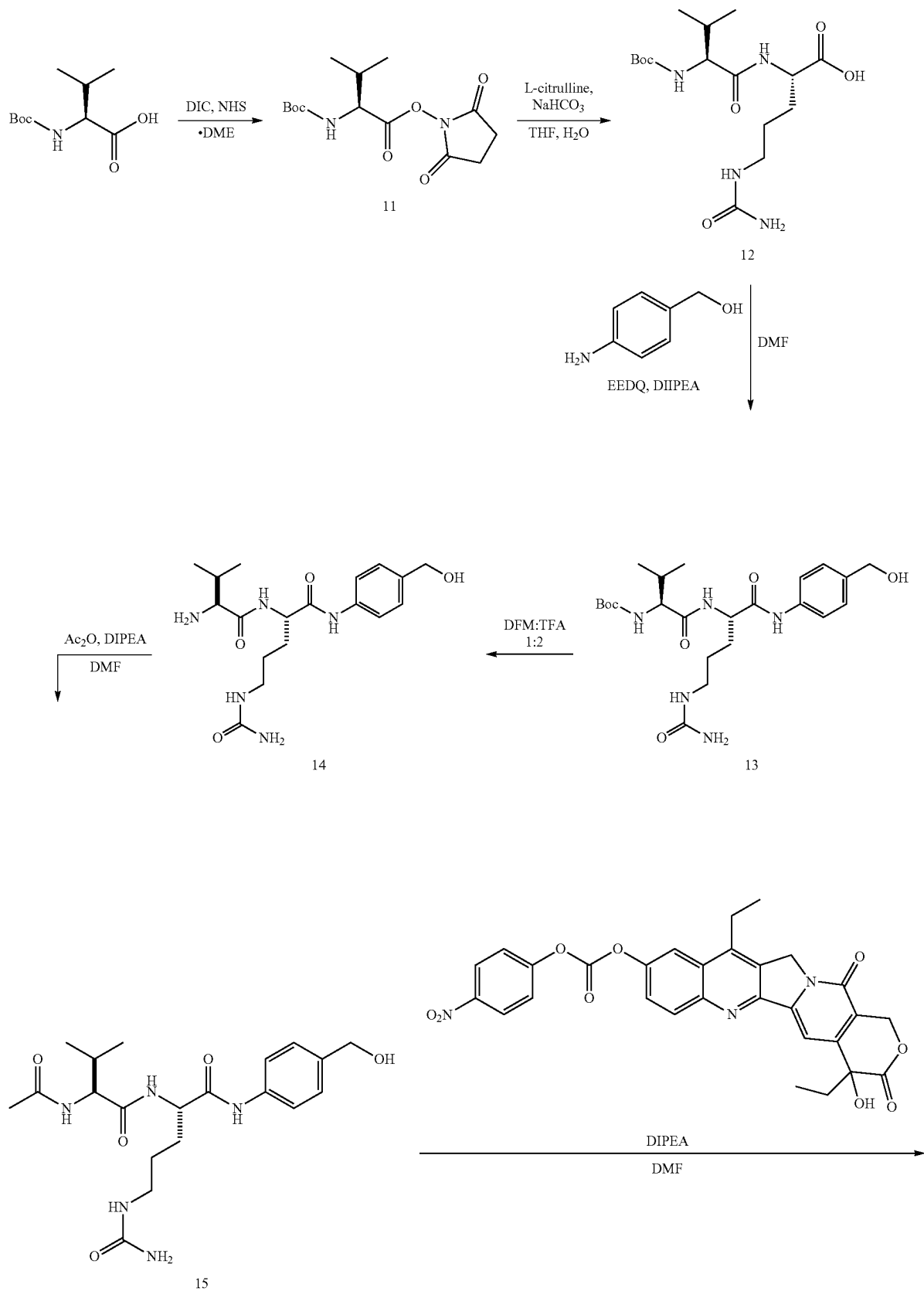
1.3. Synthesis and Confirmation of Structure of VC Linker

1.3.1. Synthesis and Confirmation of Structure of VC Linker (DD2)

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[Scheme 10]



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mmol, 1.0 eq.) of Compound 15 in 10 mL of DMF. The resulting mixture was stirred for 20 hours, and then concentrated under reduced pressure to remove a reaction solution. The concentrate was purified by column chromatography (10% MeOH in DCM) to obtain a target compound. TLC (DCM:MeOH=10:1); R_f =0.2.

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Confirmation of Structure of VC Linker (DD2)

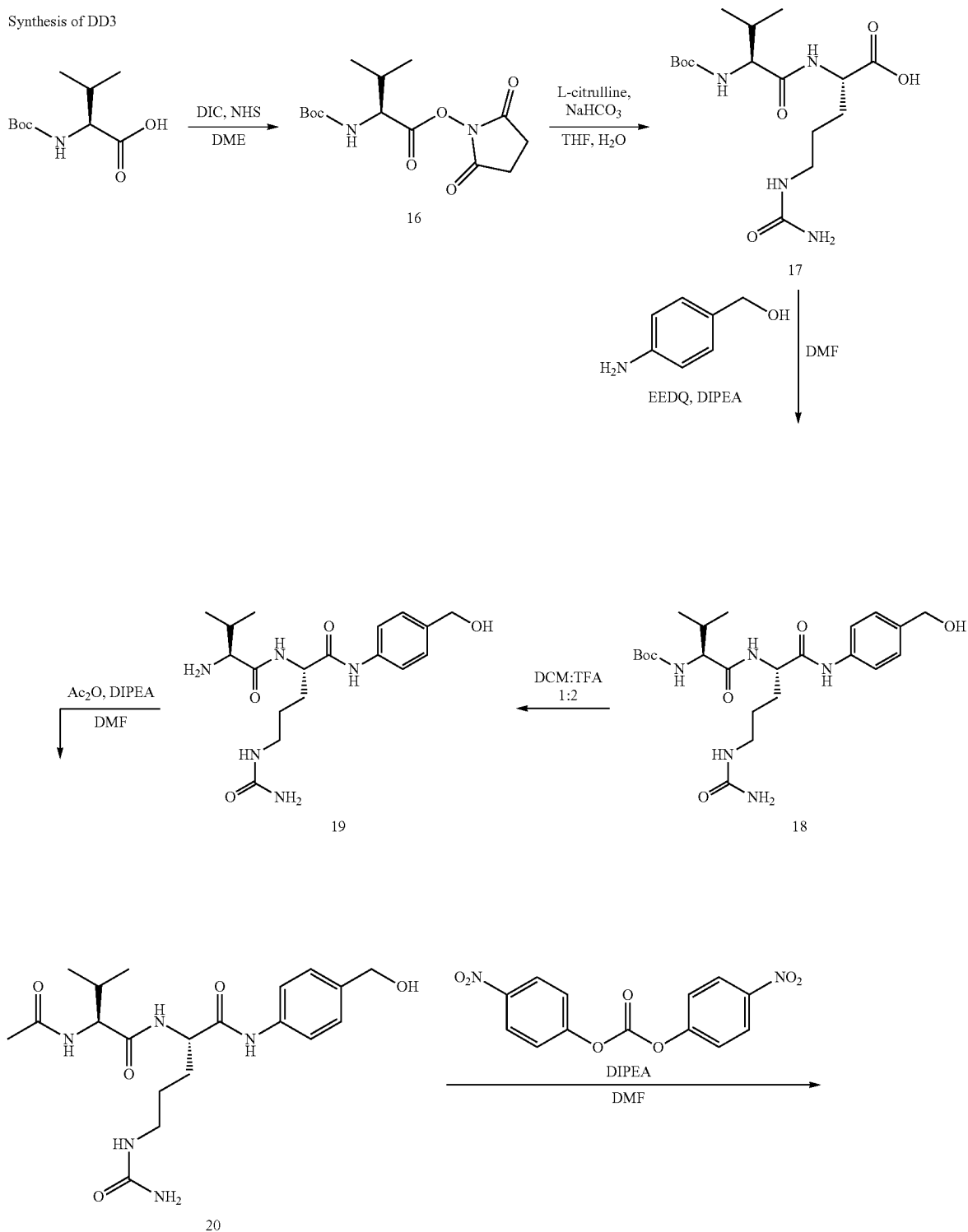
LRMS (ESI): m/z 840.4 $[M+H]^+$

The results are shown in FIG. 28.

1.3.2. Synthesis and Confirmation of Structure of VC Linker (DD3)

[Scheme 11]

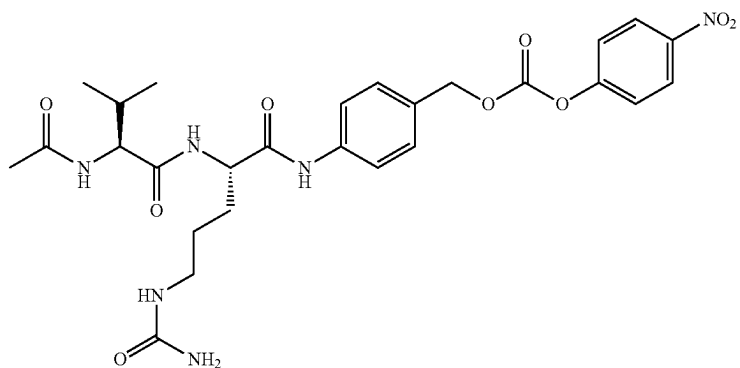
Synthesis of DD3



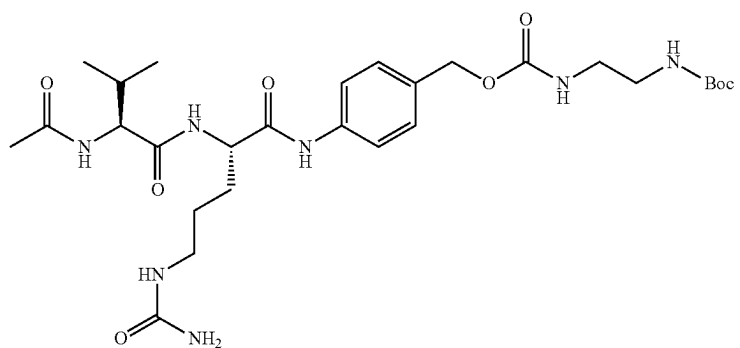
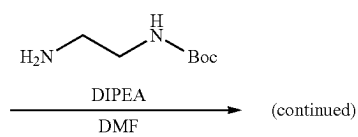
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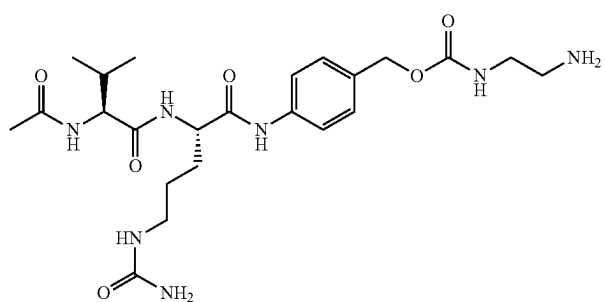
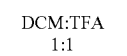
-continued



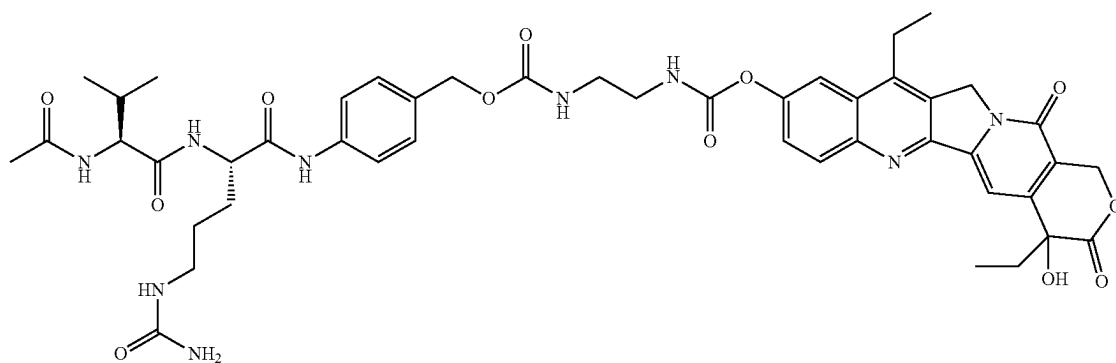
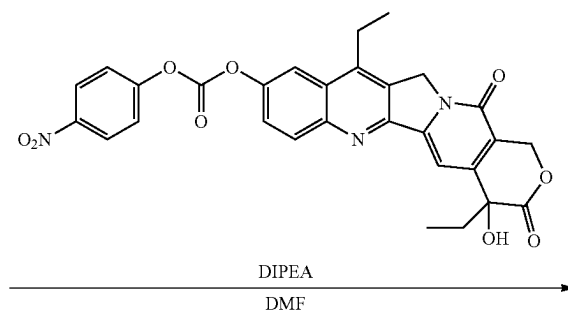
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DD3

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Synthesis of VC Linker (DD3)

Synthesis of Compound 16

5 g (14.7 mmol, 1.0 eq.) of Fmoc-Val-OH and 1.7 g (14.7 mmol, 1.0 eq.) of N-hydroxysuccinimide (NHS) were dissolved in 140 mL of dimethoxyethane (DME), and stirred. 2.5 mL (16.2 mmol, 1.1 eq.) of N,N'-diisopropylcarbodiimide (DIC) was added dropwise at 0° C., and stirred for 16 hours. The reaction solution was filtered under reduced pressure to remove floating matter, and the filtrate was concentrated under reduced pressure. The residue was dissolved in acetone, and stored at a low temperature for 4 hours in a refrigerator. Thereafter, the resulting solution was filtered under reduced pressure to remove the re-formed floating matter, and used in the next reaction without any purification (crude yield: 5.5 g, 86%). TLC (EA:Hex=1:1); $R_f=0.5$.

Synthesis of Compound 17

2.0 g (11.5 mmol, 1.0 eq.) of L-citrulline was dissolved in 100 mL of a 1:1 mixed solution of tetrahydrofuran (THF) and water, and stirred. 988 mg of sodium hydrogen carbonate was added thereto, and stirred. Thereafter, 5.0 g (11.5 mmol, 1.0 eq.) of Compound 16 was dissolved in 80 mL of acetone, added dropwise to the reaction solution, and then stirred. The resulting mixture was stirred for 21 hours, and then concentrated under reduced pressure to remove the organic solvent. An aqueous layer was washed with ethyl acetate (EA), and then titrated to pH 3 by dropwise addition of 2 N HCl. EA was added thereto so that the precipitate in an organic layer was extracted. Then, the organic layer was dried over a saturated saline solution and sodium sulfate, and used in the next reaction without any purification (crude yield: 5.6 g, 98%). TLC (DCM:MeOH=10:1, one drop of formic acid); $R_f=0.1$.

Synthesis of Compound 18

50 mL of 10% piperidine in N,N-dimethylformamide (DMF) was added dropwise to 2.84 g (5.72 mmol, 1.0 eq.) of Compound 17, and stirred. After 4 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the residue was dissolved in water, and filtered under reduced pressure to remove the formed floating matter. An aqueous layer was concentrated to obtain a target compound, which was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); $R_f=0.01$.

Synthesis of Compound 19

54 mg (0.22 mmol, 1.0 eq.) of Compound 18 was dissolved in 2 mL of DMF, and 0.05 mL (0.264 mmol, 1.2 eq.) of N,N-diisopropylethylamine (DIPEA) was added dropwise thereto. Compound 15 was completely dissolved by adding 2 mL of water, and 0.05 mL (0.528 mmol, 2.4 eq.) of an acetic anhydride was added dropwise thereto at room temperature. After 3 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the concentrate was purified by reversed-phase column chromatography to obtain a target compound. TLC (DCM:MeOH=10:1); $R_f=0.05$.

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Synthesis of Compound 20

1 g (3.16 mmol, 1.0 eq.) of Compound 19 was dissolved in 30 mL of a 2:1 mixed solution of DCM and methanol, and 868 mg (3.48 mmol, 1.1 eq.) of N-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ) was added thereto, and stirred. 451 mg (3.66 mmol, 1.16 eq.) of 4-aminobenzyl alcohol was added thereto, and stirred for 5 hours. The resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the concentrate was purified by column chromatography (10% MeOH in DCM) to obtain 419 mg of a target compound (yield: 32%). TLC (DCM:MeOH=10:1); $R_f=0.1$.

Synthesis of Compound 21

240 mg (0.55 mmol, 1.0 eq.) of Compound 20 was dissolved in 10 mL of DMF, and 0.3 mL (1.65 mmol, 3.0 eq.) of DIPEA was added dropwise thereto, and stirred. Thereafter, a reaction was carried out by adding 509 mg (1.65 mmol, 3.0 eq.) of bis-(4-aminophenyl) carbonate. After 3 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and diethyl ether was added thereto to precipitate a target compound, which was filtered under reduced pressure. The obtained target compound was used in the next reaction without any purification.

Synthesis of Compound 22

33 mg (0.06 mmol, 1.0 eq.) of Compound 21 was dissolved in 5 mL of DMF, and 16 mg (0.08 mmol, 1.3 eq.) of 1-[(tert-butoxycarbonyl)amino]-2-aminoethane and 0.02 mL (0.08 mmol, 1.3 eq.) of DIPEA were added dropwise thereto, and stirred. The resulting mixture was stirred for 7 hours, and then concentrated under reduced pressure to remove a reaction solution, and diethyl ether was added thereto to precipitate a target compound, which was filtered under reduced pressure. The obtained target compound was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); $R_f=0.1$.

Synthesis of Compound 23

17 mg (0.03 mmol, 1.0 eq.) of Compound 22 was dissolved in 1 mL of DCM, and 1 mL of trifluoroacetic acid (TFA) was then added dropwise thereto at 0° C. A reaction was carried out by stirring the resulting mixture at 0° C. for 30 minutes, followed by stirring at room temperature for 30 minutes. Thereafter, the mixture was concentrated under reduced pressure to remove a reaction solution such as DCM, and this concentration was then repeated three times. The concentrated matter was dried under high vacuum, and purified by reversed-phase column chromatography to obtain 13 mg of a target compound. TLC (DCM:MeOH=10:1); $R_f=0.01$.

Synthesis of Compound DD3

0.01 mL (0.04 mmol, 1.5 eq.) of DIPEA was added dropwise to a solution obtained by dissolving 13 mg (0.03 mmol, 1.0 eq.) of Compound 23 in 3 mL of DMF, and 22 mg (0.04 mmol, 1.5 eq.) of phenol-activated SN38 was added

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thereto. The resulting mixture was stirred for an hour, and then concentrated under reduced pressure to remove a reaction solution. The concentrate was purified by column chromatography (10% MeOH in DCM) to obtain a target compound. TLC (DCM:MeOH=10:1); R_f =0.2.

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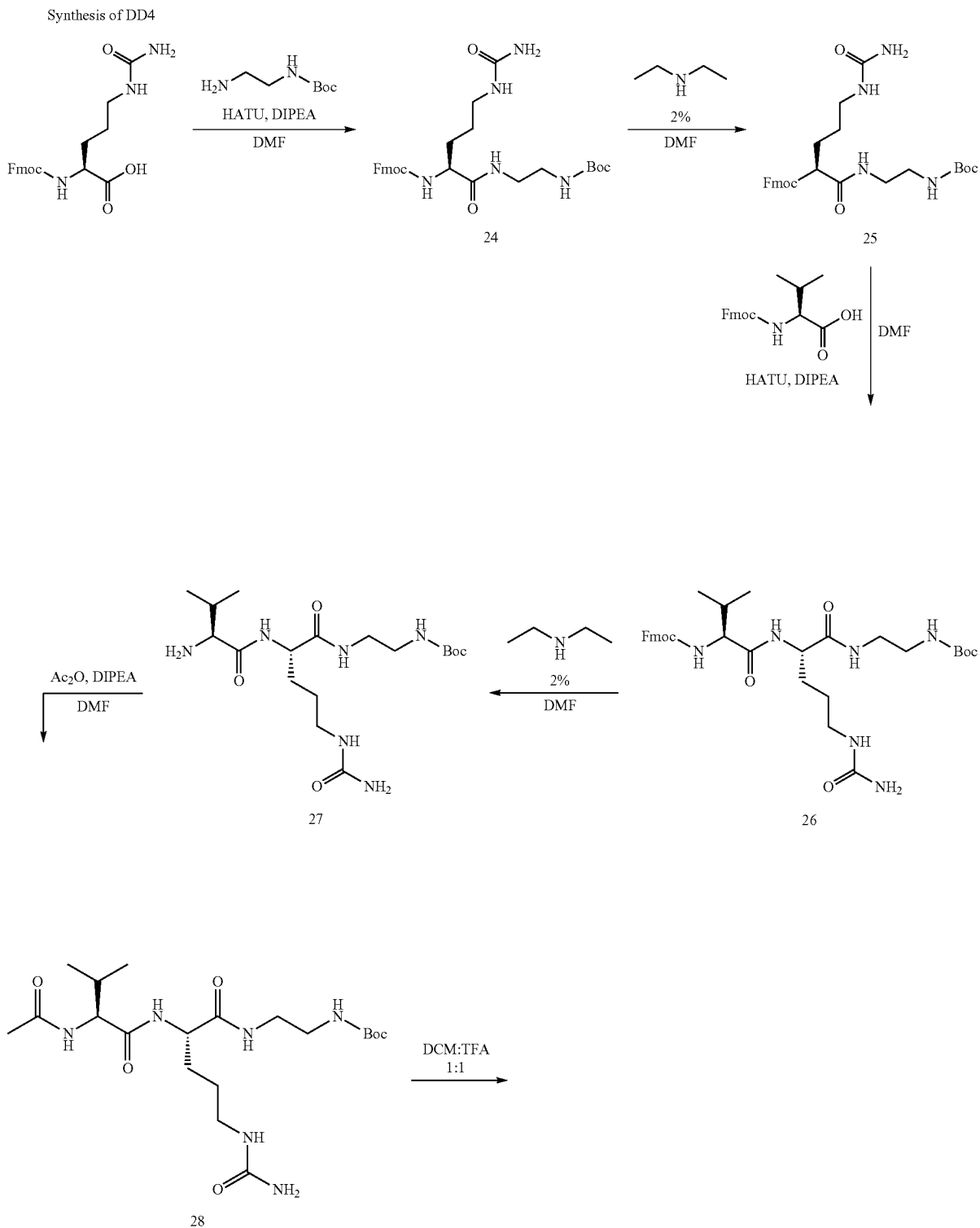
Confirmation of Structure of VC Linker (DD3)

LRMS (ESI): m/z 926.4 $[M+H]^+$

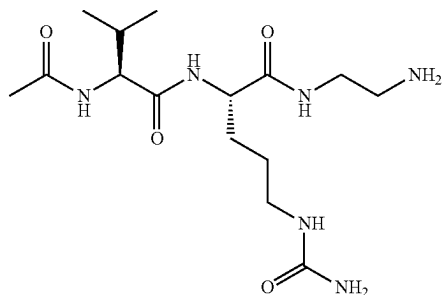
The results are shown in FIG. 29.

1.3.3. Synthesis and Confirmation of Structure of VC Linker (DD4)

[Scheme 12]

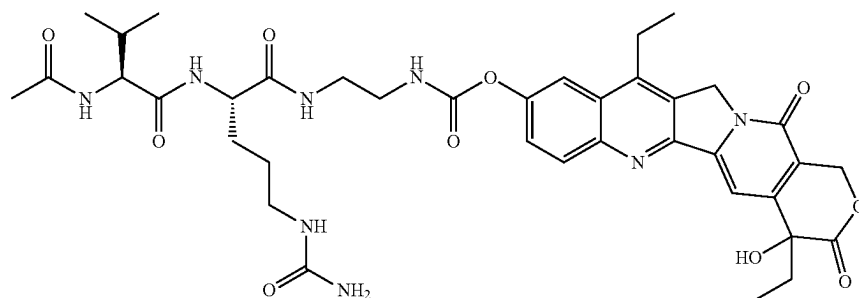
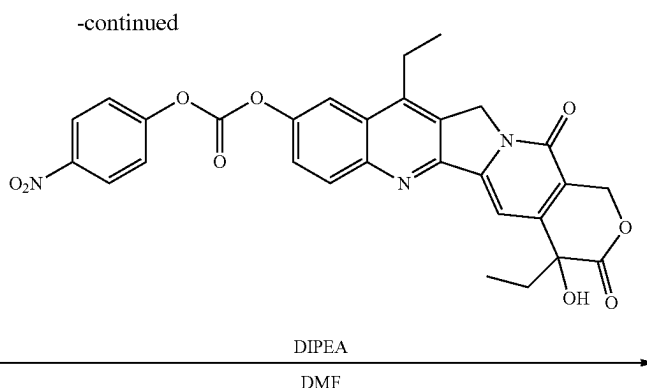


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DD4

Synthesis of VC Linker (DD4)

Synthesis of Compound 24

5 g (14.7 mmol, 1.0 eq.) of Fmoc-Val-OH and 1.7 g (14.7 mmol, 1.0 eq.) of N-hydroxysuccinimide (NHS) were dissolved in 140 mL of dimethoxyethane (DME), and stirred. 2.5 mL (16.2 mmol, 1.1 eq.) of N,N'-diisopropylcarbodiimide (DIC) was added dropwise at 0° C., and stirred for 16 hours. The reaction solution was filtered under reduced pressure to remove floating matter, and the filtrate was concentrated under reduced pressure. The residue was dissolved in acetone, and stored at a low temperature for 4 hours in a refrigerator. Thereafter, the resulting solution was filtered under reduced pressure to remove the re-formed floating matter, and used in the next reaction without any purification (crude yield: 5.5 g, 86%). TLC (EA:Hex=1:1); $R_f=0.5$.

Synthesis of Compound 25

2.0 g (11.5 mmol, 1.0 eq.) of L-citrulline was dissolved in 100 mL of a 1:1 mixed solution of tetrahydrofuran (THF) and water, and stirred. 988 mg of sodium hydrogen carbon-

ate was added thereto, and stirred. Thereafter, 5.0 g of Compound 21 (11.5 mmol, 1.0 eq.) was dissolved in 80 mL of acetone, added dropwise to the reaction solution, and then stirred. The resulting mixture was stirred for 21 hours, and then concentrated under reduced pressure to remove the organic solvent. An aqueous layer was washed with ethyl acetate (EA), and then titrated to pH 3 by slow dropwise addition of 2 N HCl. EA was added thereto so that the precipitate in an organic layer was extracted. Then, the organic layer was dried over a saturated saline solution and sodium sulfate, and used in the next reaction without any purification (crude yield: 5.6 g, 98%). TLC (DCM: MeOH=10:1, one drop of formic acid); $R_f=0.1$.

Synthesis of Compound 26

50 mL of 10% piperidine in N,N-dimethylformamide (DMF) was added dropwise to 2.84 g (5.72 mmol, 1.0 eq.) of Compound 25, and stirred. After 4 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the residue was dissolved in water, and filtered under reduced pressure to remove the formed floating matter. An aqueous layer was concentrated to obtain

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a target compound, which was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); $R_f=0.01$.

Synthesis of Compound 27

54 mg (0.22 mmol, 1.0 eq.) of Compound 26 was dissolved in 2 mL of DMF, and 0.05 mL (0.264 mmol, 1.2 eq.) of N,N-diisopropylethylamine (DIPEA) was added dropwise thereto. Compound 23 was completely dissolved by adding 2 mL of water, and 0.05 mL (0.528 mmol, 2.4 eq.) of an acetic anhydride was added dropwise thereto at room temperature. After 3 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the concentrate was purified by reversed-phase column chromatography to obtain a target compound. TLC (DCM:MeOH=10:1); $R_f=0.05$.

Synthesis of Compound 28

300 mg (0.95 mmol, 1.0 eq.) of Compound 27 was dissolved in 6 mL of DMF, and 550 mg (1.43 mmol, 1.5 eq.) of 1-[bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxide hexafluorophosphate (HATU) was then added thereto. Thereafter, 0.25 mL (1.43 mmol, 1.5 eq.) of DIPEA was added dropwise, and 230 mg (1.43 mmol, 1.5 eq.) of 1-[(tert-butoxycarbonyl)amino]-2-aminoethane was dissolved in 3.5 mL of DMF, and added dropwise at room temperature. After 7 hours, the reaction solution was diluted with ethyl acetate (EA), and an organic layer was washed with a saturated sodium hydrogen carbonate solution. Then,

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the organic layer was dried over a saturated saline solution and sodium sulfate, and filtered to obtain a target compound. The obtained target compound was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); $R_f=0.5$.

Synthesis of Compound 29

Compound 28 was dissolved in a 2:1 mixed solution of DCM and TFA, and a reaction was then carried out at 0° C. Thereafter, the reaction mixture was concentrated under reduced pressure to remove a reaction solution such as DCM, and this concentration was then repeated three times. The residue was dried under high vacuum, and purified by reversed-phase column chromatography to obtain 370 mg of a target compound. TLC (DCM:MeOH=10:1); $R_f=0.01$.

Synthesis of Compound DD4

0.03 mL (0.17 mmol, 1.2 eq.) of DIPEA was added dropwise to a solution obtained by dissolving 50 mg (0.14 mmol, 1.0 eq.) of Compound 26 in 3 mL of DMF, and 107 mg (0.21 mmol, 1.5 eq.) of phenol-activated SN38 was added thereto. The resulting mixture was stirred for an hour, and then concentrated under reduced pressure to remove a reaction solution. The concentrate was purified by column chromatography (10% MeOH in DCM) to obtain a target compound. TLC (DCM:MeOH=10:1); $R_f=0.1$.

Confirmation of Structure of VC Linker (DD4)

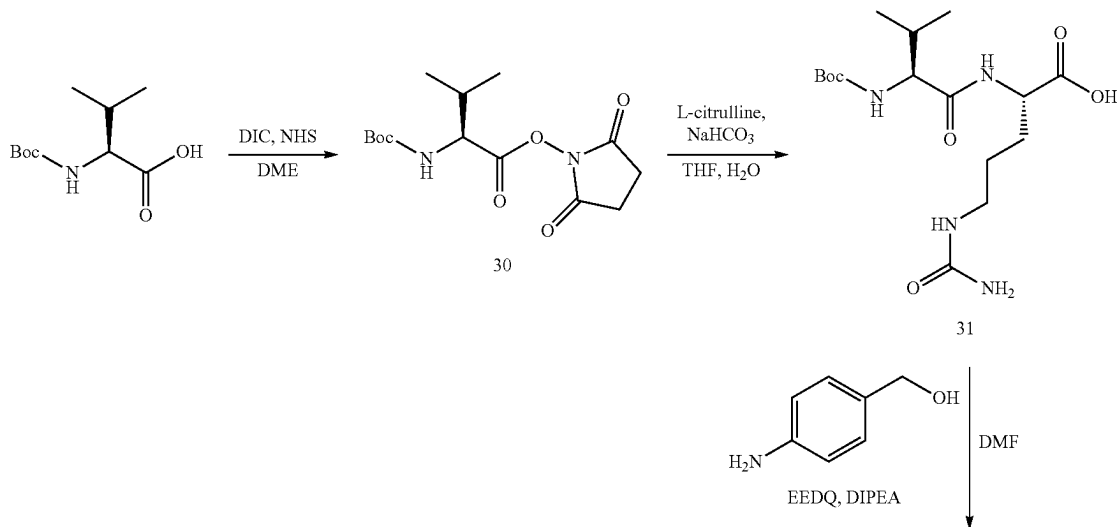
LRMS (ESI): m/z 777.3 $[M+H]^+$

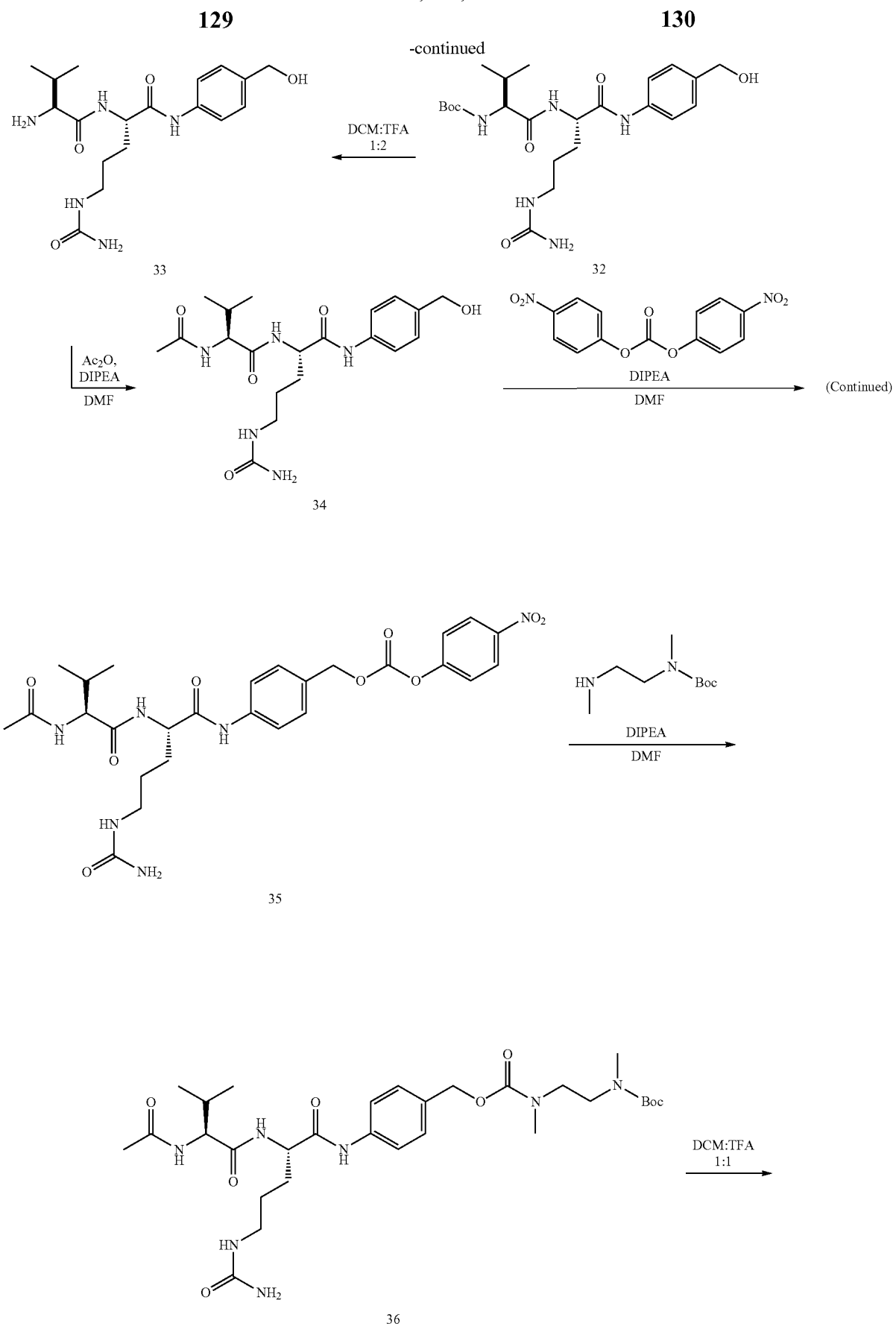
The results are shown in FIG. 30.

1.3.4. Synthesis and Confirmation of Structure of VC Linker (DD5)

[Scheme 13]

Synthesis of DD5

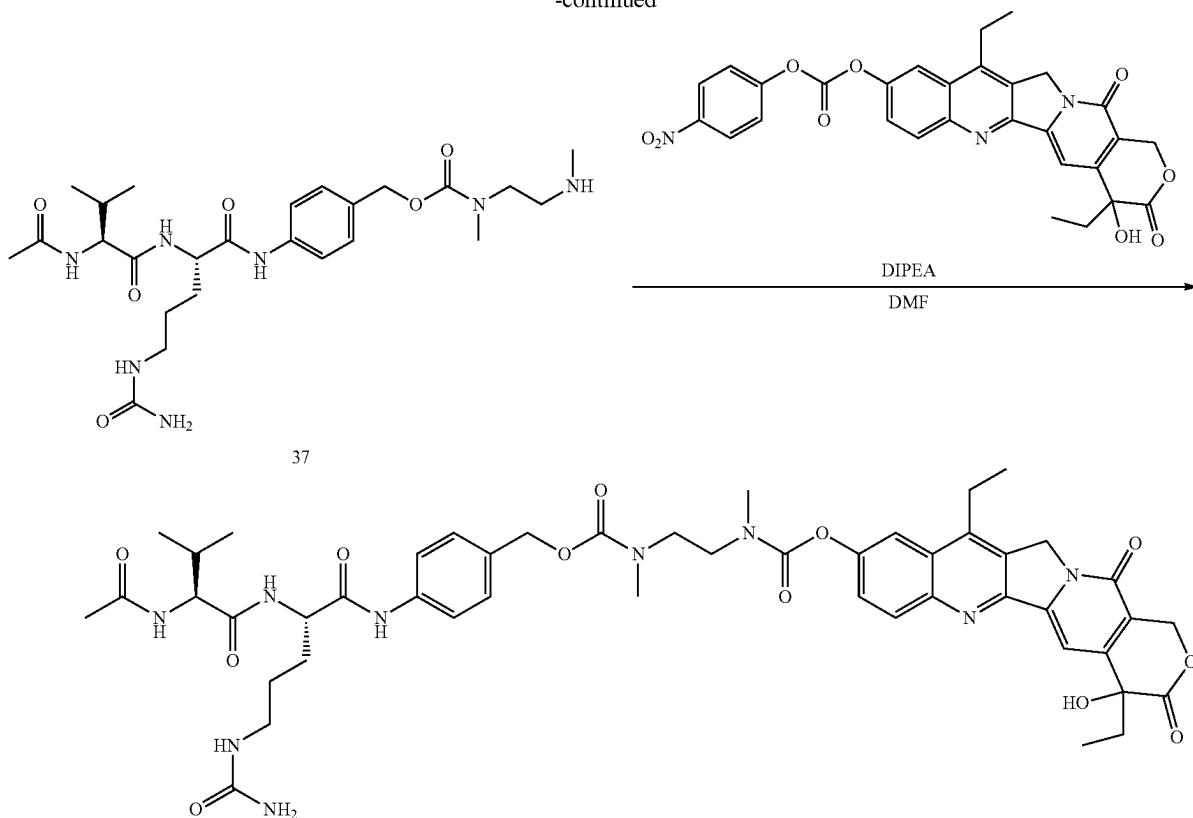




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-continued



DD5

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Synthesis of VC Linker (DD5)

Synthesis of Compound 30

5 g (14.7 mmol, 1.0 eq.) of Fmoc-Val-OH and 1.7 g (14.7 mmol, 1.0 eq.) of N-hydroxysuccinimide (NHS) were dissolved in 140 mL of dimethoxyethane (DME), and stirred. 2.5 mL (16.2 mmol, 1.1 eq.) of N,N'-diisopropylcarbodiimide (DIC) was added dropwise at 0° C., and stirred for 16 hours. The reaction solution was filtered under reduced pressure to remove floating matter, and the filtrate was concentrated under reduced pressure. The residue was dissolved in acetone, and stored at a low temperature for 4 hours in a refrigerator. Thereafter, the resulting solution was filtered under reduced pressure to remove the re-formed floating matter, and used in the next reaction without any purification (crude yield: 5.5 g, 86%). TLC (EA:Hex=1:1); $R_f=0.5$.

Synthesis of Compound 31

2.0 g (11.5 mmol, 1.0 eq.) of L-citrulline was dissolved in 100 mL of a 1:1 mixed solution of tetrahydrofuran (THF) and water, and stirred. 988 mg of sodium hydrogen carbonate was added thereto, and stirred. Thereafter, 5.0 g (11.5 mmol, 1.0 eq.) of Compound 30 was dissolved in 80 mL of acetone, added dropwise to the reaction solution, and then stirred. The resulting mixture was stirred for 21 hours, and then concentrated under reduced pressure to remove the organic solvent. An aqueous layer was washed with ethyl acetate (EA), and then titrated to pH 3 by slow dropwise

addition of 2 N HCl. EA was added thereto so that the precipitate in an organic layer was extracted. Then, the organic layer was dried over a saturated saline solution and sodium sulfate, and used in the next reaction without any purification (crude yield: 5.6 g, 98%). TLC (DCM: MeOH=10:1, one drop of formic acid); $R_f=0.1$.

Synthesis of Compound 32

50 mL of 10% piperidine in N,N-dimethylformamide (DMF) was added dropwise to 2.84 g (5.72 mmol, 1.0 eq.) of Compound 31, and stirred. After 4 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the residue was dissolved in water, and filtered under reduced pressure to remove the formed floating matter. An aqueous layer was concentrated to obtain a target compound, which was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); $R_f=0.01$.

Synthesis of Compound 33

54 mg (0.22 mmol, 1.0 eq.) of Compound 29 was dissolved in 2 mL of DMF, and 0.05 mL (0.264 mmol, 1.2 eq.) of N,N-diisopropylethylamine (DIPEA) was added dropwise thereto. Compound 32 was completely dissolved by adding 2 mL of water, and 0.05 mL (0.528 mmol, 2.4 eq.) of an acetic anhydride was added dropwise thereto at room temperature. After 3 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the concentrate was purified by reversed-phase

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column chromatography to obtain a target compound. TLC (DCM:MeOH=10:1); R_f =0.05.

Synthesis of Compound 34

1 g (3.16 mmol, 1.0 eq.) of Compound 33 was dissolved in 30 mL of a 2:1 mixed solution of DCM and methanol, and 868 mg (3.48 mmol, 1.1 eq.) of N-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ) was added thereto, and stirred. 451 mg (3.66 mmol, 1.16 eq.) of 4-aminobenzyl alcohol was added thereto, and stirred for 5 hours. The resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the concentrate was purified by column chromatography (10% MeOH in DCM) to obtain 419 mg of a target compound (yield: 32%). TLC (DCM:MeOH=10:1); R_f =0.1.

Synthesis of Compound 35

240 mg (0.55 mmol, 1.0 eq.) of Compound 34 was dissolved in 10 mL of DMF, and 0.3 mL (1.65 mmol, 3.0 eq.) of DIPEA was added dropwise thereto, and stirred. Thereafter, a reaction was carried out by adding 509 mg (1.65 mmol, 3.0 eq.) of bis-(4-aminophenyl) carbonate. After 3 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and diethyl ether was added thereto to precipitate a target compound, which was filtered under reduced pressure. The obtained target compound was used in the next reaction without any purification.

Synthesis of Compound 36

33 mg (0.06 mmol, 1.0 eq.) of Compound 35 was dissolved in 5 mL of DMF, and 16 mg (0.08 mmol, 1.3 eq.) of N-[(tert-butoxy)carbonyl]-N,N'-dimethylethylenediamine and 0.02 mL (0.08 mmol, 1.3 eq.) of DIPEA were added dropwise thereto, and stirred. The resulting mixture was stirred for 7 hours, and then concentrated under reduced

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pressure to remove a reaction solution, and diethyl ether was added thereto to precipitate a target compound, which was filtered under reduced pressure. The obtained target compound was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); R_f =0.1.

Synthesis of Compound 37

17 mg (0.03 mmol, 1.0 eq.) of Compound 36 was dissolved in 1 mL of DCM, and 1 mL of trifluoroacetic acid (TFA) was then added dropwise thereto at 0° C. A reaction was carried out by stirring the resulting mixture at 0° C. for 30 minutes, followed by stirring at room temperature for 30 minutes. Thereafter, the mixture was concentrated under reduced pressure to remove a reaction solution such as DCM, and this concentration was then repeated three times. The concentrated matter was dried under high vacuum, and purified by reversed-phase column chromatography to obtain 13 mg of a target compound. TLC (DCM:MeOH=10:1); R_f =0.01.

Synthesis of Compound DD5

0.01 mL (0.04 mmol, 1.5 eq.) of DIPEA was added dropwise to a solution obtained by dissolving 13 mg (0.03 mmol, 1.0 eq.) of Compound 37 in 3 mL of DMF, and 22 mg (0.04 mmol, 1.5 eq.) of phenol-activated SN38 was added thereto. The resulting mixture was stirred for an hour, and then concentrated under reduced pressure to remove a reaction solution. The concentrate was purified by column chromatography (10% MeOH in DCM) to obtain a target compound. TLC (DCM:MeOH=10:1); R_f =0.2.

3-4-2: Confirmation of Structure of VC Linker (DD5)

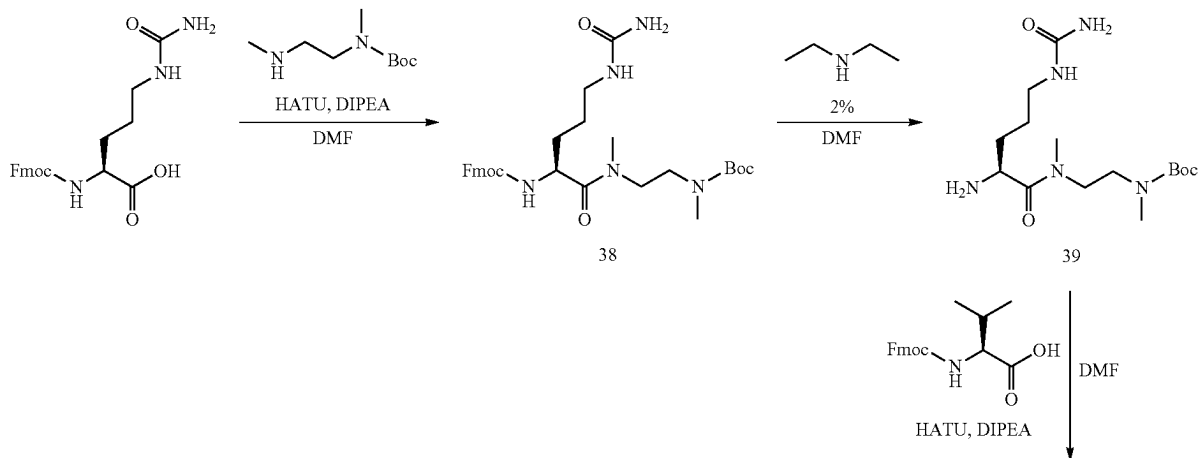
LRMS (ESI): m/z 954.4 $[M+H]^+$

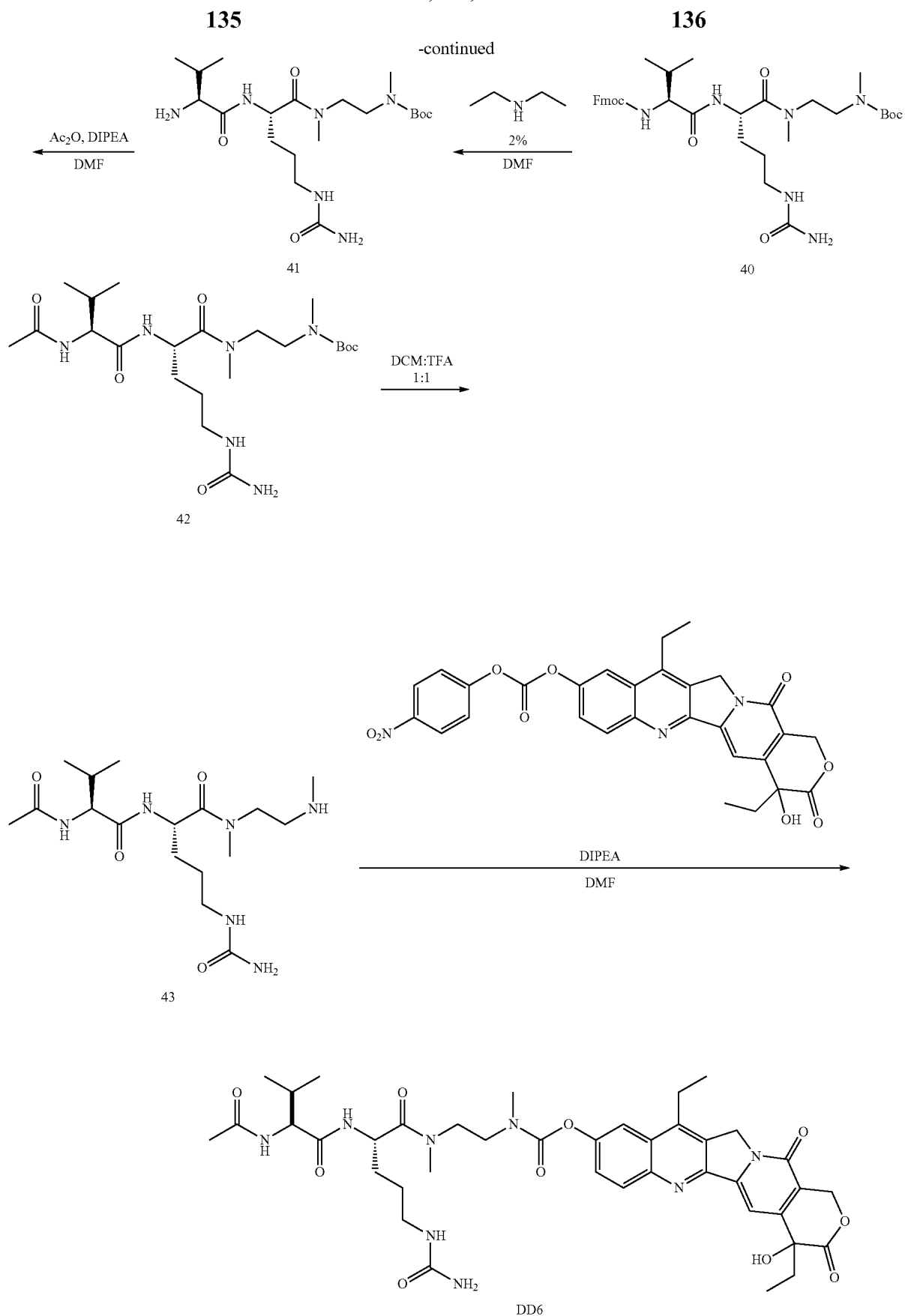
The results are shown in FIG. 31.

1.3.5. Synthesis and Confirmation of Structure of VC Linker (DD6)

[Scheme 14]

Synthesis of DD6





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Synthesis of VC Linker (DD6)

Synthesis of Compound 38

5 g (14.7 mmol, 1.0 eq.) of Fmoc-Val-OH and 1.7 g (14.7 mmol, 1.0 eq.) of N-hydroxysuccinimide (NHS) were dissolved in 140 mL of dimethoxyethane (DME), and stirred. 2.5 mL (16.2 mmol, 1.1 eq.) of N,N'-diisopropylcarbodiimide (DIC) was added dropwise at 0° C., and stirred for 16 hours. The reaction solution was filtered under reduced pressure to remove floating matter, and the filtrate was concentrated under reduced pressure. The residue was dissolved in acetone, and stored at a low temperature for 4 hours in a refrigerator. Thereafter, the resulting solution was filtered under reduced pressure to remove the re-formed floating matter, and used in the next reaction without any purification (crude yield: 5.5 g, 86%). TLC (EA:Hex=1:1); R_f =0.5.

Synthesis of Compound 39

2.0 g (11.5 mmol, 1.0 eq.) of L-citrulline was dissolved in 100 mL of a 1:1 mixed solution of tetrahydrofuran (THF) and water, and stirred. 988 mg of sodium hydrogen carbonate was added thereto, and stirred. Thereafter, 5.0 g (11.5 mmol, 1.0 eq.) of Compound 38 was dissolved in 80 mL of acetone, added dropwise to the reaction solution, and then stirred. The resulting mixture was stirred for 21 hours, and then concentrated under reduced pressure to remove the organic solvent. An aqueous layer was washed with ethyl acetate (EA), and then titrated to pH 3 by slow dropwise addition of 2 N HCl. EA was added thereto so that the precipitate in an organic layer was extracted. Then, the organic layer was dried over a saturated saline solution and sodium sulfate, and used in the next reaction without any purification (crude yield: 5.6 g, 98%). TLC (DCM:MeOH=10:1, one drop of formic acid); R_f =0.1.

Synthesis of Compound 40

50 mL of 10% piperidine in N,N-dimethylformamide (DMF) was added dropwise to 2.84 g (5.72 mmol, 1.0 eq.) of Compound 39, and stirred. After 4 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the residue was dissolved in water, and filtered under reduced pressure to remove the formed floating matter. An aqueous layer was concentrated to obtain a target compound, which was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); R_f =0.01.

Synthesis of Compound 41

54 mg (0.22 mmol, 1.0 eq.) of Compound 40 was dissolved in 2 mL of DMF, and 0.05 mL (0.264 mmol, 1.2 eq.) of N,N-diisopropylethylamine (DIPEA) was added drop-

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wise thereto. Compound 37 was completely dissolved by adding 2 mL of water, and 0.05 mL (0.528 mmol, 2.4 eq.) of an acetic anhydride was added dropwise thereto at room temperature. After 3 hours, the resulting mixture was concentrated under reduced pressure to remove a reaction solution, and the concentrate was purified by reversed-phase column chromatography to obtain a target compound. TLC (DCM:MeOH=10:1); R_f =0.05.

Synthesis of Compound 42

300 mg (0.95 mmol, 1.0 eq.) of Compound 41 was dissolved in 6 mL of DMF, and 550 mg (1.43 mmol, 1.5 eq.) of 1-[bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxide hexafluorophosphate (HATU) was then added thereto. Thereafter, 0.25 mL (1.43 mmol, 1.5 eq.) of DIPEA was added dropwise, and 230 mg (1.43 mmol, 1.5 eq.) of N-[(tert-butoxy)carbonyl]-N,N'-dimethylethylenediamine was dissolved in 3.5 mL of DMF, and added dropwise at room temperature. After 7 hours, the reaction solution was diluted with ethyl acetate (EA), and an organic layer was washed with a saturated sodium hydrogen carbonate solution. Then, the organic layer was dried over a saturated saline solution and sodium sulfate, and filtered to obtain a target compound. The obtained target compound was used in the next reaction without any purification. TLC (DCM:MeOH=10:1); R_f =0.5.

Synthesis of Compound 43

Compound 42 was dissolved in a 2:1 mixed solution of DCM and TFA, and a reaction was then carried out at 0° C. Thereafter, the reaction mixture was concentrated under reduced pressure to remove a reaction solution such as DCM, and this concentration was then repeated three times. The residue was dried under high vacuum, and purified by reversed-phase column chromatography to obtain 370 mg of a target compound. TLC (DCM:MeOH=10:1); R_f =0.01.

Synthesis of Compound DD6

0.03 mL (0.17 mmol, 1.2 eq.) of DIPEA was added dropwise to a solution obtained by dissolving 50 mg (0.14 mmol, 1.0 eq.) of Compound 43 in 3 mL of DMF, and 107 mg (0.21 mmol, 1.5 eq.) of phenol-activated SN38 was added thereto. The resulting mixture was stirred for an hour, and then concentrated under reduced pressure to remove a reaction solution. The concentrate was purified by column chromatography (10% MeOH in DCM) to obtain a target compound. TLC (DCM:MeOH=10:1); R_f =0.1.

Confirmation of Structure of VC Linker (DD6)

LRMS (ESI): m/z 805.5 [M+H⁺]

The results are shown in FIG. 32.

2. Preparation of Agent for Transferring Site-Specific First Click-Chemistry Functional Group to Antibody (H₁-L₂-SSFI)

2.1. Synthesis and Confirmation of Structure of Compound I-SSFI (where Xa₁ is Lysine)



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Synthesis Method of Compound I-SSFI (Lys)

Synthesis of Compound I-SSFI was carried out in DMF, and 3 equivalents of DIPEA and 8.4 μmol of Compound I were dissolved in 7.3 μmol of SSFI, and stirred to introduce Compound I into SSFI.

After the termination of the reaction was confirmed, the reaction solution was concentrated, and purification of Compound I-SSFI by preparative-HPLC was attempted. After the purification, the Compound I-SSFI was freeze-dried to obtain a target compound, the amount of which was confirmed to be 12 mg (purity: >95% up (H PLC), and yield: 71%).

Confirmation of Structure of Compound I-SSFI

Measuring equipment: Waters Quattro Premier Xe

Calculated molecular weight: 2246.99 g/mol

Measured molecular weight ($M/2+H$)²⁺: 1124.90 g/mol

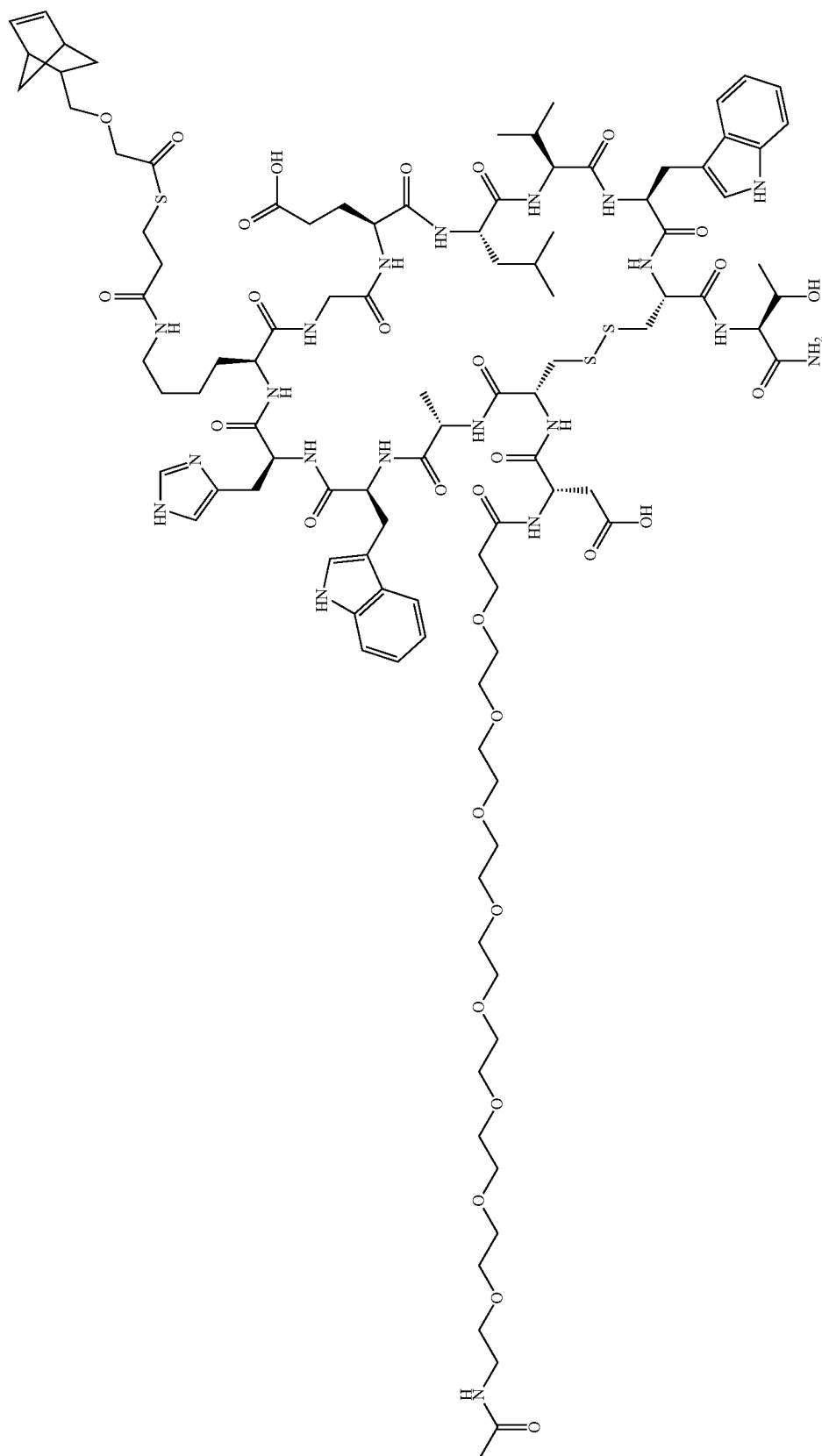
The results are shown in FIG. 33.

2.2. Synthesis and Confirmation of Structure of Compound II-SSFI (Wherein Xa_1 is Lysine)

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Exact Mass: 2261.01
II-FcBP(61,ys)

145

Synthesis Method of Compound II-SSFI (Lys)

Synthesis of Compound II-SSFI was carried out in DMF, and 3 equivalents of DIPEA and 8.4 μmol of Compound II were dissolved in 7.3 μmol of SSFI, and stirred to introduce Compound II into SSFI.

After the termination of the reaction was confirmed, the reaction solution was concentrated, and purification of Compound II-SSFI by preparative-HPLC was attempted. After the purification, the Compound II-SSFI was freeze-dried to obtain a target compound, the amount of which was confirmed to be 13 mg (purity: >95% up (HPLC), and yield: 78%).

Confirmation of Structure of Compound II-SSFI (Lys)

Measuring equipment: Ultraflexxtreme (Bruker)

Measuring matrix: CHCA (α -Cyano-4-hydroxycinnamic acid) & DHB (2,5-Dihydroxybenzoic acid)

Calculated molecular weight: 2261.01 g/mol

Measured molecular weight (M/2+H)²⁺: 2263.26 g/mol

The results are shown in FIG. 34.

2.3. Synthesis and Confirmation of Structure of Compound III-SSFI (Wherein Xa₁ is Lysine) 20

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Synthesis Method of Compound III-SSFI (Lys)

Synthesis of Compound III-SSFI was carried out in DMF, and 3 equivalents of DIPEA and 8.4 μmol of Compound III were dissolved in 7.3 μmol of SSFI, and stirred to introduce Compound III into SSFI.

After the termination of the reaction was confirmed, the reaction solution was concentrated, and purification of Compound III-SSFI by preparative-H PLC was attempted. After the purification, the Compound III-SSFI was freeze-dried to obtain a target compound, the amount of which was confirmed to be 12 mg (purity: >95% up (HPLC), and yield: 75%).

Confirmation of Structure of Compound III-SSFI (Lys)

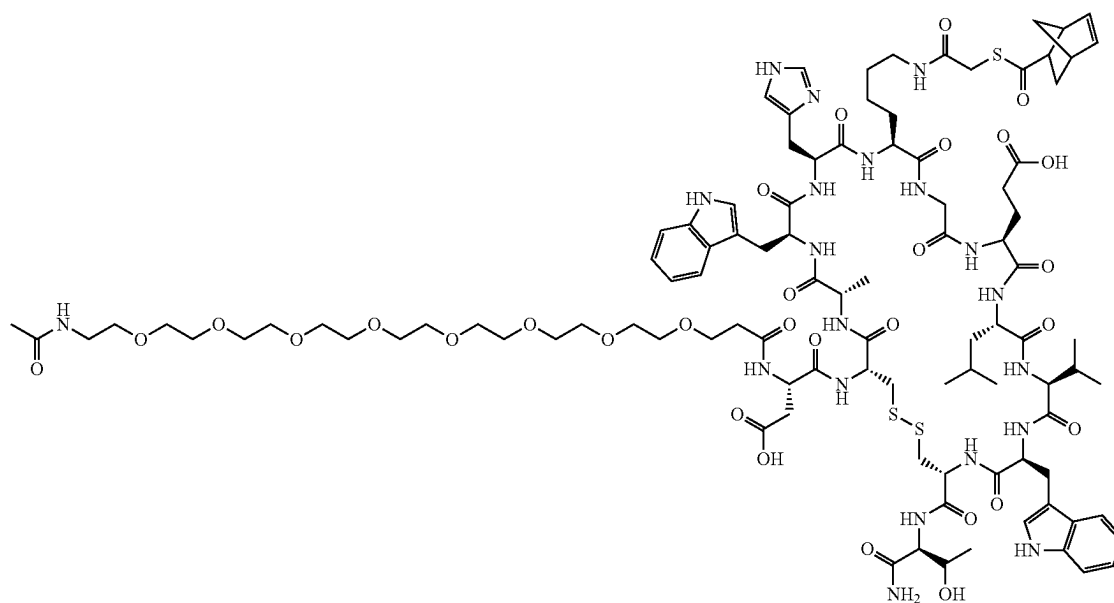
Measuring equipment: Waters Quattro Premier Xe

Calculated molecular weight: 2202.97 g/mol

Measured molecular weight $(M/2+H)^{2+}$: 1103.64 g/mol

The results are shown in FIG. 35.

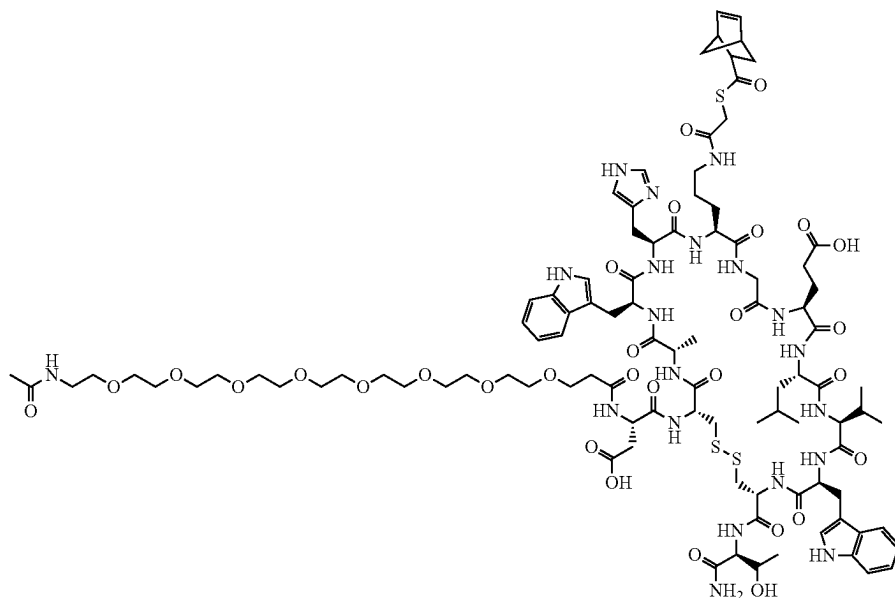
2.4. Synthesis and Confirmation of Structure of Compound III-SSFI (Wherein Xa₁ is Ornithine)



Exact Mass: 2202.97
III-FcBP(6Lys)

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Exact Mass: 2188.95

Synthesis Method of Compound III-SSFI (Orn)

Synthesis of Compound III-SSFI was carried out in DMF, and 3 equivalents of DIPEA and 8.4 μmol of Compound III were dissolved in 7.3 μmol of SSFI, and stirred to introduce Compound III into SSFI.

After the termination of the reaction was confirmed, the reaction solution was concentrated, and purification of Compound III-SSFI by preparative-HPLC was attempted. After the purification, the Compound III-SSFI was freeze-dried to obtain a target compound, the amount of which was

confirmed to be 14 mg (purity: >95% up (HPLC), and yield: 87%).

Confirmation of Structure of Compound III-SSFI (Orn)

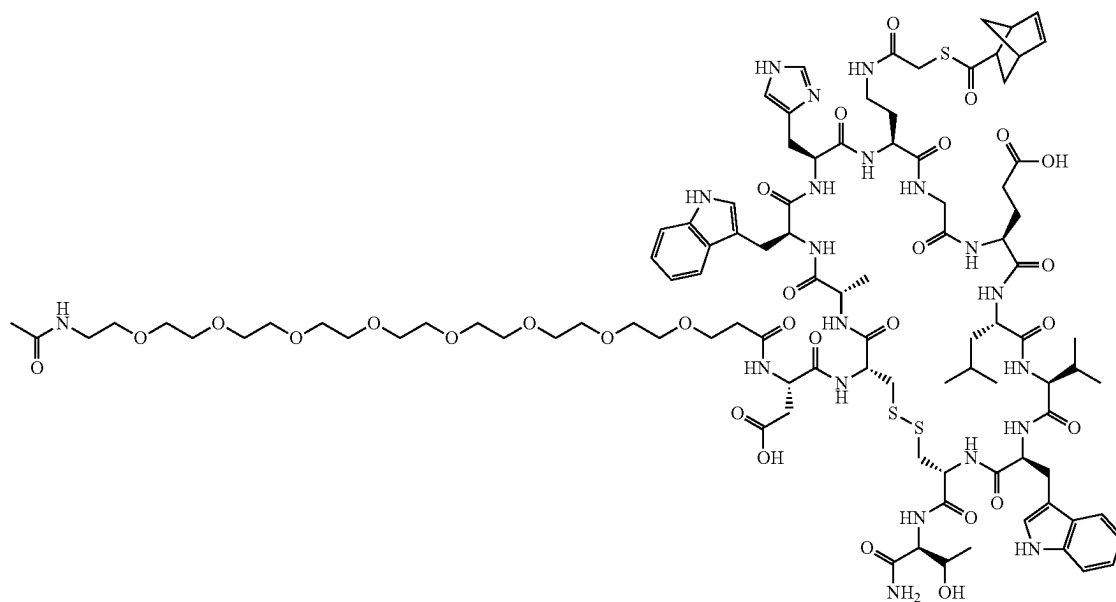
Measuring equipment: Waters Quattro Premier Xe

Calculated molecular weight: 2188.97 g/mol

Measured molecular weight ($M/2+H$)²⁺: 1096.64 g/mol

The results are shown in FIG. 36.

2.5. Synthesis and Confirmation of Structure of Compound III-SSFI (Wherein X_{a1} is 2,4-diaminobutanoic acid (Dab))



Exact Mass: 2174.94
III-FcBP(6Dab)

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Synthesis Method of Compound III-SSFI (Dab)

Synthesis of Compound III-SSFI was carried out in DMF, and 3 equivalents of DIPEA and 8.4 μmol of Compound III were dissolved in 7.3 μmol of SSFI, and stirred to introduce Compound III into SSFI.

After the termination of the reaction was confirmed, the reaction solution was concentrated, and purification of Compound III-SSFI by preparative-HPLC was attempted. After the purification, the Compound III-SSFI was freeze-dried to obtain a target compound, the amount of which was confirmed to be 12 mg (purity: >95% up (HPLC), and yield: 75%).

Confirmation of Structure of Compound III-SSFI (Dab)

Measuring equipment: Waters Quattro Premier Xe

Calculated molecular weight: 2174.94 g/mol

Measured molecular weight ($M/2+H$)²⁺: 1089.94 g/mol

The results are shown in FIG. 37.

2.6. Synthesis and Confirmation of Structure of Compound III-SSFI (Wherein Xa₁ is 2,3-diaminopropionic acid (Dap))

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Synthesis Method of Compound III-SSFI (Dap)

Synthesis of Compound III-SSFI was carried out in DMF, and 3 equivalents of DIPEA and 8.4 μmol of Compound III were dissolved in 7.3 μmol of SSFI, and stirred to introduce Compound III into SSFI.

After the termination of the reaction was confirmed, the reaction solution was concentrated, and purification of Compound III-SSFI by preparative-HPLC was attempted. After the purification, the Compound III-SSFI was freeze-dried to obtain a target compound, the amount of which was confirmed to be 12 mg (purity: >95% up (HPLC), and yield: 75%).

Confirmation of Structure of Compound III-SSFI (Dap)

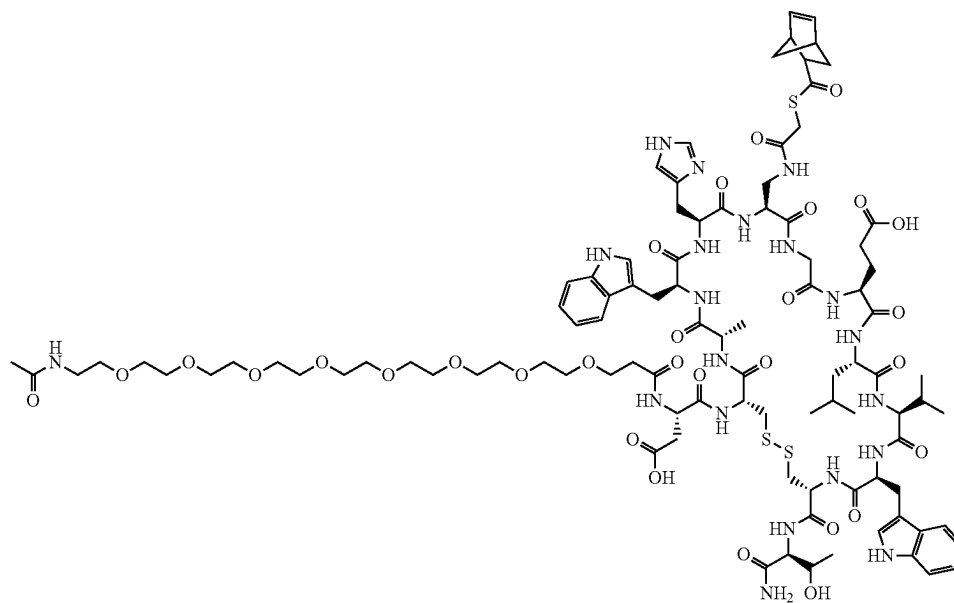
Measuring equipment: Waters Quattro Premier Xe

Calculated molecular weight: 2160.92 g/mol

Measured molecular weight ($M/2+H$)²⁺: 1082.34 g/mol

The results are shown in FIG. 38.

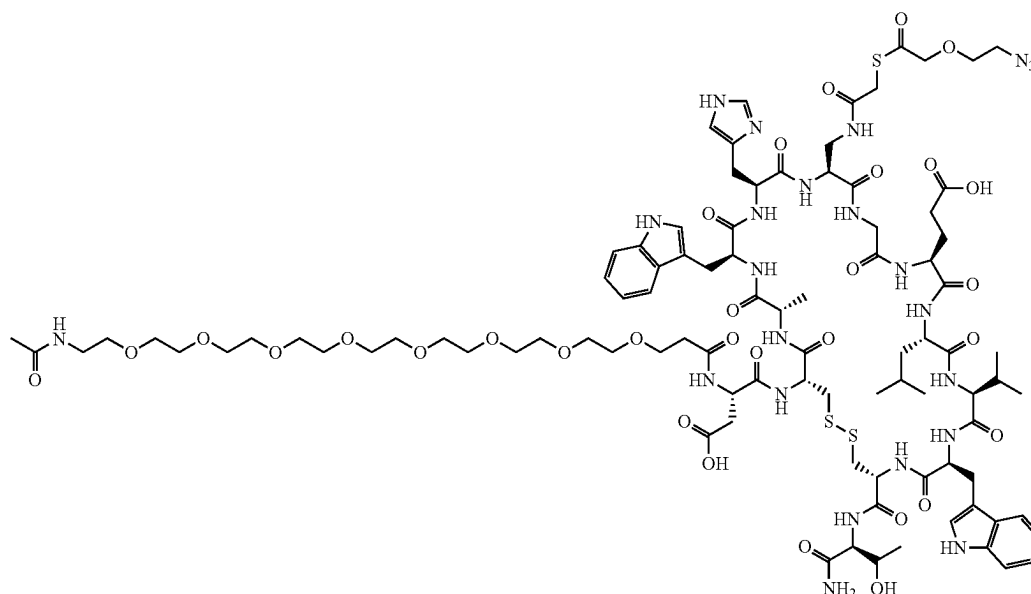
2.7. Synthesis and Confirmation of Structure of Compound IV-SSFI (Wherein Xa₁ is Dap)



Exact Mass: 2160.92
III-FcBP(6Dap)

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Exact Mass: 2167.90
IV-FcBP(6Dap)

Synthesis Method of Compound IV-SSFI (Dap)

Synthesis of Compound IV-SSFI was carried out in DMF, and 3 equivalents of DIPEA and 8.4 μmol of Compound IV were dissolved in 7.3 μmol of SSFI, and stirred to introduce Compound IV into SSFI.

After the termination of the reaction was confirmed, the reaction solution was concentrated, and purification of Compound IV-SSFI by preparative-HPLC was attempted. After the purification, the Compound IV-SSFI was freeze-dried to obtain a target compound, the amount of which was confirmed to be 12 mg (purity: >95% up (HPLC), and yield: 76%).

Confirmation of Structure of Compound IV-SSFI (Dap)

Measuring equipment: Waters Quattro Premier Xe

Calculated molecular weight: 2167.90 g/mol

Measured molecular weight ($M/2+H$)²⁺: 1085.60 g/mol

The results are shown in FIG. 39.

3. Synthesis and Confirmation of Structure of Antibody Containing First Click-Chemistry Functional Group in Site-Specific Manner

3.1. Synthesis of Antibody-Norbornene

Synthesis method of Trastuzumab-Norbornene (1)

Introduction Reaction Using Compound I-SSFI (Lys)

Synthesis of Ab (246/248 Lys)-norbornene was carried out in a buffer using Compound I-SSFI (Lys). A 1×PBS buffer (0.01% Tween 20, pH 7.4) was used as the reaction buffer. To introduce norbornene at a certain site of an antibody (trastuzumab, 4 mg/mL), 8 equivalents of Compound I-SSFI (Lys) was added per one antibody, and reacted at room temperature for 72 hours, and the reaction monitoring and termination were confirmed by HIC-HPLC. As the binding of norbornene to the antibody proceeds, the peaks on the chromatogram were observed to shift from 4 minutes to 4.5 minutes and 5 minutes, thereby confirming a degree of progression of the reaction. The Ab-norbornene conjugate was dialyzed three times using a 1×PBS buffer (pH 7.4), and purified using a size exclusion chromatography technique.

Observation of the binding reaction between trastuzumab and Compound I-SSFI (Lys) according to reaction time is shown in FIG. 40.

Synthesis Method of Trastuzumab-Norbornene (2)

Introduction Reaction Using Compound II-SSFI (Lys)

Synthesis of Ab (246/248 Lys)-norbornene was carried out in a buffer using Compound II-SSFI (Lys). A 1×PBS buffer (0.01% Tween 20, pH 7.4) was used as the reaction buffer. To introduce norbornene at a certain site of an antibody (trastuzumab, 4 mg/mL), 8 equivalents of Compound II-SSFI (Lys) was added per one antibody, and reacted at room temperature for 72 hours, and the reaction monitoring and termination were confirmed by HIC-HPLC. As the binding of norbornene to the antibody proceeds, the peaks on the chromatogram were observed to shift from 4 minutes to 4.5 minutes, thereby confirming a degree of progression of the reaction. The Ab-norbornene conjugate was dialyzed three times using a 1×PBS buffer (pH 7.4), and purified using a size exclusion chromatography technique.

Observation of the binding reaction between trastuzumab and Compound II-SSFI (Lys) according to reaction time is shown in FIG. 41.

Synthesis Method of Trastuzumab-Norbornene (3)

Introduction Reaction Using Compound III-SSFI (Dap, Dab, Orn, or Lys)

Synthesis of Ab (246/248 Lys)-norbornene was carried out in a buffer using Compound III-SSFI (Lys). A 1×PBS buffer (0.01% Tween 20, pH 7.4) was used as the reaction buffer. To introduce norbornene at a certain site of an antibody (Herceptin or trastuzumab, 4 mg/mL), 10 equivalents of Compound III-SSFI (Dap, Dab, Orn, or Lys) was added per one antibody, and reacted at room temperature for 28 hours, and the reaction monitoring and termination were confirmed by HIC-HPLC. As the binding of norbornene to the antibody proceeds, the peaks on the chromatogram were observed to shift from 6.2 minutes to 6.4 to 6.7 minutes, thereby confirming a degree of progression of the reaction. The Ab-norbornene conjugate was dialyzed using a 1×PBS buffer (pH 7.4), and purified using a size exclusion chromatography technique.

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The reaction between Compound III-SSFI (Dap, Dab, Orn, or Lys) and the antibody is shown in FIG. 42, and the structure of the final product (i.e., Ab(246/248)-Norbornene) is shown in FIG. 43.

Observation of the binding reaction between trastuzumab and Compound III-SSFI (Dap) according to reaction time is shown in FIG. 44.

Observation of the binding reaction between trastuzumab and Compound III-SSFI (Dab) according to reaction time is shown in FIG. 45.

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−0.10 was considered to be a system error in the high-mass molecular weight analysis). The observed mass spectrum of the Ides_F(ab')₂ was detected, but the mass value of one charge was not shifted due to the complexity of the respective mass spectra, thereby making impossible to obtain the corresponding mass value.

The spectra of the complex having an increased molecular weight due to the antibody-norbornene binding are shown in FIGS. 49 and 50, and changes in molecular weights due to the norbornene conjugation are as listed in Table 12.

TABLE 12

Changes in molecular weights of antibody-norbornene conjugates				
Sample		Theoretical mass (peptide full seq.)	Observed mass	Δ mass (Obs-Theo) (continued-)
Trastuzumab	Ides Fc/2	23916.08	25231.10	1915.02
Trastuzumab-norbornene conjugate	Ides Fc/2	23916.08	25353.53	1437.45
(-continued) N-glycan (G0F/G0F)	C-term (−2K)	Norbornene conjugate	Δ mass	
1445.00	128	ND	−1.98	(G0F/−K)
1445.00	128	122	−1.55	(G0F/−K)

Observation of the binding reaction between trastuzumab and Compound III-SSFI (Orn) according to reaction time is shown in FIG. 46.

Observation of the binding reaction between trastuzumab and Compound III-SSFI (Lys) according to reaction time is shown in FIG. 47.

3.2. Synthesis of Antibody-Azide

Synthesis method of Trastuzumab-Azide (4)

Introduction Reaction Using Compound IV-SSFI (Dap)

Synthesis of Ab (246/248 Lys)-azide was carried out in a buffer using Compound IV-SSFI (Dap). A 1×PBS buffer (0.01% Tween 20, pH 7.4) was used as the reaction buffer. To introduce an azide at a certain site of an antibody (trastuzumab, 4 mg/mL), 10 equivalents of Compound IV-SSFI (Dap) was added per one antibody, and reacted at room temperature for 24 hours, and the reaction monitoring and termination were confirmed by HIC-HPLC. As the binding of norbornene to the antibody proceeds, the peaks on the chromatogram were observed to shift from 6 minutes to 6.3 minutes, thereby confirming a degree of progression of the reaction. The Ab-norbornene conjugate was dialyzed three times using a 1×PBS buffer (pH 7.4), and purified using a size exclusion chromatography technique.

Observation of the binding reaction between trastuzumab and Compound IV-SSFI (Dap) according to reaction time is shown in FIG. 48.

3.3. Confirmation of Structure and Site-Specific Binding of Trastuzumab-Norbornene Conjugate

An antibody-norbornene complex in which a norbornene linker was bound to an antibody via Compound III-SSFI (Dap) was confirmed by mass spectrometry. It was determined to which site of the antibody the norbornene was bound via F(ab')₂ and Fc/2 after the antibody-norbornene complex was treated with an IdeS enzyme. It was confirmed that the Ides Fc/2-norbornene complex was found to have an "Obs-Theo" value of 2991.68, a value which is an exact match in consideration of the combined value of the mass value of N-glycan, the value of a loss of one lysine residue, and the value of one norbornene molecule, which corresponded to the molecular weight of 120 Da, (a value of

Also, the norbornene binding site of the anti body-norbornene complex was confirmed through comparison of the MS/MS spectrum with trastuzumab. As a result, it was confirmed that the norbornene molecule was bound to a K248 site of the sequence LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKF (SEQ ID NO: 26). The MS/MS chromatograms are shown FIGS. 51 to 54, and the sequence matching results based on the MS/MS spectra are shown in FIG. 55.

The conditions for the analysis are as follows:

UPLC conditions

Measuring equipment: Acquity UPLC I-Class system

1. Column: Thermo MAbPac™ RP (2.1 mm×5 mm)

2. Column Temp.: 60° C.

3. Mobile phase

A. 0.1% Formic acid in water

B. 0.1% Formic acid in acetonitrile

Mass spectroscopy conditions

Measuring equipment: LTQ Elite (Thermo)

Source type: HESI

Capillary Temp.: 320° C.

Source heater temp.: 300° C.

Sheath gas flow: 40.00

Aux gas flow: 20.00

Sweep gas flow: 5.00

Source voltage (KV): 4.00

FTMS resolution: 120,000

FTMS mass range: 400 to 4,000

3.4. Confirmation of Structure of Trastuzumab-Azide

The antibody-azide complex in which an azide linker was bound to an antibody via Compound IV-SSFI (Dap) was confirmed using mass spectrometry. A molecular weight increase of 249 Da was confirmed when two molecules of an azide compound were bound to trastuzumab. The measured mass spectrum of trastuzumab is shown in FIG. 56, and the mass spectrum of the antibody-azide complex to which two azide molecules were bound is shown in FIG. 57.

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Measuring equipment: Ultraflex III (TOF/TOF)
 Analysis mode: Linear mode
 Polarity: Positive
 Detection: m/z 2,000 to 300,000
 Laser repetition rate: 100 Hz
 Number of shots: 1,000 shots
 Deflection: On, 5,000 Da

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Voltage: Ion Source kV, Ion Source II 23.00 kV, Lens 9.00 kV

Calculated molecular weight: 148,271 g/mol

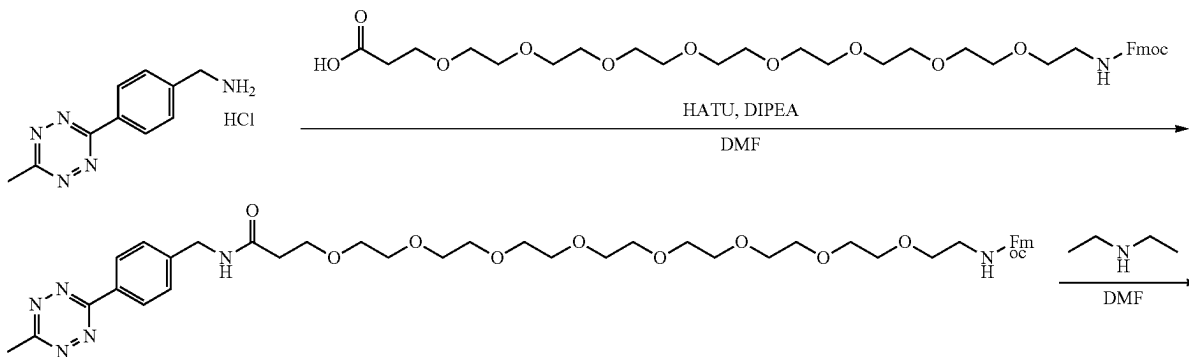
Measured molecular weight: 148,262 g/mol

5 4. Synthesis and Confirmation of Structure of Antibody-Payload Conjugate

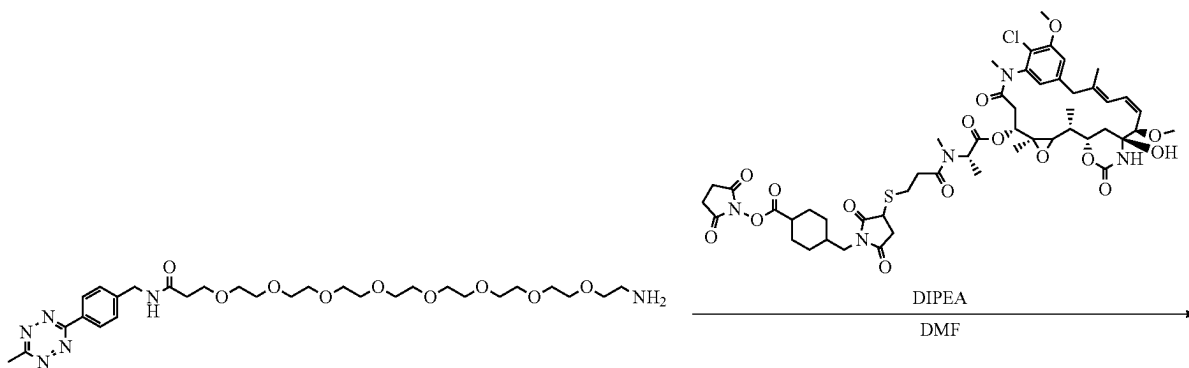
4.1. Synthesis Method and Confirmation of Structure of Payload

[Scheme 15]

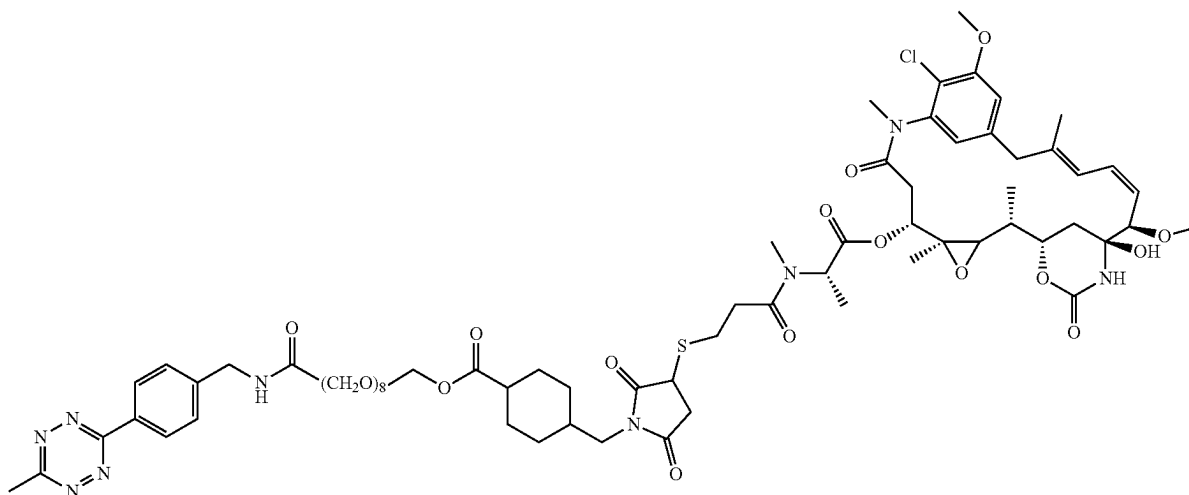
Synthesis of Tetrazine-DM1



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42



Tetrazine-DM1

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Synthesis Method of Tetrazine-DM1

Synthesis of Compound 41

0.53 g (0.8 mmol, 1.0 eq.) of Fmoc-PEG₈-OH was dissolved in 7 mL of DMF, and then stirred. Thereafter, 0.38 g (1.0 mmol, 1.25 eq.) of HATU and 0.3 mL (1.7 mmol, 2.1 eq.) of DIPEA were added dropwise thereto. 0.205 g (0.86 mmol, 1.1 eq.) of methyltetrazine-amine-HCl was added dropwise, and 0.3 mL (1.7 mmol, 2.1 eq.) of DIPEA was added dropwise thereto. The resulting mixture was stirred for 4 hours, and then concentrated under reduced pressure to remove the solvent. The mixture was purified by column chromatography (5% MeOH in DCM) to obtain a target compound. TLC (DCM:MeOH=20:1); R_f=0.3.

Synthesis of Compound 42

0.527 g (0.62 mmol, 1.0 eq.) of Compound 41 was dissolved in 6 mL of DMF, and 0.3 mL of diethylamine was added dropwise thereto. The resulting mixture was stirred for 2.5 hours, and then concentrated under reduced pressure to remove the solvent. The mixture was purified by column chromatography (DCM:MeOH:NH₄OH=80:25:2.5) to obtain a target compound, the amount of which was confirmed to be 0.273 g (yield: 70%). TLC (DCM:MeOH:NH₄OH=80:25:2.5); R_f=0.1.

Synthesis of Tetrazine-DM1

0.355 g (0.57 mmol, 1.5 eq.) of Compound 42 was dissolved in 25 mL of DMF, and then stirred. 0.405 g (0.38 mmol, 1.0 eq.) of DM1-SMCC-NHS was added thereto, and DIPEA was added dropwise until the pH of the mixture reached pH 9.0. Thereafter, the mixture was stirred for 2 hours. The reaction solution was concentrated under reduced pressure, and then purified by column chromatography (10% MeOH in DCM) to obtain a target compound, the amount of which was confirmed to be 0.434 g (yield: 73%). TLC (DCM:MeOH=10:1); R_f=0.5.

Confirmation of Structure of Tetrazine-DM1

LRMS (ESI): m/z 782.6 [M+2H⁺]

The results are shown in FIG. 58.

4.2. Synthesis and Confirmation of Structure of Antibody-Payload Conjugate

4.2.1. Antibody-Norbornene-Tetrazine-DM1

Synthesis Method of Trastuzumab-Norbornene-Tetrazine-DM1 (1)

Introduction Reaction of Tetrazine-DM1 into Trastuzumab-Norbornene Using Compound III-SSFI (Dap)

An antibody-payload conjugate was constructed using trastuzumab into which two norbornene molecules were introduced via Compound III-SSFI (Dap). A reaction was carried out using 25 mL of the antibody-payload conjugate at a concentration of 4.5 mg/mL, and conjugation of a tetrazine-PEG8-DM1 drug was attempted for a biorthogonal reaction (i.e., biorthogonal chemistry) with norbornene conjugated to the antibody. 40 Equivalents of the drug was used per one antibody, and the conjugation reaction was performed at room temperature for 24 hours in a 20 mM histidine acetate solution (pH 5.5). The conjugation reaction was monitored by HIC-HPLC, and it was observed that the peaks appearing at 7.7 minutes after only a norbornene molecule was bound to trastuzumab were shifted to 8.7 minutes (DAR 1) and 10.5 minutes (DAR 2) as the tetrazine-

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PEG8-DM1 drug reacted with the antibody-norbornene linker. As a result, formation of the antibody-payload conjugate was observed.

The structure of the product 'antibody-payload conjugate' is shown in FIG. 59, and the reaction monitoring by HIC-HPLC is shown in FIG. 60 (Table 13).

TABLE 13

Reaction monitoring by HIC-HPLC for formation of antibody-payload conjugate	
Retention time	Composition
6.6 min	Trastuzumab
7.7 min	Trastuzumab-norbornene conjugate
8.7 min	Trastuzumab-DM1 conjugate (1 site, DAR1)
10.5 min	Trastuzumab-DM1 conjugate (2 site, DAR2)

Confirmation of Trastuzumab-Norbornene-Tetrazine-DM1 Structure (2)

An antibody-drug conjugate in which tetrazine-DM1 was bound to Trastuzumab-Norbornene was confirmed by mass spectrometry. It was determined to which site of the antibody the tetrazine-DM1 was bound via F(ab')₂ and Fc/2 after the antibody-drug conjugate was treated with an IdeS enzyme. It was confirmed that the Ides Fc/2-DM1 complex was found to have an "Obs-Theo" value of 1673.5, indicating that the drug was bound to the conjugate in consideration of the combined value of one tetrazine-DM1 molecule, which corresponded to the molecular weight of 1,688.80 Da (a dehydration reaction was observed in the molecular weight analysis of the DM1-series drug). The observed mass spectrum of the Ides_F(ab')₂ was detected, but the mass value of one charge was not shifted due to the complexity of the respective mass spectra, thereby making impossible to obtain the corresponding mass value.

The spectra of the complex having an increased molecular weight due to the trastuzumab-norbornene-tetrazine-DM1 binding are shown in FIGS. 61 and 62.

UPLC Conditions

Measuring equipment: Acquity UPLC I-Class system

1. Column: Thermo MABPacTM RP (2.1 mm×5 mm)

2. Column Temp: 60° C.

3. Mobile phase

A. 0.1% Formic acid in water

B. 0.1% Formic acid in acetonitrile

Mass Spectroscopy Conditions

Measuring equipment: LTQ Elite (Thermo)

Source type: HESI

Capillary Temp.: 320° C.

Source heater temp.: 300° C.

Sheath gas flow: 40.00

Aux gas flow: 20.00

Sweep gas flow: 5.00

Source voltage (KV): 4.00

FTMS resolution: 120,000

FTMS mass range: 400 to 4,000

5. FcRn Binding Analysis of Antibody-Drug Conjugate

The antibody-drug conjugate constructed in this patent was named 'AbClick®Pro.'

To check whether or not the antibody-drug conjugate (see FIG. 59; AbClick®Pro (NC, DM1)) had an antibody recycling function, the binding affinity of the conjugate for a neonatal Fc receptor (FcRn), which is closely associated with the in vivo antibody recycling, was analyzed. As one of the biomolecular interaction systems, Octet (model: Octet QK384) was used to determine the reaction affinity of FcRn

for the constructed ADC. As a difference in reflection of light caused by a surface change, which occurs according to a degree of reaction between molecules attached to a surface of a sensor and molecules in a sample, was recognized as being free of a label, kinetic analysis was performed. Trastuzumab, Kadcyla, and AbClick®Pro (NC, DM1) were used as the samples to measure binding affinities for FcRn under a condition of pH 6.0. As a result, it was confirmed that the measured KD values were 1.12×10^{-7} M, 1.12×10^{-7} M, and 8.84×10^{-8} M, respectively, indicating that AbClick®Pro had a similar or superior binding affinity, compared to Kadcyla (Table 14).

TABLE 14

FcRn binding analysis of antibody-drug conjugate			
pH 6.0	FcRn binding		
	KD_Kinetics (M)	Ka (1/Ms)	Kd (1/S)
Trastuzumab	1.12×10^{-7}	3.31×10^5	3.69×10^{-2}
Kadcyla	1.12×10^{-7}	2.10×10^5	2.37×10^{-2}
*AbClick®Pro	8.84×10^{-8}	2.26×10^5	1.99×10^{-2}

6. Antigen Binding Analysis of Antibody-Drug Conjugate

It was determined through enzyme-linked immunosorbent assay (ELISA) analysis whether or not a Her2 protein was bound to the antibody-drug conjugate. The antibody-norbornene complex constructed using trastuzumab and Compound III-SSFI (Dap) was used as the antibody-drug conjugate for measurement of antigen binding affinity. Structures of three drugs to be bound to the antibody-norbornene complex are shown in FIG. 63.

The dissociation constants (Kd) of Herceptin (trastuzumab), Kadcyla, AbClick®Pro (NC, DM1), AbClick®Pro (VC, DM1), and AbClick®Pro (VC, MMAE) from the antigen were confirmed to be approximately 98.6 pM, approximately 94.8 pM, approximately 121.4 pM, approximately 137.0 pM, and approximately 117.3 pM, respectively. Based on the results, it was confirmed that the binding site of the drug did not affect the binding affinity of the constructed ADC for the antigen.

A graph of measuring the antigen binding affinities of the antibody-drug conjugates is shown in FIG. 64 (Table 15).

TABLE 15

Antigen binding analysis of antibody-drug conjugate	
Antibody	K _D Value (binding affinity, pM)
Herceptin	98.6
Kadcyla	94.7
AbClick®Pro (NC, DM1)	121.4
AbClick®Pro (VC, DM1)	137.0
AbClick®Pro (VC, MMAE)	117.3

7. Confirmation of Serum Stability of Antibody-Drug Conjugate

It was determined through enzyme-linked immunosorbent assay (ELISA) analysis whether or not the antibody-drug conjugate was stable in serum. The stabilities of the antibody-drug conjugates in rat, rabbit and human sera were tested at room temperature. It was confirmed that a pattern of an intact ADC (i.e., drug-bound ADC) residual level of the self-constructed ADC "AbClick®Pro (NC, DM1)" over time was consistent with that of the Kadcyla® group.

The results of serum stability analysis of the antibody-drug conjugate are shown in FIGS. 65 to 67.

8. Evaluation of Medicinal Effect of Antibody-Drug Conjugate (AbClick®Pro) at Cellular Level (In Vitro Cytotoxicity Test)

Medicinal effects of the three antibody-drug conjugates (ADC, AbClick®Pro series) using trastuzumab were evaluated according to a level of a target marker in cancer cells, and a cytotoxicity test was performed using NCI-N87 and MDA-MB-468 as a positive cell line expressing the target antigen (Her2) 'cell line' and a negative cell line expressing BT474, respectively. The three antibody-drug conjugates (AbClick®Pro (NC, DM1), AbClick®Pro (VC, DM1), and AbClick®Pro (VC, MMAE)) constructed in the Her2-over-expressing cells NCI-N87 were observed after the cells were treated with different concentrations of the antibody-drug conjugates. As a result, it was confirmed that the antibody-drug conjugates had IC50 values of 207.7 ng/mL, 93.9 ng/mL, and 57.7 ng/mL, indicating that the antibody-drug conjugates had an excellent anti-cancer effect. The three antibody-drug conjugates (AbClick®Pro (NC, DM1), AbClick®Pro (VC, DM1), and AbClick®Pro (VC, MMAE)) constructed in another Her2-overexpressing cell line BT474 were observed after the cells were treated with different concentrations of the antibody-drug conjugates. As a result, it was confirmed that the antibody-drug conjugates had IC50 values of 47.7 ng/mL, 44.6 ng/mL, and 1.09 ng/mL, indicating that the antibody-drug conjugates had an excellent anti-cancer effect. The utility of the AbClick®Pro (VC, MMAE) as the novel ADC was confirmed because the AbClick®Pro (VC, MMAE) was observed to have an equal or superior apoptotic effect, compared to Kadcyla as the commercially available Herceptin ADC.

The test results are shown in FIGS. 68 to 70.

9. Evaluation of Medicinal Effect of Antibody-Drug Conjugate (AbClick®Pro) at Animal Level (In Vitro Cytotoxicity Test)

A target antigen-overexpressing NCI-N87 gastric cancer cell line was subcutaneously transplanted into mice to establish a xenograft model, and the mice were divided into 4 groups to perform an anti-cancer efficacy test on the materials administered. Because the BALB/c nude mice used in this test were deficient in T cells, cancer cells were easily transplanted into the BALB/c nude mice. Therefore, the BALB/c nude mice were used as a model suitable for an anti-cancer efficacy test using a rodent.

(a) Preparation of Cell Line

One vial of a human tumor cell line (NCI-N87 cell line) was put into a cell culture flask containing an RPMI1640 medium (Gibco, 22400-089) supplemented with heat-inactivated 10% fetal bovine serum (FBS; Gibco, 10082-742), and cultured at 37° C. in a 5% CO₂ incubator. The cell culture medium was washed with PBS, and diluted 10-fold with 2.5% trypsin-EDTA (Gibco, 15090). Thereafter, the diluted cell culture medium was added to separate the cells, and the cells were centrifuged (at 1,000 rpm for 5 minutes) to discard a supernatant. A fresh medium was added to the cell pellets to obtain a cell suspension. The viability of the cells was confirmed using a microscope, and the cells were diluted to a concentration of 1.25×10^7 cells/mL with a solution obtained by mixing a medium and Matrigel at a mixing ratio of 1:1 to prepare a cell line.

(b) Transplantation of Cell Line

A cell line was prepared using a method described in the section "4. 3) (4) Preparation of cell line." The cells were re-suspended and homogenized to prepare a cell line, and the prepared cell line was immediately administered to animals. For transplantation of the cell line, the back region of the animal was sterilized with 70% alcohol, the dorsal skin was

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pulled with the thumb and forefinger to form a space between the skin and muscles. Thereafter, a syringe with a 26 gauge needle was stuck in a subcutaneous pocket between the thumb and forefinger, and the cell line was then subcutaneously administered at a dose of 2.5×10^6 cells/0.2 mL/head from the front of the animal. During an acclimation period, healthy animals were selected, and subjected to cell line inoculation. When the size of a tumor in the cell line-transplanted site reached approximately 100 to 150 mm³, the mice were divided according to the size of a ranked tumor so that the tumor sizes in each group were distributed as uniformly as possible.

(c) Configuration of Experimental Groups and Determination of Dose and Administration Method

Cell line=NCI-N87

Mouse type=BALB/c nude (CAnN.Cg-Foxn1nu/CrljOri)

Number per group=5

Administration method=Intravenous injection (using a 26-gauge needle syringe)

Dose=5 mg/kg

Number of administration=Once/3 day, administered three times

Observation period=5 weeks

Group 1: PBS; Group 2: Herceptin (trastuzumab); Group 3: Kadcyla; Group 4 AbClick®Pro(NC, DM1); Group 5: AbClick®Pro(VC, DM1); and Group 6: AbClick®Pro(VC, MMAE)

(d) Observation and Inspection Items

General Symptoms

During the administration and observation periods, the type of general symptoms (including death), the symptom onset date, and the severity of symptoms were observed once a day, and recorded for each individual. The individuals whose general symptoms worsened were quarantined.

Body Weight

The body weight was measured on the grouping date or the start date for administration of the test substance, and then measured twice a week.

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Measurement of Tumor Size

The tumor size was measured twice a week for 5 weeks from the start date for administration of the test substance. The major- and minor-axis lengths of a tumor were measured using calipers, and the tumor size was calculated according to the following equation.

Tumor size= $ab^2/2$ (a: a major-axis length; and b: a minor-axis length)

(e) Results

The mice in the groups into which phosphate buffered saline (PBS) and Herceptin (trastuzumab) were injected did not show a tendency to suppress the growth of tumor for an observation period of 5 weeks. It was confirmed that the AbClick®Pro-series antibody-drug conjugates constructed according to the present invention has a superior ability to suppress the growth of tumor compared to Herceptin, indicating that the antibody-drug conjugates according to the present invention successfully function as the ADC.

The related results are shown in FIGS. 71 and 72.

10. Pharmacokinetics (PK) Test of Antibody-Drug Conjugate

To analyze the pharmacokinetic characteristics of the antibody-drug conjugate (ADC) manufactured using the AbClick®Pro, an enzyme-linked immunosorbent assay (ELISA) was performed to confirm the pharmacokinetics of the antibody-drug conjugate. The manufactured ADC had an AbClick®Pro (NC, DM1) structure as shown in FIG. 59, and comparative validation with Kadcyla was carried out. Rats (n=3) were used for an animal model. In this case, the rats were intravenously injected with 5 mg/kg of ADC, and then monitored over time. It was confirmed that the intact ADC form (an intact ADC to which a drug was bound) of the administered AbClick®Pro (NC, DM1) was reduced over time. It was judged that the intact ADC had similar or improved drug characteristics, compared to Kadcyla.

The results are shown in FIG. 73.

This application contains references to amino acid sequences and/or nucleic acid sequences which have been submitted concurrently herewith as the sequence listing entitled "000072uscoa_SequenceListing.XML", file size 80.4 kilobytes (KB), created on 14 Nov. 2023. The aforementioned sequence listing is hereby incorporated by reference in its entirety pursuant to 37 C.F.R. § 1.52(e)(5).

SEQUENCE LISTING

```
Sequence total quantity: 46
SEQ ID NO: 1          moltype = AA  length = 17
FEATURE              Location/Qualifiers
source                1..17
                     mol_type = protein
                     organism = synthetic construct

SEQUENCE: 1
GPSVFLFPPK PKDTLMI

SEQ ID NO: 2          moltype = AA  length = 9
FEATURE              Location/Qualifiers
source                1..9
                     mol_type = protein
                     organism = synthetic construct

SEQUENCE: 2
AWHLGELVW

SEQ ID NO: 3          moltype = AA  length = 9
FEATURE              Location/Qualifiers
VARIANT              1
                     note = not C or absent
VARIANT              9
                     note = W, F, or naphthylalanine
source                1..9
                     mol_type = protein
                     organism = synthetic construct
```

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9

-continued

MOD_RES	9	
	note = naphthylalanine	
MOD_RES	4	
	note = alanine variant comprising D3X3 linked to the methyl sidechain, wherein D3 is a C1-3 alkylene and X3 is NH3, OH, or SH	
VARIANT	6	
	note = E, N	
SEQUENCE: 3 XXHXGXLVX		9
SEQ ID NO: 4 FEATURE REGION	moltype = AA length = 9 Location/Qualifiers 1..9 note = Synthetic sequence	
MOD_RES	4	
	note = alanine variant comprising D3-X3 linked to the methyl sidechain, wherein D3 is a C1-3 alkylene and X3 is-NH3,-OH, or -SH	
VARIANT	9	
	note = W, F, or naphthylalanine	
source	1..9 mol_type = protein organism = synthetic construct	
MOD_RES	9	
	note = naphthylalanine	
VARIANT	6	
	note = E, N	
SEQUENCE: 4 AWHXGXLVX		9
SEQ ID NO: 5 FEATURE VARIANT	moltype = AA length = 11 Location/Qualifiers 3 note = not C	
VARIANT	7	
	note = E, N	
MOD_RES	5	
	note = alanine variant comprising D3-X3 linked to the methyl side chain, wherein D3 is a C1-3 alkylene and X3 is-NH3,-OH, or -SH	
VARIANT	10	
	note = W, F, or naphthylalanine	
source	1..11 mol_type = protein organism = synthetic construct	
MOD_RES	10	
	note = naphthylalanine	
SITE	11	
	note = D-Proline	
VARIANT	2	
	note = not C	
SEQUENCE: 5 PXXHXGXLVX P		11
SEQ ID NO: 6 FEATURE source	moltype = AA length = 17 Location/Qualifiers 1..17 mol_type = protein organism = synthetic construct	
VARIANT	1	
	note = not C or absent	
VARIANT	3	
	note = not C	
VARIANT	5	
	note = not C	
VARIANT	6	
	note = not C	
MOD_RES	8	
	note = alanine variant comprising D3-X3 linked to the methyl side chain, wherein D3 is a C1-3 alkylene and X3 is-NH3,-OH, or -SH	
VARIANT	10	
	note = E or N	
VARIANT	13	
	note = W, F, or naphthylalanine	
MOD_RES	13	
	note = naphthylalanine	
VARIANT	15	

-continued

VARIANT	note = not C 16	
VARIANT	note = not C or absent 17	
VARIANT	note = not C or absent 2	
SEQUENCE: 6 XXXCXXHXGX LVXCXXX	note = not C or absent	17
SEQ ID NO: 7 FEATURE VARIANT	moltype = AA length = 15 Location/Qualifiers 4..5	
MOD_RES	note = not C 12	
VARIANT	note = naphthylalanine 9	
VARIANT	note = E or N 12	
SITE	note = W, F, or naphthylalanine 15	
source	note = D-Proline 1..15	
MOD_RES	mol_type = protein organism = synthetic construct 7	
	note = alanine variant comprising D3-X3 linked to the methyl side chain, wherein D3 is a C1-3 alkylene and X3 is-NH3,-OH, or -SH	
SEQUENCE: 7 PDCXXHXGXL VXCTP		15
SEQ ID NO: 8 FEATURE VARIANT	moltype = AA length = 15 Location/Qualifiers 4..5	
VARIANT	note = not C 9	
VARIANT	note = E or N 12	
source	note = W, F, or naphthylalanine 1..15	
MOD_RES	mol_type = protein organism = synthetic construct 7	
	note = alanine variant comprising D3-X3 linked to the methyl side chain, wherein D3 is a C1-3 alkylene and X3 is-NH3,-OH, or -SH	
MOD_RES	12	
SEQUENCE: 8 CDCXXHXGXL VXCTC	note = naphthylalanine	15
SEQ ID NO: 9 FEATURE VARIANT	moltype = AA length = 13 Location/Qualifiers 3..4	
VARIANT	note = not C 8	
VARIANT	note = E or N 11	
source	note = W, F, or naphthylalanine 1..13	
MOD_RES	mol_type = protein organism = synthetic construct 6	
	note = alanine variant comprising D3-X3 linked to the methyl side chain, wherein D3 is a C1-3 alkylene and X3 is-NH3, -OH, or -SH	
MOD_RES	11	
SEQUENCE: 9 DCXXHXGXLV XCT	note = naphthylalanine	13
SEQ ID NO: 10 FEATURE MOD_RES	moltype = AA length = 13 Location/Qualifiers 6	
	note = alanine variant comprising D3-X3 linked to the methyl side chain, wherein D3 is a C1-3 alkylene and X3 is-NH3,-OH, or -SH	
source	1..13	

-continued

	mol_type = protein organism = synthetic construct	
SEQUENCE: 10 DCAWHXGELV WCT		13
SEQ ID NO: 11 FEATURE VARIANT MOD_RES	moltype = AA length = 9 Location/Qualifiers 1..2 note = not C 4 note = alanine variant comprising D3-X3-C(=O)-D2-X1-(C=O)-D1-R1 linked to the methyl side chain, wherein D3 is a C1-3 alkylene, X3 is-NH2-, -O-, or -S-, D2 is alkylene, alkenylene, or alkynylene, X1 is an element more electronegative than carbon, D1 is alkylene, alkenylene, or alkynylene alkylene, alkenylene, or alkynylene, and R1 is a first chemical functional group	
VARIANT	6 note = E or N	
source	1..9 mol_type = protein organism = synthetic construct	
MOD_RES	9 note = naphthylalanine	
VARIANT	9 note = W, F, or naphthylalanine	
SEQUENCE: 11 XXHXGXLVX		9
SEQ ID NO: 12 FEATURE VARIANT	moltype = AA length = 17 Location/Qualifiers 1 note = not C or absent	
VARIANT	3 note = not C	
VARIANT	5..6 note = not C	
MOD_RES	8 note = alanine variant comprising D3-X3-C(=O)-D2-X1-(C=O)-D1-R1' linked to the methyl side chain, wherein D3 is a C1-3 alkylene, X3 is-NH2-, -O-, or -S-, D2 is alkylene, alkenylene, or alkynylene, X1 is an element more electronegative than carbon, D1 is alkylene, alkenylene, or alkynylene alkylene, alkenylene, or alkynylene, and R1' is a first chemical functional group	
VARIANT	10 note = E or N	
VARIANT	15 note = not C	
source	1..17 mol_type = protein organism = synthetic construct	
VARIANT	2 note = not C or absent	
VARIANT	16 note = not C or absent	
VARIANT	17 note = not C or absent	
MOD_RES	13 note = naphthylalanine	
VARIANT	13 note = W, F, or naphthylalanine	
SEQUENCE: 12 XXXCXXHXGX LVXCXXX		17
SEQ ID NO: 13 FEATURE MOD_RES	moltype = AA length = 13 Location/Qualifiers 6 note = alanine variant comprising D3-X3-C(=O)-D2-X1-(C=O)-D1-R1 linked to the methyl side chain, wherein D3 is a C1-3 alkylene, X3 is-NH2-, -O-, or -S-, D2 is alkylene, alkenylene, or alkynylene, X1 is an element more electronegative than carbon, D1 is alkylene, alkenylene, or alkynylene alkylene, alkenylene, or alkynylene, and R1 is a first chemical functional group	
source	1..13 mol_type = protein organism = synthetic construct	
SEQUENCE: 13		

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DCAWHXGELV WCT		13
SEQ ID NO: 14	moltype = AA length = 9	
FEATURE	Location/Qualifiers	
VARIANT	1..2	
	note = not C	
VARIANT	6	
	note = E or N	
source	1..9	
	mol_type = protein	
	organism = synthetic construct	
MOD_RES	9	
	note = naphthylalanine	
VARIANT	9	
	note = W, F, or naphthylalanine	
MOD_RES	4	
	note = alanine variant comprising	
	D3-X3-C(=O)-D2-X1-(C=O)-D1-R1 linked to the methyl side	
	chain, wherein D3 is a C1-3 alkylene, X3 is-NH2-, -O-, or	
	-S-, D2 is alkylene, alkenylene, or alkynylene, X1 is an	
	element more electronegative than carbon, D1 is alkylene,	
	alkenylene, or alkynylene alkylene, alkenylene, or	
	alkynylene, and R1 is a first chemical functional group	
SEQUENCE: 14		
XXHXGXLVX		9
SEQ ID NO: 15	moltype = AA length = 17	
FEATURE	Location/Qualifiers	
VARIANT	1	
	note = not C or absent	
VARIANT	3	
	note = not C	
VARIANT	5..6	
	note = not C	
MOD_RES	8	
	note = alanine variant comprising	
	D3-X3-C(=O)-D2-X1-(C=O)-D1-R1' linked to the methyl side	
	chain, wherein D3 is a C1-3 alkylene, X3 is-NH2-, -O-, or	
	-S-, D2 is alkylene, alkenylene, or alkynylene, X1 is an	
	element more electronegative than carbon, D1 is alkylene,	
	alkenylene, or alkynylene alkylene, alkenylene, or	
	alkynylene, and H1 is a first click-chemistry functional	
	group	
VARIANT	13	
	note = W, F, or naphthylalanine	
VARIANT	10	
	note = E or N	
VARIANT	16	
	note = not C or absent	
source	1..17	
	mol_type = protein	
	organism = synthetic construct	
VARIANT	2	
	note = not C or absent	
MOD_RES	13	
	note = naphthylalanine	
VARIANT	15	
	note = not C	
VARIANT	17	
	note = not C or absent	
SEQUENCE: 15		
XXXCXXHXGX LVXCXXX		17
SEQ ID NO: 16	moltype = AA length = 13	
FEATURE	Location/Qualifiers	
MOD_RES	6	
	note = alanine variant comprising	
	D3-X3-C(=O)-D2-X1-(C=O)-D1-R1' linked to the methyl side	
	chain, wherein D3 is a C1-3 alkylene, X3 is-NH2-, -O-, or	
	-S-, D2 is alkylene, alkenylene, or alkynylene, X1 is an	
	element more electronegative than carbon, D1 is alkylene,	
	alkenylene, or alkynylene alkylene, alkenylene, or	
	alkynylene, and H1 is a first click-chemistry functional	
	group	
source	1..13	
	mol_type = protein	
	organism = synthetic construct	
SEQUENCE: 16		
DCAWHXGELV WCT		13

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SEQ ID NO: 17      moltype = AA  length = 17
FEATURE           Location/Qualifiers
source            1..17
                  mol_type = protein
                  organism = synthetic construct
MOD_RES           10
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and R1 is a first chemical functional group
SEQUENCE: 17
GPSVFLFPPX PKDTLMI                                     17

SEQ ID NO: 18      moltype = AA  length = 17
FEATURE           Location/Qualifiers
source            1..17
                  mol_type = protein
                  organism = synthetic construct
MOD_RES           12
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and R1 is a first chemical functional group
SEQUENCE: 18
GPSVFLFPPK PXDTLMI                                     17

SEQ ID NO: 19      moltype = AA  length = 17
FEATURE           Location/Qualifiers
source            1..17
                  mol_type = protein
                  organism = synthetic construct
MOD_RES           10
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and R1 is a first chemical functional group
MOD_RES           12
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and R1 is a first chemical functional group
SEQUENCE: 19
GPSVFLFPPX PXDTLMI                                     17

SEQ ID NO: 20      moltype = AA  length = 17
FEATURE           Location/Qualifiers
source            1..17
                  mol_type = protein
                  organism = synthetic construct
MOD_RES           10
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and H1 is a first click-chemistry
                    functional group
SEQUENCE: 20
GPSVFLFPPX PKDTLMI                                     17

SEQ ID NO: 21      moltype = AA  length = 17
FEATURE           Location/Qualifiers
source            1..17
                  mol_type = protein
                  organism = synthetic construct
MOD_RES           12
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and H1 is a first click-chemistry
                    functional group
SEQUENCE: 21
GPSVFLFPPK PXDTLMI                                     17

SEQ ID NO: 22      moltype = AA  length = 17
FEATURE           Location/Qualifiers
MOD_RES           10
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and H1 is a first click-chemistry
                    functional group
MOD_RES           12
                  note = lysine variant comprising -C(=O)-D1-R1, linked to
                    the lysine side chain, wherein D1 is alkylene, alkenylene,
                    or alkynylene, and H1 is a first click-chemistry
                    functional group

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source	1..17	
	mol_type = protein	
	organism = synthetic construct	
SEQUENCE: 22		
GPSVFLFPPX PXDTLMI		17
SEQ ID NO: 23	moltype = AA length = 17	
FEATURE	Location/Qualifiers	
MOD_RES	10	
	note = lysine variant comprising C(=O)-D1-B-Cm, linked to the lysine side chain, wherein D1 is alkylene, alkenylene, or alkynylene, and B is formed by a click chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group, and Cm is a cargo moiety	
source	1..17	
	mol_type = protein	
	organism = synthetic construct	
SEQUENCE: 23		
GPSVFLFPPX PKDTLMI		17
SEQ ID NO: 24	moltype = AA length = 17	
FEATURE	Location/Qualifiers	
MOD_RES	12	
	note = lysine variant comprising -C(=O)-D1-B-Cm, linked to the lysine side chain, wherein D1 is alkylene, alkenylene, or alkynylene, and B is formed by a click chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group, and Cm is a cargo moiety	
source	1..17	
	mol_type = protein	
	organism = synthetic construct	
SEQUENCE: 24		
GPSVFLFPPK PXDTLMI		17
SEQ ID NO: 25	moltype = AA length = 17	
FEATURE	Location/Qualifiers	
MOD_RES	10	
	note = lysine variant comprising -C(=O)-D1-B-Cm, linked to the lysine side chain, wherein D1 is alkylene, alkenylene, or alkynylene, and B is formed by a click chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group, and Cm is a cargo moiety	
MOD_RES	12	
	note = lysine variant comprising -C(=O)-D1-B-Cm, linked to the lysine side chain, wherein D1 is alkylene, alkenylene, or alkynylene, and B is formed by a click chemistry reaction between a first click-chemistry functional group and a second click-chemistry functional group, and Cm is a cargo moiety	
source	1..17	
	mol_type = protein	
	organism = synthetic construct	
SEQUENCE: 25		
GPSVFLFPPX PXDTLMI		17
SEQ ID NO: 26	moltype = AA length = 34	
FEATURE	Location/Qualifiers	
source	1..34	
	mol_type = protein	
	organism = synthetic construct	
SEQUENCE: 26		
LFPPKPKDTL MISRTPEVTC VVVDVSHEDP EVKF		34
SEQ ID NO: 27	moltype = AA length = 10	
FEATURE	Location/Qualifiers	
source	1..10	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 27		
TWKTSRISIF		10
SEQ ID NO: 28	moltype = AA length = 11	
FEATURE	Location/Qualifiers	
source	1..11	
	mol_type = protein	
	organism = Synthetic construct	

-continued

SEQUENCE: 28 FGRLVSSAIR Y		11
SEQ ID NO: 29 FEATURE source	moltype = AA length = 12 Location/Qualifiers 1..12 mol_type = protein organism = Synthetic construct	
SEQUENCE: 29 EPIHRSTLTA LL		12
SEQ ID NO: 30 FEATURE source	moltype = AA length = 4 Location/Qualifiers 1..4 mol_type = protein organism = Synthetic construct	
SEQUENCE: 30 APAR		4
SEQ ID NO: 31 FEATURE source	moltype = AA length = 6 Location/Qualifiers 1..6 mol_type = protein organism = Synthetic construct	
SEQUENCE: 31 HWRGWV		6
SEQ ID NO: 32 FEATURE source	moltype = AA length = 6 Location/Qualifiers 1..6 mol_type = protein organism = Synthetic construct	
SEQUENCE: 32 HYFKFD		6
SEQ ID NO: 33 FEATURE source	moltype = AA length = 6 Location/Qualifiers 1..6 mol_type = protein organism = Synthetic construct	
SEQUENCE: 33 HFRRHL		6
SEQ ID NO: 34 FEATURE source	moltype = AA length = 6 Location/Qualifiers 1..6 mol_type = protein organism = Synthetic construct	
VARIANT	3 note = X is citrulline	
SEQUENCE: 34 HWXGWV		6
SEQ ID NO: 35 FEATURE source	moltype = AA length = 5 Location/Qualifiers 1..5 mol_type = protein organism = Synthetic construct	
SEQUENCE: 35 DDAAG		5
SEQ ID NO: 36 FEATURE source	moltype = AA length = 4 Location/Qualifiers 1..4 mol_type = protein organism = Synthetic construct	
SEQUENCE: 36 DAAG		4
SEQ ID NO: 37 FEATURE source	moltype = AA length = 9 Location/Qualifiers 1..9 mol_type = protein organism = Synthetic construct	
VARIANT	1 note = X is Ser(Ala)	
VARIANT	8 note = X is lactic acid	
SEQUENCE: 37		

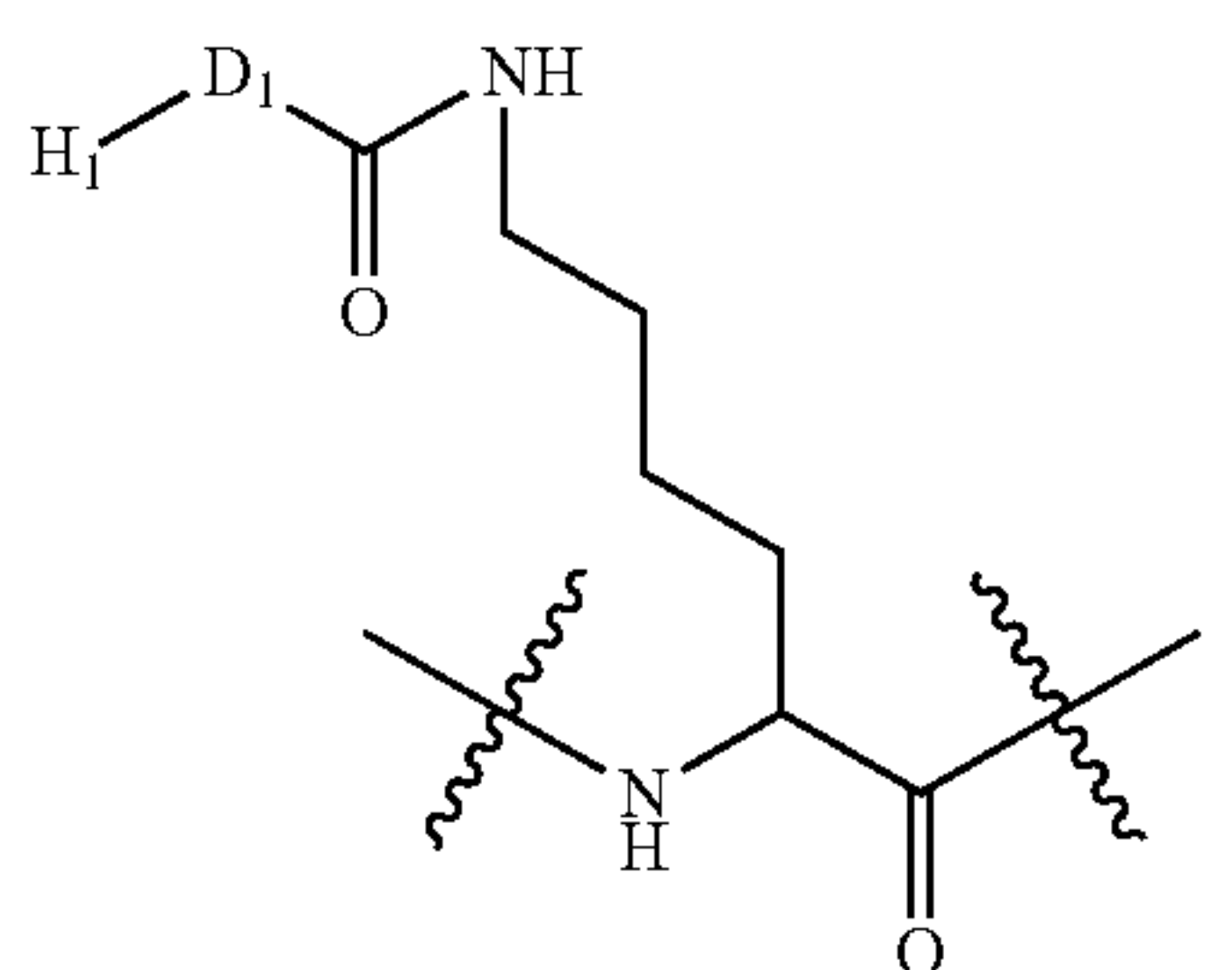
-continued

XRWHYFKXE		9
SEQ ID NO: 38	moltype = AA length = 9	
FEATURE	Location/Qualifiers	
source	1..9	
	mol_type = protein	
	organism = Synthetic construct	
VARIANT	1	
	note = X is Dap(Ala)	
VARIANT	8	
	note = X is lactic acid	
SEQUENCE: 38		
XRWHYFKXE		9
SEQ ID NO: 39	moltype = AA length = 7	
FEATURE	Location/Qualifiers	
source	1..7	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 39		
MWFRHYK		7
SEQ ID NO: 40	moltype = AA length = 8	
FEATURE	Location/Qualifiers	
source	1..8	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 40		
NKFRGKYK		8
SEQ ID NO: 41	moltype = AA length = 8	
FEATURE	Location/Qualifiers	
source	1..8	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 41		
NARKFYKG		8
SEQ ID NO: 42	moltype = AA length = 8	
FEATURE	Location/Qualifiers	
source	1..8	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 42		
FYWHCLDE		8
SEQ ID NO: 43	moltype = AA length = 8	
FEATURE	Location/Qualifiers	
source	1..8	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 43		
FYCHWALE		8
SEQ ID NO: 44	moltype = AA length = 8	
FEATURE	Location/Qualifiers	
source	1..8	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 44		
FYCHTIDE		8
SEQ ID NO: 45	moltype = AA length = 4	
FEATURE	Location/Qualifiers	
source	1..4	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 45		
RRGW		4
SEQ ID NO: 46	moltype = AA length = 7	
FEATURE	Location/Qualifiers	
source	1..7	
	mol_type = protein	
	organism = Synthetic construct	
SEQUENCE: 46		
KHRFNKD		7

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What is claimed is:

1. A modified antibody or its fragment comprising bio-orthogonal click chemistry functional group,
wherein the modified antibody comprises one or more modified heavy chain,
wherein each of the modified heavy chain comprises a modified lysine residue at 248 position of heavy chain amino acid sequence, wherein the 248 position is determined by an EU numbering system, and
wherein the modified lysine residue comprises a bio-orthogonal click chemistry functional group at terminal end, and
wherein the modified lysine residue does not comprise a terminal bio-reactive functional group selected from terminal hydroxy group, terminal carboxyl group, terminal thiol group, terminal amino group, and terminal aldehyde group.
2. The modified antibody of claim 1, wherein the modified lysine residue has following structure:



wherein H_1 is a group having the bio-orthogonal click chemistry functional group, and
wherein D_1 is bond, or is selected from C_{1-4} alkylene, C_{2-4} alkenylene, C_{2-4} alkynylene and C_{3-8} cycloalkylene.

3. The modified antibody of claim 2, wherein D_1 is bond, or is selected from unsubstituted C_{1-4} alkylene, unsubstituted C_{2-4} alkenylene, unsubstituted C_{2-4} alkynylene and unsubstituted C_{3-8} cycloalkylene.

4. The modified antibody of claim 1, wherein the modified heavy chain comprises an amino acid sequence having a structure of formula 8-2:

G-P-S-V-F-L-F-P-P-K-P-(K')-D-T-L-M-I (SEQ ID NO: 21),

[formula 8-2]

wherein K' is the modified lysine residue.

5. The modified antibody of claim 2, wherein D_1 comprises one or more heteroatoms.

6. The modified antibody of claim 2, wherein D_1 is $-[CH_2]_a-O-[CH_2]_b-$,
wherein each of a and b is independently selected from 0 to 4, and each of a and b is integer, and
wherein sum of a and b is 0 or more and 4 or less.

7. The modified antibody of claim 2, wherein D_1 is $-CH_2CH_2OCH_2-$ or $-CH_2OCH_2-$.

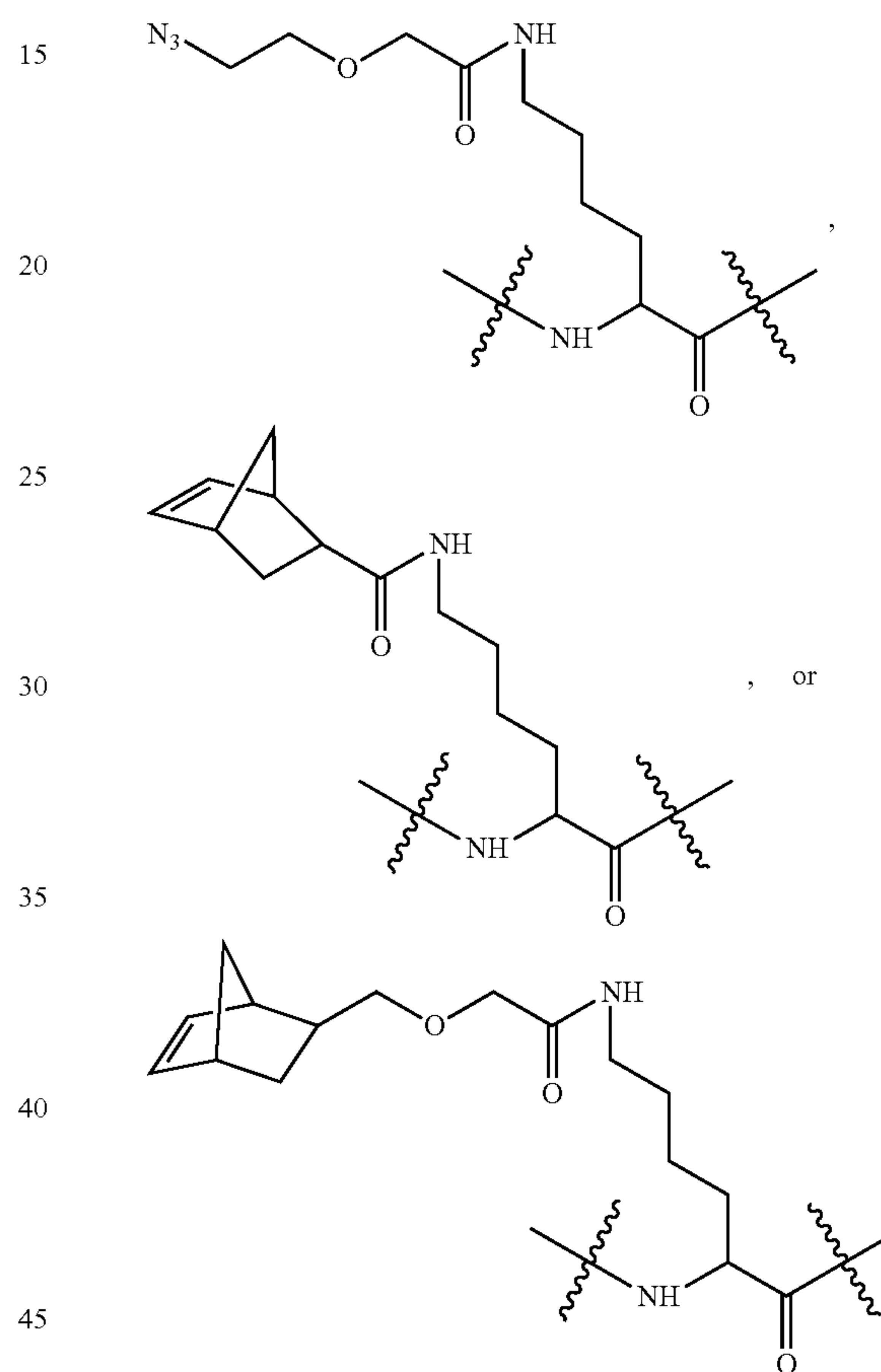
8. The modified antibody of claim 1, wherein the modified antibody comprises two modified heavy chains.

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9. The modified antibody of claim 1, wherein the bio-orthogonal click chemistry functional group is a group capable of participating in a click chemistry reaction selected from Huisgen 1,3-dipolar cycloaddition and Diels-Alder reaction.

10. The modified antibody of claim 1, wherein the bio-orthogonal click chemistry functional group is selected from azide group, tetrazine group, dibenzocyclooctyne group, and norbornene group.

11. The modified antibody of claim 1, wherein the modified lysine residue has following structure:



12. The modified antibody of claim 3, wherein the bio-orthogonal click chemistry group is a group capable of participating in a click chemistry reaction selected from Huisgen 1,3-dipolar cycloaddition and Diels-Alder reaction.

13. The modified antibody of claim 3, wherein the bio-orthogonal click chemistry functional group is selected from the group consisting of an azide group, a tetrazine group, a dibenzocyclooctyne group, and a norbornene group.

* * * * *