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PROVISIONAL APPLICATION FOR PATENT COVER SHEET – Page 1 of 2

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Additional inventors are being named on the _____ separately numbered sheets attached hereto.		
TITLE OF THE INVENTION (500 characters max):		
Novel Heterodimeric Proteins		
Direct all correspondence to: CORRESPONDENCE ADDRESS ☒ The address corresponding to Customer Number: 67374 OR ☐ Firm or Individual Name Address City State Zip Country Telephone Email		
ENCLOSED APPLICATION PARTS (check all that apply)		
☒ Application Data Sheet. See 37 CFR 1.76 ☐ CD(s), Number of CDs _____ ☒ Drawing(s) Number of Sheets <u>36</u> ☐ Other (specify) _____ ☒ Specification (e.g. description of the invention) Number of Pages <u>71</u>		
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METHOD OF PAYMENT OF THE FILING FEE AND APPLICATION SIZE FEE FOR THIS PROVISIONAL APPLICATION FOR PATENT		
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PROVISIONAL APPLICATION COVER SHEET
Page 2 of 2

PTO/SB/16 (09-11)

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SIGNATURE Robin M. Silva Date 13 March 2013

TYPED or PRINTED NAME Robin M. Silva REGISTRATION NO. 38304
(if appropriate)

TELEPHONE 415-442-1000 Docket Number: 067461-5161-PR02

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Application Data Sheet 37 CFR 1.76		Attorney Docket Number	067461-5161-PR02
		Application Number	
Title of Invention	Novel Heterodimeric Proteins		
<p>The application data sheet is part of the provisional or nonprovisional application for which it is being submitted. The following form contains the bibliographic data arranged in a format specified by the United States Patent and Trademark Office as outlined in 37 CFR 1.76.</p> <p>This document may be completed electronically and submitted to the Office in electronic format using the Electronic Filing System (EFS) or the document may be printed and included in a paper filed application.</p>			

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☐ Portions or all of the application associated with this Application Data Sheet may fall under a Secrecy Order pursuant to 37 CFR 5.2 (Paper filers only. Applications that fall under Secrecy Order may not be filed electronically.)	
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Application Data Sheet 37 CFR 1.76		Attorney Docket Number	067461-5161-PR02
		Application Number	
Title of Invention	Novel Heterodimeric Proteins		

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For further information see 37 CFR 1.33(a).

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Application Information:

Title of the Invention	Novel Heterodimeric Proteins		
Attorney Docket Number	067461-5161-PR02	Small Entity Status Claimed	☒
Application Type	Provisional		
Subject Matter	Utility		
Suggested Class (if any)		Sub Class (if any)	
Suggested Technology Center (if any)			
Total Number of Drawing Sheets (if any)	36	Suggested Figure for Publication (if any)	

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Application Data Sheet 37 CFR 1.76		Attorney Docket Number	067461-5161-PR02
		Application Number	
Title of Invention	Novel Heterodimeric Proteins		

Publication Information:

☐	Request Early Publication (Fee required at time of Request 37 CFR 1.219)
☐	Request Not to Publish. I hereby request that the attached application not be published under 35 U.S.C. 122(b) and certify that the invention disclosed in the attached application has not and will not be the subject of an application filed in another country, or under a multilateral international agreement, that requires publication at eighteen months after filing.

Representative Information:

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Prior Application Status		Remove	
Application Number	Continuity Type	Prior Application Number	Filing Date (YYYY-MM-DD)
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Application Data Sheet 37 CFR 1.76		Attorney Docket Number	067461-5161-PR02
		Application Number	
Title of Invention	Novel Heterodimeric Proteins		

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Application Data Sheet 37 CFR 1.76		Attorney Docket Number	067461-5161-PR02
		Application Number	
Title of Invention	Novel Heterodimeric Proteins		

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Non-Applicant Assignee Information:

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Application Data Sheet 37 CFR 1.76		Attorney Docket Number	067461-5161-PR02		
		Application Number			
Title of Invention	Novel Heterodimeric Proteins				
First Name	Robin M.	Last Name	Silva	Registration Number	38304
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NOVEL HETERODIMERIC PROTEINS

Priority

This application claims priority to U.S. 61/752,349, filed on January 14, 2013 and U.S. 61/764,954, filed on February 14, 2013 which are incorporated herein in their entirety.

Incorporation of Related Applications

The following applications are incorporated by reference in their entirety, USSNs 61/302,707, 61/732,813, 61/598,686, 61/441,552, 61/311,472, 61/450,457, 61/545,498, 61/368,969, 61/391,509, 61/391,515, 61/439,263, 61/368,962 and 61/593,846, 61/515,745, 61/752,349, 13/194,904, 13/568,028, 13/648,951, 12/875,015, as well as US Publication Nos. 2009/0163,699 and 2006/0024298;

TECHNICAL FIELD

[0001] The present invention describes novel immunoglobulin compositions that simultaneously co-engage antigens, where both of the antigens are bound monovalently. The novel immunoglobulins described preferably utilize heterodimeric Fc regions. Methods of using the novel immunoglobulin compositions, particularly for therapeutic purposes, are also described herein.

BACKGROUND OF THE INVENTION

[0002] Antibody-based therapeutics have been used successfully to treat a variety of diseases, including cancer and autoimmune/inflammatory disorders. Yet improvements to this class of drugs are still needed, particularly with respect to enhancing their clinical efficacy. One avenue being explored is the engineering of additional and novel antigen binding sites into antibody-based drugs such that a single immunoglobulin molecule co-engages two different antigens. Such non-native or alternate antibody formats that engage two different antigens are often referred to as bispecifics. Because the considerable diversity of the antibody variable region (Fv) makes it possible to produce an Fv that recognizes virtually any molecule, the typical approach to bispecific generation is the introduction of new variable regions into the antibody.

[0003] A number of alternate antibody formats have been explored for bispecific targeting (Chames & Baty, 2009, mAbs 1[6]:1-9; Holliger & Hudson, 2005, Nature Biotechnology 23[9]:1126-1136; and Kontermann, 2012 MAb 4(2):182, all of which are expressly incorporated herein by reference). Initially, bispecific antibodies were made by fusing two cell

lines that each produced a single monoclonal antibody (Milstein et al., 1983, Nature 305:537-540). Although the resulting hybridoma or quadroma did produce bispecific antibodies, they were only a minor population, and extensive purification was required to isolate the desired antibody. An engineering solution to this was the use of antibody fragments to make bispecifics. Because such fragments lack the complex quaternary structure of a full length antibody, variable light and heavy chains can be linked in single genetic constructs. Antibody fragments of many different forms have been generated, including diabodies, single chain diabodies, tandem scFv's, and Fab₂ bispecifics (Chames & Baty, 2009, mAbs 1[6]:1-9; Holliger & Hudson, 2005, Nature Biotechnology 23[9]:1126-1136; expressly incorporated herein by reference). While these formats can be expressed at high levels in bacteria and may have favorable penetration benefits due to their small size, they clear rapidly in vivo and can present manufacturing obstacles related to their production and stability. A principal cause of these drawbacks is that antibody fragments typically lack the constant region of the antibody with its associated functional properties, including larger size, high stability, and binding to various Fc receptors and ligands that maintain long half-life in serum (i.e., the neonatal Fc receptor FcRn) or serve as binding sites for purification (i.e., protein A and protein G).

[0004] More recent work has attempted to address the shortcomings of fragment-based bispecifics by engineering dual binding into full length antibody-like formats (Wu et al., 2007, Nature Biotechnology 25[11]:1290-1297; USSN12/477,711; Michaelson et al., 2009, mAbs 1[2]:128-141; PCT/US2008/074693; Zuo et al., 2000, Protein Engineering 13[5]:361-367; USSN09/865,198; Shen et al., 2006, J Biol Chem 281[16]:10706-10714; Lu et al., 2005, J Biol Chem 280[20]:19665-19672; PCT/US2005/025472; and Kontermann, 2012 MAb 4(2):182, all of which are expressly incorporated herein by reference). These formats overcome some of the obstacles of the antibody fragment bispecifics, principally because they contain an Fc region. One significant drawback of these formats is that, because they build new antigen binding sites on top of the homodimeric constant chains, binding to the new antigen is always bivalent.

[0005] For many antigens that are attractive as co-targets in a therapeutic bispecific format, the desired binding is monovalent rather than bivalent. For many immune receptors, cellular activation is accomplished by cross-linking of a monovalent binding interaction. The mechanism of cross-linking is typically mediated by antibody/antigen immune complexes, or via effector cell to target cell engagement. For example, the low affinity Fc gamma receptors (FcγRs) such as FcγRIIa, FcγRIIb, and FcγRIIIa bind monovalently to the antibody Fc region. Monovalent binding does not activate cells expressing these FcγRs; however, upon immune complexation or cell-to-cell contact, receptors are cross-linked and clustered on the cell

surface, leading to activation. For receptors responsible for mediating cellular killing, for example Fc γ RIIIa on natural killer (NK) cells, receptor cross-linking and cellular activation occurs when the effector cell engages the target cell in a highly avid format (Bowles & Weiner, 2005, J Immunol Methods 304:88-99, expressly incorporated by reference). Similarly, on B cells the inhibitory receptor Fc γ RIIb downregulates B cell activation only when it engages into an immune complex with the cell surface B-cell receptor (BCR), a mechanism that is mediated by immune complexation of soluble IgG's with the same antigen that is recognized by the BCR (Heyman 2003, Immunol Lett 88[2]:157-161; Smith and Clatworthy, 2010, Nature Reviews Immunology 10:328-343; expressly incorporated by reference). As another example, CD3 activation of T-cells occurs only when its associated T-cell receptor (TCR) engages antigen-loaded MHC on antigen presenting cells in a highly avid cell-to-cell synapse (Kuhns et al., 2006, Immunity 24:133-139). Indeed nonspecific bivalent cross-linking of CD3 using an anti-CD3 antibody elicits a cytokine storm and toxicity (Perruche et al., 2009, J Immunol 183[2]:953-61; Chatenoud & Bluestone, 2007, Nature Reviews Immunology 7:622-632; expressly incorporated by reference). Thus for practical clinical use, the preferred mode of CD3 co-engagement for redirected killing of targets cells is monovalent binding that results in activation only upon engagement with the co-engaged target.

[0006] Thus while bispecifics generated from antibody fragments suffer biophysical and pharmacokinetic hurdles, a drawback of those built with full length antibody -like formats is that they engage co-target antigens multivalently in the absence of the primary target antigen, leading to nonspecific activation and potentially toxicity. The present invention solves this problem by introducing a novel bispecific format that enables the co-engagement of distinct target antigens.

SUMMARY OF THE INVENTION

[0007] Accordingly, in one aspect, the invention provides heterodimeric antibodies comprising a first heavy chain comprising a first Fc domain and a single chain Fv region that binds a first antigen. The heterodimeric antibodies also comprise a second heavy chain comprising a second Fc domain, a first variable heavy chain and a first variable light chain, wherein the first and second Fc domains are different.

[0008] In a further aspect, the pIs of the first heavy chain and the second heavy chain are at least 0.5 logs apart.

[0009] In an additional aspect, the heterodimeric antibodies of the invention further comprise one or two additional antigen binding domains that can be either or both of an scFv and a Fab.

[0010] In a further aspect, the heterodimeric antibodies of the invention comprise heterodimerization variants according to Figures 4, 5, 6, 20, 21 and 22.

[0011] In an additional aspect, the heterodimeric antibodies comprise pl variants in each monomer, with monomer one are ISO(-): I199T/N203D/K274Q/R355Q/N384S/K392N/V397M/Q419E/DEL44 and the pl substitutions in monomer 2 are ISO(+RR): Q196K/I199T/P217R/P228R/N276K.

[0012] In additional aspects, the present invention provides heterodimeric antibodies wherein one of the heavy chains comprises a variant selected from the group consisting of S119E, K133E, K133Q, R133E (in case of IgG2-4), R133Q (in case of IgG2-4), T164E, K205E, K205Q, N208D, K210E, K210Q, K274E, K320E, K322E, K326E, K334E, R355E, K392E, Deletion of K447, adding peptide DEDE at the c-terminus, G137E, N203D, K274Q, R355Q, K392N and Q419E, 349A, 349C, 349E, 349I, 349K, 349S, 349T, 349W, 351E, 351K, 354C, 356K, 357K, 364C, 364D, 364E, 364F, 364G, 364H, 364R, 364T, 364Y, 366D, 366K, 366S, 366W, 366Y, 368A, 368E, 368K, 368S, 370C, 370D, 370E, 370G, 370R, 370S, 370V, 392D, 392E, 394F, 394S, 394W, 394Y, 395T, 395V, 396T, 397E, 397S, 397T, 399K, 401K, 405A, 405S, 407T, 407V, 409D, 409E, 411D, 411E, 411K, 439D, 349C/364E, 349K/351K, 349K/351K/394F, 349K/354C, 349K/394F, 349K/394F/401K, 349K/394Y, 349K/401K, 349K/405A, 349T/351E/411E, 349T/394F, 349T/394F/401K, 349T/394F/411E, 349T/405A, 349T/411E, 351E/364D, 351E/364D/405A, 351E/364E, 351E/366D, 351K/364H/401K, 351K/366K, 364D/370G, 364D/394F, 364E/405A, 364E/405S, 364E/411E, 364E/411E/405A, 364H/394F, 364H/401K, 364H/401K/405A, 364H/405A, 364H/405A/411E, 364Y/370R, 370E/411E, 370R/411K, 395T/397S/405A and 397S/405A.

[0013] The present invention describes methods for generating the novel compositions of the invention. The present invention describes purification methods for the immunoglobulins herein, particularly methods for separating heterodimeric and homodimeric protein species. Also described are methods of testing the immunoglobulins herein, including in vitro and in vitro experiments.

[0014] The present invention provides isolated nucleic acids encoding the novel immunoglobulin compositions described herein. The present invention provides vectors comprising said nucleic acids, optionally, operably linked to control sequences. The present invention provides host cells containing the vectors, and methods for producing and optionally recovering the immunoglobulin compositions.

[0015] The present invention provides compositions comprising immunoglobulin polypeptides described herein, and a physiologically or pharmaceutically acceptable carrier or diluent.

[0016] The present invention contemplates therapeutic and diagnostic uses for the immunoglobulin polypeptides disclosed herein.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures for bottle cap

[0017] Figure 1. Illustration of the “triple F” format for bispecific immunoglobulins. Figure 1A shows a scFv-Fc format. Figure 1C depicts a more standard bispecific format, also utilizing the pl variants of the invention (and optionally and independently the other heterodimerization variants). Figure 1B shows the “triple F” format (sometimes also referred to as the “bottle-opener” configuration; (and optionally and independently the other heterodimerization variants).

[0018] Figures 2A-2E depict a variety of “higher multispecificity” embodiments of the present invention. Figure 2A shows a “triple F” configuration with another scFv attached to the Fab monomer (this one, along with Figure 2D, has a greater molecular weight differential as well). Figure 2B depicts a “triple F” with another scFv attached to the scFv monomer. Figure 2C depicts a “three scFv” format. Figure 2D depicts an additional Fab attached to the Fab monomer. Figure 2E depicts a Fab hooked to one of the scFv monomers.

[0019] Figures 3A-3E show additional varieties of “higher multispecificity” embodiments of the “triple F” format, all with one monomer comprising an scFv and all of which have molecular weight differentials which can be exploited for purification of the heterodimers). Figure 3A shows a “Fab-Fv” format with binding to two different antigens, with Figure 3B depicting the “Fab-Fv” format with binding to a single antigen (e.g. bivalent binding to antigen 1). Figures 3C and 3D depicts a “Fv-Fab” format with similar bivalent or monovalent additional antigen binding. Figure 11E depicts one monomer with a CH1-CL attached to the second scFv.

[0020] Figure 4A and 4B show novel steric variants. As will be understood by those in the art, the first column of each table represents “corresponding” monomer pairs or “sets”: that is, monomer 1 has 405A and the corresponding steric variant is 394F. Again, these can be optionally and independently combined, as long as some “strandedness” is maintained.

[0021] Figure 5 depicts heterodimerization variants that find particular use in the present invention.

[0022] Figure 6 depicts novel pl heterodimerization variants of particular use in the creation of heterodimeric proteins. As will be understood by those in the art, the first row of each table represents “corresponding” monomer pairs or sets. Note that the I199T and V397M variants are “neutral” (e.g. not for the purposes of altering pl of one of the

monomers). Rather, they contribute to the reduction of immunogenicity by helping preserve a close epitope; e.g. I199T follows pl variants N203D or Q196K, and V397M follows K392N. Thus these two variants can optionally and independently be removed from any of the sequences above.

[0023] Figure 7. Amino acid sequences for an anti-CD19 Fab x anti-CD3 scFv “triple F” embodiment, showing the HC-Fab sequence, the HC-scFv sequence, and the light chain. Amino acid substitutions made to allow for efficient purification of the desired “triple F” heterodimer over the undesired dual scFv-Fc and mAb homodimers are underlined.

[0024] Figure 8. Amino acid sequences for the humanized anti-CD3 variable regions finding particular use in the invention (with CDRs underlined).

[0025] Figure 9. Production of XENP11874, a “triple F” bispecific with an anti-CD19 Fab and anti-CD3 scFv. In Fig. 9A, the ion exchange purification of the desired “triple F” heterodimer from the undesired dual scFv-Fc and mAb homodimers is shown. The purity of the “triple F” fraction was checked by IEF gel, shown in Fig. 9B. Finally, SEC was used to confirm the homogenous size of the “triple F” product (see Fig. 9C).

[0026] Figure 10. XENP11874, anti-CD19 Fab x anti-CD3 scFv “triple F” bispecific, potentially recruits T cells for B cell depletion.

[0027] Figure 11. The production of XENP11924, a “triple F” bispecific with an anti-CD19 Fab and anti-CD3 scFv. In Fig. 11A, the ion exchange purification of the desired “triple F” heterodimer from the undesired dual scFv-Fc and mAb homodimers is shown. The purity of the “triple F” fraction was checked by IEF gel, shown in Fig. 11B. Finally, SEC was used to confirm the homogenous size of the “triple F” product (see Fig. 11C).

[0028] Figure 12. XENP11924, anti-CD19 Fab x anti-CD3 scFv “triple F” bispecific, potentially recruits T cells for the killing of the Raji tumor cell line.

[0029] Figure 13. Amino acid sequences for the anti-CD38 Fab x anti-CD3 scFv “triple F” embodiment, showing the HC-Fab sequence, the HC-scFv sequence, and the light chain. Amino acid substitutions made to allow for efficient purification of the desired “triple F” heterodimer over the undesired dual scFv-Fc and mAb homodimers are underlined.

[0030] Figure 14. The production of XENP11925, a “triple F” bispecific with an anti-CD38 Fab and anti-CD3 scFv. In Fig. 14A, the ion exchange purification of the desired “triple F” heterodimer from the undesired dual scFv-Fc and mAb homodimers is shown. The purity of the “triple F” fraction was checked by IEF gel, shown in Fig. 14B. Finally, SEC was used to confirm the homogenous size of the “triple F” product (see Fig. 14C).

[0031] Figure 15. XENP11925, anti-CD38 Fab x anti-CD3 scFv “triple F” bispecific, potentially recruits T cells for the killing of the RPMI8226 tumor cell line.

[0032] Figure 16. Amino acid sequences for embodiments of the anti-CD19 Fab x anti-CD3 scFv “triple F” bispecific, showing the HC-Fab sequence, the HC-scFv sequence, and the light chain for each bispecific. Amino acid substitutions made to “skew” the formation toward desired “triple F” heterodimer over undesired dual scFv-Fc and mAb homodimers are underlined.

[0033] Figure 17. Isoelectric focusing (IEF) gel providing analysis of various “triple F” bispecifics. The desired “triple F” heterodimer has an apparent pI of 8.4 (middle band). Note the increase in heterodimer yield for XENP12152 and XENP12155.

[0034] Figure 18. Analytical ion exchange chromatography providing analysis of various “triple F” bispecifics. Yield of the desired “triple F” heterodimer is boxed. Note the increase in heterodimer yield for XENP12155.

[0035] Figure 19 shows the literature pIs of the 20 amino acids. It should be noted that the listed pIs are calculated as free amino acids; the actual pI of any side chain in the context of a protein is different, and thus this list is used to show pI trends and not absolute numbers for the purposes of the invention.

[0036] Figure 20 . Data table of exemplary pI-engineered variants listing:

XenP#	the internal reference number
Name (HC)	heavy chain sequence designation
SEQ ID NO (HC)	corresponding SEQ ID NO of the heavy chain sequence
Name (LC)	light chain sequence designation
SEQ ID NO (LC)	corresponding SEQ ID NO of the light chain sequence
Calc. pI	calculated pI value for the entire antibody sequence, including heavy and light chain Fv + constant domains, with the Fv of bevacizumab and the constant domains as defined in the table
#KR	number of Lys or Arg residues in IgG1 with the Fv of bevacizumab and the constant domains as defined in the table
Delta KR (vs. WT)	change in the number of Lys or Arg residues relative to IgG1 wild-type sequence of bevacizumab
#DE	number of Asp or Glu residues in IgG1 with the Fv of bevacizumab and the constant domains as defined in the table
Delta DE (vs. WT)	change in the number of Asp or Glu acid residues relative to IgG1 wild-type sequence of bevacizumab
Charge state	derived from the total number of Lys and Arg minus the total number of Asp and Glu residues, assuming a pH of 7
# HC Mutations vs IgG1	number of mutations in the heavy chain constant domain as compared to IgG1

# LC Mutations vs IgG1	number of mutations in the light chain constant domain as compared to IgG1
Total # of Mutations	total number of mutations in the heavy chain and light chain constant domains as compared to IgG1

[0037] It should be noted that Figure 20 has SEQ ID NO:s that are associated with the sequence listing filed in USSN 13/648,951, and are hereby expressly incorporated by reference.

[0001] Figure 21. depicts a number of pl variants that can additionally find use in the present invention, which is a list of all possible reduced pl variants created from isotypic substitutions of IgG1-4. Shown are the pl values for the three expected species as well as the average delta pl between the heterodimer and the two homodimer species present when the variant heavy chain is transfected with IgG1-WT heavy chain.

[0038] Figure 22 List of all possible increased pl variants created from isotypic substitutions of IgG1-4. Shown are the pl values for the three expected species as well as the average delta pl between the heterodimer and the two homodimer species present when the variant heavy chain is transfected with IgG1-WT heavy chain.

[0039] Figure 23 is an additional list of potential heterodimerization variants for use in the present invention.

[0040] Figure 24 depicts a matrix of possible combinations of heterodimerization formats, heterodimerization variants (separated into pl variants and steric variants (which includes charge pair variants), Fc variants, FcRn variants and combinations. Legend A are suitable FcRn variants: 434A, 434S, 428L, 308F, 259I, 428L/434S, 259I/308F, 436I/428L, 436I or V/434S, 436V/428L, 252Y, 252Y/254T/256E and 259I/308F/428L. That is, the Triple F format of Figure 1B can have any of these FcRn variants. For clarity, as each heavy chain is different, FcRn variants (as well as the Fc variants) can reside on one or both monomers. Legend B are suitable Fc variants: 236A, 239D, 239E, 332E, 332D, 239D/332E, 267D, 267E, 328F, 267E/328F, 236A/332E, 239D/332E/330Y, 239D, 332E/330L, 236R, 328R, 236R/328R, 236N/267E, 243L, 298A and 299T. (Note, additional suitable Fc variants are found in Figure 41 of US 2006/0024298, the figure and legend of which are hereby incorporated by reference in their entirety). Legend C are suitable pl variants, and these, for brevity are imported from Figure 23, again with the understanding that there is a "strandedness" to pl variants. Legend D are suitable steric variants (including charge pair variants); again, for brevity are imported from Figure 21, again with the understanding that there is a "strandedness" to steric variants. Legend E reflects the following possible combinations, again, with each variant being independently and optionally combined from

the appropriate source Legend: 1) pl variants plus FcRn variants; 2) pl variants plus Fc variants; 3) pl variants plus FcRn variants plus Fc variants; 4) steric variants plus FcRn variants; 5) steric variants plus Fc variants; 6) steric variants plus FcRn variants plus Fc variants; 7) pl variants plus steric variants plus FcRn variants; 8) pl variants plus steric variants plus Fc variants; 9) pl variants plus steric variants plus FcRn variants plus Fc variants; and 10) pl variants plus steric variants.

[0041] Figures 1-76 of USSN 61/593846 and the associated legends and discussion in the specification are hereby incorporated by reference.

[0042] Figure 2 and 17 of USSN 61/778,157, inclusive of all the sequences including the optimized CD3 sequences are expressly incorporated herein by reference.

DETAILED DESCRIPTION OF THE INVENTION

I. Overview

[0043] The present invention is directed to novel constructs to provide bispecific antibodies (or, as discussed below, trispecific or tetraspecific antibodies can also be made). An ongoing problem in antibody technologies is the desire for “bispecific” (and/or multispecific) antibodies that bind to two (or more) different antigens simultaneously, in general thus allowing the different antigens to be brought into proximity and resulting in new functionalities and new therapies. In general, these antibodies are made by including genes for each heavy and light chain into the host cells. This generally results in the formation of the desired heterodimer (A-B), as well as the two homodimers (A-A and B-B). However, a major obstacle in the formation of multispecific antibodies is the difficulty in purifying the heterodimeric antibodies away from the homodimeric antibodies and/or biasing the formation of the heterodimer over the formation of the homodimers.

[0044] The present invention is generally directed to the creation of heterodimeric proteins such as antibodies that can co-engage antigens in several ways, relying on amino acid variants in the constant regions that are different on each chain to promote heterodimeric formation and/or allow for ease of purification of heterodimers over the homodimers.

[0045] Thus, the present invention is directed to novel immunoglobulin compositions that co-engage at least a first and a second antigen. First and second antigens of the invention are herein referred to as antigen-1 and antigen-2 respectively. One heavy chain of the antibody contains an single chain Fv (“scFv”, as defined below) and the other heavy chain is a “regular” FAb format, comprising a variable heavy chain and a light chain. This structure is sometimes referred to herein as “triple F” format (scFv-FAb-Fc) or the “bottle-opener” format, due to a rough visual similarity to a bottle-opener (see Figure 1). The two chains are brought

together by the use of amino acid variants in the constant regions (e.g. the Fc domain and/or the hinge region) that promote the formation of heterodimeric antibodies as is described more fully below.

[0046] There are several distinct advantages to the present “triple F” format. As is known in the art, antibody analogs relying on two scFv constructs often have stability and aggregation problems, which can be alleviated in the present invention by the addition of a “regular” heavy and light chain pairing. In addition, as opposed to formats that rely on two heavy chains and two light chains, there is no issue with the incorrect pairing of heavy and light chains (e.g. heavy 1 pairing with light 2, etc.)

[0047] There are a number of mechanisms that can be used to generate the heterodimers of the present invention. In addition, as will be appreciated by those in the art and described more fully below, these mechanisms can be combined to ensure high heterodimerization.

[0048] One mechanism is generally referred to in the art as “knobs and holes” (“KIH”), referring to amino acid engineering that creates steric influences to favor heterodimeric formation and disfavor homodimeric formation can also optionally be used; this is sometimes referred to as “knobs and holes”, as described in USSN 61/596,846, Ridgway et al., Protein Engineering 9(7):617 (1996); Atwell et al., J. Mol. Biol. 1997 270:26; US Patent No. 8,216,805, all of which are hereby incorporated by reference in their entirety. Figure 4, further described below, identifies a number of “monomer A – monomer B” pairs that rely on “knobs and holes”. In addition, as described in Merchant et al., Nature Biotech. 16:677 (1998), these “knobs and hole” mutations can be combined with disulfide bonds to skew formation to heterodimerization. Some of these variants are shown in Figures 16, 20, 21 and 22.

[0002] An additional mechanism that finds use in the generation of heterodimers is sometimes referred to as “electrostatic steering” as described in Gunasekaran et al., J. Biol. Chem. 285(25):19637 (2010), hereby incorporated by reference in its entirety. This is sometimes referred to herein as “charge pairs”. In this embodiment, electrostatics are used to skew the formation towards heterodimerization. As those in the art will appreciate, these may also have an effect on pl, and thus on purification, and thus could in some cases also be considered pl variants. However, as these were generated to force heterodimerization and were not used as purification tools, they are classified as “steric variants”. These include, but are not limited to, D221E/P228E/L368E paired with D221R/P228R/K409R (e.g. these are “monomer corresponding sets) and C220E/P228E/368E paired with C220R/E224R/P228R/K409R.

[0049] In the present invention, there are two basic mechanisms that can lead to ease of purifying heterodimeric proteins; one relies on the use of pI variants, such that each monomer has a different pI, thus allowing the isoelectric purification of A-A, A-B and B-B dimeric proteins. Alternatively, the “triple F” format also allows separation on the basis of size. As is further outlined below, it is also possible to “skew” the formation of heterodimers over homodimers, as is generally outlined below.

[0050] In the present invention that utilizes pI as a separation mechanism to create the heterodimeric Triple F format, amino acid variants can be introduced into one or both of the monomer polypeptides; that is, the pI of one of the monomers (referred to herein for simplicity as “monomer A”) can be engineered away from monomer B, or both monomer A and B change be changed, with the pI of monomer A increasing and the pI of monomer B decreasing. As is outlined more fully below, the pI changes of either or both monomers can be done by removing or adding a charged residue (e.g. a neutral amino acid is replaced by a positively or negatively charged amino acid residue, e.g. glycine to glutamic acid), changing a charged residue from positive or negative to the opposite charge (aspartic acid to lysine) or changing a charged residue to a neutral residue (e.g. loss of a charge; lysine to serine). A number of these variants are shown in Figures 6, 20 and 21.

[0051] Accordingly, in this embodiment of the present invention provides for creating a sufficient change in pI in at least one of the monomers such that heterodimers can be separated from homodimers. As will be appreciated by those in the art, and as discussed further below, this can be done by using a “wild type” heavy chain constant region and a variant region that has been engineered to either increase or decrease its pI (wt A-+B or wt A - -B), or by increasing one region and decreasing the other region (A+ -B- or A- B+). It should be noted that in this discussion it does not matter which monomer comprises the scFv and which the Fab.

[0052] Thus, in general, a component of the present invention are amino acid variants in the constant regions of antibodies that are directed to altering the isoelectric point (pI) of at least one, if not both, of the monomers of a dimeric protein to form “pI heterodimers” (when the protein is an antibody, these are referred to as “pI antibodies”) by incorporating amino acid substitutions (“pI variants” or “pI substitutions”) into one or both of the monomers. As shown herein, the separation of the heterodimers from the two homodimers can be accomplished if the pIs of the two monomers differ by as little as 0.1 pH unit, with 0.2, 0.3, 0.4 and 0.5 or greater all finding use in the present invention.

[0053] Furthermore, as will be appreciated by those in the art and outlined herein, heterodimers can be separated from homodimers on the basis of size. For example, as

shown in Figure 1, heterodimers with two scFvs (Figure 1A) can be separated by those of the “triple F” format (figure 1B) and a bispecific mAb (Figure 1C). This can be further exploited in higher valency with additional antigen binding sites being utilized. For example, in as shown in Figure 2D, one monomer will have two Fab fragments and the other will have one scFv, resulting in a differential in size and thus molecular weight. Other examples are shown in Figures 2 and 3.

[0054] In addition, as will be appreciated by those in the art and outlined herein, the format outlined herein can be expanded to provide trispecific and tetraspecific antibodies as well. In this embodiment, some variations of which are depicted in Figures 2 and 3, it will be recognized that it is possible that some antigens are bound divalently (e.g. two antigen binding sites to a single antigen; for example, A and B could be part of a typical bivalent association and C and D can be optionally present and optionally the same or different). As will be appreciated, any combination of Fab and scFvs can be utilized to achieve the desired result and combinations.

[0055] In the case where pl variants are used to achieve heterodimerization, by using the constant region(s) of the heavy chain(s), a more modular approach to designing and purifying multispecific proteins, including antibodies, is provided. In addition, in some embodiments, the possibility of immunogenicity resulting from the pl variants is significantly reduced by importing pl variants from different IgG isotypes such that pl is changed without introducing significant immunogenicity. Thus, an additional problem to be solved is the elucidation of low pl constant domains with high human sequence content, e.g. the minimization or avoidance of non-human residues at any particular position.

[0056] In one embodiment, the heterodimeric antibody provides for monovalent engagement of one antigen using a scFv and monovalent engagement of the other antigen using a FAb. As outlined below, this format can also be varied; in some embodiments, there is monovalent engagement of three antigens, divalent engagement of one antigen and monovalent engagement of a second antigen (e.g. A and C are to the same antigen and B is to a different antigen), etc.

[0057] A side benefit that can occur with this pl engineering is also the extension of serum half-life and increased FcRn binding. That is, as described in USSN 13/194,904 (incorporated by reference in its entirety), lowering the pl of antibody constant domains (including those found in antibodies and Fc fusions) can lead to longer serum retention in vivo. These pl variants for increased serum half life also facilitate pl changes for purification.

[0058] In addition to all or part of a variant heavy constant domain, one or both of the monomers may contain one or two fusion partners, such that the heterodimers form multivalent proteins. As is generally depicted in the Figures, the fusion partners are depicted as A, B, C and D, with all combinations possible. In general, A, B, C and D are selected such that the heterodimer is at least bispecific or bivalent in its ability to interact with additional proteins. In the context of the present “triple F” format, generally A and B are an scFv and a Fv (as will be appreciated, either monomer can contain the scFv and the other the Fv/Fab) and then optionally one or two additional fusion partners.

[0059] Accordingly, the present invention provides novel constructs to produce multivalent antibodies.

[0060] Furthermore, as outlined herein, additional amino acid variants may be introduced into the bispecific antibodies of the invention, to add additional functionalities. For example, amino acid changes within the Fc region can be added (either to one monomer or both) to facilitate increased ADCC or CDC (e.g. altered binding to Fcγ receptors); to allow or increase yield of the addition of toxins and drugs (e.g. for ADC), as well as to increase binding to FcRn and/or increase serum half-life of the resulting molecules. As is further described herein and as will be appreciated by those in the art, any and all of the variants outlined herein can be optionally and independently combined with other variants.

II. Definitions

[0061] In order that the application may be more completely understood, several definitions are set forth below. Such definitions are meant to encompass grammatical equivalents.

[0062] By “ablation” herein is meant a decrease or removal of activity. Thus for example, “ablating FcγR binding” means the Fc region amino acid variant has less than 50% starting binding as compared to an Fc region not containing the specific variant, with less than 70-80-90-95-98% loss of activity being preferred, and in general, with the activity being below the level of detectable binding in a Biacore assay. Of particular use in the ablation of FcγR binding is the double variant 236R/328R.

[0063] By “ADCC” or “antibody dependent cell-mediated cytotoxicity” as used herein is meant the cell-mediated reaction wherein nonspecific cytotoxic cells that express FcγRs recognize bound antibody on a target cell and subsequently cause lysis of the target cell. ADCC is correlated with binding to FcγRIIIa; increased binding to FcγRIIIa leads to an increase in ADCC activity.

[0064] By “ADCP” or antibody dependent cell-mediated phagocytosis as used herein is meant the cell-mediated reaction wherein nonspecific cytotoxic cells that express FcγRs recognize bound antibody on a target cell and subsequently cause phagocytosis of the target cell.

[0065] By “modification” herein is meant an amino acid substitution, insertion, and/or deletion in a polypeptide sequence or an alteration to a moiety chemically linked to a protein. For example, a modification may be an altered carbohydrate or PEG structure attached to a protein. By “amino acid modification” herein is meant an amino acid substitution, insertion, and/or deletion in a polypeptide sequence. For clarity, unless otherwise noted, the amino acid modification is always to an amino acid coded for by DNA, e.g. the 20 amino acids that have codons in DNA and RNA.

[0066] By “amino acid substitution” or “substitution” herein is meant the replacement of an amino acid at a particular position in a parent polypeptide sequence with a different amino acid. In particular, in some embodiments, the substitution is to an amino acid that is not naturally occurring at the particular position, either not naturally occurring within the organism or in any organism. For example, the substitution E272Y refers to a variant polypeptide, in this case an Fc variant, in which the glutamic acid at position 272 is replaced with tyrosine. For clarity, a protein which has been engineered to change the nucleic acid coding sequence but not change the starting amino acid (for example exchanging CGG (encoding arginine) to CGA (still encoding arginine) to increase host organism expression levels) is not an “amino acid substitution”; that is, despite the creation of a new gene encoding the same protein, if the protein has the same amino acid at the particular position that it started with, it is not an amino acid substitution.

[0067] By “amino acid insertion” or “insertion” as used herein is meant the addition of an amino acid sequence at a particular position in a parent polypeptide sequence. For example, -233E or 233E designates an insertion of glutamic acid after position 233 and before position 234. Additionally, -233ADE or A233ADE designates an insertion of AlaAspGlu after position 233 and before position 234.

[0068] By “amino acid deletion” or “deletion” as used herein is meant the removal of an amino acid sequence at a particular position in a parent polypeptide sequence. For example, E233- or E233# designates a deletion of glutamic acid at position 233. Additionally, EDA233- or EDA233# designates a deletion of the sequence GluAspAla that begins at position 233.

[0069] By “variant protein” or “protein variant”, or “variant” as used herein is meant a protein that differs from that of a parent protein by virtue of at least one amino acid modification. Protein variant may refer to the protein itself, a composition comprising the

protein, or the amino sequence that encodes it. Preferably, the protein variant has at least one amino acid modification compared to the parent protein, e.g. from about one to about seventy amino acid modifications, and preferably from about one to about five amino acid modifications compared to the parent. As described below, in some embodiments the parent polypeptide, for example an Fc parent polypeptide, is a human wild type sequence, such as the Fc region from IgG1, IgG2, IgG3 or IgG4, although human sequences with variants can also serve as "parent polypeptides". The protein variant sequence herein will preferably possess at least about 80% identity with a parent protein sequence, and most preferably at least about 90% identity, more preferably at least about 95-98-99% identity. Variant protein can refer to the variant protein itself, compositions comprising the protein variant, or the DNA sequence that encodes it. Accordingly, by "antibody variant" or "variant antibody" as used herein is meant an antibody that differs from a parent antibody by virtue of at least one amino acid modification, "IgG variant" or "variant IgG" as used herein is meant an antibody that differs from a parent IgG (again, in many cases, from a human IgG sequence) by virtue of at least one amino acid modification, and "immunoglobulin variant" or "variant immunoglobulin" as used herein is meant an immunoglobulin sequence that differs from that of a parent immunoglobulin sequence by virtue of at least one amino acid modification. "Fc variant" or "variant Fc" as used herein is meant a protein comprising an amino acid modification in an Fc domain. The Fc variants of the present invention are defined according to the amino acid modifications that compose them. Thus, for example, N434S or 434S is an Fc variant with the substitution serine at position 434 relative to the parent Fc polypeptide, wherein the numbering is according to the EU index. Likewise, M428L/N434S defines an Fc variant with the substitutions M428L and N434S relative to the parent Fc polypeptide. The identity of the WT amino acid may be unspecified, in which case the aforementioned variant is referred to as 428L/434S. It is noted that the order in which substitutions are provided is arbitrary, that is to say that, for example, 428L/434S is the same Fc variant as M428L/N434S, and so on. For all positions discussed in the present invention that relate to antibodies, unless otherwise noted, amino acid position numbering is according to the EU index. The EU index or EU index as in Kabat or EU numbering scheme refers to the numbering of the EU antibody (Edelman et al., 1969, Proc Natl Acad Sci USA 63:78-85, hereby entirely incorporated by reference.) The modification can be an addition, deletion, or substitution. Substitutions can include naturally occurring amino acids and, in some cases, synthetic amino acids. Examples include U.S. Pat. No. 6,586,207; WO 98/48032; WO 03/073238; US2004-0214988A1; WO 05/35727A2; WO 05/74524A2; J. W. Chin et al., (2002), Journal of the American Chemical Society 124:9026-9027; J. W. Chin, & P. G. Schultz, (2002), ChemBioChem 11:1135-1137; J. W. Chin, et al., (2002), PICAS United

States of America 99:11020-11024; and, L. Wang, & P. G. Schultz, (2002), Chem. 1-10, all entirely incorporated by reference.

[0070] As used herein, “protein” herein is meant at least two covalently attached amino acids, which includes proteins, polypeptides, oligopeptides and peptides. The peptidyl group may comprise naturally occurring amino acids and peptide bonds, or synthetic peptidomimetic structures, i.e. “analogs”, such as peptoids (see Simon et al., PNAS USA 89(20):9367 (1992), entirely incorporated by reference). The amino acids may either be naturally occurring or synthetic (e.g. not an amino acid that is coded for by DNA); as will be appreciated by those in the art. For example, homo-phenylalanine, citrulline, ornithine and noreleucine are considered synthetic amino acids for the purposes of the invention, and both D- and L-(R or S) configured amino acids may be utilized. The variants of the present invention may comprise modifications that include the use of synthetic amino acids incorporated using, for example, the technologies developed by Schultz and colleagues, including but not limited to methods described by Cropp & Shultz, 2004, Trends Genet. 20(12):625-30, Anderson et al., 2004, Proc Natl Acad Sci USA 101 (2):7566-71, Zhang et al., 2003, 303(5656):371-3, and Chin et al., 2003, Science 301(5635):964-7, all entirely incorporated by reference. In addition, polypeptides may include synthetic derivatization of one or more side chains or termini, glycosylation, PEGylation, circular permutation, cyclization, linkers to other molecules, fusion to proteins or protein domains, and addition of peptide tags or labels.

[0071] By “residue” as used herein is meant a position in a protein and its associated amino acid identity. For example, Asparagine 297 (also referred to as Asn297 or N297) is a residue at position 297 in the human antibody IgG1.

[0072] By “Fab” or “Fab region” as used herein is meant the polypeptide that comprises the VH, CH1, VL, and CL immunoglobulin domains. Fab may refer to this region in isolation, or this region in the context of a full length antibody, antibody fragment or Fab fusion protein. By “Fv” or “Fv fragment” or “Fv region” as used herein is meant a polypeptide that comprises the VL and VH domains of a single antibody.

[0073] By “IgG subclass modification” or “isotype modification” as used herein is meant an amino acid modification that converts one amino acid of one IgG isotype to the corresponding amino acid in a different, aligned IgG isotype. For example, because IgG1 comprises a tyrosine and IgG2 a phenylalanine at EU position 296, a F296Y substitution in IgG2 is considered an IgG subclass modification.

[0074] By “non-naturally occurring modification” as used herein is meant an amino acid modification that is not isotypic. For example, because none of the IgGs comprise a serine at

position 434, the substitution 434S in IgG1, IgG2, IgG3, or IgG4 (or hybrids thereof) is considered a non-naturally occurring modification.

[0075] By “amino acid” and “amino acid identity” as used herein is meant one of the 20 naturally occurring amino acids that are coded for by DNA and RNA.

[0076] By “effector function” as used herein is meant a biochemical event that results from the interaction of an antibody Fc region with an Fc receptor or ligand. Effector functions include but are not limited to ADCC, ADCP, and CDC.

[0077] By “IgG Fc ligand” as used herein is meant a molecule, preferably a polypeptide, from any organism that binds to the Fc region of an IgG antibody to form an Fc/Fc ligand complex. Fc ligands include but are not limited to FcγRIs, FcγRIIs, FcγRIIIa, FcRn, C1q, C3, mannan binding lectin, mannose receptor, staphylococcal protein A, streptococcal protein G, and viral FcγR. Fc ligands also include Fc receptor homologs (FcRH), which are a family of Fc receptors that are homologous to the FcγRs (Davis et al., 2002, Immunological Reviews 190:123-136, entirely incorporated by reference). Fc ligands may include undiscovered molecules that bind Fc. Particular IgG Fc ligands are FcRn and Fc gamma receptors. By “Fc ligand” as used herein is meant a molecule, preferably a polypeptide, from any organism that binds to the Fc region of an antibody to form an Fc/Fc ligand complex.

[0078] By “Fc gamma receptor”, “FcγR” or “FcγgammaR” as used herein is meant any member of the family of proteins that bind the IgG antibody Fc region and is encoded by an FcγR gene. In humans this family includes but is not limited to FcγRI (CD64), including isoforms FcγRIa, FcγRIb, and FcγRIc; FcγRII (CD32), including isoforms FcγRIIa (including allotypes H131 and R131), FcγRIIb (including FcγRIIb-1 and FcγRIIb-2), and FcγRIIc; and FcγRIII (CD16), including isoforms FcγRIIIa (including allotypes V158 and F158) and FcγRIIIb (including allotypes FcγRIIb-NA1 and FcγRIIb-NA2) (Jefferis et al., 2002, Immunol Lett 82:57-65, entirely incorporated by reference), as well as any undiscovered human FcγRs or FcγR isoforms or allotypes. An FcγR may be from any organism, including but not limited to humans, mice, rats, rabbits, and monkeys. Mouse FcγRs include but are not limited to FcγRI (CD64), FcγRII (CD32), FcγRIII (CD16), and FcγRIII-2 (CD16-2), as well as any undiscovered mouse FcγRs or FcγR isoforms or allotypes.

[0079] By “FcRn” or “neonatal Fc Receptor” as used herein is meant a protein that binds the IgG antibody Fc region and is encoded at least in part by an FcRn gene. The FcRn may be from any organism, including but not limited to humans, mice, rats, rabbits, and monkeys. As is known in the art, the functional FcRn protein comprises two polypeptides, often referred to as the heavy chain and light chain. The light chain is beta-2-microglobulin and the heavy chain is encoded by the FcRn gene. Unless otherwise noted herein, FcRn or an FcRn

protein refers to the complex of FcRn heavy chain with beta-2-microglobulin. A variety of FcRn variants used to increase binding to the FcRn receptor, and in some cases, to increase serum half-life, are discussed in Legend A of Figure 24.

[0080] By “parent polypeptide” as used herein is meant a starting polypeptide that is subsequently modified to generate a variant. The parent polypeptide may be a naturally occurring polypeptide, or a variant or engineered version of a naturally occurring polypeptide. Parent polypeptide may refer to the polypeptide itself, compositions that comprise the parent polypeptide, or the amino acid sequence that encodes it. Accordingly, by “parent immunoglobulin” as used herein is meant an unmodified immunoglobulin polypeptide that is modified to generate a variant, and by “parent antibody” as used herein is meant an unmodified antibody that is modified to generate a variant antibody. It should be noted that “parent antibody” includes known commercial, recombinantly produced antibodies as outlined below.

[0081] By “Fc fusion protein” or “immunoadhesin” herein is meant a protein comprising an Fc region, generally linked (optionally through a linker moiety, as described herein) to a different protein, such as a binding moiety to a target protein, as described herein)..

[0082] By “position” as used herein is meant a location in the sequence of a protein. Positions may be numbered sequentially, or according to an established format, for example the EU index for antibody numbering.

[0083] By “strandedness” in the context of the monomers of the heterodimeric proteins of the invention herein is meant that, similar to the two strands of DNA that “match”, heterodimerization variants are incorporated into each monomer so as to preserve the ability to “match” to form heterodimers. For example, if some pl variants are engineered into monomer A (e.g. making the pl higher) then steric variants that are “charge pairs” that can be utilized as well do not interfere with the pl variants, e.g. the charge variants that make a pl higher are put on the same “strand” or “monomer” to preserve both functionalities.

[0084] By “target antigen” as used herein is meant the molecule that is bound specifically by the variable region of a given antibody. A target antigen may be a protein, carbohydrate, lipid, or other chemical compound. A wide number of suitable target antigens are described below.

[0085] By “target cell” as used herein is meant a cell that expresses a target antigen.

[0086] By “variable region” as used herein is meant the region of an immunoglobulin that comprises one or more Ig domains substantially encoded by any of the V.kappa., V.lamda.,

and/or VH genes that make up the kappa, lambda, and heavy chain immunoglobulin genetic loci respectively.

[0087] By “wild type or WT” herein is meant an amino acid sequence or a nucleotide sequence that is found in nature, including allelic variations. A WT protein has an amino acid sequence or a nucleotide sequence that has not been intentionally modified.

Heterodimeric Proteins

[0088] The present invention is directed to the generation of multispecific, particularly bispecific binding proteins, and in particular, multispecific antibodies that have one monomer comprising an scFv and the other an Fv.

Antibody

[0089] The present invention relates to the generation of multispecific antibodies, generally therapeutic antibodies. As is discussed below, the term “antibody” is used generally. Antibodies that find use in the present invention can take on a number of formats as described herein, including traditional antibodies as well as antibody derivatives, fragments and mimetics, described below. In general, the term “antibody” includes any polypeptide that includes at least one constant domain, including, but not limited to, CH1, CH2, CH3 and CL.

[0090] Traditional antibody structural units typically comprise a tetramer. Each tetramer is typically composed of two identical pairs of polypeptide chains, each pair having one “light” (typically having a molecular weight of about 25 kDa) and one “heavy” chain (typically having a molecular weight of about 50-70 kDa). Human light chains are classified as kappa and lambda light chains. The present invention is directed to the IgG class, which has several subclasses, including, but not limited to IgG1, IgG2, IgG3, and IgG4. Thus, “isotype” as used herein is meant any of the subclasses of immunoglobulins defined by the chemical and antigenic characteristics of their constant regions. It should be understood that therapeutic antibodies can also comprise hybrids of isotypes and/or subclasses. For example, as shown in US Publication 2009/0163699, incorporated by reference, the present invention covers pl engineering of IgG1/G2 hybrids.

[0091] The amino-terminal portion of each chain includes a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition, generally referred to in the art and herein as the “Fv domain” or “Fv region”. In the variable region, three loops are gathered for each of the V domains of the heavy chain and light chain to form an antigen-binding site. Each of the loops is referred to as a complementarity-determining region (hereinafter referred to as a “CDR”), in which the variation in the amino acid sequence is most significant. “Variable” refers to the fact that certain segments of the variable region

differ extensively in sequence among antibodies. Variability within the variable region is not evenly distributed. Instead, the V regions consist of relatively invariant stretches called framework regions (FRs) of 15-30 amino acids separated by shorter regions of extreme variability called "hypervariable regions" that are each 9-15 amino acids long or longer.

[0092] Each VH and VL is composed of three hypervariable regions ("complementary determining regions," "CDRs") and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1-CDR1-FR2-CDR2-FR3-CDR3-FR4.

[0093] The hypervariable region generally encompasses amino acid residues from about amino acid residues 24-34 (LCDR1; "L" denotes light chain), 50-56 (LCDR2) and 89-97 (LCDR3) in the light chain variable region and around about 31-35B (HCDR1; "H" denotes heavy chain), 50-65 (HCDR2), and 95-102 (HCDR3) in the heavy chain variable region; Kabat et al., SEQUENCES OF PROTEINS OF IMMUNOLOGICAL INTEREST, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, Md. (1991) and/or those residues forming a hypervariable loop (e.g. residues 26-32 (LCDR1), 50-52 (LCDR2) and 91-96 (LCDR3) in the light chain variable region and 26-32 (HCDR1), 53-55 (HCDR2) and 96-101 (HCDR3) in the heavy chain variable region; Chothia and Lesk (1987) J. Mol. Biol. 196:901-917. Specific CDRs of the invention are described below.

[0094] Throughout the present specification, the Kabat numbering system is generally used when referring to a residue in the variable domain (approximately, residues 1-107 of the light chain variable region and residues 1-113 of the heavy chain variable region) (e.g., Kabat et al., supra (1991)).

[0095] The CDRs contribute to the formation of the antigen-binding, or more specifically, epitope binding site of antibodies. "Epitope" refers to a determinant that interacts with a specific antigen binding site in the variable region of an antibody molecule known as a paratope. Epitopes are groupings of molecules such as amino acids or sugar side chains and usually have specific structural characteristics, as well as specific charge characteristics. A single antigen may have more than one epitope.

[0096] The epitope may comprise amino acid residues directly involved in the binding (also called immunodominant component of the epitope) and other amino acid residues, which are not directly involved in the binding, such as amino acid residues which are effectively blocked by the specifically antigen binding peptide; in other words, the amino acid residue is within the footprint of the specifically antigen binding peptide.

[0097] Epitopes may be either conformational or linear. A conformational epitope is produced by spatially juxtaposed amino acids from different segments of the linear polypeptide chain. A linear epitope is one produced by adjacent amino acid residues in a

polypeptide chain. Conformational and nonconformational epitopes may be distinguished in that the binding to the former but not the latter is lost in the presence of denaturing solvents.

[0098] An epitope typically includes at least 3, and more usually, at least 5 or 8-10 amino acids in a unique spatial conformation. Antibodies that recognize the same epitope can be verified in a simple immunoassay showing the ability of one antibody to block the binding of another antibody to a target antigen, for example “binning.”

[0099] The carboxy-terminal portion of each chain defines a constant region primarily responsible for effector function. Kabat et al. collected numerous primary sequences of the variable regions of heavy chains and light chains. Based on the degree of conservation of the sequences, they classified individual primary sequences into the CDR and the framework and made a list thereof (see SEQUENCES OF IMMUNOLOGICAL INTEREST, 5th edition, NIH publication, No. 91-3242, E.A. Kabat et al., entirely incorporated by reference).

[0100] In the IgG subclass of immunoglobulins, there are several immunoglobulin domains in the heavy chain. By “immunoglobulin (Ig) domain” herein is meant a region of an immunoglobulin having a distinct tertiary structure. Of interest in the present invention are the heavy chain domains, including, the constant heavy (CH) domains and the hinge domains. In the context of IgG antibodies, the IgG isotypes each have three CH regions. Accordingly, “CH” domains in the context of IgG are as follows: “CH1” refers to positions 118-220 according to the EU index as in Kabat. “CH2” refers to positions 237-340 according to the EU index as in Kabat, and “CH3” refers to positions 341-447 according to the EU index as in Kabat. As shown herein and described below, the pl variants can be in one or more of the CH regions, as well as the hinge region, discussed below.

[0101] It should be noted that the sequences depicted herein start at the CH1 region, position 118; the variable regions are not included except as noted. For example, the first amino acid of SEQ ID NO: 2, while designated as position “1” in the sequence listing, corresponds to position 118 of the CH1 region, according to EU numbering.

[0102] Another type of Ig domain of the heavy chain is the hinge region. By “hinge” or “hinge region” or “antibody hinge region” or “immunoglobulin hinge region” herein is meant the flexible polypeptide comprising the amino acids between the first and second constant domains of an antibody. Structurally, the IgG CH1 domain ends at EU position 220, and the IgG CH2 domain begins at residue EU position 237. Thus for IgG the antibody hinge is herein defined to include positions 221 (D221 in IgG1) to 236 (G236 in IgG1), wherein the numbering is according to the EU index as in Kabat. In some embodiments, for example in the context of an Fc region, the lower hinge is included, with the “lower hinge” generally

referring to positions 226 or 230. As noted herein, pl variants can be made in the hinge region as well.

[0103] The light chain generally comprises two domains, the variable light domain (containing the light chain CDRs and together with the variable heavy domains forming the Fv region), and a constant light chain region (often referred to as CL or Ck).

[0104] Another region of interest for additional substitutions, outlined below, is the Fc region. By “Fc” or “Fc region” or “Fc domain” as used herein is meant the polypeptide comprising the constant region of an antibody excluding the first constant region immunoglobulin domain and in some cases, part of the hinge. Thus Fc refers to the last two constant region immunoglobulin domains of IgA, IgD, and IgG, the last three constant region immunoglobulin domains of IgE and IgM, and the flexible hinge N-terminal to these domains. For IgA and IgM, Fc may include the J chain. For IgG, the Fc domain comprises immunoglobulin domains Cy2 and Cy3 (Cy2 and Cy3) and the lower hinge region between Cy1 (Cy1) and Cy2 (Cy2). Although the boundaries of the Fc region may vary, the human IgG heavy chain Fc region is usually defined to include residues C226 or P230 to its carboxyl-terminus, wherein the numbering is according to the EU index as in Kabat. In some embodiments, as is more fully described below, amino acid modifications are made to the Fc region, for example to alter binding to one or more FcγR receptors or to the FcRn receptor.

[0105] In some embodiments, the antibodies are full length. By “full length antibody” herein is meant the structure that constitutes the natural biological form of an antibody, including variable and constant regions, including one or more modifications as outlined herein.

[0106] Alternatively, the antibodies can include a variety of structures, including, but not limited to, antibody fragments,, monoclonal antibodies, bispecific antibodies, minibodies, domain antibodies, synthetic antibodies (sometimes referred to herein as “antibody mimetics”), chimeric antibodies, humanized antibodies, antibody fusions (sometimes referred to as “antibody conjugates”), and fragments of each, respectively.

[0107] In one embodiment, the antibody is an antibody fragment, as long as it contains at least one constant domain which can be engineered to produce heterodimers, such as pl engineering. Other antibody fragments that can be used include fragments that contain one or more of the CH1, CH2, CH3, hinge and CL domains of the invention that have been pl engineered. For example, Fc fusions are fusions of the Fc region (CH2 and CH3, optionally with the hinge region) fused to another protein. A number of Fc fusions are known the art and can be improved by the addition of the heterodimerization variants of the invention. In the present case, antibody fusions can be made comprising CH1; CH1, CH2 and CH3; CH2;

CH3; CH2 and CH3; CH1 and CH3, any or all of which can be made optionally with the hinge region, utilizing any combination of heterodimerization variants described herein.

[0108] In some embodiments of the present invention, one monomer comprises a heavy chain comprising a scFV linked to an Fc domain, and the other monomer comprises a heavy chain comprising a Fab linked to an Fc domain, e.g. a “typical” heavy chain, and a light chain. By “Fab” or “Fab region” as used herein is meant the polypeptide that comprises the VH, CH1, VL, and CL immunoglobulin domains. Fab may refer to this region in isolation, or this region in the context of a full length antibody, antibody fragment or Fab fusion protein. By “Fv” or “Fv fragment” or “Fv region” as used herein is meant a polypeptide that comprises the VL and VH domains of a single antibody.

Chimeric and Humanized Antibodies

[0109] In some embodiments, the antibody can be a mixture from different species, e.g. a chimeric antibody and/or a humanized antibody. In general, both “chimeric antibodies” and “humanized antibodies” refer to antibodies that combine regions from more than one species. For example, “chimeric antibodies” traditionally comprise variable region(s) from a mouse (or rat, in some cases) and the constant region(s) from a human. “Humanized antibodies” generally refer to non-human antibodies that have had the variable-domain framework regions swapped for sequences found in human antibodies. Generally, in a humanized antibody, the entire antibody, except the CDRs, is encoded by a polynucleotide of human origin or is identical to such an antibody except within its CDRs. The CDRs, some or all of which are encoded by nucleic acids originating in a non-human organism, are grafted into the beta-sheet framework of a human antibody variable region to create an antibody, the specificity of which is determined by the engrafted CDRs. The creation of such antibodies is described in, e.g., WO 92/11018, Jones, 1986, Nature 321:522-525, Verhoeyen et al., 1988, Science 239:1534-1536, all entirely incorporated by reference. “Backmutation” of selected acceptor framework residues to the corresponding donor residues is often required to regain affinity that is lost in the initial grafted construct (US 5530101; US 5585089; US 5693761; US 5693762; US 6180370; US 5859205; US 5821337; US 6054297; US 6407213, all entirely incorporated by reference). The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region, typically that of a human immunoglobulin, and thus will typically comprise a human Fc region. Humanized antibodies can also be generated using mice with a genetically engineered immune system. Roque et al., 2004, Biotechnol. Prog. 20:639-654, entirely incorporated by reference. A variety of techniques and methods for humanizing and reshaping non-human antibodies are well known in the art (See Tsurushita & Vasquez, 2004, Humanization of Monoclonal Antibodies, Molecular Biology of B Cells, 533-545, Elsevier Science (USA), and references

cited therein, all entirely incorporated by reference). Humanization methods include but are not limited to methods described in Jones et al., 1986, Nature 321:522-525; Riechmann et al., 1988; Nature 332:323-329; Verhoeven et al., 1988, Science, 239:1534-1536; Queen et al., 1989, Proc Natl Acad Sci, USA 86:10029-33; He et al., 1998, J. Immunol. 160: 1029-1035; Carter et al., 1992, Proc Natl Acad Sci USA 89:4285-9, Presta et al., 1997, Cancer Res. 57(20):4593-9; Gorman et al., 1991, Proc. Natl. Acad. Sci. USA 88:4181-4185; O'Connor et al., 1998, Protein Eng 11:321-8, all entirely incorporated by reference. Humanization or other methods of reducing the immunogenicity of nonhuman antibody variable regions may include resurfacing methods, as described for example in Roguska et al., 1994, Proc. Natl. Acad. Sci. USA 91:969-973, entirely incorporated by reference. In one embodiment, the parent antibody has been affinity matured, as is known in the art. Structure-based methods may be employed for humanization and affinity maturation, for example as described in USSN 11/004,590. Selection based methods may be employed to humanize and/or affinity mature antibody variable regions, including but not limited to methods described in Wu et al., 1999, J. Mol. Biol. 294:151-162; Baca et al., 1997, J. Biol. Chem. 272(16):10678-10684; Rosok et al., 1996, J. Biol. Chem. 271(37): 22611-22618; Rader et al., 1998, Proc. Natl. Acad. Sci. USA 95: 8910-8915; Krauss et al., 2003, Protein Engineering 16(10):753-759, all entirely incorporated by reference. Other humanization methods may involve the grafting of only parts of the CDRs, including but not limited to methods described in USSN 09/810,510; Tan et al., 2002, J. Immunol. 169:1119-1125; De Pascalis et al., 2002, J. Immunol. 169:3076-3084, all entirely incorporated by reference.

Multispecific Antibody Constructs

[0110] As will be appreciated by those in the art and discussed more fully below, the heterodimeric fusion proteins of the present invention take on a number variety of configurations, with a preferred embodiment shown in Figure 1B as a “triple F” construct.

Heterodimeric Heavy Chain Constant Regions

[0111] Accordingly, the present invention provides heterodimeric proteins based on the use of monomers containing variant heavy chain constant regions as a first domain. By “monomer” herein is meant one half of the heterodimeric protein. It should be noted that traditional antibodies are actually tetrameric (two heavy chains and two light chains). In the context of the present invention, one pair of heavy-light chains (if applicable, e.g. if the monomer comprises an Fab) is considered a “monomer”. Similarly, a heavy chain region comprising the scFv is considered a monomer. In the case where an Fv region is one fusion partner (e.g. heavy and light chain) and a non-antibody protein is another fusion partner, each “half” is considered a monomer. Essentially, each monomer comprises sufficient heavy chain constant region to allow heterodimerization engineering, whether that be all the

constant region, e.g. CH1-hinge-CH2-CH3, the Fc region (CH2-CH3), or just the CH3 domain.

[0112] The variant heavy chain constant regions can comprise all or part of the heavy chain constant region, including the full length construct, CH1-hinge-CH2-CH3, or portions thereof, including for example CH2-CH3 or CH3 alone. In addition, the heavy chain region of each monomer can be the same backbone (CH1-hinge-CH2-CH3 or CH2-CH3) or different. N- and C-terminal truncations and additions are also included within the definition; for example, some pl variants include the addition of charged amino acids to the C-terminus of the heavy chain domain.

[0113] Thus, in general, one monomer of the present “triple F” construct is a scFv region-hinge-Fc domain) and the other is (VH-CH1-hinge- CH2-CH3 plus associated light chain), with heterodimerization variants, including steric and pl variants, Fc and FcRn variants, and additional antigen binding domains (with optional linkers) included in these regions.

[0114] In addition to the heterodimerization variants (e.g. steric and pl variants) outlined herein, the heavy chain regions may also contain additional amino acid substitutions, including changes for altering FcγR and FcRn binding as discussed below.

[0115] In addition, some monomers can utilize linkers between the variant heavy chain constant region and the fusion partner. For the scFv portion of the “bottle-opener”, standard linkers as are known in the art can be used. In the case where additional fusion partners are made (e.g. Figures 2 and 3), traditional peptide linkers can be used, including flexible linkers of glycine and serine. In some cases, the linkers for use as components of the monomer are different from those defined below for the ADC constructs, and are in many embodiments not cleavable linkers (such as those susceptible to proteases), although cleavable linkers may find use in some embodiments.

[0116] The heterodimerization variants include a number of different types of variants, including, but not limited to, steric variants (including charge variants) and pl variants, that can be optionally and independently combined with any other variants. In these embodiments, it is important to match “monomer A” with “monomer B”; that is, if a heterodimeric protein relies on both steric variants and pl variants, these need to be correctly matched to each monomer: e.g. the set of steric variants that work (1 set on monomer A, 1 set on monomer B) is combined with pl variant sets (1 set on monomer A, 1 set on monomer B), such that the variants on each monomer are designed to achieve the desired function.

[0117] It is important to note that the heterodimerization variants outlined herein (for example, including but not limited to those variants shown in Figures 4, 5 and 6), can be optionally and independently combined with any other variants, and on any other monomer.

Thus, for example, pl variants for monomer 1 from Figure 6 can be added to other heterodimerization variants for monomer 1 in Figures 5 and 6 or from monomer 2. That is, what is important for the heterodimerization is that there are “sets” of variants, one set for one monomer and one set for the other. Whether these are combined from the Figures 1 to 1 (e.g. monomer 1 listings can go together) or switched (monomer 1 pl variants with monomer 2 steric variants) is irrelevant. However, as noted herein, “strandedness” should be preserved when combinations are made as outlined above. Furthermore, for the additional Fc variants (such as for FcγR binding, FcRn binding, etc.), either monomer, or both monomers, can include any of the listed variants, independently and optionally. In some cases, both monomers have the additional variants and in some only one monomer has the additional variants, or they can be combined.

Heterodimerization Variants

[0118] The present invention provides multispecific antibody formats, on a “triple F” or “bottle opener” scaffold as depicted in Figure 1B and Figures 2 and 3.

Steric Variants

[0119] In some embodiments, the formation of heterodimers can be facilitated by the addition of steric variants. That is, by changing amino acids in each heavy chain, different heavy chains are more likely to associate to form the heterodimeric structure than to form homodimers with the same Fc amino acid sequences. Suitable steric variants are shown in Figures 4 and 5.

[0120] One mechanism is generally referred to in the art as “knobs and holes”, referring to amino acid engineering that creates steric influences to favor heterodimeric formation and disfavor homodimeric formation can also optionally be used; this is sometimes referred to as “knobs and holes”, as described in USSN 61/596,846, Ridgway et al., Protein Engineering 9(7):617 (1996); Atwell et al., J. Mol. Biol. 1997 270:26; US Patent No. 8,216,805, all of which are hereby incorporated by reference in their entirety. Figures 4 and 5, further described below, identifies a number of “monomer A – monomer B” pairs that rely on “knobs and holes”. In addition, as described in Merchant et al., Nature Biotech. 16:677 (1998), these “knobs and hole” mutations can be combined with disulfide bonds to skew formation to heterodimerization.

[0121] An additional mechanism that finds use in the generation of heterodimers is sometimes referred to as “electrostatic steering” as described in Gunasekaran et al., J. Biol. Chem. 285(25):19637 (2010), hereby incorporated by reference in its entirety. This is sometimes referred to herein as “charge pairs”. In this embodiment, electrostatics are used to skew the formation towards heterodimerization. As those in the art will appreciate, these

may also have have an effect on pI, and thus on purification, and thus could in some cases also be considered pI variants. However, as these were generated to force heterodimerization and were not used as purification tools, they are classified as “steric variants”. These include, but are not limited to, D221E/P228E/L368E paired with D221R/P228R/K409R (e.g. these are “monomer corresponding sets) and C220E/P228E/368E paired with C220R/E224R/P228R/K409R.

[0122] Additional monomer A and monomer B variants that can be combined with other variants, optionally and independently in any amount, such as pI variants outlined herein or other steric variants that are shown in Figure 37 of US 2012/0149876, the figure and legend of which are incorporated expressly by reference herein.

[0123] In some embodiments, the steric variants outlined herein can be optionally and independently incorporated with any pI variant (or other variants such as Fc variants, FcRn variants, etc.) into one or both monomers.

pI (Isoelectric point) Variants for Heterodimers

[0124] In general, as will be appreciated by those in the art, there are two general categories of pI variants: those that increase the pI of the protein (basic changes) and those that decrease the pI of the protein (acidic changes). As described herein, all combinations of these variants can be done: one monomer may be wild type, or a variant that does not display a significantly different pI from wild-type, and the other can be either more basic or more acidic. Alternatively, each monomer is changed, one to more basic and one to more acidic.

[0125] Preferred combinations of pI variants are shown in Figure 6, with additional pI variants disclosed in Figures 20, 21 and 22.

Heavy Chain Acidic pI Changes

[0126] Accordingly, when one monomer comprising a variant heavy chain constant domain is to be made more positive (e.g. lower the pI), one or more of the following substitutions can be made: S119E, K133E, K133Q, T164E, K205E, K205Q, N208D, K210E, K210Q, K274E, K320E, K322E, K326E, K334E, R355E, K392E, a deletion of K447, adding peptide DEDE at the c-terminus, G137E, N203D, K274Q, R355Q, K392N and Q419E. As outlined herein and shown in the figures, these changes are shown relative to IgG1, but all isotypes can be altered this way, as well as isotype hybrids.

[0127] In the case where the heavy chain constant domain is from IgG2-4, R133E and R133Q can also be used.

Basic pI changes

[0128] Accordingly, when one monomer comprising a variant heavy chain constant domain is to be made more negative (e.g. increase the pI), one or more of the following substitutions can be made: Q196K, P217R, P228R, N276K and H435R. As outlined herein and shown in the figures, these changes are shown relative to IgG1, but all isotypes can be altered this way, as well as isotype hybrids.

Antibody Heterodimers Light chain variants

[0129] In the case of antibody based heterodimers, e.g. where at least one of the monomers comprises a light chain in addition to the heavy chain domain, pI variants can also be made in the light chain. Amino acid substitutions for lowering the pI of the light chain include, but are not limited to, K126E, K126Q, K145E, K145Q, N152D, S156E, K169E, S202E, K207E and adding peptide DEDE at the c-terminus of the light chain. Changes in this category based on the constant lambda light chain include one or more substitutions at R108Q, Q124E, K126Q, N138D, K145T and Q199E. In addition, increasing the pI of the light chains can also be done.

Isotypic Variants

[0130] In addition, many embodiments of the invention rely on the “importation” of pI amino acids at particular positions from one IgG isotype into another, thus reducing or eliminating the possibility of unwanted immunogenicity being introduced into the variants. That is, IgG1 is a common isotype for therapeutic antibodies for a variety of reasons, including high effector function. However, the heavy constant region of IgG1 has a higher pI than that of IgG2 (8.10 versus 7.31). By introducing IgG2 residues at particular positions into the IgG1 backbone, the pI of the resulting monomer is lowered (or increased) and additionally exhibits longer serum half-life. For example, IgG1 has a glycine (pI 5.97) at position 137, and IgG2 has a glutamic acid (pI 3.22); importing the glutamic acid will affect the pI of the resulting protein. As is described below, a number of amino acid substitutions are generally required to significantly affect the pI of the variant antibody. However, it should be noted as discussed below that even changes in IgG2 molecules allow for increased serum half-life.

[0131] In other embodiments, non-isotypic amino acid changes are made, either to reduce the overall charge state of the resulting protein (e.g. by changing a higher pI amino acid to a lower pI amino acid), or to allow accommodations in structure for stability, etc. as is more further described below.

[0132] In addition, by pI engineering both the heavy and light constant domains, significant changes in each monomer of the heterodimer can be seen. As discussed herein, having

the pIs of the two monomers differ by at least 0.5 can allow separation by ion exchange chromatography or isoelectric focusing, or other methods sensitive to isoelectric point.

Calculating pI

[0133] The pI of each monomer can depend on the pI of the variant heavy chain constant domain and the pI of the total monomer, including the variant heavy chain constant domain and the fusion partner. Thus, in some embodiments, the change in pI is calculated on the basis of the variant heavy chain constant domain, using the chart in the Figures.

Alternatively, the pI of each monomer can be compared.

pI Variants that also confer better FcRn *in vivo* binding

[0134] In the case where the pI variant decreases the pI of the monomer, they can have the added benefit of improving serum retention *in vivo*.

[0135] Although still under examination, Fc regions are believed to have longer half-lives *in vivo*, because binding to FcRn at pH 6 in an endosome sequesters the Fc (Ghetie and Ward, 1997 Immunol Today. 18(12): 592-598, entirely incorporated by reference). The endosomal compartment then recycles the Fc to the cell surface. Once the compartment opens to the extracellular space, the higher pH, ~7.4, induces the release of Fc back into the blood. In mice, Dall'Acqua et al. showed that Fc mutants with increased FcRn binding at pH 6 and pH 7.4 actually had reduced serum concentrations and the same half life as wild-type Fc (Dall'Acqua et al. 2002, J. Immunol. 169:5171-5180, entirely incorporated by reference). The increased affinity of Fc for FcRn at pH 7.4 is thought to forbid the release of the Fc back into the blood. Therefore, the Fc mutations that will increase Fc's half-life *in vivo* will ideally increase FcRn binding at the lower pH while still allowing release of Fc at higher pH. The amino acid histidine changes its charge state in the pH range of 6.0 to 7.4. Therefore, it is not surprising to find His residues at important positions in the Fc/FcRn complex.

[0136] Recently it has been suggested that antibodies with variable regions that have lower isoelectric points may also have longer serum half-lives (Igawa et al., 2010 PEDS. 23(5): 385-392, entirely incorporated by reference). However, the mechanism of this is still poorly understood. Moreover, variable regions differ from antibody to antibody. Constant region variants with reduced pI and extended half-life would provide a more modular approach to improving the pharmacokinetic properties of antibodies, as described herein.

[0137] pI variants that find use in this embodiment, as well as their use for purification optimization, are disclosed in Figure 20.

Combination of Heterodimeric Variants

[0138] As will be appreciated by those in the art, all of the recited heterodimerization variants can be optionally and independently combined in any way, as long as they retain

their “strandedness” or “monomer partition”. In addition, all of these variants can be combined into any of the heterodimerization formats. See Figure 24.

[0139] In the case of pl variants, while embodiments finding particular use are shown in the Figures, other combinations can be generated, following the basic rule of altering the pl difference between two monomers to facilitate purification.

Target Antigens

[0140] The immunoglobulins of the invention may target virtually any antigens. The “triple F” format is particularly beneficial for targeting two (or more) distinct antigens. (As outlined herein, this targeting can be any combination of monovalent and divalent binding, depending on the format). Thus the immunoglobulins herein preferably co-engage two target antigens, although in some cases, three or four antigens can be monovalently engaged. Each monomer’s specificity can be selected from the lists below. While the triple F immunoglobulins described herein are particularly beneficial for targeting distinct antigens, in some cases it may be beneficial to target only one antigen. That is, each monomer may have specificity for the same antigen.

[0141] Particular suitable applications of the immunoglobulins herein are co-target pairs for which it is beneficial or critical to engage each target antigen monovalently. Such antigens may be, for example, immune receptors that are activated upon immune complexation. Cellular activation of many immune receptors occurs only by cross-linking, achieved typically by antibody/antigen immune complexes, or via effector cell to target cell engagement. For some immune receptors, for example the CD3 signaling receptor on T cells, activation only upon engagement with co-engaged target is critical, as nonspecific cross-linking in a clinical setting can elicit a cytokine storm and toxicity. Therapeutically, by engaging such antigens monovalently rather than multivalently, using the immunoglobulins herein, such activation occurs only in response to cross-linking only in the microenvironment of the primary target antigen. The ability to target two different antigens with different valencies is a novel and useful aspect of the present invention. Examples of target antigens for which it may be therapeutically beneficial or necessary to co-engage monovalently include but are not limited to immune activating receptors such as CD3, FcγRs, toll-like receptors (TLRs) such as TLR4 and TLR9, cytokine, chemokine, cytokine receptors, and chemokine receptors. In many embodiments, one of the antigen binding sites binds to CD3, and in some embodiments it is the scFv-containing monomer.

[0142] Virtually any antigen may be targeted by the immunoglobulins herein, including but not limited to proteins, subunits, domains, motifs, and/or epitopes belonging to the following list of target antigens, which includes both soluble factors such as cytokines and membrane-

bound factors, including transmembrane receptors: 17-1A, 4-1BB, 4Dc, 6-keto-PGF1a, 8-iso-PGF2a, 8-oxo-dG, A1 Adenosine Receptor, A33, ACE, ACE-2, Activin, Activin A, Activin AB, Activin B, Activin C, Activin RIA, Activin RIA ALK-2, Activin RIB ALK-4, Activin RIIA, Activin RIIB, ADAM, ADAM10, ADAM12, ADAM15, ADAM17/TACE, ADAM8, ADAM9, ADAMTS, ADAMTS4, ADAMTS5, Addressins, aFGF, ALCAM, ALK, ALK-1, ALK-7, alpha-1-antitrypsin, alpha-V/beta-1 antagonist, ANG, Ang, APAF-1, APE, APJ, APP, APRIL, AR, ARC, ART, Artemin, anti-Id, ASPARTIC, Atrial natriuretic factor, av/b3 integrin, Axl, b2M, B7-1, B7-2, B7-H, B-lymphocyte Stimulator (BlyS), BACE, BACE-1, Bad, BAFF, BAFF-R, Bag-1, BAK, Bax, BCA-1, BCAM, Bcl, BCMA, BDNF, b-ECGF, bFGF, BID, Bik, BIM, BLC, BL-CAM, BLK, BMP, BMP-2 BMP-2a, BMP-3 Osteogenin, BMP-4 BMP-2b, BMP-5, BMP-6 Vgr-1, BMP-7 (OP-1), BMP-8 (BMP-8a, OP-2), BMPR, BMPR-IA (ALK-3), BMPR-IB (ALK-6), BRK-2, RPK-1, BMPR-II (BRK-3), BMPs, b-NGF, BOK, Bombesin, Bone-derived neurotrophic factor, BPDE, BPDE-DNA, BTC, complement factor 3 (C3), C3a, C4, C5, C5a, C10, CA125, CAD-8, Calcitonin, cAMP, carcinoembryonic antigen (CEA), carcinoma-associated antigen, Cathepsin A, Cathepsin B, Cathepsin C/DPPI, Cathepsin D, Cathepsin E, Cathepsin H, Cathepsin L, Cathepsin O, Cathepsin S, Cathepsin V, Cathepsin X/Z/P, CBL, CCL, CCK2, CCL, CCL1, CCL11, CCL12, CCL13, CCL14, CCL15, CCL16, CCL17, CCL18, CCL19, CCL2, CCL20, CCL21, CCL22, CCL23, CCL24, CCL25, CCL26, CCL27, CCL28, CCL3, CCL4, CCL5, CCL6, CCL7, CCL8, CCL9/10, CCR, CCR1, CCR10, CCR10, CCR2, CCR3, CCR4, CCR5, CCR6, CCR7, CCR8, CCR9, CD1, CD2, CD3, CD3E, CD4, CD5, CD6, CD7, CD8, CD10, CD11a, CD11b, CD11c, CD13, CD14, CD15, CD16, CD18, CD19, CD20, CD21, CD22, CD23, CD25, CD27L, CD28, CD29, CD30, CD30L, CD32, CD33 (p67 proteins), CD34, CD38, CD40, CD40L, CD44, CD45, CD46, CD49a, CD52, CD54, CD55, CD56, CD61, CD64, CD66e, CD74, CD80 (B7-1), CD89, CD95, CD123, CD137, CD138, CD140a, CD146, CD147, CD148, CD152, CD164, CEACAM5, CFTR, cGMP, CINC, Clostridium botulinum toxin, Clostridium perfringens toxin, CKb8-1, CLC, CMV, CMV UL, CNTF, CNTN-1, COX, C-Ret, CRG-2, CT-1, CTACK, CTGF, CTLA-4, CX3CL1, CX3CR1, CXCL, CXCL1, CXCL2, CXCL3, CXCL4, CXCL5, CXCL6, CXCL7, CXCL8, CXCL9, CXCL10, CXCL11, CXCL12, CXCL13, CXCL14, CXCL15, CXCL16, CXCR, CXCR1, CXCR2, CXCR3, CXCR4, CXCR5, CXCR6, cytokeratin tumor-associated antigen, DAN, DCC, DcR3, DC-SIGN, Decay accelerating factor, des(1-3)-IGF-I (brain IGF-1), Dhh, digoxin, DNAM-1, Dnase, Dpp, DPPIV/CD26, Dtk, ECAD, EDA, EDA-A1, EDA-A2, EDAR, EGF, EGFR (ErbB-1), EMA, EMMPRIN, ENA, endothelin receptor, Enkephalinase, eNOS, Eot, eotaxin1, EpCAM, Ephrin B2/ EphB4, EPO, ERCC, E-selectin, ET-1, Factor IIa, Factor VII, Factor VIIIc, Factor IX, fibroblast activation protein (FAP), Fas, FcR1, FEN-1, Ferritin, FGF, FGF-19, FGF-2, FGF3, FGF-8, FGFR, FGFR-3, Fibrin, FL, FLIP, Flt-3, Flt-4, Follicle stimulating hormone, Fractalkine, FZD1, FZD2, FZD3, FZD4, FZD5, FZD6, FZD7, FZD8,

FZD9, FZD10, G250, Gas 6, GCP-2, GCSF, GD2, GD3, GDF, GDF-1, GDF-3 (Vgr-2), GDF-5 (BMP-14, CDMP-1), GDF-6 (BMP-13, CDMP-2), GDF-7 (BMP-12, CDMP-3), GDF-8 (Myostatin), GDF-9, GDF-15 (MIC-1), GDNF, GDNF, GFAP, GFRa-1, GFR-alpha1, GFR-alpha2, GFR-alpha3, GTR, Glucagon, Glut 4, glycoprotein IIb/IIIa (GP IIb/IIIa), GM-CSF, gp130, gp72, GRO, Growth hormone releasing factor, Hapten (NP-cap or NIP-cap), HB-EGF, HCC, HCMV gB envelope glycoprotein, HCMV) gH envelope glycoprotein, HCMV UL, Hemopoietic growth factor (HGF), Hep B gp120, heparanase, Her2, Her2/neu (ErbB-2), Her3 (ErbB-3), Her4 (ErbB-4), herpes simplex virus (HSV) gB glycoprotein, HSV gD glycoprotein, HGFA, High molecular weight melanoma-associated antigen (HMW-MAA), HIV gp120, HIV IIIB gp 120 V3 loop, HLA, HLA-DR, HM1.24, HMFG PEM, HRG, Hrk, human cardiac myosin, human cytomegalovirus (HCMV), human growth hormone (HGH), HVEM, I-309, IAP, ICAM, ICAM-1, ICAM-3, ICE, ICOS, IFNg, Ig, IgA receptor, IgE, IGF, IGF binding proteins, IGF-1R, IGFBP, IGF-I, IGF-II, IL, IL-1, IL-1R, IL-2, IL-2R, IL-4, IL-4R, IL-5, IL-5R, IL-6, IL-6R, IL-8, IL-9, IL-10, IL-12, IL-13, IL-15, IL-18, IL-18R, IL-23, interferon (INF)-alpha, INF-beta, INF-gamma, Inhibin, iNOS, Insulin A-chain, Insulin B-chain, Insulin-like growth factor 1, integrin alpha2, integrin alpha3, integrin alpha4, integrin alpha4/beta1, integrin alpha4/beta7, integrin alpha5 (alphaV), integrin alpha5/beta1, integrin alpha5/beta3, integrin alpha6, integrin beta1, integrin beta2, interferon gamma, IP-10, I-TAC, JE, Kallikrein 2, Kallikrein 5, Kallikrein 6, , Kallikrein 11, Kallikrein 12, Kallikrein 14, Kallikrein 15, Kallikrein L1, Kallikrein L2, Kallikrein L3, Kallikrein L4, KC, KDR, Keratinocyte Growth Factor (KGF), laminin 5, LAMP, LAP, LAP (TGF- 1), Latent TGF-1, Latent TGF-1 bp1, LBP, LDGF, LECT2, Lefty, Lewis-Y antigen, Lewis-Y related antigen, LFA-1, LFA-3, Lfo, LIF, LIGHT, lipoproteins, LIX, LKN, Lptn, L-Selectin, LT-a, LT-b, LTB4, LTBP-1, Lung surfactant, Luteinizing hormone, Lymphotoxin Beta Receptor, Mac-1, MAdCAM, MAG, MAP2, MARC, MCAM, MCAM, MCK-2, MCP, M-CSF, MDC, Mer, METALLOPROTEASES, MGDF receptor, MGMT, MHC (HLA-DR), MIF, MIG, MIP, MIP-1-alpha, MK, MMAC1, MMP, MMP-1, MMP-10, MMP-11, MMP-12, MMP-13, MMP-14, MMP-15, MMP-2, MMP-24, MMP-3, MMP-7, MMP-8, MMP-9, MPIF, Mpo, MSK, MSP, mucin (Muc1), MUC18, Muellierian-inhibitin substance, Mug, MuSK, NAIP, NAP, NCAD, N-Cadherin, NCA 90, NCAM, NCAM, Neprilysin, Neurotrophin-3,-4, or -6, Neurturin, Neuronal growth factor (NGF), NGFR, NGF-beta, nNOS, NO, NOS, Npn, NRG-3, NT, NTN, OB, OGG1, OPG, OPN, OSM, OX40L, OX40R, p150, p95, PADPr, Parathyroid hormone, PARC, PARP, PBR, PBSF, PCAD, P-Cadherin, PCNA, PDGF, PDGF, PDK-1, PECAM, PEM, PF4, PGE, PGF, PGI2, PGJ2, PIN, PLA2, placental alkaline phosphatase (PLAP), PIGF, PLP, PP14, Proinsulin, Prorelaxin, Protein C, PS, PSA, PSCA, prostate specific membrane antigen (PSMA), PTEN, PTHrp, Ptk, PTN, R51, RANK, RANKL, RANTES, RANTES, Relaxin A-chain, Relaxin B-chain, renin, respiratory syncytial virus (RSV) F, RSV Fgp, Ret, Rheumatoid factors, RLIP76, RPA2, RSK, S100, SCF/KL, SDF-1,

SERINE, Serum albumin, sFRP-3, Shh, SIGIRR, SK-1, SLAM, SLPI, SMAC, SMDF, SMOH, SOD, SPARC, Stat, STEAP, STEAP-II, TACE, TACI, TAG-72 (tumor-associated glycoprotein-72), TARC, TCA-3, T-cell receptors (e.g., T-cell receptor alpha/beta), TdT, TECK, TEM1, TEM5, TEM7, TEM8, TERT, testicular PLAP-like alkaline phosphatase, TfR, TGF, TGF-alpha, TGF-beta, TGF-beta Pan Specific, TGF-beta RI (ALK-5), TGF-beta RII, TGF-beta RIIB, TGF-beta RIIL, TGF-beta1, TGF-beta2, TGF-beta3, TGF-beta4, TGF-beta5, Thrombin, Thymus Ck-1, Thyroid stimulating hormone, Tie, TIMP, TIQ, Tissue Factor, TMEFF2, Tmpo, TMPRSS2, TNF, TNF-alpha, TNF-alpha beta, TNF-beta2, TNFc, TNF-RI, TNF-RII, TNFRSF10A (TRAIL R1 Apo-2, DR4), TNFRSF10B (TRAIL R2 DR5, KILLER, TRICK-2A, TRICK-B), TNFRSF10C (TRAIL R3 DcR1, LIT, TRID), TNFRSF10D (TRAIL R4 DcR2, TRUND), TNFRSF11A (RANK ODF R, TRANCE R), TNFRSF11B (OPG OCIF, TR1), TNFRSF12 (TWEAK R FN14), TNFRSF13B (TACI), TNFRSF13C (BAFF R), TNFRSF14 (HVEM ATAR, HveA, LIGHT R, TR2), TNFRSF16 (NGFR p75NTR), TNFRSF17 (BCMA), TNFRSF18 (GITR AITR), TNFRSF19 (TROY TAJ, TRADE), TNFRSF19L (RELT), TNFRSF1A (TNF RI CD120a, p55-60), TNFRSF1B (TNF RII CD120b, p75-80), TNFRSF26 (TNFRH3), TNFRSF3 (LTbR TNF RIIL, TNFC R), TNFRSF4 (OX40 ACT35, TXGP1 R), TNFRSF5 (CD40 p50), TNFRSF6 (Fas Apo-1, APT1, CD95), TNFRSF6B (DcR3 M68, TR6), TNFRSF7 (CD27), TNFRSF8 (CD30), TNFRSF9 (4-1BB CD137, ILA), TNFRSF21 (DR6), TNFRSF22 (DcTRAIL R2 TNFRH2), TNFRST23 (DcTRAIL R1 TNFRH1), TNFRSF25 (DR3 Apo-3, LARD, TR-3, TRAMP, WSL-1), TNFSF10 (TRAIL Apo-2 Ligand, TL2), TNFSF11 (TRANCE/RANK Ligand ODF, OPG Ligand), TNFSF12 (TWEAK Apo-3 Ligand, DR3 Ligand), TNFSF13 (APRIL TALL2), TNFSF13B (BAFF BLYS, TALL1, THANK, TNFSF20), TNFSF14 (LIGHT HVEM Ligand, LTg), TNFSF15 (TL1A/VEGI), TNFSF18 (GITR Ligand AITR Ligand, TL6), TNFSF1A (TNF-a Conectin, DIF, TNFSF2), TNFSF1B (TNF-b LTa, TNFSF1), TNFSF3 (LTb TNFC, p33), TNFSF4 (OX40 Ligand gp34, TXGP1), TNFSF5 (CD40 Ligand CD154, gp39, HIGM1, IMD3, TRAP), TNFSF6 (Fas Ligand Apo-1 Ligand, APT1 Ligand), TNFSF7 (CD27 Ligand CD70), TNFSF8 (CD30 Ligand CD153), TNFSF9 (4-1BB Ligand CD137 Ligand), TP-1, t-PA, Tpo, TRAIL, TRAIL R, TRAIL-R1, TRAIL-R2, TRANCE, transferring receptor, TRF, Trk, TROP-2, TSG, TSLP, tumor-associated antigen CA 125, tumor-associated antigen expressing Lewis Y related carbohydrate, TWEAK, TXB2, Ung, uPAR, uPAR-1, Urokinase, VCAM, VCAM-1, VECAD, VE-Cadherin, VE-cadherin-2, VEGFR-1 (flt-1), VEGF, VEGFR, VEGFR-3 (flt-4), VEGI, VIM, Viral antigens, VLA, VLA-1, VLA-4, VNR integrin, von Willebrands factor, WIF-1, WNT1, WNT2, WNT2B/13, WNT3, WNT3A, WNT4, WNT5A, WNT5B, WNT6, WNT7A, WNT7B, WNT8A, WNT8B, WNT9A, WNT9A, WNT9B, WNT10A, WNT10B, WNT11, WNT16, XCL1, XCL2, XCR1, XCR1, XEDAR, XIAP, XPD, and receptors for hormones and growth factors. To form the bispecific or trispecific antibodies of the invention, antibodies to any combination of these antigens can

be made; that is, each of these antigens can be optionally and independently included or excluded from a multispecific antibody according to the present invention.

[0143] Exemplary antigens that may be targeted specifically by the immunoglobulins of the invention include but are not limited to: CD20, CD19, Her2, EGFR, EpCAM, CD3, FcγRIIIa (CD16), FcγRIIa (CD32a), FcγRIIb (CD32b), FcγRI (CD64), Toll-like receptors (TLRs) such as TLR4 and TLR9, cytokines such as IL-2, IL-5, IL-13, IL-12, IL-23, and TNFα, cytokine receptors such as IL-2R, chemokines, chemokine receptors, growth factors such as VEGF and HGF, and the like. . To form the multispecific antibodies of the invention, antibodies to any combination of these antigens can be made; that is, each of these antigens can be optionally and independently included or excluded from a multispecific antibody according to the present invention.

[0144] Particularly preferred combinations for bispecific antibodies are an antigen-binding domain to CD3 and an antigen binding domain to CD19; an antigen-binding domain to CD3 and an antigen binding domain to CD33; an antigen-binding domain to CD3 and an antigen binding domain to CD 38. Again, in many embodiments, the CD3 binding domain is the scFv, having an exemplary sequence as depicted in the Figures and/or CD3 CDRs as outlined.

[0145] The choice of suitable target antigens and co-targets depends on the desired therapeutic application. Some targets that have proven especially amenable to antibody therapy are those with signaling functions. Other therapeutic antibodies exert their effects by blocking signaling of the receptor by inhibiting the binding between a receptor and its cognate ligand. Another mechanism of action of therapeutic antibodies is to cause receptor down regulation. Other antibodies do not work by signaling through their target antigen. The choice of co-targets will depend on the detailed biology underlying the pathology of the indication that is being treated.

[0146] Monoclonal antibody therapy has emerged as an important therapeutic modality for cancer (Weiner et al., 2010, Nature Reviews Immunology 10:317-327; Reichert et al., 2005, Nature Biotechnology 23[9]:1073-1078; herein expressly incorporated by reference). For anti-cancer treatment it may be desirable to target one antigen (antigen-1) whose expression is restricted to the cancerous cells while co-targeting a second antigen (antigen-2) that mediates some immunological killing activity. For other treatments it may be beneficial to co-target two antigens, for example two angiogenic factors or two growth factors, that are each known to play some role in proliferation of the tumor. Exemplary co-targets for oncology include but are not limited to HGF and VEGF, IGF-1R and VEGF, Her2 and VEGF, CD19 and CD3, CD20 and CD3, Her2 and CD3, CD19 and FcγRIIIa, CD20 and FcγRIIIa, Her2 and

FcγRIIIa. An immunoglobulin of the invention may be capable of binding VEGF and phosphatidylserine; VEGF and ErbB3; VEGF and PLGF; VEGF and ROBO4; VEGF and BSG2; VEGF and CDCP1; VEGF and ANPEP; VEGF and c-MET; HER-2 and ERB3; HER-2 and BSG2; HER-2 and CDCP1; HER-2 and ANPEP; EGFR and CD64; EGFR and BSG2; EGFR and CDCP1; EGFR and ANPEP; IGF1R and PDGFR; IGF1R and VEGF; IGF1R and CD20; CD20 and CD74; CD20 and CD30; CD20 and DR4; CD20 and VEGFR2; CD20 and CD52; CD20 and CD4; HGF and c-MET; HGF and NRP1; HGF and phosphatidylserine; ErbB3 and IGF1R; ErbB3 and IGF1,2; c-Met and Her-2; c-Met and NRP1; c-Met and IGF1R; IGF1,2 and PDGFR; IGF1,2 and CD20; IGF1,2 and IGF1R; IGF2 and EGFR; IGF2 and HER2; IGF2 and CD20; IGF2 and VEGF; IGF2 and IGF1R; IGF1 and IGF2; PDGFRa and VEGFR2; PDGFRa and PLGF; PDGFRa and VEGF; PDGFRa and c-Met; PDGFRa and EGFR; PDGFRb and VEGFR2; PDGFRb and c-Met; PDGFRb and EGFR; RON and c-Met; RON and MTSP1; RON and MSP; RON and CDCP1; VGFR1 and PLGF; VGFR1 and RON; VGFR1 and EGFR; VEGFR2 and PLGF; VEGFR2 and NRP1; VEGFR2 and RON; VEGFR2 and DLL4; VEGFR2 and EGFR; VEGFR2 and ROBO4; VEGFR2 and CD55; LPA and S1P; EPHB2 and RON; CTLA4 and VEGF; CD3 and EPCAM; CD40 and IL6; CD40 and IGF; CD40 and CD56; CD40 and CD70; CD40 and VEGFR1; CD40 and DR5; CD40 and DR4; CD40 and APRIL; CD40 and BCMA; CD40 and RANKL; CD28 and MAPG; CD80 and CD40; CD80 and CD30; CD80 and CD33; CD80 and CD74; CD80 and CD2; CD80 and CD3; CD80 and CD19; CD80 and CD4; CD80 and CD52; CD80 and VEGF; CD80 and DR5; CD80 and VEGFR2; CD22 and CD20; CD22 and CD80; CD22 and CD40; CD22 and CD23; CD22 and CD33; CD22 and CD74; CD22 and CD19; CD22 and DR5; CD22 and DR4; CD22 and VEGF; CD22 and CD52; CD30 and CD20; CD30 and CD22; CD30 and CD23; CD30 and CD40; CD30 and VEGF; CD30 and CD74; CD30 and CD19; CD30 and DR5; CD30 and DR4; CD30 and VEGFR2; CD30 and CD52; CD30 and CD4; CD138 and RANKL; CD33 and FTL3; CD33 and VEGF; CD33 and VEGFR2; CD33 and CD44; CD33 and DR4; CD33 and DR5; DR4 and CD137; DR4 and IGF1,2; DR4 and IGF1R; DR4 and DR5; DR5 and CD40; DR5 and CD137; DR5 and CD20; DR5 and EGFR; DR5 and IGF1,2; DR5 and IGFR, DR5 and HER-2, and EGFR and DLL4. Other target combinations include one or more members of the EGF/erb-2/erb-3 family.

[0147] Other targets (one or more) involved in oncological diseases that the immunoglobulins herein may bind include, but are not limited to those selected from the group consisting of: CD52, CD20, CD19, CD3, CD4, CD8, BMP6, IL12A, IL1A, IL1B, 1L2, IL24, INHA, TNF, TNFSF10, BMP6, EGF, FGF1, FGF10, FGF11, FGF12, FGF13, FGF14, FGF16, FGF17, FGF18, FGF19, FGF2, FGF20, FGF21, FGF22, FGF23, FGF3, FGF4, FGF5, FGF6, FGF7, FGF8, FGF9, GRP, IGF1, IGF2, IL12A, IL1A, IL1B, 1L2, INHA, TGFA,

TGFB1, TGFB2, TGFB3, VEGF, CDK2, FGF10, FGF18, FGF2, FGF4, FGF7, IGF1R, IL2, BCL2, CD164, CDKN1A, CDKN1B, CDKN1C, CDKN2A, CDKN2B, CDKN2C, CDKN3, GNRH1, IGFBP6, IL1A, IL1B, ODZ1, PAWR, PLG, TGFB11, AR, BRCA1, CDK3, CDK4, CDK5, CDK6, CDK7, CDK9, E2F1, EGFR, ENO1, ERBB2, ESR1, ESR2, IGFBP3, IGFBP6, IL2, INSL4, MYC, NOX5, NR6A1, PAP, PCNA, PRKCQ, PRKD1, PRL, TP53, FGF22, FGF23, FGF9, IGFBP3, IL2, INHA, KLK6, TP53, CHGB, GNRH1, IGF1, IGF2, INHA, INSL3, INSL4, PRL, KLK6, SHBG, NR1D1, NR1H3, NR1I3, NR2F6, NR4A3, ESR1, ESR2, NR0B1, NR0B2, NR1D2, NR1H2, NR1H4, NR112, NR2C1, NR2C2, NR2E1, NR2E3, NR2F1, NR2F2, NR3C1, NR3C2, NR4A1, NR4A2, NR5A1, NR5A2, NR6 μ l, PGR, RARB, FGF1, FGF2, FGF6, KLK3, KRT1, APOC1, BRCA1, CHGA, CHGB, CLU, COL1A1, COL6A1, EGF, ERBB2, ERK8, FGF1, FGF10, FGF11, FGF13, FGF14, FGF16, FGF17, FGF18, FGF2, FGF20, FGF21, FGF22, FGF23, FGF3, FGF4, FGF5, FGF6, FGF7, FGF8, FGF9, GNRH1, IGF1, IGF2, IGFBP3, IGFBP6, IL12A, IL1A, IL1B, IL2, IL24, INHA, INSL3, INSL4, KLK10, KLK12, KLK13, KLK14, KLK15, KLK3, KLK4, KLK5, KLK6, KLK9, MMP2, MMP9, MSMB, NTN4, ODZ1, PAP, PLAUI, PRL, PSAP, SERPINA3, SHBG, TGFA, TIMP3, CD44, CDH1, CDH10, CDH19, CDH20, CDH7, CDH9, CDH1, CDH10, CDH13, CDH18, CDH19, CDH20, CDH7, CDH8, CDH9, ROBO2, CD44, ILK, ITGA1, APC, CD164, COL6A1, MTSS1, PAP, TGFB11, AGR2, AIG1, AKAP1, AKAP2, CANT1, CAV1, CDH12, CLDN3, CLN3, CYB5, CYC1, DAB2IP, DES, DNCL1, ELAC2, ENO2, ENO3, FASN, FLJ12584, FLJ25530, GAGEB1, GAGEC1, GGT1, GSTP1, HIP1, HUMCYT2A, IL29, K6HF, KAI1, KRT2A, MIB1, PART1, PATE, PCA3, PIAS2, PIK3CG, PPID, PR1, PSCA, SLC2A2, SLC33 μ l, SLC43 μ l, STEAP, STEAP2, TPM1, TPM2, TRPC6, ANGPT1, ANGPT2, ANPEP, ECGF1, EREG, FGF1, FGF2, FIGF, FLT1, JAG1, KDR, LAMA5, NRP1, NRP2, PGF, PLXDC1, STAB 1, VEGF, VEGFC, ANGPTL3, BAI1, COL4A3, IL8, LAMA5, NRP1, NRP2, STAB 1, ANGPTL4, PECAM1, PF4, PROK2, SERPINF1, TNFAIP2, CCL11, CCL2, CXCL1, CXCL10, CXCL3, CXCL5, CXCL6, CXCL9, IFNA1, IFNB1, IFNG, IL1B, IL6, MDK, EDG1, EFNA1, EFNA3, EFNB2, EGF, EPHB4, FGFR3, HGF, IGF1, ITGB3, PDGFA, TEK, TGFA, TGFB1, TGFB2, TGFB1, CCL2, CDH5, COL1A1, EDG1, ENG, ITGAV, ITGB3, THBS1, THBS2, BAD, BAG1, BCL2, CCNA1, CCNA2, CCND1, CCNE1, CCNE2, CDH1 (E-cadherin), CDKN1B (p27Kip1), CDKN2A (p16INK4a), COL6A1, CTNNB1 (b-catenin), CTSB (cathepsin B), ERBB2 (Her-2), ESR1, ESR2, F3 (TF), FOSL1 (FRA-1), GATA3, GSN (Gelsolin), IGFBP2, IL2RA, IL6, IL6R, IL6ST (glycoprotein 130), ITGA6 (a6 integrin), JUN, KLK5, KRT19, MAP2K7 (c-Jun), MKI67 (Ki-67), NGFB (GF), NGFR, NME1 (M23A), PGR, PLAUI (uPA), PTEN, SERPINB5 (maspin), SERPINE1 (PAI-1), TGFA, THBS1 (thrombospondin-1), TIE (Tie-1), TNFRSF6 (Fas), TNFSF6 (FasL), TOP2A (topoisomerase IIA), TP53, AZGP1 (zinc-a-glycoprotein), BPAG1 (plectin), CDKN1A (p21Wap1/Cip1), CLDN7 (claudin-7), CLU (clusterin), ERBB2 (Her-2), FGF1, FLRT1 (fibronectin), GABRP (GABAa), GNAS1, ID2,

ITGA6 (a6 integrin), ITGB4 (b 4 integrin), KLF5 (GC Box BP), KRT19 (Keratin 19), KRTHB6 (hair-specific type II keratin), MACMARCKS, MT3 (metallothionein-III), MUC1 (mucin), PTGS2 (COX-2), RAC2 (p21Rac2), S100A2, SCGB1D2 (lipophilin B), SCGB2A1 (mammaglobin 2), SCGB2A2 (mammaglobin 1), SPRR1B (Spr1), THBS1, THBS2, THBS4, and TNFAIP2 (B94), RON, c-Met, CD64, DLL4, PLGF, CTLA4, phosphatidylserine, ROBO4, CD80, CD22, CD40, CD23, CD28, CD80, CD55, CD38, CD70, CD74, CD30, CD138, CD56, CD33, CD2, CD137, DR4, DR5, RANKL, VEGFR2, PDGFR, VEGFR1, MTSP1, MSP, EPHB2, EPHA1, EPHA2, EpCAM, PGE2, NKG2D, LPA, SIP, APRIL, BCMA, MAPG, FLT3, PDGFR alpha, PDGFR beta, ROR1, PSMA, PSCA, SCD1, and CD59. To form the bispecific or trispecific antibodies of the invention, antibodies to any combination of these antigens can be made; that is, each of these antigens can be optionally and independently included or excluded from a multispecific antibody according to the present invention.

[0148] Monoclonal antibody therapy has become an important therapeutic modality for treating autoimmune and inflammatory disorders (Chan & Carter, 2010, Nature Reviews Immunology 10:301-316; Reichert et al., 2005, Nature Biotechnology 23[9]:1073-1078; herein expressly incorporated by reference). Many proteins have been implicated in general autoimmune and inflammatory responses, and thus may be targeted by the immunoglobulins of the invention. Autoimmune and inflammatory targets include but are not limited to C5, CCL1 (I-309), CCL11 (eotaxin), CCL13 (mcp-4), CCL15 (MIP-1d), CCL16 (HCC-4), CCL17 (TARC), CCL18 (PARC), CCL19, CCL2 (mcp-1), CCL20 (MIP-3a), CCL21 (MIP-2), CCL23 (MPIF-1), CCL24 (MPIF-2/eotaxin-2), CCL25 (TECK), CCL26, CCL3 (MIP-1a), CCL4 (MIP-1b), CCL5 (RANTES), CCL7 (mcp-3), CCL8 (mcp-2), CXCL1, CXCL10 (IP-10), CXCL11 (1-TAC/IP-9), CXCL12 (SDF1), CXCL13, CXCL14, CXCL2, CXCL3, CXCL5 (ENA-78/LIX), CXCL6 (GCP-2), CXCL9, IL13, IL8, CCL13 (mcp-4), CCR1, CCR2, CCR3, CCR4, CCR5, CCR6, CCR7, CCR8, CCR9, CX3CR1, IL8RA, XCR1 (CCXCR1), IFNA2, IL10, IL13, IL17C, IL1A, IL1B, IL1F10, IL1F5, IL1F6, IL1F7, IL1F8, IL1F9, IL22, IL5, IL8, IL9, LTA, LTB, MIF, SCYE1 (endothelial Monocyte-activating cytokine), SPP1, TNF, TNFSF5, IFNA2, IL10RA, IL10RB, IL13, IL13RA1, IL5RA, IL9, IL9R, ABCF1, BCL6, C3, C4A, CEBPB, CRP, ICEBERG, IL1R1, IL1RN, IL8RB, LTB4R, TOLLIP, FADD, IRAK1, IRAK2, MYD88, NCK2, TNFAIP3, TRADD, TRAF1, TRAF2, TRAF3, TRAF4, TRAF5, TRAF6, ACVR1, ACVR1B, ACVR2, ACVR2B, ACVRL1, CD28, CD3E, CD3G, CD3Z, CD69, CD80, CD86, CNR1, CTLA4, CYSLTR1, FCER1A, FCER2, FCGR3A, GPR44, HAVCR2, OPRD1, P2RX7, TLR2, TLR3, TLR4, TLR5, TLR6, TLR7, TLR8, TLR9, TLR10, BLR1, CCL1, CCL2, CCL3, CCL4, CCL5, CCL7, CCL8, CCL11, CCL13, CCL15, CCL16, CCL17, CCL18, CCL19, CCL20, CCL21, CCL22, CCL23, CCL24, CCL25, CCR1, CCR2, CCR3, CCR4, CCR5, CCR6, CCR7, CCR8, CCR9, CX3CL1, CX3CR1, CXCL1, CXCL2, CXCL3, CXCL5, CXCL6, CXCL10,

CXCL11, CXCL12, CXCL13, CXCR4, GPR2, SCYE1, SDF2, XCL1, XCL2, XCR1, AMH, AMHR2, BMPR1A, BMPR1B, BMPR2, C19orf10 (IL27w), CER1, CSF1, CSF2, CSF3, DKFZp451J0118, FGF2, GFI1, IFNA1, IFNB1, IFNG, IGF1, IL1A, IL1B, IL1R1, IL1R2, IL2, IL2RA, IL2RB, IL2RG, IL3, IL4, IL4R, IL5, IL5RA, IL6, IL6R, IL6ST, IL7, IL8, IL8RA, IL8RB, IL9, IL9R, IL10, IL10RA, IL10RB, IL11, IL12RA, IL12A, IL12B, IL12RB1, IL12RB2, IL13, IL13RA1, IL13RA2, IL15, IL15RA, IL16, IL17, IL17R, IL18, IL18R1, IL19, IL20, KITLG, LEP, LTA, LTB, LTB4R, LTB4R2, LTBR, MIF, NPPB, PDGFB, TBX21, TDGF1, TGFA, TGFB1, TGFB1I1, TGFB2, TGFB3, TGFB1, TGFB1R1, TGFB1R2, TGFB1R3, TH1L, TNF, TNFRSF1A, TNFRSF1B, TNFRSF7, TNFRSF8, TNFRSF9, TNFRSF11A, TNFRSF21, TNFSF4, TNFSF5, TNFSF6, TNFSF11, VEGF, ZFPM2, and RNF110 (ZNF144). To form the bispecific or trispecific antibodies of the invention, antibodies to any combination of these antigens can be made; that is, each of these antigens can be optionally and independently included or excluded from a multispecific antibody according to the present invention.

[0149] Exemplary co-targets for autoimmune and inflammatory disorders include but are not limited to IL-1 and TNFalpha, IL-6 and TNFalpha, IL-6 and IL-1, IgE and IL-13, IL-1 and IL-13, IL-4 and IL-13, IL-5 and IL-13, IL-9 and IL-13, CD19 and FcγRIIb, and CD79 and FcγRIIb.

[0150] Immunglobulins of the invention with specificity for the following pairs of targets to treat inflammatory disease are contemplated: TNF and IL-17A; TNF and RANKL; TNF and VEGF; TNF and SOST; TNF and DKK; TNF and alphaVbeta3; TNF and NGF; TNF and IL-23p19; TNF and IL-6; TNF and SOST; TNF and IL-6R; TNF and CD-20; IgE and IL-13; IL-13 and IL23p19; IgE and IL-4; IgE and IL-9; IgE and IL-9; IgE and IL-13; IL-13 and IL-9; IL-13 and IL-4; IL-13 and IL-9; IL-13 and IL-9; IL-13 and IL-4; IL-13 and IL-23p19; IL-13 and IL-9; IL-6R and VEGF; IL-6R and IL-17A; IL-6R and RANKL; IL-17A and IL-1beta; IL-1beta and RANKL; IL-1beta and VEGF; RANKL and CD-20; IL-1alpha and IL-1beta; IL-1alpha and IL-1beta.

[0151] Pairs of targets that the immunoglobulins described herein can bind and be useful to treat asthma may be determined. In an embodiment, such targets include, but are not limited to, IL-13 and IL-1beta, since IL-1beta is also implicated in inflammatory response in asthma; IL-13 and cytokines and chemokines that are involved in inflammation, such as IL-13 and IL-9; IL-13 and IL-4; IL-13 and IL-5; IL-13 and IL-25; IL-13 and TARC; IL-13 and MDC; IL-13 and MIF; IL-13 and TGF-β; IL-13 and LHR agonist; IL-13 and CL25; IL-13 and SPRR2a; IL-13 and SPRR2b; and IL-13 and ADAM8. The immunoglobulins herein may have specificity for one or more targets involved in asthma selected from the group consisting of CSF1 (MCSF), CSF2 (GM-CSF), CSF3 (GCSF), FGF2, IFNA1, IFNB1, IFNG, histamine and

histamine receptors, IL1A, IL1B, IL2, IL3, IL4, IL5, IL6, IL7, IL8, IL9, IL10, IL11, IL12A, IL12B, IL13, IL14, IL15, IL16, IL17, IL18, IL19, KITLG, PDGFB, IL2RA, IL4R, IL5RA, IL8RA, IL8RB, IL12RB1, IL12RB2, IL13RA1, IL13RA2, IL18R1, TSLP, CCLi, CCL2, CCL3, CCL4, CCL5, CCL7, CCL8, CCL13, CCL17, CCL18, CCL19, CCL20, CCL22, CCL24, CX3CL1, CXCL1, CXCL2, CXCL3, XCLi, CCR2, CCR3, CCR4, CCR5, CCR6, CCR7, CCR8, CX3CR1, GPR2, XCR1, FOS, GATA3, JAK1, JAK3, STAT6, TBX21, TGFB1, TNF, TNFSF6, YY1, CYSLTR1, FCER1A, FCER2, LTB4R, TB4R2, LTBR, and Chitinase. To form the bispecific or trispecific antibodies of the invention, antibodies to any combination of these antigens can be made; that is, each of these antigens can be optionally and independently included or excluded from a multispecific antibody according to the present invention.

[0152] Pairs of targets involved in rheumatoid arthritis (RA) may be co-targeted by the invention, including but not limited to TNF and IL-18; TNF and IL-12; TNF and IL-23; TNF and IL-1beta; TNF and MIF; TNF and IL-17; and TNF and IL-15.

[0153] Antigens that may be targeted in order to treat systemic lupus erythematosus (SLE) by the immunoglobulins herein include but are not limited to CD-20, CD-22, CD-19, CD28, CD4, CD80, HLA-DRA, IL10, IL2, IL4, TNFRSF5, TNFRSF6, TNFSF5, TNFSF6, BLR1, HDAC4, HDAC5, HDAC7A, HDAC9, ICOSL, IGBP1, MS4A1, RGS1, SLA2, CD81, IFNB1, IL10, TNFRSF5, TNFRSF7, TNFSF5, AICDA, BLNK, GALNAC4S-6ST, HDAC4, HDAC5, HDAC7A, HDAC9, IL10, IL11, IL4, INHA, INHBA, KLF6, TNFRSF7, CD28, CD38, CD69, CD80, CD83, CD86, DPP4, FCER2, IL2RA, TNFRSF8, TNFSF7, CD24, CD37, CD40, CD72, CD74, CD79A, CD79B, CR2, ILIR2, ITGA2, ITGA3, MS4A1, ST6GALI, CDIC, CHST10, HLA-A, HLA-DRA, and NT5E.; CTLA4, B7.1, B7.2, BlyS, BAFF, C5, IL-4, IL-6, IL-10, IFN- α , and TNF- α . To form the bispecific or trispecific antibodies of the invention, antibodies to any combination of these antigens can be made; that is, each of these antigens can be optionally and independently included or excluded from a multispecific antibody according to the present invention.

[0154] The immunoglobulins herein may target antigens for the treatment of multiple sclerosis (MS), including but not limited to IL-12, TWEAK, IL-23, CXCL13, CD40, CD40L, IL-18, VEGF, VLA-4, TNF, CD45RB, CD200, IFNgamma, GM-CSF, FGF, C5, CD52, and CCR2. An embodiment includes co-engagement of anti-IL-12 and TWEAK for the treatment of MS.

[0155] One aspect of the invention pertains to immunoglobulins capable of binding one or more targets involved in sepsis, in an embodiment two targets, selected from the group consisting TNF, IL-1, MIF, IL-6, IL-8, IL-18, IL-12, IL-23, FasL, LPS, Toll-like receptors, TLR-4, tissue factor, MIP-2, ADORA2A, CASP1, CASP4, IL-10, IL-1B, NFkB1, PROC,

TNFRSF1A, CSF3, CCR3, IL1RN, MIF, NFκB1, PTAFR, TLR2, TLR4, GPR44, HMOX1, midkine, IRAK1, NFκB2, SERPINA1, SERPINE1, and TREM1. To form the bispecific or trispecific antibodies of the invention, antibodies to any combination of these antigens can be made; that is, each of these antigens can be optionally and independently included or excluded from a multispecific antibody according to the present invention.

[0156] In some cases, immunoglobulins herein may be directed against antigens for the treatment of infectious diseases.

[0157] The antibodies of the present invention are generally isolated or recombinant. "Isolated," when used to describe the various polypeptides disclosed herein, means a polypeptide that has been identified and separated and/or recovered from a cell or cell culture from which it was expressed. Ordinarily, an isolated polypeptide will be prepared by at least one purification step. An "isolated antibody," refers to an antibody which is substantially free of other antibodies having different antigenic specificities.

[0158] "Specific binding" or "specifically binds to" or is "specific for" a particular antigen or an epitope means binding that is measurably different from a non-specific interaction. Specific binding can be measured, for example, by determining binding of a molecule compared to binding of a control molecule, which generally is a molecule of similar structure that does not have binding activity. For example, specific binding can be determined by competition with a control molecule that is similar to the target.

[0159] Specific binding for a particular antigen or an epitope can be exhibited, for example, by an antibody having a KD for an antigen or epitope of at least about 10⁻⁴ M, at least about 10⁻⁵ M, at least about 10⁻⁶ M, at least about 10⁻⁷ M, at least about 10⁻⁸ M, at least about 10⁻⁹ M, alternatively at least about 10⁻¹⁰ M, at least about 10⁻¹¹ M, at least about 10⁻¹² M, or greater, where KD refers to a dissociation rate of a particular antibody-antigen interaction. Typically, an antibody that specifically binds an antigen will have a KD that is 20-, 50-, 100-, 500-, 1000-, 5,000-, 10,000- or more times greater for a control molecule relative to the antigen or epitope.

[0160] Also, specific binding for a particular antigen or an epitope can be exhibited, for example, by an antibody having a KA or Ka for an antigen or epitope of at least 20-, 50-, 100-, 500-, 1000-, 5,000-, 10,000- or more times greater for the epitope relative to a control, where KA or Ka refers to an association rate of a particular antibody-antigen interaction.

Modified Antibodies

[0161] In addition to the modifications outlined above, other modifications can be made. For example, the molecules may be stabilized by the incorporation of disulphide bridges linking the VH and VL domains (Reiter et al., 1996, Nature Biotech. 14:1239-1245, entirely

incorporated by reference). In addition, there are a variety of covalent modifications of antibodies that can be made as outlined below.

[0162] Covalent modifications of antibodies are included within the scope of this invention, and are generally, but not always, done post-translationally. For example, several types of covalent modifications of the antibody are introduced into the molecule by reacting specific amino acid residues of the antibody with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C-terminal residues.

[0163] Cysteinyl residues most commonly are reacted with α -haloacetates (and corresponding amines), such as chloroacetic acid or chloroacetamide, to give carboxymethyl or carboxyamidomethyl derivatives. Cysteinyl residues may also be derivatized by reaction with bromotrifluoroacetone, α -bromo- β -(5-imidozoyl)propionic acid, chloroacetyl phosphate, N-alkylmaleimides, 3-nitro-2-pyridyl disulfide, methyl 2-pyridyl disulfide, p-chloromercuribenzoate, 2-chloromercuri-4-nitrophenol, or chloro-7-nitrobenzo-2-oxa-1,3-diazole and the like.

[0164] In addition, modifications at cysteines are particularly useful in antibody-drug conjugate (ADC) applications, further described below. In some embodiments, the constant region of the antibodies can be engineered to contain one or more cysteines that are particularly "thiol reactive", so as to allow more specific and controlled placement of the drug moiety. See for example US Patent No. 7,521,541, incorporated by reference in its entirety herein.

[0165] Histidyl residues are derivatized by reaction with diethylpyrocarbonate at pH 5.5-7.0 because this agent is relatively specific for the histidyl side chain. Para-bromophenacyl bromide also is useful; the reaction is preferably performed in 0.1M sodium cacodylate at pH 6.0.

[0166] Lysinyl and amino terminal residues are reacted with succinic or other carboxylic acid anhydrides. Derivatization with these agents has the effect of reversing the charge of the lysinyl residues. Other suitable reagents for derivatizing alpha-amino-containing residues include imidoesters such as methyl picolinimide; pyridoxal phosphate; pyridoxal; chloroborohydride; trinitrobenzenesulfonic acid; O-methylisourea; 2,4-pentanedione; and transaminase-catalyzed reaction with glyoxylate.

[0167] Arginyl residues are modified by reaction with one or several conventional reagents, among them phenylglyoxal, 2,3-butanedione, 1,2-cyclohexanedione, and ninhydrin. Derivatization of arginine residues requires that the reaction be performed in alkaline conditions because of the high pKa of the guanidine functional group. Furthermore,

these reagents may react with the groups of lysine as well as the arginine epsilon-amino group.

[0168] The specific modification of tyrosyl residues may be made, with particular interest in introducing spectral labels into tyrosyl residues by reaction with aromatic diazonium compounds or tetranitromethane. Most commonly, N-acetylimidazole and tetranitromethane are used to form O-acetyl tyrosyl species and 3-nitro derivatives, respectively. Tyrosyl residues are iodinated using ^{125}I or ^{131}I to prepare labeled proteins for use in radioimmunoassay, the chloramine T method described above being suitable.

[0169] Carboxyl side groups (aspartyl or glutamyl) are selectively modified by reaction with carbodiimides ($\text{R}'\text{---N}=\text{C}=\text{N}\text{---R}'$), where R and R' are optionally different alkyl groups, such as 1-cyclohexyl-3-(2-morpholinyl-4-ethyl) carbodiimide or 1-ethyl-3-(4-azonia-4,4-dimethylpentyl) carbodiimide. Furthermore, aspartyl and glutamyl residues are converted to asparaginyl and glutaminyl residues by reaction with ammonium ions.

[0170] Derivatization with bifunctional agents is useful for crosslinking antibodies to a water-insoluble support matrix or surface for use in a variety of methods, in addition to methods described below. Commonly used crosslinking agents include, e.g., 1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde, N-hydroxysuccinimide esters, for example, esters with 4-azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis (succinimidylpropionate), and bifunctional maleimides such as bis-N-maleimido-1,8-octane. Derivatizing agents such as methyl-3-[(p-azidophenyl)dithio]propioimide yield photoactivatable intermediates that are capable of forming crosslinks in the presence of light. Alternatively, reactive water-insoluble matrices such as cyanomolgusogen bromide-activated carbohydrates and the reactive substrates described in U.S. Pat. Nos. 3,969,287; 3,691,016; 4,195,128; 4,247,642; 4,229,537; and 4,330,440, all entirely incorporated by reference, are employed for protein immobilization.

[0171] Glutaminyl and asparaginyl residues are frequently deamidated to the corresponding glutamyl and aspartyl residues, respectively. Alternatively, these residues are deamidated under mildly acidic conditions. Either form of these residues falls within the scope of this invention.

[0172] Other modifications include hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α -amino groups of lysine, arginine, and histidine side chains (T. E. Creighton, *Proteins: Structure and Molecular Properties*, W. H. Freeman & Co., San Francisco, pp. 79-86 [1983], entirely incorporated by reference), acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

[0173] In addition, as will be appreciated by those in the art, labels (including fluorescent, enzymatic, magnetic, radioactive, etc. can all be added to the antibodies (as well as the other compositions of the invention).

Glycosylation

[0174] Another type of covalent modification is alterations in glycosylation. In another embodiment, the antibodies disclosed herein can be modified to include one or more engineered glycoforms. By “engineered glycoform” as used herein is meant a carbohydrate composition that is covalently attached to the antibody, wherein said carbohydrate composition differs chemically from that of a parent antibody. Engineered glycoforms may be useful for a variety of purposes, including but not limited to enhancing or reducing effector function. A preferred form of engineered glycoform is afucosylation, which has been shown to be correlated to an increase in ADCC function, presumably through tighter binding to the FcγRIIIa receptor. In this context, “afucosylation” means that the majority of the antibody produced in the host cells is substantially devoid of fucose, e.g. 90-95-98% of the generated antibodies do not have appreciable fucose as a component of the carbohydrate moiety of the antibody (generally attached at N297 in the Fc region). Defined functionally, afucosylated antibodies generally exhibit at least a 50% or higher affinity to the FcγRIIIa receptor.

[0175] Engineered glycoforms may be generated by a variety of methods known in the art (Umaña et al., 1999, Nat Biotechnol 17:176-180; Davies et al., 2001, Biotechnol Bioeng 74:288-294; Shields et al., 2002, J Biol Chem 277:26733-26740; Shinkawa et al., 2003, J Biol Chem 278:3466-3473; US 6,602,684; USSN 10/277,370; USSN 10/113,929; PCT WO 00/61739A1; PCT WO 01/29246A1; PCT WO 02/31140A1; PCT WO 02/30954A1, all entirely incorporated by reference; (Potelligent® technology [Biowa, Inc., Princeton, NJ]; GlycoMAb® glycosylation engineering technology [Glycart Biotechnology AG, Zürich, Switzerland]). Many of these techniques are based on controlling the level of fucosylated and/or bisecting oligosaccharides that are covalently attached to the Fc region, for example by expressing an IgG in various organisms or cell lines, engineered or otherwise (for example Lec-13 CHO cells or rat hybridoma YB2/0 cells, by regulating enzymes involved in the glycosylation pathway (for example FUT8 [α 1,6-fucosyltransferase] and/or β 1-4- N-acetylglucosaminyltransferase III [GnTIII]), or by modifying carbohydrate(s) after the IgG has been expressed. For example, the “sugar engineered antibody” or “SEA technology” of Seattle Genetics functions by adding modified saccharides that inhibit fucosylation during production; see for example 20090317869, hereby incorporated by reference in its entirety. Engineered glycoform typically refers to the different carbohydrate or oligosaccharide; thus an antibody can include an engineered glycoform.

[0176] Alternatively, engineered glycoform may refer to the IgG variant that comprises the different carbohydrate or oligosaccharide. As is known in the art, glycosylation patterns can depend on both the sequence of the protein (e.g., the presence or absence of particular glycosylation amino acid residues, discussed below), or the host cell or organism in which the protein is produced. Particular expression systems are discussed below.

[0177] Glycosylation of polypeptides is typically either N-linked or O-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tri-peptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tri-peptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one of the sugars N-acetylgalactosamine, galactose, or xylose, to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

[0178] Addition of glycosylation sites to the antibody is conveniently accomplished by altering the amino acid sequence such that it contains one or more of the above-described tri-peptide sequences (for N-linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to the starting sequence (for O-linked glycosylation sites). For ease, the antibody amino acid sequence is preferably altered through changes at the DNA level, particularly by mutating the DNA encoding the target polypeptide at preselected bases such that codons are generated that will translate into the desired amino acids.

[0179] Another means of increasing the number of carbohydrate moieties on the antibody is by chemical or enzymatic coupling of glycosides to the protein. These procedures are advantageous in that they do not require production of the protein in a host cell that has glycosylation capabilities for N- and O-linked glycosylation. Depending on the coupling mode used, the sugar(s) may be attached to (a) arginine and histidine, (b) free carboxyl groups, (c) free sulfhydryl groups such as those of cysteine, (d) free hydroxyl groups such as those of serine, threonine, or hydroxyproline, (e) aromatic residues such as those of phenylalanine, tyrosine, or tryptophan, or (f) the amide group of glutamine. These methods are described in WO 87/05330 and in Aplin and Wriston, 1981, CRC Crit. Rev. Biochem., pp. 259-306, both entirely incorporated by reference.

[0180] Removal of carbohydrate moieties present on the starting antibody (e.g. post-translationally) may be accomplished chemically or enzymatically. Chemical deglycosylation requires exposure of the protein to the compound trifluoromethanesulfonic acid, or an

equivalent compound. This treatment results in the cleavage of most or all sugars except the linking sugar (N-acetylglucosamine or N-acetylgalactosamine), while leaving the polypeptide intact. Chemical deglycosylation is described by Hakimuddin et al., 1987, Arch. Biochem. Biophys. 259:52 and by Edge et al., 1981, Anal. Biochem. 118:131, both entirely incorporated by reference. Enzymatic cleavage of carbohydrate moieties on polypeptides can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura et al., 1987, Meth. Enzymol. 138:350, entirely incorporated by reference. Glycosylation at potential glycosylation sites may be prevented by the use of the compound tunicamycin as described by Duskin et al., 1982, J. Biol. Chem. 257:3105, entirely incorporated by reference. Tunicamycin blocks the formation of protein-N-glycoside linkages.

[0181] Another type of covalent modification of the antibody comprises linking the antibody to various nonproteinaceous polymers, including, but not limited to, various polyols such as polyethylene glycol, polypropylene glycol or polyoxyalkylenes, in the manner set forth in, for example, 2005-2006 PEG Catalog from Nektar Therapeutics (available at the Nektar website) US Patents 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337, all entirely incorporated by reference. In addition, as is known in the art, amino acid substitutions may be made in various positions within the antibody to facilitate the addition of polymers such as PEG. See for example, U.S. Publication No. 2005/0114037A1, entirely incorporated by reference.

Additional Fc Variants for Additional Functionality

[0182] In addition to pl amino acid variants, there are a number of useful Fc amino acid modification that can be made for a variety of reasons, including, but not limited to, altering binding to one or more FcγR receptors, altered binding to FcRn receptors, etc.

[0183] Accordingly, the proteins of the invention can include amino acid modifications, including the heterodimerization variants outlined herein, which includes the pl variants

FcγR Variants

[0184] Accordingly, there are a number of useful Fc substitutions that can be made to alter binding to one or more of the FcγR receptors. Substitutions that result in increased binding as well as decreased binding can be useful. For example, it is known that increased binding to Fc RIIIa generally results in increased ADCC (antibody dependent cell-mediated cytotoxicity; the cell-mediated reaction wherein nonspecific cytotoxic cells that express FcγRs recognize bound antibody on a target cell and subsequently cause lysis of the target cell). Similarly, decreased binding to FcγRIIb (an inhibitory receptor) can be beneficial as well in some circumstances. Amino acid substitutions that find use in the present invention include those listed in USSNs 11/124,620 (particularly Figure 41), 11/174,287, 11/396,495,

11/538,406, all of which are expressly incorporated herein by reference in their entirety and specifically for the variants disclosed therein. Particular variants that find use include, but are not limited to, 236A, 239D, 239E, 332E, 332D, 239D/332E, 267D, 267E, 328F, 267E/328F, 236A/332E, 239D/332E/330Y, 239D, 332E/330L and 299T.

[0185] In addition, there are additional Fc substitutions that find use in increased binding to the FcRn receptor and increased serum half life, as specifically disclosed in USSN 12/341,769, hereby incorporated by reference in its entirety, including, but not limited to, 434S, 428L, 308F, 259I, 428L/434S, 259I/308F, 436I/428L, 436I or V/434S, 436V/428L and 259I/308F/428L.

Linkers

[0186] The present invention optionally provides linkers as needed, for example in the addition of additional antigen binding sites, as depicted for example in Figure 2, where “the other end” of the molecule contains additional antigen binding components. In addition, as outlined below, linkers are optionally also used in antibody drug conjugate (ADC) systems. When used to join the components of the central mAb-Fv constructs, the linker is generally a polypeptide comprising two or more amino acid residues joined by peptide bonds and are used to link one or more of the components of the present invention. Such linker polypeptides are well known in the art (see e.g., Holliger, P., et al. (1993) Proc. Natl. Acad. Sci. USA 90:6444-6448; Poljak, R. J., et al. (1994) Structure 2:1121-1123). A variety of linkers may find use in some embodiments described herein. As will be appreciated by those in the art, there are at least three different linker types used in the present invention.

[0187] “Linker” herein is also referred to as “linker sequence”, “spacer”, “tethering sequence” or grammatical equivalents thereof. Homo- or hetero-bifunctional linkers as are well known (see, 1994 Pierce Chemical Company catalog, technical section on cross-linkers, pages 155-200, incorporated entirely by reference). A number of strategies may be used to covalently link molecules together. These include, but are not limited to polypeptide linkages between N- and C-termini of proteins or protein domains, linkage via disulfide bonds, and linkage via chemical cross-linking reagents. In one aspect of this embodiment, the linker is a peptide bond, generated by recombinant techniques or peptide synthesis. The linker peptide may predominantly include the following amino acid residues: Gly, Ser, Ala, or Thr. The linker peptide should have a length that is adequate to link two molecules in such a way that they assume the correct conformation relative to one another so that they retain the desired activity. In one embodiment, the linker is from about 1 to 50 amino acids in length, preferably about 1 to 30 amino acids in length. In one embodiment, linkers of 1 to 20 amino acids in length may be used. Useful linkers include glycine-serine polymers, including for example (GS)_n, (GSGGS)_n, (GGGS)_n, and (GGGS)_n, where n is an integer of at least one, glycine-

alanine polymers, alanine-serine polymers, and other flexible linkers. Alternatively, a variety of nonproteinaceous polymers, including but not limited to polyethylene glycol (PEG), polypropylene glycol, polyoxyalkylenes, or copolymers of polyethylene glycol and polypropylene glycol, may find use as linkers, that is may find use as linkers.

[0188] Other linker sequences may include any sequence of any length of CL/CH1 domain but not all residues of CL/CH1 domain; for example the first 5-12 amino acid residues of the CL/CH1 domains. Linkers can be derived from immunoglobulin light chain, for example C κ or C λ . Linkers can be derived from immunoglobulin heavy chains of any isotype, including for example C γ 1, C γ 2, C γ 3, C γ 4, C α 1, C α 2, C δ , C ϵ , and C μ . Linker sequences may also be derived from other proteins such as Ig-like proteins (e.g. TCR, FcR, KIR), hinge region-derived sequences, and other natural sequences from other proteins.

Antibody-Drug Conjugates

[0189] In some embodiments, the multispecific antibodies of the invention are conjugated with drugs to form antibody-drug conjugates (ADCs). In general, ADCs are used in oncology applications, where the use of antibody-drug conjugates for the local delivery of cytotoxic or cytostatic agents allows for the targeted delivery of the drug moiety to tumors, which can allow higher efficacy, lower toxicity, etc. An overview of this technology is provided in Ducry et al., Bioconjugate Chem., 21:5-13 (2010), Carter et al., Cancer J. 14(3):154 (2008) and Senter, Current Opin. Chem. Biol. 13:235-244 (2009), all of which are hereby incorporated by reference in their entirety.

[0190] Thus the invention provides multispecific antibodies conjugated to drugs. Generally, conjugation is done by covalent attachment to the antibody, as further described below, and generally relies on a linker, often a peptide linkage (which, as described below, may be designed to be sensitive to cleavage by proteases at the target site or not). In addition, as described above, linkage of the linker-drug unit (LU-D) can be done by attachment to cysteines within the antibody. As will be appreciated by those in the art, the number of drug moieties per antibody can change, depending on the conditions of the reaction, and can vary from 1:1 to 10:1 drug:antibody. As will be appreciated by those in the art, the actual number is an average.

[0191] Thus the invention provides multispecific antibodies conjugated to drugs. As described below, the drug of the ADC can be any number of agents, including but not limited to cytotoxic agents such as chemotherapeutic agents, growth inhibitory agents, toxins (for example, an enzymatically active toxin of bacterial, fungal, plant, or animal origin, or fragments thereof), or a radioactive isotope (that is, a radioconjugate) are provided. In other embodiments, the invention further provides methods of using the ADCs.

[0192] Drugs for use in the present invention include cytotoxic drugs, particularly those which are used for cancer therapy. Such drugs include, in general, DNA damaging agents, anti-metabolites, natural products and their analogs. Exemplary classes of cytotoxic agents include the enzyme inhibitors such as dihydrofolate reductase inhibitors, and thymidylate synthase inhibitors, DNA intercalators, DNA cleavers, topoisomerase inhibitors, the anthracycline family of drugs, the vinca drugs, the mitomycins, the bleomycins, the cytotoxic nucleosides, the pteridine family of drugs, diynenes, the podophyllotoxins, dolastatins, maytansinoids, differentiation inducers, and taxols.

[0193] Members of these classes include, for example, methotrexate, methopterin, dichloromethotrexate, 5-fluorouracil, 6-mercaptopurine, cytosine arabinoside, melphalan, leurosine, leurosideine, actinomycin, daunorubicin, doxorubicin, mitomycin C, mitomycin A, caminomycin, aminopterin, tallysomyin, podophyllotoxin and podophyllotoxin derivatives such as etoposide or etoposide phosphate, vinblastine, vincristine, vindesine, taxanes including taxol, taxotere retinoic acid, butyric acid, N8-acetyl spermidine, camptothecin, calicheamicin, esperamicin, ene-diynes, duocarmycin A, duocarmycin SA, calicheamicin, camptothecin, maytansinoids (including DM1), monomethylauristatin E (MMAE), monomethylauristatin F (MMAF), and maytansinoids (DM4) and their analogues.

[0194] Toxins may be used as antibody-toxin conjugates and include bacterial toxins such as diphtheria toxin, plant toxins such as ricin, small molecule toxins such as geldanamycin (Mandler et al (2000) J. Nat. Cancer Inst. 92(19):1573-1581; Mandler et al (2000) Bioorganic & Med. Chem. Letters 10:1025-1028; Mandler et al (2002) Bioconjugate Chem. 13:786-791), maytansinoids (EP 1391213; Liu et al., (1996) Proc. Natl. Acad. Sci. USA 93:8618-8623), and calicheamicin (Lode et al (1998) Cancer Res. 58:2928; Hinman et al (1993) Cancer Res. 53:3336-3342). Toxins may exert their cytotoxic and cytostatic effects by mechanisms including tubulin binding, DNA binding, or topoisomerase inhibition.

[0195] Conjugates of a multispecific antibody and one or more small molecule toxins, such as a maytansinoids, dolastatins, auristatins, a trichothecene, calicheamicin, and CC1065, and the derivatives of these toxins that have toxin activity, are contemplated.

Maytansinoids

[0196] Maytansine compounds suitable for use as maytansinoid drug moieties are well known in the art, and can be isolated from natural sources according to known methods, produced using genetic engineering techniques (see Yu et al (2002) PNAS 99:7968-7973), or maytansinol and maytansinol analogues prepared synthetically according to known methods. As described below, drugs may be modified by the incorporation of a functionally active group such as a thiol or amine group for conjugation to the antibody.

[0197] Exemplary maytansinoid drug moieties include those having a modified aromatic ring, such as: C-19-dechloro (U.S. Pat. No. 4,256,746) (prepared by lithium aluminum hydride reduction of ansamycin P2); C-20-hydroxy (or C-20-demethyl) +/-C-19-dechloro (U.S. Pat. Nos. 4,361,650 and 4,307,016) (prepared by demethylation using *Streptomyces* or *Actinomyces* or dechlorination using LAH); and C-20-demethoxy, C-20-acyloxy (–OCOR), +/-dechloro (U.S. Pat. No. 4,294,757) (prepared by acylation using acyl chlorides) and those having modifications at other positions

[0198] Exemplary maytansinoid drug moieties also include those having modifications such as: C-9-SH (U.S. Pat. No. 4,424,219) (prepared by the reaction of maytansinol with H₂S or P₂S₅); C-14-alkoxymethyl(demethoxy/CH₂OR) (U.S. Pat. No. 4,331,598); C-14-hydroxymethyl or acyloxymethyl (CH₂OH or CH₂OAc) (U.S. Pat. No. 4,450,254) (prepared from *Nocardia*); C-15-hydroxy/acyloxy (U.S. Pat. No. 4,364,866) (prepared by the conversion of maytansinol by *Streptomyces*); C-15-methoxy (U.S. Pat. Nos. 4,313,946 and 4,315,929) (isolated from *Trewia nudiflora*); C-18-N-demethyl (U.S. Pat. Nos. 4,362,663 and 4,322,348) (prepared by the demethylation of maytansinol by *Streptomyces*); and 4,5-deoxy (U.S. Pat. No. 4,371,533) (prepared by the titanium trichloride/LAH reduction of maytansinol).

[0199] Of particular use are DM1 (disclosed in US Patent No. 5,208,020, incorporated by reference) and DM4 (disclosed in US Patent No. 7,276,497, incorporated by reference). See also a number of additional maytansinoid derivatives and methods in 5,416,064, WO/01/24763, 7,303,749, 7,601,354, USSN 12/631,508, WO02/098883, 6,441,163, 7,368,565, WO02/16368 and WO04/1033272, all of which are expressly incorporated by reference in their entirety.

[0200] ADCs containing maytansinoids, methods of making same, and their therapeutic use are disclosed, for example, in U.S. Pat. Nos. 5,208,020; 5,416,064; 6,441,163 and European Patent EP 0 425 235 B1, the disclosures of which are hereby expressly incorporated by reference. Liu et al., Proc. Natl. Acad. Sci. USA 93:8618-8623 (1996) described ADCs comprising a maytansinoid designated DM1 linked to the monoclonal antibody C242 directed against human colorectal cancer. The conjugate was found to be highly cytotoxic towards cultured colon cancer cells, and showed antitumor activity in an in vivo tumor growth assay.

[0201] Chari et al., Cancer Research 52:127-131 (1992) describe ADCs in which a maytansinoid was conjugated via a disulfide linker to the murine antibody A7 binding to an antigen on human colon cancer cell lines, or to another murine monoclonal antibody TA.1 that binds the HER-2/neu oncogene. The cytotoxicity of the TA.1-maytansinoid conjugate was tested in vitro on the human breast cancer cell line SK-BR-3, which expresses 3x10⁵

HER-2 surface antigens per cell. The drug conjugate achieved a degree of cytotoxicity similar to the free maytansinoid drug, which could be increased by increasing the number of maytansinoid molecules per antibody molecule. The A7-maytansinoid conjugate showed low systemic cytotoxicity in mice.

Auristatins and Dolastatins

[0202] In some embodiments, the ADC comprises a multispecific antibody conjugated to dolastatins or dolostatin peptidic analogs and derivatives, the auristatins (U.S. Pat. Nos. 5,635,483; 5,780,588). Dolastatins and auristatins have been shown to interfere with microtubule dynamics, GTP hydrolysis, and nuclear and cellular division (Woyke et al (2001) *Antimicrob. Agents and Chemother.* 45(12):3580-3584) and have anticancer (U.S. Pat. No. 5,663,149) and antifungal activity (Pettit et al (1998) *Antimicrob. Agents Chemother.* 42:2961-2965). The dolastatin or auristatin drug moiety may be attached to the antibody through the N (amino) terminus or the C (carboxyl) terminus of the peptidic drug moiety (WO 02/088172).

[0203] Exemplary auristatin embodiments include the N-terminus linked monomethylauristatin drug moieties DE and DF, disclosed in "Senter et al, Proceedings of the American Association for Cancer Research, Volume 45, Abstract Number 623, presented Mar. 28, 2004 and described in United States Patent Publication No. 2005/0238648, the disclosure of which is expressly incorporated by reference in its entirety.

[0204] An exemplary auristatin embodiment is MMAE (see US Patent No. 6,884,869 expressly incorporated by reference in its entirety).

[0205] Another exemplary auristatin embodiment is MMAF (see US 2005/0238649, 5,767,237 and 6,124,431, expressly incorporated by reference in their entirety).

[0206] Additional exemplary embodiments comprising MMAE or MMAF and various linker components (described further herein) have the following structures and abbreviations (wherein Ab means antibody and p is 1 to about 8):

[0207] Typically, peptide-based drug moieties can be prepared by forming a peptide bond between two or more amino acids and/or peptide fragments. Such peptide bonds can be prepared, for example, according to the liquid phase synthesis method (see E. Schroder and K. Lubke, "The Peptides", volume 1, pp 76-136, 1965, Academic Press) that is well known in the field of peptide chemistry. The auristatin/dolastatin drug moieties may be prepared according to the methods of: U.S. Pat. No. 5,635,483; U.S. Pat. No. 5,780,588; Pettit et al (1989) *J. Am. Chem. Soc.* 111:5463-5465; Pettit et al (1998) *Anti-Cancer Drug Design* 13:243-277; Pettit, G. R., et al. *Synthesis*, 1996, 719-725; Pettit et al (1996) *J. Chem. Soc. Perkin Trans. 1* 5:859-863; and Doronina (2003) *Nat Biotechnol* 21(7):778-784.

Calicheamicin

[0208] In other embodiments, the ADC comprises an antibody of the invention conjugated to one or more calicheamicin molecules. For example, Mylotarg is the first commercial ADC drug and utilizes calicheamicin γ 1 as the payload (see US Patent No. 4,970,198, incorporated by reference in its entirety). Additional calicheamicin derivatives are described in US Patent Nos. 5,264,586, 5,384,412, 5,550,246, 5,739,116, 5,773,001, 5,767,285 and 5,877,296, all expressly incorporated by reference. The calicheamicin family of antibiotics are capable of producing double-stranded DNA breaks at sub-picomolar concentrations. For the preparation of conjugates of the calicheamicin family, see U.S. Pat. Nos. 5,712,374, 5,714,586, 5,739,116, 5,767,285, 5,770,701, 5,770,710, 5,773,001, 5,877,296 (all to American Cyanamid Company). Structural analogues of calicheamicin which may be used include, but are not limited to, γ 1I, α 2I, α 2I, N-acetyl- γ 1I, PSAG and θ 1I (Hinman et al., Cancer Research 53:3336-3342 (1993), Lode et al., Cancer Research 58:2925-2928 (1998) and the aforementioned U.S. patents to American Cyanamid). Another anti-tumor drug that the antibody can be conjugated is QFA which is an antifolate. Both calicheamicin and QFA have intracellular sites of action and do not readily cross the plasma membrane. Therefore, cellular uptake of these agents through antibody mediated internalization greatly enhances their cytotoxic effects.

Duocarmycins

[0209] CC-1065 (see 4,169,888, incorporated by reference) and duocarmycins are members of a family of antitumor antibiotics utilized in ADCs. These antibiotics appear to work through sequence-selectively alkylating DNA at the N3 of adenine in the minor groove, which initiates a cascade of events that result in apoptosis.

[0210] Important members of the duocarmycins include duocarmycin A (US Patent No. 4,923,990, incorporated by reference) and duocarmycin SA (U.S. Pat. No. 5,101,038, incorporated by reference), and a large number of analogues as described in US Patent Nos. 7,517,903, 7,691,962, 5,101,038; 5,641,780; 5,187,186; 5,070,092; 5,070,092; 5,641,780; 5,101,038; 5,084,468, 5,475,092, 5,585,499, 5,846,545, WO2007/089149, WO2009/017394A1, 5,703,080, 6,989,452, 7,087,600, 7,129,261, 7,498,302, and 7,507,420, all of which are expressly incorporated by reference.

Other Cytotoxic Agents

[0211] Other antitumor agents that can be conjugated to the antibodies of the invention include BCNU, streptozocin, vincristine and 5-fluorouracil, the family of agents known collectively LL-E33288 complex described in U.S. Pat. Nos. 5,053,394, 5,770,710, as well as esperamicins (U.S. Pat. No. 5,877,296).

[0212] Enzymatically active toxins and fragments thereof which can be used include diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, Aleurites fordii proteins, dianthin proteins, Phytolaca americana proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcin, crotin, sapaonaria officinalis inhibitor, gelonin, mitogellin, restrictocin, phenomycin, enomycin and the tricothecenes. See, for example, WO 93/21232 published Oct. 28, 1993.

[0213] The present invention further contemplates an ADC formed between an antibody and a compound with nucleolytic activity (e.g., a ribonuclease or a DNA endonuclease such as a deoxyribonuclease; DNase).

[0214] For selective destruction of the tumor, the antibody may comprise a highly radioactive atom. A variety of radioactive isotopes are available for the production of radioconjugated antibodies. Examples include At211, I131, I125, Y90, Re186, Re188, Sm153, Bi212, P32, Pb212 and radioactive isotopes of Lu.

[0215] The radio- or other labels may be incorporated in the conjugate in known ways. For example, the peptide may be biosynthesized or may be synthesized by chemical amino acid synthesis using suitable amino acid precursors involving, for example, fluorine-19 in place of hydrogen. Labels such as Tc99m or I123, Re186, Re188 and In111 can be attached via a cysteine residue in the peptide. Yttrium-90 can be attached via a lysine residue. The IODOGEN method (Fraker et al (1978) Biochem. Biophys. Res. Commun. 80: 49-57 can be used to incorporate Iodine-123. "Monoclonal Antibodies in Immunoscintigraphy" (Chatal, CRC Press 1989) describes other methods in detail.

[0216] For compositions comprising a plurality of antibodies, the drug loading is represented by p, the average number of drug molecules per Antibody. Drug loading may range from 1 to 20 drugs (D) per Antibody. The average number of drugs per antibody in preparation of conjugation reactions may be characterized by conventional means such as mass spectroscopy, ELISA assay, and HPLC. The quantitative distribution of Antibody-Drug-Conjugates in terms of p may also be determined.

[0217] In some instances, separation, purification, and characterization of homogeneous Antibody-Drug-conjugates where p is a certain value from Antibody-Drug-Conjugates with other drug loadings may be achieved by means such as reverse phase HPLC or electrophoresis. In exemplary embodiments, p is 2, 3, 4, 5, 6, 7, or 8 or a fraction thereof.

[0218] The generation of Antibody-drug conjugate compounds can be accomplished by any technique known to the skilled artisan. Briefly, the Antibody-drug conjugate compounds

can include a multispecific antibody as the Antibody unit, a drug, and optionally a linker that joins the drug and the binding agent.

[0219] A number of different reactions are available for covalent attachment of drugs and/or linkers to binding agents. This is can be accomplished by reaction of the amino acid residues of the binding agent, for example, antibody molecule, including the amine groups of lysine, the free carboxylic acid groups of glutamic and aspartic acid, the sulfhydryl groups of cysteine and the various moieties of the aromatic amino acids. A commonly used non-specific methods of covalent attachment is the carbodiimide reaction to link a carboxy (or amino) group of a compound to amino (or carboxy) groups of the antibody. Additionally, bifunctional agents such as dialdehydes or imidoesters have been used to link the amino group of a compound to amino groups of an antibody molecule.

[0220] Also available for attachment of drugs to binding agents is the Schiff base reaction. This method involves the periodate oxidation of a drug that contains glycol or hydroxy groups, thus forming an aldehyde which is then reacted with the binding agent. Attachment occurs via formation of a Schiff base with amino groups of the binding agent. Isothiocyanates can also be used as coupling agents for covalently attaching drugs to binding agents. Other techniques are known to the skilled artisan and within the scope of the present invention.

[0221] In some embodiments, an intermediate, which is the precursor of the linker, is reacted with the drug under appropriate conditions. In other embodiments, reactive groups are used on the drug and/or the intermediate. The product of the reaction between the drug and the intermediate, or the derivatized drug, is subsequently reacted with an multispecific antibody of the invention under appropriate conditions.

[0222] It will be understood that chemical modifications may also be made to the desired compound in order to make reactions of that compound more convenient for purposes of preparing conjugates of the invention. For example a functional group e.g. amine, hydroxyl, or sulfhydryl, may be appended to the drug at a position which has minimal or an acceptable effect on the activity or other properties of the drug

Linker Units

[0223] Typically, the antibody-drug conjugate compounds comprise a Linker unit between the drug unit and the antibody unit. In some embodiments, the linker is cleavable under intracellular or extracellular conditions, such that cleavage of the linker releases the drug unit from the antibody in the appropriate environment. For example, solid tumors that secrete certain proteases may serve as the target of the cleavable linker; in other embodiments, it is

the intracellular proteases that are utilized. In yet other embodiments, the linker unit is not cleavable and the drug is released, for example, by antibody degradation in lysosomes.

[0224] In some embodiments, the linker is cleavable by a cleaving agent that is present in the intracellular environment (for example, within a lysosome or endosome or caveolea). The linker can be, for example, a peptidyl linker that is cleaved by an intracellular peptidase or protease enzyme, including, but not limited to, a lysosomal or endosomal protease. In some embodiments, the peptidyl linker is at least two amino acids long or at least three amino acids long or more.

[0225] Cleaving agents can include, without limitation, cathepsins B and D and plasmin, all of which are known to hydrolyze dipeptide drug derivatives resulting in the release of active drug inside target cells (see, e.g., Dubowchik and Walker, 1999, Pharm. Therapeutics 83:67-123). Peptidyl linkers that are cleavable by enzymes that are present in CD38-expressing cells. For example, a peptidyl linker that is cleavable by the thiol-dependent protease cathepsin-B, which is highly expressed in cancerous tissue, can be used (e.g., a Phe-Leu or a Gly-Phe-Leu-Gly linker (SEQ ID NO: X)). Other examples of such linkers are described, e.g., in U.S. Pat. No. 6,214,345, incorporated herein by reference in its entirety and for all purposes.

[0226] In some embodiments, the peptidyl linker cleavable by an intracellular protease is a Val-Cit linker or a Phe-Lys linker (see, e.g., U.S. Pat. No. 6,214,345, which describes the synthesis of doxorubicin with the val-cit linker).

[0227] In other embodiments, the cleavable linker is pH-sensitive, that is, sensitive to hydrolysis at certain pH values. Typically, the pH-sensitive linker hydrolyzable under acidic conditions. For example, an acid-labile linker that is hydrolyzable in the lysosome (for example, a hydrazone, semicarbazone, thiosemicarbazone, cis-aconitic amide, orthoester, acetal, ketal, or the like) may be used. (See, e.g., U.S. Pat. Nos. 5,122,368; 5,824,805; 5,622,929; Dubowchik and Walker, 1999, Pharm. Therapeutics 83:67-123; Neville et al., 1989, Biol. Chem. 264:14653-14661.) Such linkers are relatively stable under neutral pH conditions, such as those in the blood, but are unstable at below pH 5.5 or 5.0, the approximate pH of the lysosome. In certain embodiments, the hydrolyzable linker is a thioether linker (such as, e.g., a thioether attached to the therapeutic agent via an acylhydrazone bond (see, e.g., U.S. Pat. No. 5,622,929).

[0228] In yet other embodiments, the linker is cleavable under reducing conditions (for example, a disulfide linker). A variety of disulfide linkers are known in the art, including, for example, those that can be formed using SATA (N-succinimidyl-5-acetylthioacetate), SPDP (N-succinimidyl-3-(2-pyridylthio)propionate), SPDB (N-succinimidyl-3-(2-

pyridyldithio)butyrate) and SMPT (N-succinimidyl-oxycarbonyl-alpha-methyl-alpha-(2-pyridyldithio)toluene)-, SPDB and SMPT. (See, e.g., Thorpe et al., 1987, Cancer Res. 47:5924-5931; Wawrzynczak et al., In Immunoconjugates: Antibody Conjugates in Radioimagery and Therapy of Cancer (C. W. Vogel ed., Oxford U. Press, 1987. See also U.S. Pat. No. 4,880,935.)

[0229] In other embodiments, the linker is a malonate linker (Johnson et al., 1995, Anticancer Res. 15:1387-93), a maleimidobenzoyl linker (Lau et al., 1995, Bioorg-Med-Chem. 3(10):1299-1304), or a 3'-N-amide analog (Lau et al., 1995, Bioorg-Med-Chem. 3(10):1305-12).

[0230] In yet other embodiments, the linker unit is not cleavable and the drug is released by antibody degradation. (See U.S. Publication No. 2005/0238649 incorporated by reference herein in its entirety and for all purposes).

[0231] In many embodiments, the linker is self-immolative. As used herein, the term "self-immolative Spacer" refers to a bifunctional chemical moiety that is capable of covalently linking together two spaced chemical moieties into a stable tripartite molecule. It will spontaneously separate from the second chemical moiety if its bond to the first moiety is cleaved. See for example, WO 2007059404A2, WO06110476A2, WO05112919A2, WO2010/062171, WO09/017394, WO07/089149, WO 07/018431, WO04/043493 and WO02/083180, which are directed to drug-cleavable substrate conjugates where the drug and cleavable substrate are optionally linked through a self-immolative linker and which are all expressly incorporated by reference.

[0232] Often the linker is not substantially sensitive to the extracellular environment. As used herein, "not substantially sensitive to the extracellular environment," in the context of a linker, means that no more than about 20%, 15%, 10%, 5%, 3%, or no more than about 1% of the linkers, in a sample of antibody-drug conjugate compound, are cleaved when the antibody-drug conjugate compound presents in an extracellular environment (for example, in plasma).

[0233] Whether a linker is not substantially sensitive to the extracellular environment can be determined, for example, by incubating with plasma the antibody-drug conjugate compound for a predetermined time period (for example, 2, 4, 8, 16, or 24 hours) and then quantitating the amount of free drug present in the plasma.

[0234] In other, non-mutually exclusive embodiments, the linker promotes cellular internalization. In certain embodiments, the linker promotes cellular internalization when conjugated to the therapeutic agent (that is, in the milieu of the linker-therapeutic agent moiety of the antibody-drug conjugate compound as described herein). In yet other

embodiments, the linker promotes cellular internalization when conjugated to both the auristatin compound and the multispecific antibodies of the invention.

[0235] A variety of exemplary linkers that can be used with the present compositions and methods are described in WO 2004-010957, U.S. Publication No. 2006/0074008, U.S. Publication No. 20050238649, and U.S. Publication No. 2006/0024317 (each of which is incorporated by reference herein in its entirety and for all purposes).

Drug Loading

[0236] Drug loading is represented by p and is the average number of Drug moieties per antibody in a molecule. Drug loading (" p ") may be 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more moieties (D) per antibody, although frequently the average number is a fraction or a decimal. Generally, drug loading of from 1 to 4 is frequently useful, and from 1 to 2 is also useful. ADCs of the invention include collections of antibodies conjugated with a range of drug moieties, from 1 to 20. The average number of drug moieties per antibody in preparations of ADC from conjugation reactions may be characterized by conventional means such as mass spectroscopy and, ELISA assay.

[0237] The quantitative distribution of ADC in terms of p may also be determined. In some instances, separation, purification, and characterization of homogeneous ADC where p is a certain value from ADC with other drug loadings may be achieved by means such as electrophoresis.

[0238] For some antibody-drug conjugates, p may be limited by the number of attachment sites on the antibody. For example, where the attachment is a cysteine thiol, as in the exemplary embodiments above, an antibody may have only one or several cysteine thiol groups, or may have only one or several sufficiently reactive thiol groups through which a linker may be attached. In certain embodiments, higher drug loading, e.g. $p > 5$, may cause aggregation, insolubility, toxicity, or loss of cellular permeability of certain antibody-drug conjugates. In certain embodiments, the drug loading for an ADC of the invention ranges from 1 to about 8; from about 2 to about 6; from about 3 to about 5; from about 3 to about 4; from about 3.1 to about 3.9; from about 3.2 to about 3.8; from about 3.2 to about 3.7; from about 3.2 to about 3.6; from about 3.3 to about 3.8; or from about 3.3 to about 3.7. Indeed, it has been shown that for certain ADCs, the optimal ratio of drug moieties per antibody may be less than 8, and may be about 2 to about 5. See US 2005-0238649 A1 (herein incorporated by reference in its entirety).

[0239] In certain embodiments, fewer than the theoretical maximum of drug moieties are conjugated to an antibody during a conjugation reaction. An antibody may contain, for example, lysine residues that do not react with the drug-linker intermediate or linker reagent,

as discussed below. Generally, antibodies do not contain many free and reactive cysteine thiol groups which may be linked to a drug moiety; indeed most cysteine thiol residues in antibodies exist as disulfide bridges. In certain embodiments, an antibody may be reduced with a reducing agent such as dithiothreitol (DTT) or tricarboylethylphosphine (TCEP), under partial or total reducing conditions, to generate reactive cysteine thiol groups. In certain embodiments, an antibody is subjected to denaturing conditions to reveal reactive nucleophilic groups such as lysine or cysteine.

[0240] The loading (drug/antibody ratio) of an ADC may be controlled in different ways, e.g., by: (i) limiting the molar excess of drug-linker intermediate or linker reagent relative to antibody, (ii) limiting the conjugation reaction time or temperature, (iii) partial or limiting reductive conditions for cysteine thiol modification, (iv) engineering by recombinant techniques the amino acid sequence of the antibody such that the number and position of cysteine residues is modified for control of the number and/or position of linker-drug attachments (such as thioMab or thioFab prepared as disclosed herein and in WO2006/034488 (herein incorporated by reference in its entirety)).

[0241] It is to be understood that where more than one nucleophilic group reacts with a drug-linker intermediate or linker reagent followed by drug moiety reagent, then the resulting product is a mixture of ADC compounds with a distribution of one or more drug moieties attached to an antibody. The average number of drugs per antibody may be calculated from the mixture by a dual ELISA antibody assay, which is specific for antibody and specific for the drug. Individual ADC molecules may be identified in the mixture by mass spectroscopy and separated by HPLC, e.g. hydrophobic interaction chromatography.

[0242] In some embodiments, a homogeneous ADC with a single loading value may be isolated from the conjugation mixture by electrophoresis or chromatography.

Methods of Determining Cytotoxic Effect of ADCs

[0243] Methods of determining whether a Drug or Antibody-Drug conjugate exerts a cytostatic and/or cytotoxic effect on a cell are known. Generally, the cytotoxic or cytostatic activity of an Antibody Drug conjugate can be measured by: exposing mammalian cells expressing a target protein of the Antibody Drug conjugate in a cell culture medium; culturing the cells for a period from about 6 hours to about 5 days; and measuring cell viability. Cell-based in vitro assays can be used to measure viability (proliferation), cytotoxicity, and induction of apoptosis (caspase activation) of the Antibody Drug conjugate.

[0244] For determining whether an Antibody Drug conjugate exerts a cytostatic effect, a thymidine incorporation assay may be used. For example, cancer cells expressing a target antigen at a density of 5,000 cells/well of a 96-well plated can be cultured for a 72-hour

period and exposed to 0.5 μ Ci of 3H-thymidine during the final 8 hours of the 72-hour period. The incorporation of 3H-thymidine into cells of the culture is measured in the presence and absence of the Antibody Drug conjugate.

[0245] For determining cytotoxicity, necrosis or apoptosis (programmed cell death) can be measured. Necrosis is typically accompanied by increased permeability of the plasma membrane; swelling of the cell, and rupture of the plasma membrane. Apoptosis is typically characterized by membrane blebbing, condensation of cytoplasm, and the activation of endogenous endonucleases. Determination of any of these effects on cancer cells indicates that an Antibody Drug conjugate is useful in the treatment of cancers.

[0246] Cell viability can be measured by determining in a cell the uptake of a dye such as neutral red, trypan blue, or ALAMAR™ blue (see, e.g., Page et al., 1993, Intl. J. Oncology 3:473-476). In such an assay, the cells are incubated in media containing the dye, the cells are washed, and the remaining dye, reflecting cellular uptake of the dye, is measured spectrophotometrically. The protein-binding dye sulforhodamine B (SRB) can also be used to measure cytotoxicity (Skehan et al., 1990, J. Natl. Cancer Inst. 82:1107-12).

[0247] Alternatively, a tetrazolium salt, such as MTT, is used in a quantitative colorimetric assay for mammalian cell survival and proliferation by detecting living, but not dead, cells (see, e.g., Mosmann, 1983, J. Immunol. Methods 65:55-63).

[0248] Apoptosis can be quantitated by measuring, for example, DNA fragmentation. Commercial photometric methods for the quantitative in vitro determination of DNA fragmentation are available. Examples of such assays, including TUNEL (which detects incorporation of labeled nucleotides in fragmented DNA) and ELISA-based assays, are described in Biochemica, 1999, no. 2, pp. 34-37 (Roche Molecular Biochemicals).

[0249] Apoptosis can also be determined by measuring morphological changes in a cell. For example, as with necrosis, loss of plasma membrane integrity can be determined by measuring uptake of certain dyes (e.g., a fluorescent dye such as, for example, acridine orange or ethidium bromide). A method for measuring apoptotic cell number has been described by Duke and Cohen, Current Protocols in Immunology (Coligan et al. eds., 1992, pp. 3.17.1-3.17.16). Cells also can be labeled with a DNA dye (e.g., acridine orange, ethidium bromide, or propidium iodide) and the cells observed for chromatin condensation and margination along the inner nuclear membrane. Other morphological changes that can be measured to determine apoptosis include, e.g., cytoplasmic condensation, increased membrane blebbing, and cellular shrinkage.

[0250] The presence of apoptotic cells can be measured in both the attached and "floating" compartments of the cultures. For example, both compartments can be collected

by removing the supernatant, trypsinizing the attached cells, combining the preparations following a centrifugation wash step (e.g., 10 minutes at 2000 rpm), and detecting apoptosis (e.g., by measuring DNA fragmentation). (See, e.g., Piazza et al., 1995, Cancer Research 55:3110-16).

[0251] In vivo, the effect of a therapeutic composition of the multispecific antibody of the invention can be evaluated in a suitable animal model. For example, xenogenic cancer models can be used, wherein cancer explants or passaged xenograft tissues are introduced into immune compromised animals, such as nude or SCID mice (Klein et al., 1997, Nature Medicine 3: 402-408). Efficacy can be measured using assays that measure inhibition of tumor formation, tumor regression or metastasis, and the like.

[0252] The therapeutic compositions used in the practice of the foregoing methods can be formulated into pharmaceutical compositions comprising a carrier suitable for the desired delivery method. Suitable carriers include any material that when combined with the therapeutic composition retains the anti-tumor function of the therapeutic composition and is generally non-reactive with the patient's immune system. Examples include, but are not limited to, any of a number of standard pharmaceutical carriers such as sterile phosphate buffered saline solutions, bacteriostatic water, and the like (see, generally, Remington's Pharmaceutical Sciences 16th Edition, A. Osal., Ed., 1980).

Antibody Compositions for In Vivo Administration

[0253] Formulations of the antibodies used in accordance with the present invention are prepared for storage by mixing an antibody having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers (Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. [1980]), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e.g. Zn-

protein complexes); and/or non-ionic surfactants such as TWEEN™, PLURONICS™ or polyethylene glycol (PEG).

[0254] The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. For example, it may be desirable to provide antibodies with other specificities. Alternatively, or in addition, the composition may comprise a cytotoxic agent, cytokine, growth inhibitory agent and/or small molecule antagonist. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

[0255] The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. (1980).

[0256] The formulations to be used for in vivo administration should be sterile, or nearly so. This is readily accomplished by filtration through sterile filtration membranes.

[0257] Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, e.g. films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919), copolymers of L-glutamic acid and .gamma. ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT™ (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods.

[0258] When encapsulated antibodies remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37°C, resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies can be devised for stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues,

lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

Administrative modalities

[0259] The antibodies and chemotherapeutic agents of the invention are administered to a subject, in accord with known methods, such as intravenous administration as a bolus or by continuous infusion over a period of time, by intramuscular, intraperitoneal, intracerebrospinal, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. Intravenous or subcutaneous administration of the antibody is preferred.

Treatment modalities

[0260] In the methods of the invention, therapy is used to provide a positive therapeutic response with respect to a disease or condition. By "positive therapeutic response" is intended an improvement in the disease or condition, and/or an improvement in the symptoms associated with the disease or condition. For example, a positive therapeutic response would refer to one or more of the following improvements in the disease: (1) a reduction in the number of neoplastic cells; (2) an increase in neoplastic cell death; (3) inhibition of neoplastic cell survival; (5) inhibition (i.e., slowing to some extent, preferably halting) of tumor growth; (6) an increased patient survival rate; and (7) some relief from one or more symptoms associated with the disease or condition.

[0261] Positive therapeutic responses in any given disease or condition can be determined by standardized response criteria specific to that disease or condition. Tumor response can be assessed for changes in tumor morphology (i.e., overall tumor burden, tumor size, and the like) using screening techniques such as magnetic resonance imaging (MRI) scan, x-radiographic imaging, computed tomographic (CT) scan, bone scan imaging, endoscopy, and tumor biopsy sampling including bone marrow aspiration (BMA) and counting of tumor cells in the circulation.

[0262] In addition to these positive therapeutic responses, the subject undergoing therapy may experience the beneficial effect of an improvement in the symptoms associated with the disease.

[0263] Thus for B cell tumors, the subject may experience a decrease in the so-called B symptoms, i.e., night sweats, fever, weight loss, and/or urticaria. For pre-malignant conditions, therapy with an multispecific therapeutic agent may block and/or prolong the time before development of a related malignant condition, for example, development of multiple myeloma in subjects suffering from monoclonal gammopathy of undetermined significance (MGUS).

[0264] An improvement in the disease may be characterized as a complete response. By “complete response” is intended an absence of clinically detectable disease with normalization of any previously abnormal radiographic studies, bone marrow, and cerebrospinal fluid (CSF) or abnormal monoclonal protein in the case of myeloma.

[0265] Such a response may persist for at least 4 to 8 weeks, or sometimes 6 to 8 weeks, following treatment according to the methods of the invention. Alternatively, an improvement in the disease may be categorized as being a partial response. By “partial response” is intended at least about a 50% decrease in all measurable tumor burden (i.e., the number of malignant cells present in the subject, or the measured bulk of tumor masses or the quantity of abnormal monoclonal protein) in the absence of new lesions, which may persist for 4 to 8 weeks, or 6 to 8 weeks.

[0266] Treatment according to the present invention includes a “therapeutically effective amount” of the medicaments used. A “therapeutically effective amount” refers to an amount effective, at dosages and for periods of time necessary, to achieve a desired therapeutic result.

[0267] A therapeutically effective amount may vary according to factors such as the disease state, age, sex, and weight of the individual, and the ability of the medicaments to elicit a desired response in the individual. A therapeutically effective amount is also one in which any toxic or detrimental effects of the antibody or antibody portion are outweighed by the therapeutically beneficial effects.

[0268] A “therapeutically effective amount” for tumor therapy may also be measured by its ability to stabilize the progression of disease. The ability of a compound to inhibit cancer may be evaluated in an animal model system predictive of efficacy in human tumors.

[0269] Alternatively, this property of a composition may be evaluated by examining the ability of the compound to inhibit cell growth or to induce apoptosis by in vitro assays known to the skilled practitioner. A therapeutically effective amount of a therapeutic compound may decrease tumor size, or otherwise ameliorate symptoms in a subject. One of ordinary skill in the art would be able to determine such amounts based on such factors as the subject's size, the severity of the subject's symptoms, and the particular composition or route of administration selected.

[0270] Dosage regimens are adjusted to provide the optimum desired response (e.g., a therapeutic response). For example, a single bolus may be administered, several divided doses may be administered over time or the dose may be proportionally reduced or increased as indicated by the exigencies of the therapeutic situation. Parenteral compositions may be formulated in dosage unit form for ease of administration and

uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subjects to be treated; each unit contains a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier.

[0271] The specification for the dosage unit forms of the present invention are dictated by and directly dependent on (a) the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active compound for the treatment of sensitivity in individuals.

[0272] The efficient dosages and the dosage regimens for the multispecific antibodies used in the present invention depend on the disease or condition to be treated and may be determined by the persons skilled in the art.

[0273] An exemplary, non-limiting range for a therapeutically effective amount of an multispecific antibody used in the present invention is about 0.1-100 mg/kg, such as about 0.1-50 mg/kg, for example about 0.1-20 mg/kg, such as about 0.1-10 mg/kg, for instance about 0.5, about such as 0.3, about 1, or about 3 mg/kg. In another embodiment, he antibody is administered in a dose of 1 mg/kg or more, such as a dose of from 1 to 20 mg/kg, e.g. a dose of from 5 to 20 mg/kg, e.g. a dose of 8 mg/kg.

[0274] A medical professional having ordinary skill in the art may readily determine and prescribe the effective amount of the pharmaceutical composition required. For example, a physician or a veterinarian could start doses of the medicament employed in the pharmaceutical composition at levels lower than that required in order to achieve the desired therapeutic effect and gradually increase the dosage until the desired effect is achieved.

[0275] In one embodiment, the multispecific antibody is administered by infusion in a weekly dosage of from 10 to 500 mg/kg such as of from 200 to 400 mg/kg Such administration may be repeated, e.g., 1 to 8 times, such as 3 to 5 times. The administration may be performed by continuous infusion over a period of from 2 to 24 hours, such as of from 2 to 12 hours.

[0276] In one embodiment, the multispecific antibody is administered by slow continuous infusion over a long period, such as more than 24 hours, if required to reduce side effects including toxicity.

[0277] In one embodiment the multispecific antibody is administered in a weekly dosage of from 250 mg to 2000 mg, such as for example 300 mg, 500 mg, 700 mg, 1000 mg, 1500 mg or 2000 mg, for up to 8 times, such as from 4 to 6 times. The administration may be performed by continuous infusion over a period of from 2 to 24 hours, such as of from 2 to

12 hours. Such regimen may be repeated one or more times as necessary, for example, after 6 months or 12 months. The dosage may be determined or adjusted by measuring the amount of compound of the present invention in the blood upon administration by for instance taking out a biological sample and using anti-idiotypic antibodies which target the antigen binding region of the multispecific antibody.

[0278] In a further embodiment, the multispecific antibody is administered once weekly for 2 to 12 weeks, such as for 3 to 10 weeks, such as for 4 to 8 weeks.

[0279] In one embodiment, the multispecific antibody is administered by maintenance therapy, such as, e.g., once a week for a period of 6 months or more.

[0280] In one embodiment, the multispecific antibody is administered by a regimen including one infusion of an multispecific antibody followed by an infusion of an multispecific antibody conjugated to a radioisotope. The regimen may be repeated, e.g., 7 to 9 days later.

[0281] As non-limiting examples, treatment according to the present invention may be provided as a daily dosage of an antibody in an amount of about 0.1-100 mg/kg, such as 0.5, 0.9, 1.0, 1.1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 40, 45, 50, 60, 70, 80, 90 or 100 mg/kg, per day, on at least one of day 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40, or alternatively, at least one of week 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 after initiation of treatment, or any combination thereof, using single or divided doses of every 24, 12, 8, 6, 4, or 2 hours, or any combination thereof.

[0282] In some embodiments the multispecific antibody molecule thereof is used in combination with one or more additional therapeutic agents, e.g. a chemotherapeutic agent. Non-limiting examples of DNA damaging chemotherapeutic agents include topoisomerase I inhibitors (e.g., irinotecan, topotecan, camptothecin and analogs or metabolites thereof, and doxorubicin); topoisomerase II inhibitors (e.g., etoposide, teniposide, and daunorubicin); alkylating agents (e.g., melphalan, chlorambucil, busulfan, thiotepa, ifosfamide, carmustine, lomustine, semustine, streptozocin, decarbazine, methotrexate, mitomycin C, and cyclophosphamide); DNA intercalators (e.g., cisplatin, oxaliplatin, and carboplatin); DNA intercalators and free radical generators such as bleomycin; and nucleoside mimetics (e.g., 5-fluorouracil, capecitabine, gemcitabine, fludarabine, cytarabine, mercaptopurine, thioguanine, pentostatin, and hydroxyurea).

[0283] Chemotherapeutic agents that disrupt cell replication include: paclitaxel, docetaxel, and related analogs; vincristine, vinblastin, and related analogs; thalidomide, lenalidomide, and related analogs (e.g., CC-5013 and CC-4047); protein tyrosine kinase inhibitors (e.g.,

imatinib mesylate and gefitinib); proteasome inhibitors (e.g., bortezomib); NF- κ B inhibitors, including inhibitors of I κ B kinase; antibodies which bind to proteins overexpressed in cancers and thereby downregulate cell replication (e.g., trastuzumab, rituximab, cetuximab, and bevacizumab); and other inhibitors of proteins or enzymes known to be upregulated, over-expressed or activated in cancers, the inhibition of which downregulates cell replication.

[0284] In some embodiments, the antibodies of the invention can be used prior to, concurrent with, or after treatment with Velcade® (bortezomib).

[0285] All cited references are herein expressly incorporated by reference in their entirety.

[0286] Whereas particular embodiments of the invention have been described above for purposes of illustration, it will be appreciated by those skilled in the art that numerous variations of the details may be made without departing from the invention as described in the appended claims.

Examples

[0287] Examples are provided below to illustrate the present invention. These examples are not meant to constrain the present invention to any particular application or theory of operation. For all constant region positions discussed in the present invention, numbering is according to the EU index as in Kabat (Kabat et al., 1991, Sequences of Proteins of Immunological Interest, 5th Ed., United States Public Health Service, National Institutes of Health, Bethesda, entirely incorporated by reference). Those skilled in the art of antibodies will appreciate that this convention consists of nonsequential numbering in specific regions of an immunoglobulin sequence, enabling a normalized reference to conserved positions in immunoglobulin families. Accordingly, the positions of any given immunoglobulin as defined by the EU index will not necessarily correspond to its sequential sequence.

EXAMPLE 1. Prototype “triple F” bispecific antibody.

[0288] The present invention describes novel immunoglobulin compositions that co-engage a first and second antigen. One heavy chain of the antibody contains a single chain Fv (“scFv”, as defined below) and the other heavy chain is a “regular” Fab format, comprising a variable heavy chain and a light chain (see Fig. 1). This structure is sometimes referred to herein as “triple F” format (scFv-Fab-Fc). The two chains are brought together by the dimeric Fc region (see Fig. 2). The Fc region can be modified by amino acid substitution to allow for efficient purification of the “triple F” heterodimer. Further, the Fc region can be modified by amino acid substitution to promote the formation of the “triple F” heterodimer. Examples of Fc substitutions are described more fully below.

[0289] Fc substitutions can be included in the “triple F” format to allow for efficient purification of the desired “triple F” heterodimer over the undesired dual scFv-Fc and mAb homodimers. An example of this is in the inclusion of Fc substitutions that alter the isoelectric point (pI) of each monomer so that each monomer has a different pI. In this case the desired “triple F” heterodimer will have a different pI than that of the undesired dual scFv-Fc and mAb homodimers, thus facilitating isoelectric purification of the “triple F” heterodimer (e.g., anionic exchange columns, cationic exchange columns). These substitutions also aid in the determination and monitoring of any contaminating dual scFv-Fc and mAb homodimers post-purification (e.g., IEF gels, cIEF, and analytical IEX columns). See Fig. 3 for a list of substitutions that can be made in Fc monomer 1 and Fc monomer 2 to allow for efficient purification of the desired “triple F” heterodimer.

[0290] Fc substitutions can be included in the “triple F” format to “skew” the formation toward the desired “triple F” heterodimer over the undesired dual scFv-Fc and mAb homodimers. For example, see Fig. 4 for a list of substitutions that can be made in Fc monomer 1 and Fc monomer 2 to “skew” production toward the “triple F” heterodimer. Amino acid substitutions listed in Fig. 3 and Fig. 4 can be combined, leading to an increased yield of “triple F” heterodimer that can be easily purified away from any contaminating dual scFv-Fc and mAb homodimers.

[0291] After optimization of an scFv domain for inclusion in the “triple F” format, an optimized scFv domain can be coupled with a variety of standard antibody heavy chains in a convenient fashion. For example, an anti-CD3 scFv for recruiting T cell cytotoxicity can be coupled with a variety of anti-tumor antigen antibody heavy chains (e.g., those binding CD5, CD20, CD30, CD40, CD33, CD38, EGFR, EpCAM, Her2, HM1.24, or other tumor antigen). Further examples of optimized scFv domains that can be conveniently coupled with standard antibody heavy chains include anti-CD16 scFv for natural killer cell cytotoxicity; anti-CD32b scFv for inhibitory activity (here the coupled antibody heavy chain would bind, e.g., CD19, CD40, CD79a, CD79b, or other immune receptors); and anti-transferrin receptor scFv, anti-insulin receptor, or anti-LRP1 for transport across the blood-brain barrier.

EXAMPLE 2. Multi-specific antibodies derived from the “triple F” format.

[0292] Multi-specific antibodies can be constructed by attaching additional scFv or Fab domains that bind a third antigen to the C-terminus of one of the “triple F” heavy chains. See Fig. 5 for examples. Alternatively, the C-terminal scFv or Fab may bind the first or second antigen, thus conferring bivalency and an increase in overall binding affinity for that antigen.

[0293] Multi-specific antibodies can also be constructed by coupling the scFv-Fc heavy chain of the “triple F” format may with rearranged antibody heavy chains as depicted in Fig.

6. Such rearranged heavy chains may include an additional Fv region that binds a third antigen or an additional Fv region that binds the first antigen or second antigen, thus conferring bivalency and an increase in overall binding affinity for that antigen.

EXAMPLE 3. Anti-CD19 Fab x anti-CD3 scFv “triple F” bispecific.

[0294] Amino acid sequences for anti-CD19 Fab x anti-CD3 scFv “triple F” bispecifics are listed in Fig. 7. Amino acid substitutions made to allow for efficient purification of the desired “triple F” heterodimer over the undesired dual scFv-Fc and mAb homodimers are underlined. Amino acid sequences for preferred humanized anti-CD3 variable regions are listed in Fig. 8 (with CDRs underlined). Some examples of expression and purification of the desired “triple F” species and its bioactivity are given below.

[0295] The production of XENP11874, a “triple F” bispecific with an anti-CD19 Fab and anti-CD3 scFv, is outlined in Fig. 9. In Fig. 9A, the ion exchange purification of the desired “triple F” heterodimer from the undesired dual scFv-Fc and mAb homodimers is shown. The purity of the “triple F” fraction was checked by IEF gel, shown in Fig. 9B. Finally, SEC was used to confirm the homogenous size of the “triple F” product (see Fig. 9C).

[0296] XENP11874, anti-CD19 Fab x anti-CD3 scFv “triple F” bispecific, was shown to have potent bioactivity. The ability of XENP11874 to potentially recruit T cells for B cell depletion is shown in Fig. 10.

[0297] The production of XENP11924, a “triple F” bispecific with an anti-CD19 Fab and anti-CD3 scFv, is outlined in Fig. 11. In Fig. 11A, the ion exchange purification of the desired “triple F” heterodimer from the undesired dual scFv-Fc and mAb homodimers is shown. The purity of the “triple F” fraction was checked by IEF gel, shown in Fig. 11B. Finally, SEC was used to confirm the homogenous size of the “triple F” product (see Fig. 11C).

[0298] XENP11924, anti-CD19 Fab x anti-CD3 scFv “triple F” bispecific, was shown to have potent bioactivity. The ability of XENP11924 to potentially recruit T cells for the killing of the Raji tumor cell line is shown in Fig. 12.

EXAMPLE 4. Anti-CD38 Fab x anti-CD3 scFv “triple F” bispecific.

[0299] Amino acid sequences for anti-CD38 Fab x anti-CD3 scFv “triple F” bispecifics are listed in Fig. 13. Amino acid substitutions made to allow for efficient purification of the desired “triple F” heterodimer over the undesired dual scFv-Fc and mAb homodimers are underlined. Some examples of expression and purification of the desired “triple F” species and its bioactivity are given below.

[0300] The production of XENP11925, a “triple F” bispecific with an anti-CD38 Fab and anti-CD3 scFv, is outlined in Fig. 14. In Fig. 14A, the ion exchange purification of the desired “triple F” heterodimer from the undesired dual scFv-Fc and mAb homodimers is shown. The purity of the “triple F” fraction was checked by IEF gel, shown in Fig. 14B. Finally, SEC was used to confirm the homogenous size of the “triple F” product (see Fig. 14C).

[0301] XENP11925, anti-CD38 Fab x anti-CD3 scFv “triple F” bispecific, was shown to have potent bioactivity. The ability of XENP11925 to potently recruit T cells for the killing of the RPMI8226 tumor cell line is shown in Fig. 15.

CLAIMS

We claim:

1. A heterodimeric antibody comprising:
 - a) a first heavy chain comprising:
 - i) a first Fc domain; and
 - ii) a single chain Fv region that binds a first antigen; and
 - b) a second heavy chain comprising:
 - i) a second Fc domain;
 - ii) a first variable heavy chain; and
 - iii) a first variable light chain;wherein said first and second Fc domains are different.
2. A heterodimeric antibody according to claim 1 wherein the pIs of said first heavy chain and said second heavy chain are at least 0.5 logs apart.
3. A heterodimeric antibody according to claim 1 or 2 further comprising an additional antigen binding domain.
4. A heterodimeric antibody according to claim 3 wherein said additional antigen binding domain is selected from the group consisting of an scFv and a Fab.
5. A heterodimeric antibody according to any of claims 1 to 4 wherein said scFv region binds to CD3 and said first heavy and light chains form a domain that binds to an antigen selected from the group consisting of CD5, CD20, CD30, CD33, CD38, CD40, EGFR, EpCAM, Her2, HM1.24.
6. A heterodimeric antibody according to any one of claims 1 -5 wherein said antibody has a structure selected from Figure 1B, 2A, 2B, 2C, 2D, 2D 3A, 3B, 3C, 3D and 3E.
7. A heterodimeric antibody according to any of claims 1-6 wherein the first Fc domain and said second Fc domain comprises a set of corresponding variants selected from Figure 4.
8. A heterodimeric antibody according to any of claims 1-7 wherein the first Fc domain and said second Fc domain comprises a set of corresponding variants selected from Figure 5.

9. A heterodimeric antibody according to any of claims 1-8 wherein the first Fc domain and said second Fc domain comprises a set of corresponding variants selected from Figure 6.
10. A heterodimeric antibody according to any previous claim wherein the pl substitutions in monomer one are ISO(-): I199T/N203D/K274Q/R355Q/N384S/K392N/V397M/Q419E/DEL44 and the pl substitutions in monomer 2 are ISO(+RR): Q196K/I199T/P217R/P228R/N276K.
11. A heterodimeric antibody according to any previous claim further comprising a conjugated drug.
12. A composition comprising a heterodimeric antibody according to any previous claim and a carrier.
13. A nucleic acid encoding the first heavy chain of any previous claim.
14. A nucleic acid encoding the second heavy chain of any previous claim.
15. A nucleic acid encoding the second light chain of any previous claim.
16. A nucleic acid encoding the third light chain of any previous claim.
17. An expression vector comprising a nucleic acid according to any of claims 13-16.
18. A host cell comprising a nucleic acid of any of claims 13-16.
19. A host cell comprising the expression vector of claim 17.
20. A method of making an antibody according to any of claims 1-11 comprising
 - a) Culturing a host cell comprising:
 - i) a nucleic acid encoding said first heavy chain; and
 - ii) a nucleic acid encoding said second heavy chain;
 - b) under conditions wherein said nucleic acids are expressed.
21. A method of purifying a heterodimeric antibody comprising:
 - a) providing an antibody according to any of claims 1-11;
 - b) loading said composition onto an ion exchange column; and
 - c) collecting the heterodimeric fraction.

22. A method of treating an individual in need thereof comprising administering the composition of any of claims 1-12.

Fcs

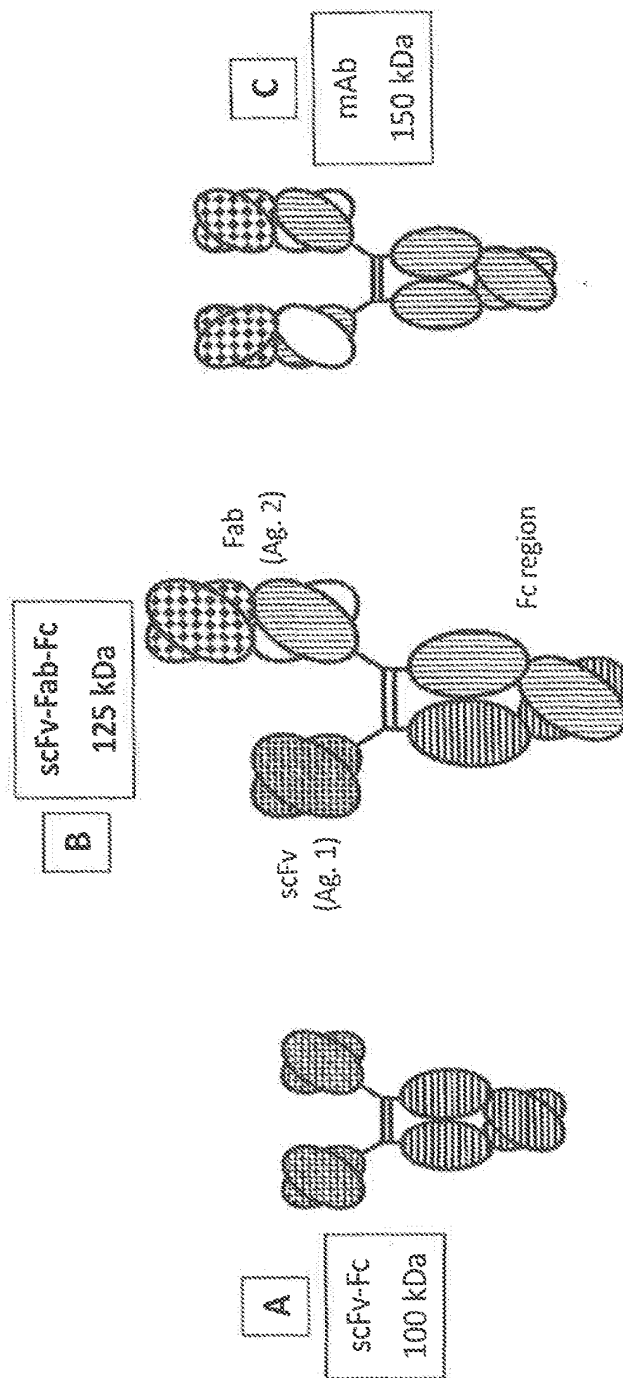
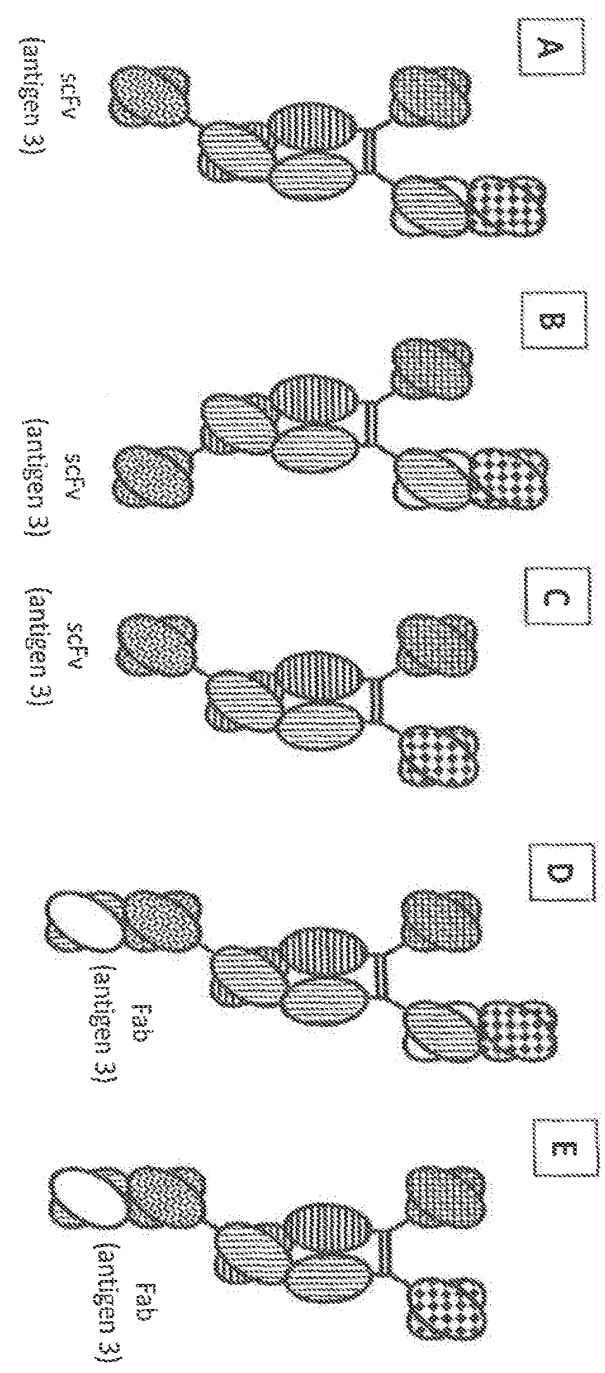
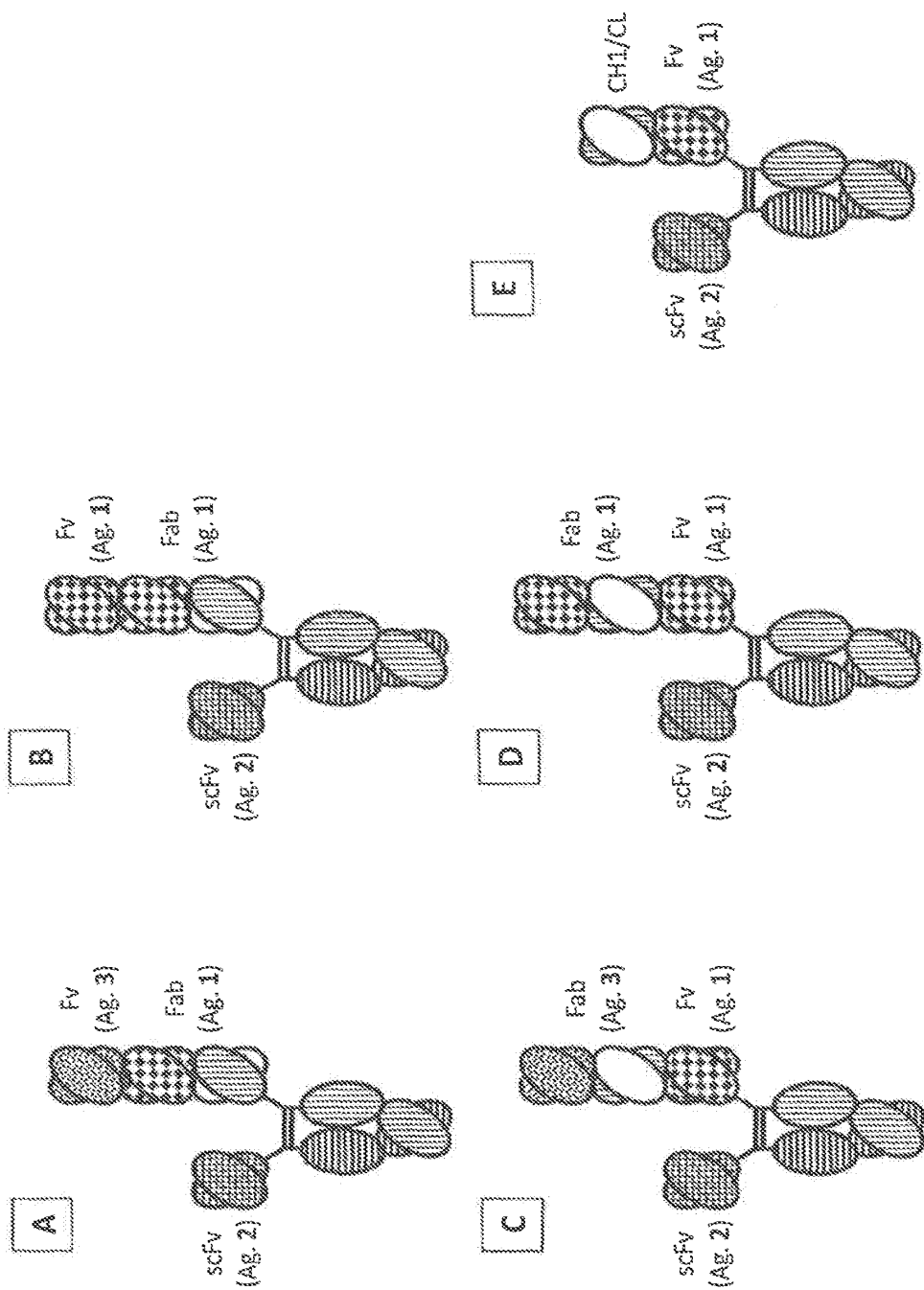


Figure 1

Fig 2
Figure 2





Handwritten notes: "3" and "5" with arrows pointing to the diagrams.

Figure 3A Preferred steric variants that favor Fc heterodimerization.

Monomer 1	Monomer 2
F405A	T394F
S364D	Y349K
S364E	L368K
S364E	Y349K
S364F	K370G
S364H	Y349K
S364H	Y349T
S364Y	K370G
T411K	K370E
V397S/F405A	T394F
K370R/T411K	K370E/T411E
L351E/S364D	Y349K/L351K
L351E/S364E	Y349K/L351K
L351E/T366D	L351K/T366K
P395T/V397S/F405A	T394F
S364D/K370G	S364Y/K370R
S364D/T394F	Y349K/F405A
S364E/F405A	Y349K/T394F
S364E/F405S	Y349K/T394Y
S364E/T411E	Y349K/D401K
S364H/D401K	Y349T/T411E
S364H/F405A	Y349T/T394F
S364H/T394F	Y349T/F405A
Y349C/S364E	Y349K/S354C
L351E/S364D/F405A	Y349K/L351K/T394F
L351K/S364H/D401K	Y349T/L351E/T411E
S364E/T411E/F405A	Y349K/T394F/D401K
S364H/D401K/F405A	Y349T/T394F/T411E
S364H/F405A/T411E	Y349T/T394F/D401K
Y349T	S364H
T394F	F405A
Y349T/T394F	S364H/F405A
K370E	T411K
K370E/T411D	T411K
K370E/T411E	K370R/T411K
L368E/K409E	L368K
Y349T/T411E	S364H/D401K
Y349T/T394F/S354C	S364H/F405A/Y349C
T411E	D401K
T411E	D401R/T411R
Q347E/K360E	Q347R
L368E	S364K
L368E/K370S	S364K
L368E/K370T	S364K
L368E/D401R	S364K
L368E/D401N	S364K
L368E	E357S/S364K
L368E	S364K/K409E
L368E	S364K/K409V
L368D	S364K

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Figure 298 4 B Specifically preferred steric variants that favor Fc heterodimerization.

Variant 1	Variant 2
F405A	T394F
S364D	Y349K
S364E	Y349K
S364H	Y349T
L351K	L351E
D401K	T411E
S364D/T394F	Y349K/F405A
S364E/F405A	Y349K/T394F
S364H/D401K	Y349T/T411E
S364H/F405A	Y349T/T394F
S364H/T394F	Y349T/F405A
L351K/S364H/D401K	Y349T/L351E/T411E
S364H/D401K/F405A	Y349T/T394F/T411E
S364H/F405A/T411E	Y349T/T394F/D401K
Y349T	S364H
T394F	F405A
Y349T/T394F	S364H/F405A
K370E	T411K
K370E/T411D	T411K
K370E/T411E	K370R/T411K
L368E/K409E	L368K
Y349T/T411E	S364H/D401K
Y349T/T394F/S354C	S364H/F405A/Y349C
T411E	D401K
T411E	D401R/T411R
Q347E/K360E	Q347R
L368E	S364K
L368E/K370S	S364K
L368E/K370T	S364K
L368E/D401R	S364K
L368E/D401N	S364K
L368E	E357S/S364K
L368E	S364K/K409E
L368E	S364K/K409V
L368D	S364K

FIGURE 5
PREFERRED HETERODIMERIZATION VARIANTS

Fc monomer 1 substitutions	Fc monomer 2 substitutions
Y407T	T366Y
F405A	T394W
T366Y/F405A	T394W/Y407T
Y407A	T366W
T366S/L368A/Y407V	T366W
T366S/L368A/Y407V/Y349C	T366W/S354C
K392D/K409D	E356K/D399K
K370D/K392D/K409D	E356K/E357K/D399K
Y349T	S364H
T394F	F405A
Y349T/T394F	S364H/F405A
K370E	T411K
K370E/T411D	T411K
K370E/T411E	K370R/T411K
L368E/K409E	L368K
Y349T/T411E	S364H/D401K
Y349T/T394F/S354C	S364H/F405A/Y349C
T411E	D401K
T411E	D401R/T411R
Q347E/K360E	Q347R
L368E	S364K
I199T/N203D/K247Q/R355Q/N384S/ K392N/V397M/Q419E/K447_ (deletion of K447)	Q196K/I199T/P217R/P228R/N276K
I199T/N203D/K247Q/R355Q/N384S /K392N/V397M/Q419E/K447_	Q196K/I199T/N276K
N384S/K392N/V397M/Q419E	N276K
D221E/P228E/L368E	D221R/P228R/K409R
C220E/P228E/L368E	C220R/E224R/P228R/K409R
F405L	K409R
T366I/K392M/T394W	F405A/Y407V
T366V/K409F	L351Y/Y407A
T366A/K392E/K409F/T411E	D399R/S400R/Y407A
Y349T	S364H
T394F	F405A
Y349T/T394F	S364H/F405A
K370E	T411K
K370E/T411D	T411K
K370E/T411E	K370R/T411K
L368E/K409E	L368K
Y349T/T411E	S364H/D401K
Y349T/T394F/S354C	S364H/F405A/Y349C
T411E	D401K
T411E	D401R/T411R

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Figure 5 (cont.)

Q347E/K360E	Q347R
L368E	S364K
L368E/K370S	S364K
L368E/K370T	S364K
L368E/D401R	S364K
L368E/D401N	S364K
L368E	E357S/S364K
L368E	S364K/K409E
L368E	S364K/K409V
L368D	S364K

Figure 821

Fc monomer 1 substitutions	Fc monomer 2 substitutions
I199T/N203D/K247Q/R355Q/N384S/K392N/V397M/Q419E/K447_	Q196K/I199T/P217R/P228R/N276K
I199T/N203D/K247Q/R355Q/N384S/K392N/V397M/Q419E/K447_	Q196K/I199T/N276K
N203D/K247Q/R355Q/N384S/K392N/V397M/Q419E/K447_	Q196K/P217R/P228R/N276K
N203D/K247Q/R355Q/N384S/K392N/V397M/Q419E/K447_	Q196K/N276K
N203D/K247Q/R355Q/N384S/K392N/Q419E/K447_	Q196K/P217R/P228R/N276K
N203D/K247Q/R355Q/N384S/K392N/Q419E/K447_	Q196K/N276K

Figure 7

XENP11874_HC-Fab

EVQLVESGGGLVQPGGSLKLSCAASGYTFTSYVMHWVRQAPGKGLEWIGYINPYNDGTYNEKFQGRVTLSDDK
SISTAYMELSSLRSEDTAMYVCARGTYXYGTRVFDYWGQGTTLTVTVSSASTKGPSVFPLAPSSKSTSGGTAALGC
LVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVFPSSSLGTQTYICNVNHPKSTKVDKKVEPK
SCDKTHTCPPCPAPELLRGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKTKPR
REQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKARPAPIEKTIISKAKGQPREPOVYITLPPSREEMTKNQVSLT
CLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSEFFLYSKLTVDKSRWQQGNVFCSCVMHEALHNYTQKS
LSLSPGK

XENP11874_HC-scFv

EVQLVESGGGLVQPGGSLRLSCAASGYTFNTYAMNWVRQAPGKGLEWVGRIIRSKYNNYATYYADSVKGEFTISR
DDSKNTLYLQMNSLRADDTAVYYCVRSNGFNSYVSNFAYWGQGTTLTVTVSSGGGGSGGGGSGGGGSGQAVVTQEP
SLTVSPGGTVTLTCSSTGAVTTSNYANWVQXKPGQAPRGLIGGTNKPAPGVPARFSGSLLGSKAALTLSGAQP
EDHAAYYCALWYSNLWVFGGGTKLTVLERKSSDKTHTCPRCPPAPELLRGPSVFLFPPKPKDTLMISRTPEVTCV
VVDVSHEDPEVKFKWYVDGVEVHNAKTKPRREQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKARPAPIEKTI
ISKAKGQPREPOVYITLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSEFFLYSKL
TVDKSRWQQGNVFCSCVMHEALHNYTQKSLSLSPGK

XENP11874_LC

DIVMTQSPATLSLSPGERATLSCRSSKSLQNVNGNTYLYWFQXKPGQSPQLLIYRMSNLNGGVPRFSGSGSGT
EFTLTITSSLEPEDFAVYYCMQHLEYPITFGAGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCCLINNFYPRE
AKVQWKVDNALQSGNSQESVTEQDSKDSSTYSLSSTLTLSKADYEKHKVYACEVTHRQGLSSPVTKSFNRGEC

XENP11924_HC-Fab

EVQLVESGGGLVQPGGSLKLSCAASGYTFTSYVMHWVRQAPGKGLEWIGYINPYNDGTYNEKFQGRVTISSDK
SISTAYMELSSLRSEDTAMYVCARGTYXYGTRVFDYWGQGTTLTVTVSSASTKGPSVFPLAPSSKSTSGGTAALGC
LVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVFPSSSLGTQTYTCNVDPKPKSTKVDKKVEPK
SCDKTHTCPPCPAPELLRGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVQFNWYVDGVEVHNAKTKPR
REQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKARPAPIEKTIISKAKGQPREPOVYITLPPSQEEMTKNQVSLT
CLVKGFYPSDIAVEWESSGQPENNYNTTTPMQLSDGSEFFLYSKLTVDKSRWQEGNVFCSCVMHEALHNYTQKS
LSLSPG

Figure 7 (continued)

XENP11924_HC-scFv

EVQLVESGGGLVQPGGSLRLSCAASGFTFTNTYAMNWVRQAPCKGLEWVGRIRSKYNNYATYYADSVKGRFTISR
DDSKNTLYLQMNSLRAEDTAVYYCVRHGNFGNCSYVSWFAYWCQGTLLVTVSSGGGGSGGGGSGGGGSQAVVTQEP
SLTVSPGGTVTLTCGSETGAVTTSNYANWVQKPGQAPRGLIGGTNKRAPGVPARFSGSLLGCKAALTLSGAQP
EDEAAYYCALWYSHLWVFGGGTKLTVLERKSSDKTNTCPRCPAPELLRGPSVFLFPPKPKDTLMISRTPEVTCV
VVDVSHEDPEVKFKWYVDGVEVHNAKTRFREEQYNSTYRVVSVLTVYLHQDWLNGKEYCKVKSNKARPAPIEKTI
SKAKGQPREPQVYITLPSRREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKL
TVDKSKRWQGCNVFSCSVMHEALHNHYTQKSLSLSPGK

XENP11924_LC

DIIVMTQSPATLSLSPGERATLSCRSSKSLQNVNGNTYLYWFQKPKGQSPQLLIYKMSNLNSGVPDRFGSGSGGT
EFTLTITSSLEPEDFAVYYCMQHLEYPIITFCAGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPRE
AKVQWVKVDNALQSGNSQESVTEQDSKDSSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSPVTKSFNBGEC

Figure 8

Humanized Anti-CD3 VH with Kabat CDRs underlined

EVQLVESGGGLVQPGGSLRLSCAASGFTFNTYAMNWRQAPGKGLEWVGRIIRSKYNNYATYYADSVKGRFTISR
DDSKNTLYLQMNSLRAEDTAVYYCVRHGNFGNSYVSWFAYWGQGTLLTVSS

Humanized Anti-CD3 VL with Kabat CDRs underlined

QAVVTQEPSTLVSPGGTVTLTCGSSTGAVTTSNYANWVQOKPGQAPRGLIGGTNKKRPFQVPAFSGSLLGGKAA
LTLGAQPEDEAEYYCALWYSNLWVFGGGTKLTVL

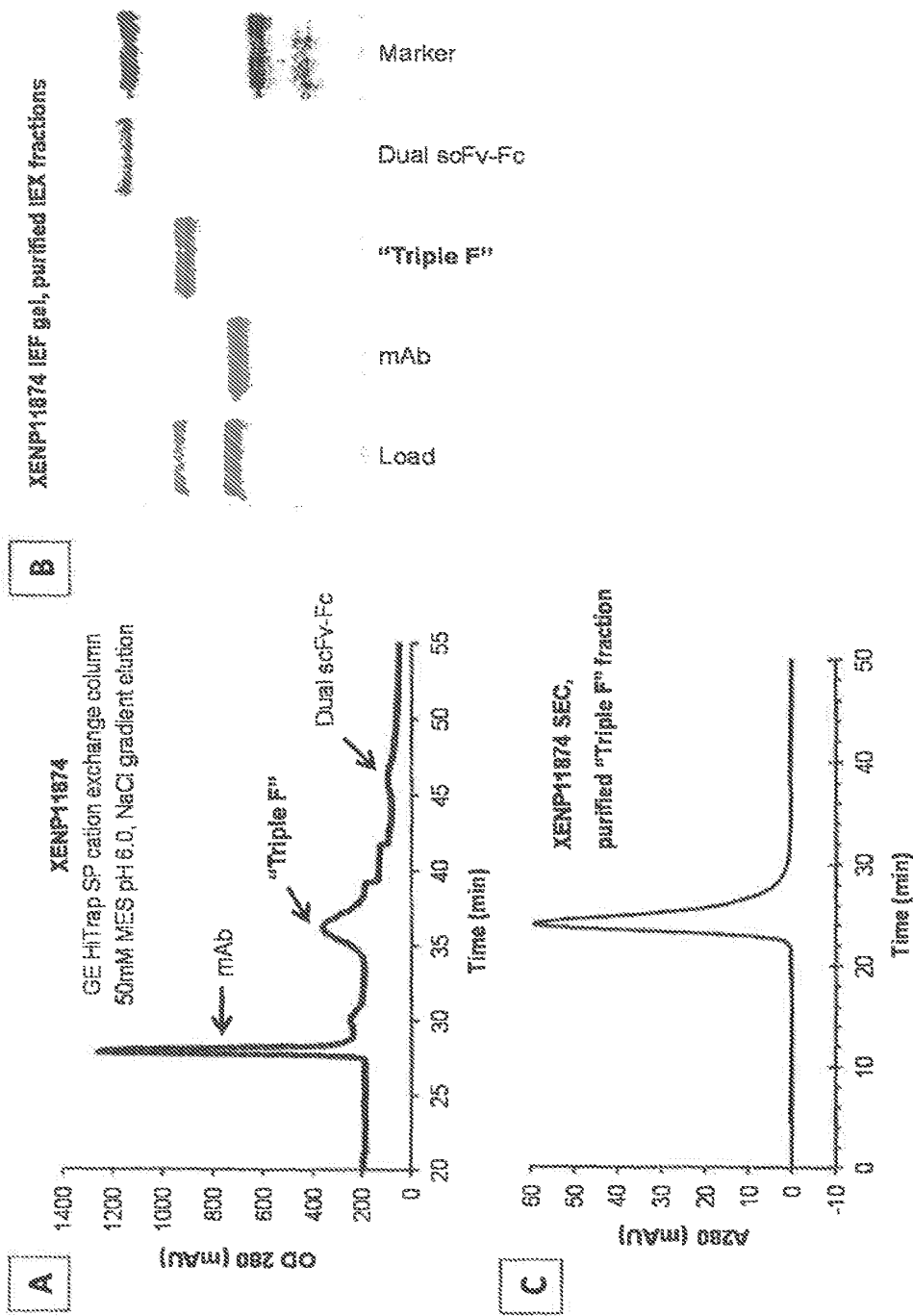
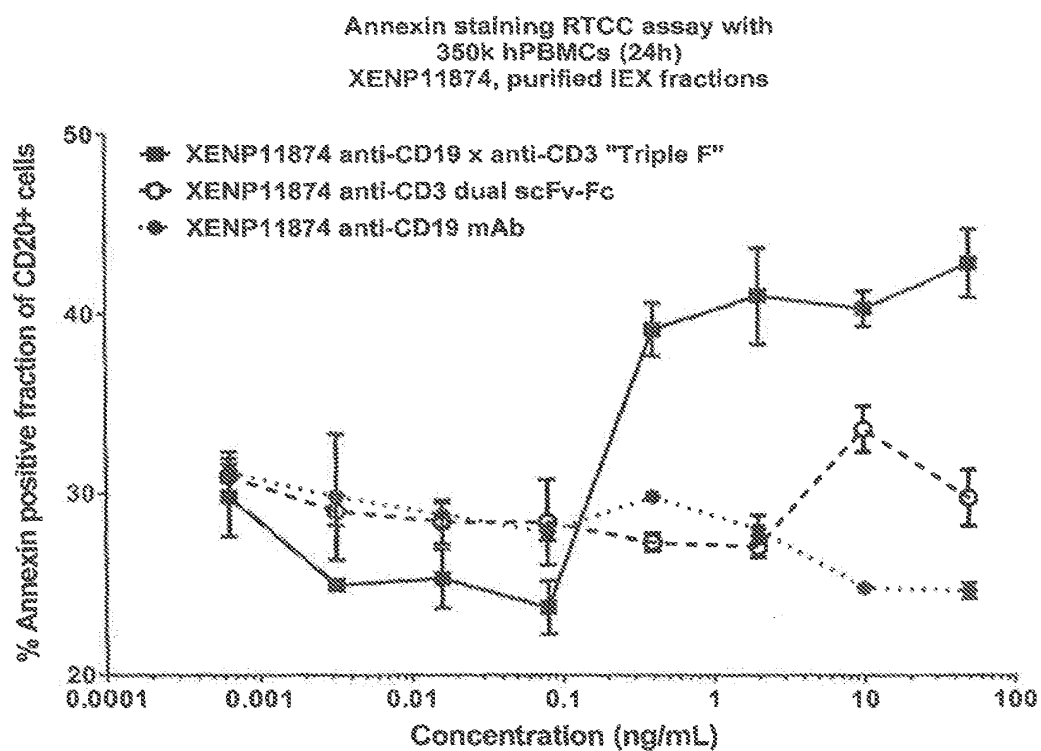


Figure 9

Figure 10



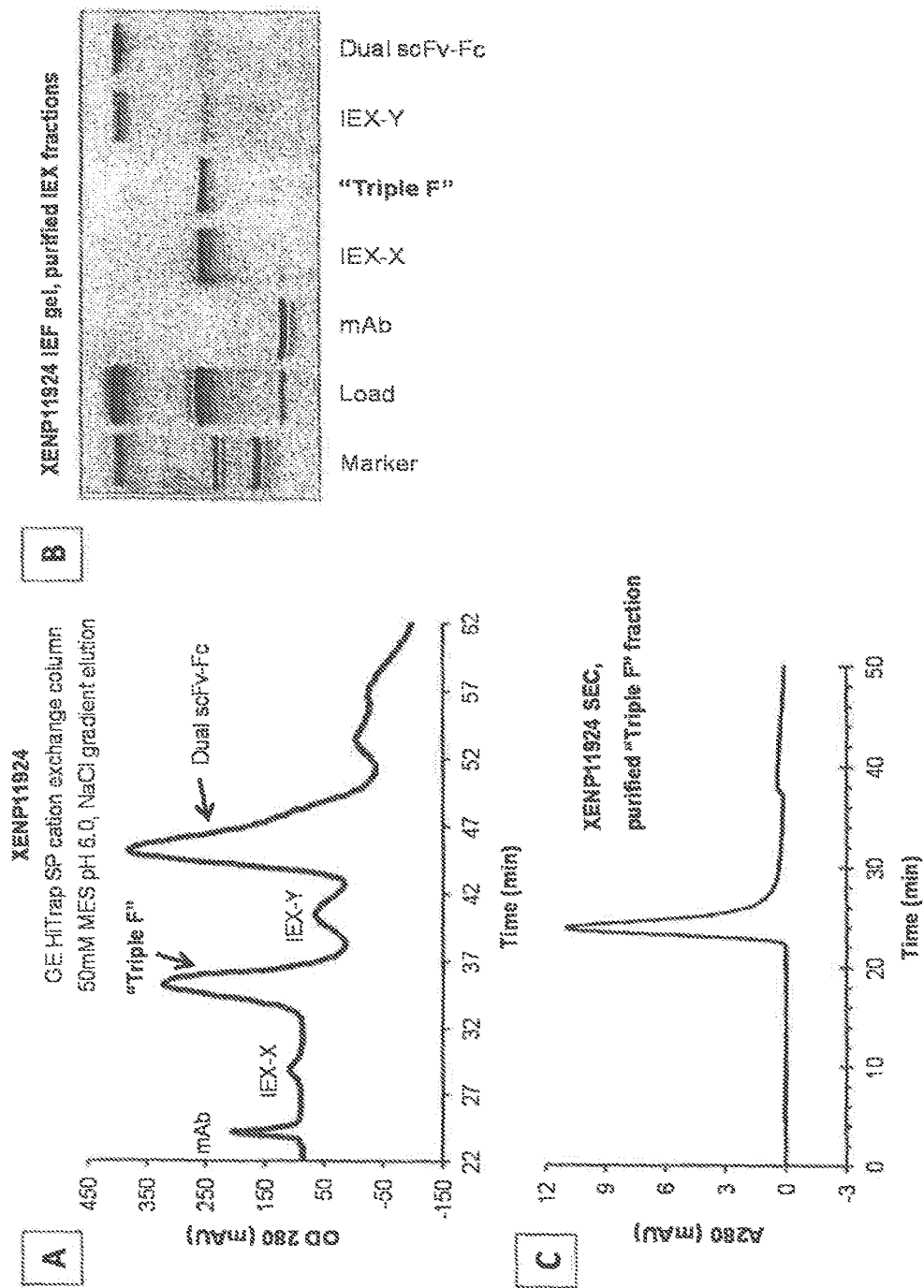


Figure 11

Figure 12

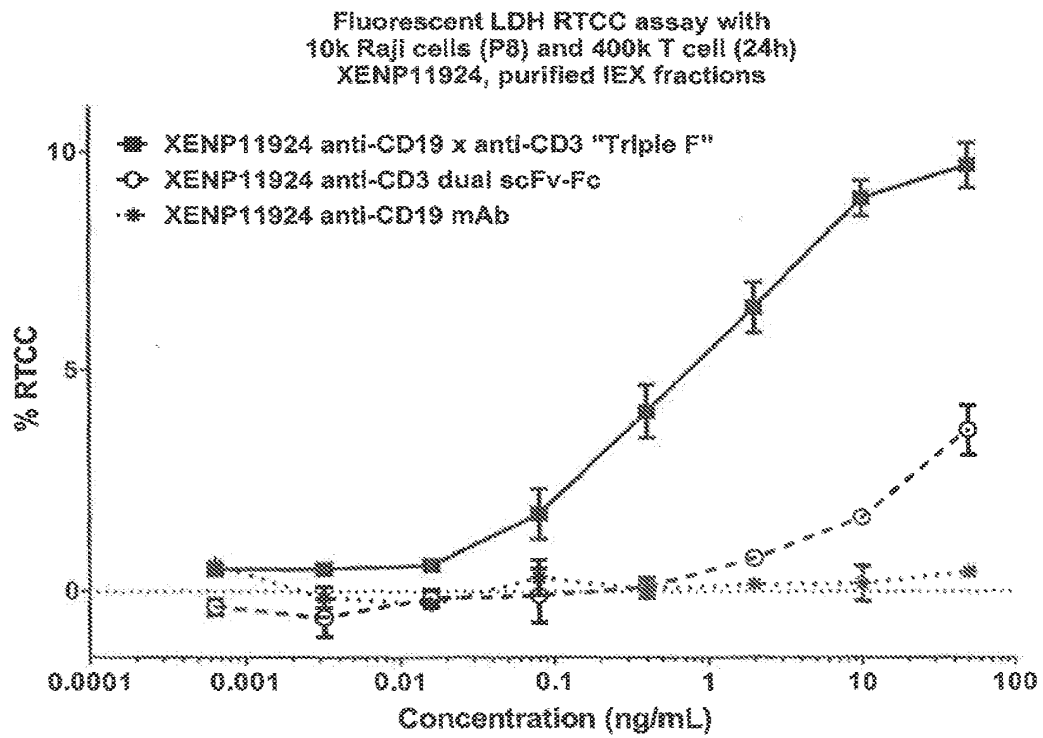


Figure 13

XENP11925_HC-Fab

EVQLVESGGGLVQPGGSLRLSCAASGFD~~F~~SRSWMNWVRQAPGKGLEWVSEINFDSSSTINYATSVKGRFTISRD
SKNTLYLQMNSLPAEDTAVYYCARYGNWFFPYWGQGTFLVTVSSASTKGPSVFPFLAPSSKSTSGGTAALGCLVKDY
FPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSELGTQTYTCNVDPKPSNTKVDKKVEPKSCDKT
HTCPPCPAPELIRGSPSVFLFPPPKFDTLMISPTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHNAKTKPREEQYN
STYRVVSVLTVFLQD~~W~~LNGKEYKCKVSNKARPAPTEKTI~~S~~KAKGQPREPQVYTLPPS~~Q~~EEMTKNQVSLTCLVKG
FYPSDIAVEWES~~S~~SGQFENNYNTTTPMLDSUGSFFLYSKLTVDKSRWQ~~Q~~GNVFS~~C~~SVMH~~E~~ALHNRYTQKSLSLSP
G

XENP11925_HC-scFv

EVQLVESGGGLVQPGGSLRLSCAASGFTFNTYAMN~~W~~VRQAPGKGLEWVGRIKSKYNNYATYYADSVKGRFTISR
DDSKNTLYLQMNSLPAEDTAVYYCVPHGNFGNSYVSWFAYWGQGTFLVTVSSGGGGSGGGGSGGGGSGQAVVTQEP
SLTVSPGGTVELTCCSSTGAVTTSNYANWVQKPGQAPRGLIGGTNKRAPGVPAFFSGSLLGCKAALTLSGAQF
EDEAEYYCALWYSLNWVFGGGTKLTVLERKSSDKTHTCP~~R~~CPAPELIRGSPSVFLFPPPKFDTLMISPTPEVTCV
VVDVSHEDPEVKF~~K~~WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVFLQD~~W~~LNGKEYKCKVSNKARPAPTEKTI
SKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQFENNYKTTPPVLD~~S~~UGSFFLYSKL
TVDKSRWQ~~Q~~GNVFS~~C~~SVMH~~E~~ALHNRYTQKSLSLSPGK

XENP11925_LC

DIIVMTQSPSSLSASVGDRVTITCRASQNVDTNVAWYQQKPGQSPKALIYSASYPYSGVPDRFTGSGSGTDFTLT
ISSLQPEDFATYFCQQYDSYPLTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQW
KVNALQS~~G~~NSQESVTEQDSK~~D~~STYSLSS~~T~~LTLSKADYEKHKVYACEVTRQGLSSPVTKSFN~~P~~GEC

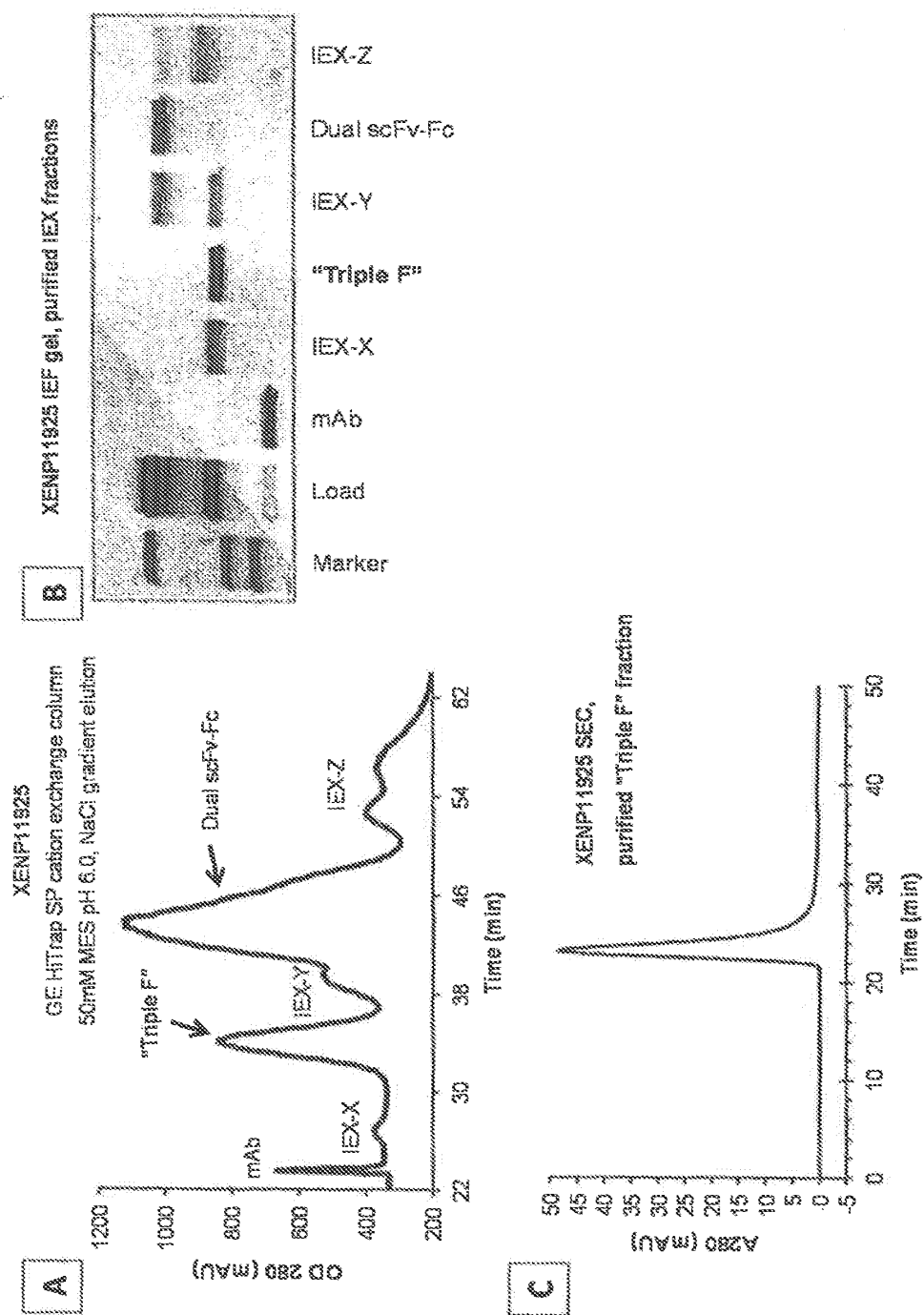


Figure 14

Figure 15

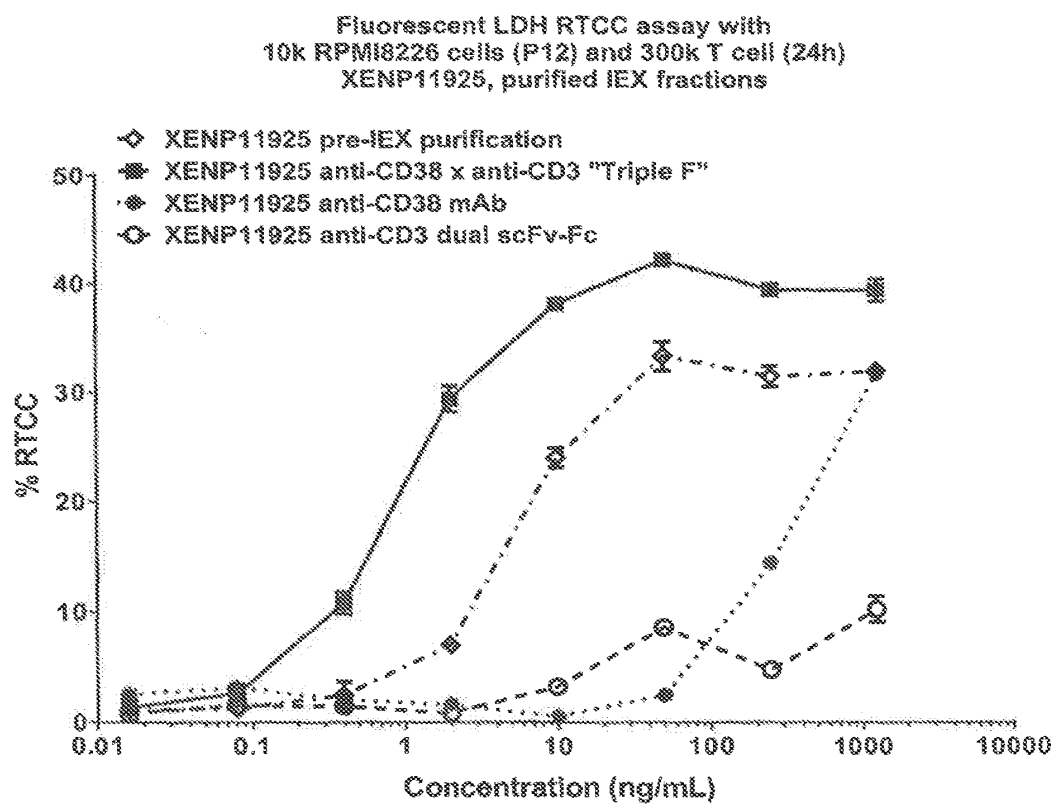


Figure 2

F5 16

XENP11924 Heavy Chain HC-Fab

EVQLVESGGGLVQPGGSLRLSCAASGYTFSTSYVMHWVRQAPGKGLEWIGYINPYNDGTTYNEKFKQGRVTI
SSDKSISTAYMELSSLRSEDTAMYYCARGTYYVGRVFDYWGQGTTLTVTVSSASTKGPSVFPLAPSSKSTSS
GGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVFSSSLGTQTYTCNVDPKPK
SNTKVDKKVEPKSCDKTHTCTPPCPAPPELLRGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVQFN
WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKARPAPIEKTIISKAKGQPREP
QVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESSGQPENNYNTTPPMLDSDGSEFFLYSKLTVDKSR
WQEGNVFSCSVMEALHNHYTQKSLSLSPG

XENP11924 Heavy Chain HC-scFv

EVQLVESGGGLVQPGGSLRLSCAASGYTFNTYAMNWVRQAPGKGLEWVGRIRSKYNNYATYYADSVKGRF
TISRDDSKNTLYLQMNSLRAEDTAVYYCVRHGNFGNSYVSWFAYWGQGTTLTVTVSSGGGGSGGGGSGGGGS
QAVVTQEPSTLVSPGGTVTLTSGSSTGAVTTSNYANWVQOKPGQAPRGLIGGTNKRAPGVPAFSGSLLG
GKAALTLSGAQPEDEAEYYCALWYSNLWVFGGGTKLTVLERKSSDKTHTCPRCPPAPPELLRGPSVFLFPPK
PKDTLMISRTPEVTCVVDVSHEDPEVKFKWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNG
KEYKCKVSNKARPAPIEKTIISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQ
PENNYKTTTPVLDSDGSEFFLYSKLTVDKSPWQQGNVFSCSVMEALHNHYTQKSLSLSPGK

XENP12152 Heavy Chain HC-Fab (T411E)

EVQLVESGGGLVQPGGSLRLSCAASGYTFSTSYVMHWVRQAPGKGLEWIGYINPYNDGTTYNEKFKQGRVTI
SSDKSISTAYMELSSLRSEDTAMYYCARGTYYVGRVFDYWGQGTTLTVTVSSASTKGPSVFPLAPSSKSTSS
GGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVFSSSLGTQTYTCNVDPKPK
SNTKVDKKVEPKSCDKTHTCTPPCPAPPELLRGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVQFN
WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKARPAPIEKTIISKAKGQPREP
QVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESSGQPENNYNTTPPMLDSDGSEFFLYSKLEVDKSR
WQEGNVFSCSVMEALHNHYTQKSLSLSPG

XENP12152 Heavy Chain HC-scFv (D401K)

EVQLVESGGGLVQPGGSLRLSCAASGYTFNTYAMNWVRQAPGKGLEWVGRIRSKYNNYATYYADSVKGRF
TISRDDSKNTLYLQMNSLRAEDTAVYYCVRHGNFGNSYVSWFAYWGQGTTLTVTVSSGGGGSGGGGSGGGGS
QAVVTQEPSTLVSPGGTVTLTSGSSTGAVTTSNYANWVQOKPGQAPRGLIGGTNKRAPGVPAFSGSLLG
GKAALTLSGAQPEDEAEYYCALWYSNLWVFGGGTKLTVLERKSSDKTHTCPRCPPAPPELLRGPSVFLFPPK
PKDTLMISRTPEVTCVVDVSHEDPEVKFKWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNG
KEYKCKVSNKARPAPIEKTIISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQ
PENNYKTTTPVLDSDGSEFFLYSKLTVDKSRWQQGNVFSCSVMEALHNHYTQKSLSLSPGK

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Figure 2 (continued)

XENP12155 Heavy Chain HC-Fab (L368E)

EVQLVESGGGLVKPGGSLKLSCAASGYTFTSYVMHWVRQAPGKGLEWIGYINPYNDGTYNEKFQGRVTI
SSDKSISTAYMELSSLRSEDTAMYYCAPGTYYYGTRVFDYWGQGTLLTVTVSSASTKGPSEVFPPLAPSSKSTS
GGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSVTVTPSSSLGTQTYTCNVDHKP
SNTKVDKRVKVEPKSCDKTHTCPPCPAPELLRGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFN
WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVQLHQLDNLNGKEYKCKVSNKARPAIEKTIKAKGQPREP
QVYTLPPSQEEMTKNQVSLTCEVKGFYPSDIAVEWESGQPENNYNTTPPMLDSGGSFFLYSKLTVDKSR
WQEGNVFSCSVMHREALHNYTQKSLSLSPG

XENP12155 Heavy Chain HC-scFv (S364K)

EVQLVESGGGLVQPGGSLRLSCAASGFTFTNTYAMNWVRQAPGKGLEWVGRIRSKYNNYATYYADSVKGRF
TISRDDSKNTLYLQMNSLRADTAIVYCVRHGNEFGNSYVSWFAYWGQGTLLTVTVSSGGGGSGGGSGGGGS
QAVVTQEPSTLVSPGGTVTLTCSSTGAVTTSNYANWVQKPGQAPRGLIGGTNKRAPGVPAKFSGLLG
GKAALTLSGAQPEDAEYICALWYSNLWVFGGGTKLTVLERKSSDKTHTCPRCPAPELLRGPSVFLFPPK
PKDTLMISRTPEVTCVVVDVSHEDPEVKFKWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVQLHQLDNLNG
KEYKCKVSNKARPAIEKTIKAKGQPPPEPVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQ
PENNYKTTFPVLDSGGSFFLYSKLTVDKSRWQQGNVFSCSVMHREALHNYTQKSLSLSPGK

XENP11924, XENP12152, XENP12155 Light Chain

DIVMTQSPATLSLSPGERATLSCRSSKSLQNVNGNTYLYWFQQKPGQSPQLLIYRMSNINSGVPDRFSGS
GSGTEFTLTITSSLEPEDFAVYYCMQHLEYPIITFGAGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCCL
LNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPV
TKSFNRGEC

Figure 3

F5
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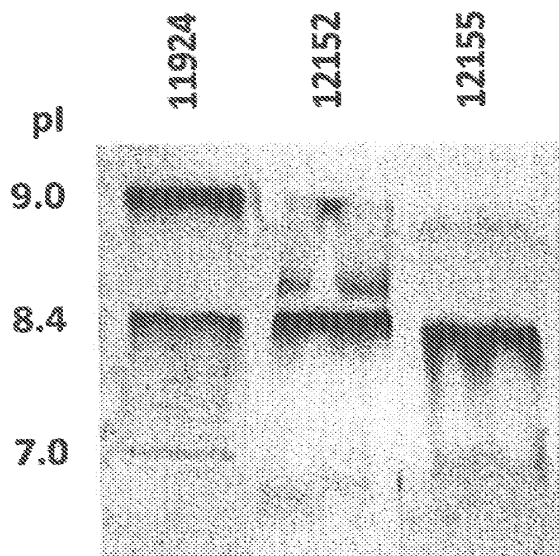
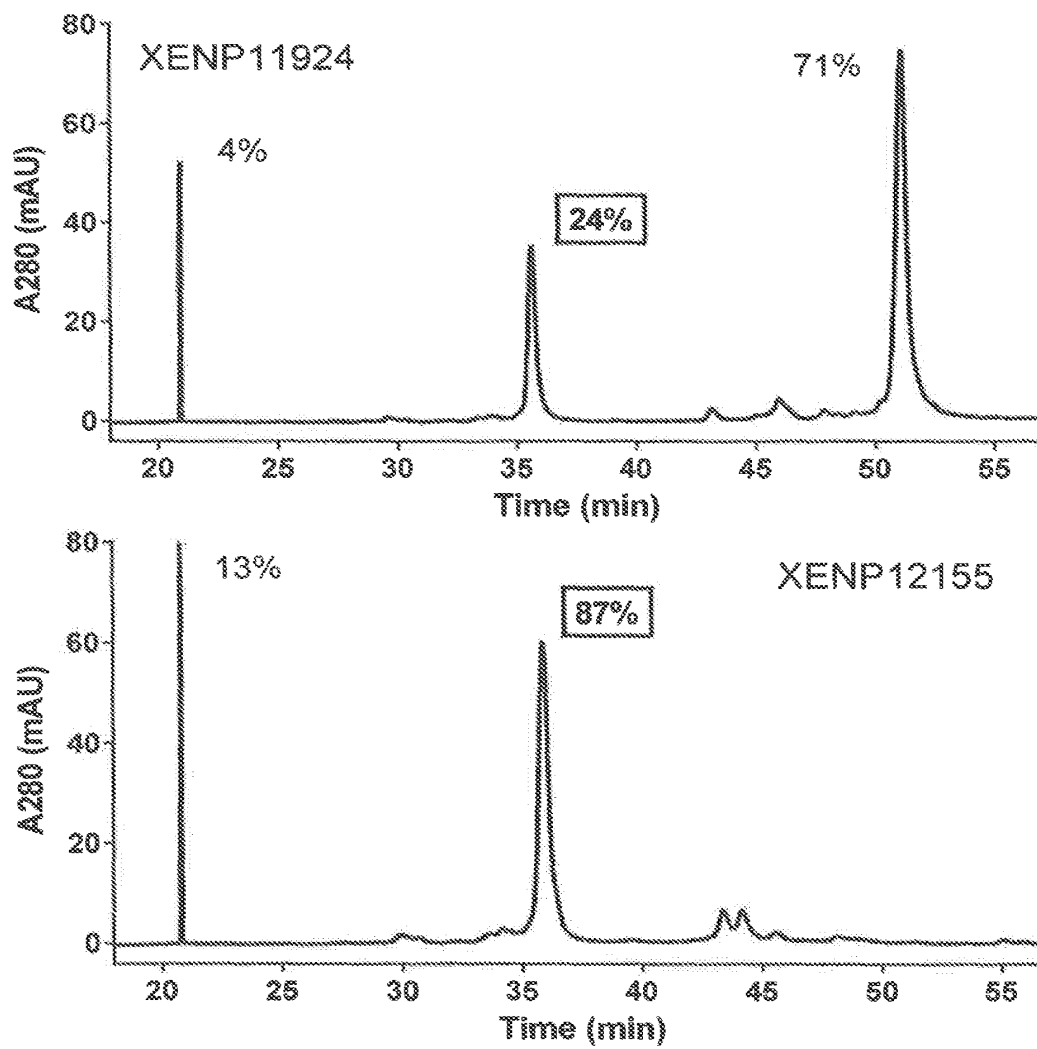


Figure 4

Fig 18



19
24
Figure 26

AMINO ACID	pI
Alanine Ala A	6.00
Arginine Arg R	11.15
Asparagine Asn N	5.41
Aspartic acid Asp D	2.77
Cysteine Cys C	5.02
Glutamic acid Glu E	3.22
Glutamine Gln Q	5.65
Glycine Gly G	5.97
Histidine His H	7.47
Isoleucine Ile I	5.94
Leucine Leu L	5.98
Lysine Lys K	9.89
Methionine Met M	5.74
Phenylalanine Phe F	5.48
Proline Pro P	6.30
Serine Ser S	5.68
Threonine Thr T	5.64
Tryptophan Trp W	5.89
Tyrosine Tyr Y	5.66
Valine Val V	5.96

FIGURE 27A

Xenp#	Name (NC)	SEQ ID NO (NC)	Name (LC)	SEQ ID NO (LC)	Calc. pI	# WA	Delta KR (vs. WT)	# DE	Delta DE (vs. WT)	Change State	# MC Mutations vs IgG1	# LC Mutations vs IgG1	Total # of Mutations
XENP010447	IgG1-WT	2	Ck-WT	112	8.10	122	0	116	0	0	27	0	27
XENP008384	IgG2-WT	3	Ck-WT	113	7.31	113	-4	118	2	0	27	0	27
MA	IgG3-WT	4	Ck-WT	114	8.11	120	-2	114	-2	0	22	0	22
MA	IgG4-WT	5	Ck-WT	114	8.11	120	-2	114	-2	0	22	0	22
XENP010340	IgG1/2-MC	6	Ck-WT	114	8.11	120	-2	114	-2	0	22	0	22
XENP003653	IgG1-43A5	50	Ck-WT	115	8.1	122	0	116	0	0	1	0	1
XENP003653	IgG2-43A5	51	Ck-WT	116	7.31	118	-4	118	2	0	20	0	20
XENP003653	IgG1-C11-pH6	7	Ck-WT	116	6.21	116	-6	126	12	-18	0	0	0
XENP003653	IgG1-WT	2	Ck-pH6	117	6.21	116	-6	128	12	-18	0	0	0
XENP003653	IgG1-C11-pH6	192	Ck-pH6	117	6.49	110	-12	140	24	-30	6	6	12
XENP003653	IgG1-C11-pH6	52	Ck-pH6	117	6.49	110	-12	140	24	-30	7	6	13
XENP003653	IgG1-C11-pH6	53	Ck-pH6	117	6.49	110	-12	140	24	-30	8	6	14
XENP010009	IgG1-WT	2	Ck-pH3	120	8.68	116	-8	122	6	-6	0	3	3
XENP010009	IgG1-WT	2	Ck-pH6-DEDE1	121	5.85	116	-8	126	20	-20	0	10	10
XENP010009	IgG2-WT	3	Ck-pH3	122	6.16	112	-10	124	8	-12	27	3	30
XENP010009	IgG2-WT	3	Ck-pH6	123	5.88	112	-10	130	14	-18	27	6	33
XENP010009	IgG2-WT	3	Ck-pH6-DEDE1	124	5.58	112	-10	138	22	-26	27	10	37
XENP010009	IgG2-WT	13	Ck-WT	125	6.20	110	-12	122	6	-12	19	0	19
XENP010009	IgG2-WT	14	Ck-WT	126	6.20	110	-12	122	6	-12	19	0	19
XENP010009	IgG2-WT	15	Ck-WT	127	6.16	110	-12	122	6	-12	19	0	19
XENP010009	IgG2-WT	15	Ck-pH3	128	5.63	104	-19	126	12	-24	18	3	22
XENP010009	IgG2-WT	15	Ck-pH6	129	5.43	104	-19	134	18	-30	18	6	25
XENP010009	IgG2-WT	15	Ck-pH6-DEDE1	130	5.23	104	-19	142	26	-38	19	10	29
XENP010009	IgG2-WT	16	Ck-WT	131	5.79	110	-12	130	14	-20	22	0	22

Figure 37B.20 (cont.)

XenP#	Name (HC)	SEQ ID NO (HC)	Name (LC)	SEQ ID NO (LC)	Calc. pI	# KR	Delta KR (vs. WT)	# DE	Delta DE (vs. WT)	Change State	# HC Mutations vs IgG1	# LC Mutations vs IgG1	Total # of Mutations
XENP010104	p1-hs01(NF-VE-DEDE)	16	Ck-p1(3)	132	5.37	104	-13	136	20	-32	22	3	25
XENP010105	p1-hs01(NF-VE-DEDE)	16	Ck-p1(6)	133	5.22	104	-15	142	26	-36	22	8	28
XENP010106	p1-hs01(NF-VE-DEDE)	16	Ck-p1(6-DEDE)	134	5.07	104	-18	150	34	-46	22	10	32
XENP010107	lg51-p1(7)	25	Ck-p1(4)	135	5.31	109	-10	136	20	-36	7	4	11
XENP010108	lg51-p1(11)	26	Ck-p1(4)	136	4.96	92	-22	144	26	-50	14	4	18
XENP010109	lg51/2-p1(7)	27	Ck-p1(4)	137	5.36	109	-30	134	18	-46	17	4	21
XENP010110	lg51/2-p1(11)	28	Ck-p1(4)	145	5.01	92	-22	142	28	-46	21	4	25
XENP010117	lg52-p1(6)-Neurosp. to DE	56	CX-N151D-N155E	148	6.39	123	0	138	12	-6	3	3	6
XENP010119	lg51-p1(6)-KR-to Neutral	57	CX-K135Q-K145Q	139	6.58	110	-12	116	0	-6	3	3	6
XENP010119	lg51-p1(6)-KR-to DE	58	CX-K135E-K145E	140	5.92	110	-12	128	12	-18	3	3	6
XENP010125	lg50-p1-hs02	19	Ck-WT	141	6.27	110	-12	120	4	-10	28	0	28
XENP010125	lg50-p1-hs02	22	Ck-WT	142	6.20	110	-12	122	6	-12	19	0	19
XENP010125	lg50-p1-hs02-4545	62	Ck-WT	143	6.27	110	-12	130	4	-10	23	0	23
XENP010125	lg50-p1-hs02-4245	63	Ck-WT	144	6.20	110	-12	122	6	-12	28	0	28
XENP010125	lg50-p1-hs02	19	Ck-p1(4)	145	6.55	102	-20	128	12	-26	28	4	32
XENP010125	lg50-p1-hs02	22	Ck-p1(4)	145	6.54	102	-20	130	14	-28	19	4	23
XENP010125	lg50-p1-hs02-4245	62	Ck-p1(4)	145	6.55	102	-20	128	12	-26	29	4	33
XENP010125	lg50-p1-hs02-4245	63	Ck-p1(4)	145	6.54	103	-20	130	14	-28	28	4	32
XENP010125	lg50-p1-hs02-722K	70	any light chain	170	6.31	112	-10	122	8	-10	18	0	18
XENP010125	lg50-p1-hs02-722K	71	any light chain	171	6.31	112	-10	122	6	-10	18	0	18
XENP010125	lg50-p1-hs02-259Y	72	any light chain	172	6.20	110	-12	122	0	-12	18	0	18
XENP010125	lg50-p1-hs02-303Y	73	any light chain	173	6.20	110	-12	122	0	-12	18	0	18
XENP010125	lg50-p1-hs02-309I	74	any light chain	174	6.20	110	-12	122	0	-12	18	0	18

Figure 22C 20 (cont.)

Xen#	Name (HC)	SEQ ID NO (HC)	Name (LC)	SEQ ID NO (LC)	Cole. PI	# KR	Delta KR (vs. WT)	# DE	Delta DE (vs. WT)	Charge State	# MC Mutations vs. IgG1	# LC Mutations vs. IgG1	Total # of Mutations
XENP010276	IgG-p1-hc3-356A	75	any light chain	175	6.20	110	-12	122	6	-12	18	0	18
XENP010275	IgG-p1-hc3-355Q	76	any light chain	176	6.31	112	-10	122	6	-10	18	0	18
XENP010272	IgG-p1-hc3-354M	77	any light chain	177	6.20	110	-12	122	6	-12	18	0	18
XENP010273	IgG-p1-hc3-352K	78	any light chain	178	6.31	112	-10	122	6	-10	18	0	18
XENP010274	IgG-p1-hc3-357V	79	any light chain	179	6.20	110	-12	122	6	-12	18	0	18
XENP010275	IgG-p1-hc3-419Q	80	any light chain	180	6.31	110	-12	120	4	-10	16	0	16
XENP010276	IgG-p1-hc3- 236K/330Y	81	any light chain	181	6.20	110	-12	122	6	-12	17	0	17
XENP010277	IgG-p1-hc3- 394N/391K/397V	82	any light chain	182	6.31	112	-10	122	6	-10	16	0	16
XENP010278	IgG-p1-hc3-137G	83	any light chain	183	6.31	110	-12	120	4	-10	18	0	18
XENP010279	IgG-p1-hc3-136G	84	any light chain	184	6.20	110	-12	122	6	-12	18	0	18
XENP010280	IgG-p1-hc3-192S	85	any light chain	185	6.20	110	-12	122	6	-12	18	0	18
XENP010281	IgG-p1-hc3-193I	86	any light chain	186	6.20	110	-12	122	6	-12	18	0	18
XENP010282	IgG-p1-hc3-199I	87	any light chain	187	6.20	110	-12	122	6	-12	18	0	18
XENP010283	IgG-p1-hc3-203N	88	any light chain	188	6.31	110	-12	120	4	-10	18	0	18
XENP010284	IgG-p1-hc3-214K	89	any light chain	189	6.31	112	-10	122	6	-10	18	0	18
XENP010285	IgG-p1-hc3- 137Q/138G	90	any light chain	190	6.31	110	-12	120	4	-10	17	0	17
XENP010286	IgG-p1-hc3-SL	91	Ch-WT	194	6.20	110	-12	122	6	-12	17	0	17
XENP010287	IgG-p1-hc3- 199Y/203M	91	any light chain	195			-12		4	-10	17	0	17
XENP010288	IgG-p1-hc3- 214K/222K	92	any light chain	196	6.31	110	-8	120	6	-8	17	0	17
XENP010289	IgG-p1-hc3- 136G/193S/193K	93	any light chain	197	6.44	114	-12	122	6	-12	16	0	16
XENP010290	IgG-p1-hc3- 137Q/138G/192K	94	any light chain	198	6.20	110	-12	122	4	-10	16	0	16

FIGURE 3-22- 20 (cont.)

Gene#	Name (HC)	SEQ ID NO (HC)	Name (LC)	SEQ ID NO (LC)	Calc. pI	# KR	Delta KR (vs. WT)	# DE	Delta DE (vs. WT)	Change State	# HC Mutations vs IgG1	# LC Mutations vs IgG1	Total # of Mutations
XENP010469	lgG-p1-hc03	22	Cx-hc03	246	5.92	106	-16	124	8	-16	19	3	22
	lgG-p1-hc03	22	Cx-hc04	149	5.83	104	-18	124	8	-20	19	4	23
	lgG-p1-hc03	22	Cx-hc05	150	5.75	104	-18	126	10	-22	19	5	24
	lgG-p1-hc03	22	Cx-hc06	151	5.68	104	-18	128	12	-24	19	6	25
XENP010475	Hc01-igG2-CHL1						-2		4	0	16	0	16
	igG1-CH2-CH4	55	any light chain	199	7.30	120	-12	120	6	-12	9	10	9
	igG-p1-hc03		Cx-WT	152			-12						
	charges-only	24			6.30	110	-12	122	4	-10	16	0	16
XENP010487	igG-p1-hc02		Cx-WT	152			-12						
	charges-only	21			6.27	110	0	120	4	2	9	0	9
	Hc01-igG2-CHL1												
	igG1-Hinge-CH2-C												
XENP010499	H1	161	any light chain	200	7.67	122	-12	120	6	-12	16	0	16
	igG-p1-hc03-SL	98	Cx-WT	152			-12						
	C345				6.20	110	-12	122	6	-12	19	0	19
	igG-p1-hc08-SL	97	Cx-WT	152			-12						
XENP010501	K28/A245				6.20	110	-12	122	6	-12	17	0	17
	Hc01-p1-hc03-SL3												
	HC/N19X5/F193L												
	N4345	152	any light chain	201	6.20	110	-12	122	4	-10	16	0	16
XENP010468	Hc01-p1-hc03-EL3												
	TS/S135G/N152N/												
	F129L_M245	153	any light chain	202	6.31	110	-12	120	4	-10	26	0	26
	igG-p1-hc02-SL	23	Cx-WT	152	6.27	110	-12	120	4	-10	27	0	27
XENP010469	igG-p1-hc02-SL	98	Cx-WT	152			-12						
	K245				6.27	110		120					

Figure 3-20 (cont.)

Xenp#	Name (HO)	SEQ ID NO (HO)	Name (LC)	SEQ ID NO (LC)	Calc. pl	# KR	Delta KR (vs. WT)	# DE (vs. WT)	Charge State	# HC Mutations vs IgG1	# LC Mutations vs IgG1	Total # of Mutations
XENP010473	igc-pl-iso2	131	CK-WT	152	6.20	110	-12	6	-12	19	0	19
	charges-only-4345	131	CK-WT	153	6.20	110	-12	4	-10	19	0	19
	igc-pl-iso2	132	CK-WT	152	6.27	110	-2	4	0	17	0	17
	charges-only-4345	132	CK-WT	153	6.27	110	-2	4	0	17	0	17
XENP010474	HOLD_igc2_CH1_L	164	any light chain	203	7.30	120	0	4	2	19	0	19
	igc1_CH2_CH3_M4	164	any light chain	203	7.30	120	0	4	2	19	0	19
XENP010475	HOLD_igc2_CH1_L	165	any light chain	204	7.87	122	-22	20	-36	8	4	12
	igc1_HHPR_CH2_C	165	CK-pl(4)	135	5.34	100	-22	18	-36	18	4	22
XENP010476	igc1_pl(7)-4345	25	CK-pl(4)	135	5.36	100	-22	18	-36	18	4	22
XENP010477	igc1/2_pl(7)-4345	166	CK-pl(4)	135	5.36	100	-12	4	-10	23	0	23
XENP010478	HOLD_pl-iso1/2_c	167	any light chain	205	6.37	110	-16	9	-18	17	3	20
	charges-only	167	CK-iso(3)	148	5.92	106	-16	8	-20	17	4	21
	igc-pl-iso3-SL	23	CK-iso(4)	149	5.83	104	-18	10	-22	17	5	22
	igc-pl-iso3-SL	23	CK-iso(4)	150	5.75	104	-16	8	-18	18	3	21
	igc-pl-iso3-SL	98	CK-iso(3)	148	5.92	106	-18	8	-20	18	4	22
	igc-pl-iso3-SL	98	CK-iso(4)	149	5.83	104	-18	10	-22	18	5	23
	igc-pl-iso3-SL	99	CK-iso(4)	150	5.75	104	-18	10	-22	18	5	23
	igc-pl-iso3-SL	99	CK-iso(4)	150	5.75	104	-18	10	-22	18	5	23
	igc-pl-iso3-SL	99	CK-iso(4)	150	5.75	104	-18	10	-22	18	5	23
	igc-pl-iso3-SL	99	CK-iso(4)	150	5.75	104	-18	10	-22	18	5	23

Figure 3FF 20 (cont.)

Xenp#	Name (HC)	SEQ ID NO (HC)	Name (LC)	SEQ ID NO (LC)	Calc. pI	# KR	Delta KR (vs. WT)	# DE	Delta DE (vs. WT)	Charge State	# HC Mutations vs IgG1	# LC Mutations vs IgG1	Total # of Mutations
6191103	185-pH-1803-SL	97	CK-180(3)	148	5.92	106	-18	124	8	-18	18	3	22
6221103	185-pH-1803-SL	97	CK-180(3)	148	5.92	106	-18	124	8	-20	19	4	23
6231103	185-pH-1803-SL	97	CK-180(4)	149	5.83	104	-18	124	10	-22	19	5	24
6281103	185-pH-1803-SL	97	CK-180(5)	150	5.75	104	-20	126	14	-28	18	8	22
6341103	185-pH-1803-SL	98	CK-180(4)	149	5.54	102	-18	130	10	-32	20	5	25
6351103	185-pH-1803-SL	99	CK-180(5)	150	5.75	104	-18	126	8	-20	27	8	32
6361103	185-pH-1803-SL	65	CK-180(5)	150	5.78	104	-18	124					

Figure S8

Variant	# of sub(s)	pl / pl	pl / WT	WT / WT	avg delta pl
G137E/N203D/K274Q/R355Q/K392N/Q419E/K447_	7	6.43	7.14	8.02	-0.79
G137E/N203D/K274Q/R355Q/K392N/Q419E	6	6.58	7.30	8.02	-0.72
G137E/N203D/K274Q/R355Q/K392N/K447_	6	6.58	7.30	8.02	-0.72
G137E/N203D/K274Q/R355Q/Q419E/K447_	6	6.58	7.30	8.02	-0.72
G137E/N203D/K274Q/K392N/Q419E/K447_	6	6.58	7.30	8.02	-0.72
G137E/N203D/R355Q/K392N/Q419E/K447_	6	6.58	7.30	8.02	-0.72
G137E/K274Q/R355Q/K392N/Q419E/K447_	6	6.58	7.30	8.02	-0.72
N203D/K274Q/R355Q/K392N/Q419E/K447_	6	6.58	7.30	8.02	-0.72
G137E/N203D/K274Q/R355Q/K392N	5	6.76	7.46	8.02	-0.63
G137E/N203D/K274Q/R355Q/Q419E	5	6.76	7.46	8.02	-0.63
G137E/N203D/K274Q/R355Q/K447_	5	6.76	7.46	8.02	-0.63
G137E/N203D/K274Q/K392N/Q419E	5	6.76	7.46	8.02	-0.63
G137E/N203D/K274Q/K392N/K447_	5	6.76	7.46	8.02	-0.63
G137E/N203D/K274Q/Q419E/K447_	5	6.76	7.46	8.02	-0.63
G137E/N203D/R355Q/K392N/Q419E	5	6.76	7.46	8.02	-0.63
G137E/N203D/R355Q/K392N/K447_	5	6.76	7.46	8.02	-0.63
G137E/N203D/R355Q/Q419E/K447_	5	6.76	7.46	8.02	-0.63
G137E/N203D/K392N/Q419E/K447_	5	6.76	7.46	8.02	-0.63
G137E/K274Q/R355Q/K392N/Q419E	5	6.76	7.46	8.02	-0.63
G137E/K274Q/R355Q/K392N/K447_	5	6.76	7.46	8.02	-0.63
G137E/K274Q/R355Q/Q419E/K447_	5	6.76	7.46	8.02	-0.63
G137E/K274Q/K392N/Q419E/K447_	5	6.76	7.46	8.02	-0.63
G137E/R355Q/K392N/Q419E/K447_	5	6.76	7.46	8.02	-0.63
N203D/K274Q/R355Q/K392N/Q419E	5	6.76	7.46	8.02	-0.63
N203D/K274Q/R355Q/K392N/K447_	5	6.76	7.46	8.02	-0.63
N203D/K274Q/R355Q/Q419E/K447_	5	6.76	7.46	8.02	-0.63
N203D/K274Q/K392N/Q419E/K447_	5	6.76	7.46	8.02	-0.63
N203D/R355Q/K392N/Q419E/K447_	5	6.76	7.46	8.02	-0.63
K274Q/R355Q/K392N/Q419E/K447_	5	6.76	7.46	8.02	-0.63
G137E/N203D/K274Q/R355Q	4	7.00	7.61	8.02	-0.51
G137E/N203D/K274Q/K392N	4	7.00	7.61	8.02	-0.51
G137E/N203D/K274Q/Q419E	4	7.00	7.61	8.02	-0.51
G137E/N203D/K274Q/K447_	4	7.00	7.61	8.02	-0.51
G137E/N203D/R355Q/K392N	4	7.00	7.61	8.02	-0.51
G137E/N203D/R355Q/Q419E	4	7.00	7.61	8.02	-0.51
G137E/N203D/R355Q/K447_	4	7.00	7.61	8.02	-0.51
G137E/N203D/K392N/Q419E	4	7.00	7.61	8.02	-0.51
G137E/N203D/K392N/K447_	4	7.00	7.61	8.02	-0.51
G137E/N203D/Q419E/K447_	4	7.00	7.61	8.02	-0.51
G137E/K274Q/R355Q/K392N	4	7.00	7.61	8.02	-0.51
G137E/K274Q/R355Q/Q419E	4	7.00	7.61	8.02	-0.51
G137E/K274Q/R355Q/K447_	4	7.00	7.61	8.02	-0.51
G137E/K274Q/K392N/Q419E	4	7.00	7.61	8.02	-0.51
G137E/K274Q/K392N/K447_	4	7.00	7.61	8.02	-0.51
G137E/K274Q/Q419E/K447_	4	7.00	7.61	8.02	-0.51
G137E/R355Q/K392N/Q419E	4	7.00	7.61	8.02	-0.51
G137E/R355Q/K392N/K447_	4	7.00	7.61	8.02	-0.51
G137E/R355Q/Q419E/K447_	4	7.00	7.61	8.02	-0.51
G137E/K392N/Q419E/K447_	4	7.00	7.61	8.02	-0.51
N203D/K274Q/R355Q/K392N	4	7.00	7.61	8.02	-0.51
N203D/K274Q/R355Q/Q419E	4	7.00	7.61	8.02	-0.51
N203D/K274Q/R355Q/K447_	4	7.00	7.61	8.02	-0.51
N203D/K274Q/K392N/Q419E	4	7.00	7.61	8.02	-0.51

Figure 21 (cont.)

N203D/K274Q/K392N/K447_	4	7.00	7.61	8.02	-0.51
N203D/K274Q/Q419E/K447_	4	7.00	7.61	8.02	-0.51
N203D/R355Q/K392N/Q419E	4	7.00	7.61	8.02	-0.51
N203D/R355Q/K392N/K447_	4	7.00	7.61	8.02	-0.51
N203D/R355Q/Q419E/K447_	4	7.00	7.61	8.02	-0.51
N203D/K392N/Q419E/K447_	4	7.00	7.61	8.02	-0.51
K274Q/R355Q/K392N/Q419E	4	7.00	7.61	8.02	-0.51
K274Q/R355Q/K392N/K447_	4	7.00	7.61	8.02	-0.51
K274Q/R355Q/Q419E/K447_	4	7.00	7.61	8.02	-0.51
K274Q/K392N/Q419E/K447_	4	7.00	7.61	8.02	-0.51
R355Q/K392N/Q419E/K447_	4	7.00	7.61	8.02	-0.51
G137E/N203D/K274Q	3	7.30	7.74	8.02	-0.36
G137E/N203D/R355Q	3	7.30	7.74	8.02	-0.36
G137E/N203D/K392N	3	7.30	7.74	8.02	-0.36
G137E/N203D/Q419E	3	7.30	7.74	8.02	-0.36
G137E/N203D/K447_	3	7.30	7.74	8.02	-0.36
G137E/K274Q/R355Q	3	7.30	7.74	8.02	-0.36
G137E/K274Q/K392N	3	7.30	7.74	8.02	-0.36
G137E/K274Q/Q419E	3	7.30	7.74	8.02	-0.36
G137E/K274Q/K447_	3	7.30	7.74	8.02	-0.36
G137E/R355Q/K392N	3	7.30	7.74	8.02	-0.36
G137E/R355Q/Q419E	3	7.30	7.74	8.02	-0.36
G137E/R355Q/K447_	3	7.30	7.74	8.02	-0.36
G137E/K392N/Q419E	3	7.30	7.74	8.02	-0.36
G137E/K392N/K447_	3	7.30	7.74	8.02	-0.36
G137E/Q419E/K447_	3	7.30	7.74	8.02	-0.36
N203D/K274Q/R355Q	3	7.30	7.74	8.02	-0.36
N203D/K274Q/K392N	3	7.30	7.74	8.02	-0.36
N203D/K274Q/Q419E	3	7.30	7.74	8.02	-0.36
N203D/K274Q/K447_	3	7.30	7.74	8.02	-0.36
N203D/R355Q/K392N	3	7.30	7.74	8.02	-0.36
N203D/R355Q/Q419E	3	7.30	7.74	8.02	-0.36
N203D/R355Q/K447_	3	7.30	7.74	8.02	-0.36
N203D/K392N/Q419E	3	7.30	7.74	8.02	-0.36
N203D/K392N/K447_	3	7.30	7.74	8.02	-0.36
N203D/Q419E/K447_	3	7.30	7.74	8.02	-0.36
K274Q/R355Q/K392N	3	7.30	7.74	8.02	-0.36
K274Q/R355Q/Q419E	3	7.30	7.74	8.02	-0.36
K274Q/R355Q/K447_	3	7.30	7.74	8.02	-0.36
K274Q/K392N/Q419E	3	7.30	7.74	8.02	-0.36
K274Q/K392N/K447_	3	7.30	7.74	8.02	-0.36
K274Q/Q419E/K447_	3	7.30	7.74	8.02	-0.36
R355Q/K392N/Q419E	3	7.30	7.74	8.02	-0.36
R355Q/K392N/K447_	3	7.30	7.74	8.02	-0.36
R355Q/Q419E/K447_	3	7.30	7.74	8.02	-0.36
K392N/Q419E/K447_	3	7.30	7.74	8.02	-0.36
G137E/N203D	2	7.61	7.85	8.02	-0.21
G137E/K274Q	2	7.61	7.85	8.02	-0.21
G137E/R355Q	2	7.61	7.85	8.02	-0.21
G137E/K392N	2	7.61	7.85	8.02	-0.21
G137E/Q419E	2	7.61	7.85	8.02	-0.21
G137E/K447_	2	7.61	7.85	8.02	-0.21
N203D/K274Q	2	7.61	7.85	8.02	-0.21
N203D/R355Q	2	7.61	7.85	8.02	-0.21
N203D/K392N	2	7.61	7.85	8.02	-0.21
N203D/Q419E	2	7.61	7.85	8.02	-0.21
N203D/K447_	2	7.61	7.85	8.02	-0.21

Figure 21 (cont.)

K274Q/R355Q	2	7.61	7.85	8.02	-0.21
K274Q/K392N	2	7.61	7.85	8.02	-0.21
K274Q/Q419E	2	7.61	7.85	8.02	-0.21
K274Q/K447_	2	7.61	7.85	8.02	-0.21
R355Q/K392N	2	7.61	7.85	8.02	-0.21
R355Q/Q419E	2	7.61	7.85	8.02	-0.21
R355Q/K447_	2	7.61	7.85	8.02	-0.21
K392N/Q419E	2	7.61	7.85	8.02	-0.21
K392N/K447_	2	7.61	7.85	8.02	-0.21
Q419E/K447_	2	7.61	7.85	8.02	-0.21
G137E	1	7.85	7.94	8.02	-0.09
N203D	1	7.85	7.94	8.02	-0.09
K274Q	1	7.85	7.94	8.02	-0.09
R355Q	1	7.85	7.94	8.02	-0.09
K392N	1	7.85	7.94	8.02	-0.09
Q419E	1	7.85	7.94	8.02	-0.09
K447	1	7.85	7.94	8.02	-0.09

Figure 59

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Variant	# of subs	pl / pl	pl / WT	WT / WT	avg delta pl
Q196K/P217R/P228R/N276K/H435R	5	8.53	8.32	8.02	0.25
Q196K/P217R/P228R/N276K	4	8.45	8.27	8.02	0.22
Q196K/P217R/P228R/H435R	4	8.46	8.28	8.02	0.22
Q196K/P217R/N276K/H435R	4	8.45	8.27	8.02	0.22
Q196K/P228R/N276K/H435R	4	8.45	8.27	8.02	0.22
P217R/P228R/N276K/H435R	4	8.46	8.28	8.02	0.22
Q196K/P217R/P228R	3	8.37	8.22	8.02	0.17
Q196K/P217R/N276K	3	8.37	8.22	8.02	0.17
Q196K/P217R/H435R	3	8.37	8.22	8.02	0.17
Q196K/P228R/N276K	3	8.37	8.22	8.02	0.17
Q196K/P228R/H435R	3	8.37	8.22	8.02	0.17
Q196K/N276K/H435R	3	8.37	8.22	8.02	0.17
P217R/P228R/N276K	3	8.37	8.22	8.02	0.17
P217R/P228R/H435R	3	8.37	8.22	8.02	0.16
P217R/N276K/H435R	3	8.37	8.22	8.02	0.17
P228R/N276K/H435R	3	8.37	8.22	8.02	0.17
Q196K/P217R	2	8.27	8.16	8.02	0.13
Q196K/P228R	2	8.27	8.16	8.02	0.13
Q196K/N276K	2	8.27	8.16	8.02	0.13
Q196K/H435R	2	8.27	8.16	8.02	0.13
P217R/P228R	2	8.28	8.16	8.02	0.13
P217R/N276K	2	8.27	8.16	8.02	0.13
P217R/H435R	2	8.28	8.16	8.02	0.13
P228R/N276K	2	8.27	8.16	8.02	0.13
P228R/H435R	2	8.28	8.16	8.02	0.13
N276K/H435R	2	8.27	8.16	8.02	0.13
Q196K	1	8.16	8.10	8.02	0.07
P217R	1	8.16	8.10	8.02	0.07
P228R	1	8.16	8.10	8.02	0.07
N276K	1	8.16	8.10	8.02	0.07
H435R	1	8.16	8.10	8.02	0.07

Figure 1
HETERODIMERIZATION VARIANTS

	IgG1	IgG2	IgG3	IgG4	Variants
196	Q	Q	Q	K	K
199	I	T	T	T	T
203	N	D	N	D	D
217	P	R	L	S	R
220	C	C	P	G	E,R (IgG2)
221	D	—	LGD	—	E,R (IgG1)
222	K	V	T	—	E,R (IgG2)
223	T	—	T	—	D,E,R,K
225	T	—	T	P	D,E,R,K
228	P	P	RCPEPK SCDTPP PCPRCP EPKSCD TPPPCP RCPEPK SCDTPP PCPR	S	D,E,R,K
247	P	P	P	P	Q
276	N	N	K	N	K
340	K	K	K	K	E,Q
345	E	E	E	E	K
	Q	Q	Q	Q	E,K,R
349	Y	Y	Y	Y	A,C,D,E,I,K,S,T,W
350	T	T	T	T	I
351	L	L	L	L	E,K,V,Y
354	S	S	S	S	C
355	R	R	R	Q	E,Q
356	D	E	E	E	K,L,R
357	E	E	E	E	K,R,Q,T
	K	K	K	K	D,E
362	Q	Q	Q	Q	E,K
	S	S	S	S	C,D,E,F,G,H,K,R,T,Y
366	T	T	T	T	A,D,I,K,L,M,S,V,W,Y
	L	L	L	L	A,D,E,K,S all but C,P
370	K	K	K	K	C,D,E,G,R,S,T,V all but C,P
371	G	G	G	G	D
384	N	N	S	N	S
390	N	N	N	N	D,E,K,R
392	K	K	N	K	C,D,E,F,L,M,N
394	T	T	T	T	F,S,V,W,Y

Figure 23 (cont.)

	IgG1	IgG2	IgG3	IgG4	Variants
395	P	P	P	P	T,V
396	P	P	P	P	T,V
397	V	M	M	V	M,S,T
399	D	D	D	D	all but C,P C,K,R
400	S	S	S	S	A,D,E,K,R
401	D	D	D	D	K,N,R
405	F	F	F	F	L, all but C,P A,F,L,M,S,T,V
407	Y	Y	Y	Y	T,V all but C,P A,L,M,V
409	K	K	K	R	R all but C,P D,E,F,K,L,M,V,W
	T	T	T	T	D,E,K,L,N,R,S
419	Q	Q	Q	E	E
439	K	K	K	K	D,E

FIGURE 24

COMBINATIONS OF VARIANTS INTO HETERODIMERIZATION FORMATS

Heterodimerization format	FcRn variants Monomer 1 and/or Monomer 2	Fc variants Monomer 1 and/or Monomer 2	pl variants	Steric variants (including charge pairs)	Combinations (See Legend E)
"bottle opener" scFv-Fab-Fc (Figure 1B)	See Legend A	See Legend B	See Legend C	See Legend D	1), 2), 3), 4), 5), 6), 7), 8), 9) and 10)
Trimeric bottle opener (Figure 5A)	See Legend A	See Legend B	See Legend C	See Legend D	1), 2), 3), 4), 5), 6), 7), 8), 9) and 10)
Trimeric bottle opener (scFv- Fab-Fv-scFv; Figure 5A)	See Legend A	See Legend B	See Legend C	See Legend D	1), 2), 3), 4), 5), 6), 7), 8), 9) and 10)
Trimeric bottle opener (scFv- Fab-Fv-scFv; Figure 5B)	See Legend A	See Legend B	See Legend C	See Legend D	1), 2), 3), 4), 5), 6), 7), 8), 9) and 10)
Trimeric bottle opener (scFv- Fab-Fv-scFv; Figure 5C)	See Legend A	See Legend B	See Legend C	See Legend D	1), 2), 3), 4), 5), 6), 7), 8), 9) and 10)
Trimeric bottle opener (scFv- Fab-Fv-Fab; Figure 5D)	See Legend A	See Legend B	See Legend C	See Legend D	1), 2), 3), 4), 5), 6), 7), 8), 9) and 10)
Trimeric bottle opener (scFv- Fab-Fv-Fab; Figure 5E)	See Legend A	See Legend B	See Legend C	See Legend D	1), 2), 3), 4), 5), 6), 7), 8), 9) and 10)

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Provisional Filing Fees				
Description	Fee Code	Quantity	Amount	Sub-Total in USD(\$)
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Provisional Application Filing Fee	2005	1	125	125
Pages:				
Claims:				
Miscellaneous-Filing:				
Petition:				
Patent-Appeals-and-Interference:				
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Extension-of-Time:				

Description	Fee Code	Quantity	Amount	Sub-Total in USD(\$)
Miscellaneous:				
Total in USD (\$)				125

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Deposit Account	500310
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Document Number	Document Description	File Name	File Size(Bytes)/ Message Digest	Multi Part /.zip	Pages (if appl.)
1		5161_PR02_Cover_ADS.pdf	610869 1f7be0390907b04cf6ddb280edb0315d091c8290	yes	8
	Multipart Description/PDF files in .zip description				
	Document Description		Start	End	
	Provisional Cover Sheet (SB16)		1	2	
	Application Data Sheet		3	8	
Warnings:					
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2		5161_PR02_Specification_Claims.pdf	298881 123fa599a57863ba0ea090883927b29cdd8bc8cf	yes	71
	Multipart Description/PDF files in .zip description				
	Document Description		Start	End	
	Specification		1	68	
	Claims		69	71	
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3	Drawings-only black and white line drawings	5161_PR02_figs.pdf	3111471 10fbc9a76fc6c2dc5d5365678c40ad95cc4da6d7	no	36
Warnings:					
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4	Fee Worksheet (SB06)	fee-info.pdf	29511 bb723e2399f6265b7b4c6f3c7d18f834d5a238db	no	2
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