

QV
431
A1
P76

<http://www.proteinscience.org>

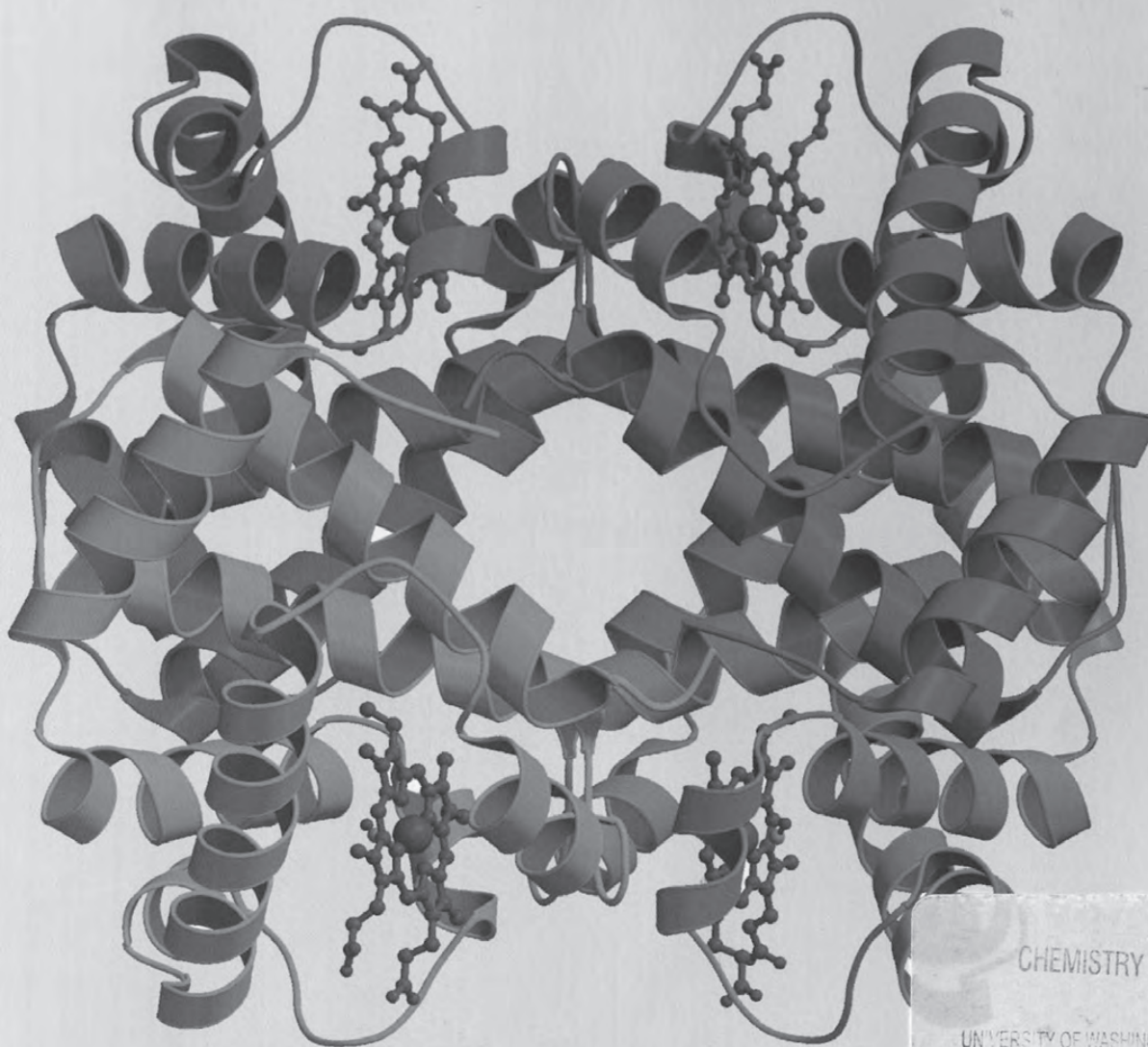


Protein Science

A PUBLICATION OF THE PROTEIN SOCIETY

Sustained in part with the support of
The American Society of Biochemistry and Molecular Biology

Volume 10, No. 9 September 2001



COLD SPRING HARBOR LABORATORY PRESS

CHEMISTRY

UNIVERSITY OF WASHINGTON

AUG 30 2001

LIBRARIES

Protein Science

ISSN 0961-8368

A PUBLICATION OF THE PROTEIN SOCIETY

Sustained in Part with the Support of The American Society for Biochemistry and Molecular Biology

EDITOR-IN-CHIEF

Mark Hermodson
Purdue University

FOUNDING EDITOR

Hans Neurath
University of Washington

ASSOCIATE EDITORS

Ralph A. Bradshaw
University of California, Irvine

Stephen H. Bryant
National Institutes of Health

Brian W. Matthews
University of Oregon

Carol V. Robinson
University of Oxford

ASSISTANT EDITOR

Michael Zierler

ELECTRONIC PUBLISHING COORDINATOR

Stephen H. White
University of California, Irvine

BOOK REVIEW COORDINATOR

Richard N. Perham
University of Cambridge

PRODUCTION MANAGER

Denise Weiss

PRODUCTION EDITOR

Peri C. Zeenkoy

EDITORIAL ADVISORY BOARD

Ruth Hogue Angeletti
Albert Einstein College of Medicine

Cheryl H. Arrowsmith
University of Toronto

David Baker
University of Washington

Wolfgang Baumeister
Max-Planck Institut für Biochemie

Hagan Bayley
Texas A&M University

David A. Bernlohr
University of Minnesota

Peer Bork
EMBL, Heidelberg

James U. Bowie
University of California, Los Angeles

Axel T. Brünger
Stanford University

A.L. Burlingame
University of California, San Francisco

Robert Cohen
University of Iowa

Valerie Daggett
University of Washington

Ken A. Dill
University of California, San Francisco

Russell F. Doolittle
University of California, San Diego

Joseph J. Falke
University of Colorado

Angelo Fontana
University of Padua

Julie D. Forman-Kay
University of Toronto

Michel E. Goldberg
Pasteur Institute

David P. Goldenberg
University of Utah

F. Ulrich Hartl
Max-Planck Institut für Biochemie, Martinsried

Steven Henikoff
Fred Hutchinson Cancer Research Center

Jan Hermans
University of North Carolina

Arthur Horwich
Yale University

Tony E. Hugli
La Jolla Institute for Molecular Medicine

Donald F. Hunt
University of Virginia, Charlottesville

Louise N. Johnson
University of Oxford

Jeffery W. Kelly
The Scripps Research Institute

Jack F. Kirsch
University of California, Berkeley

Werner Kühlbrandt
Max-Planck Institut für Biophysik, Frankfurt a. M.

Kunihiko Kuwajima
University of Tokyo

Robert Liddington
University of Leicester

Mark O. Lively
Wake Forest University

Erwin London
SUNY, Stony Brook

Matthias Mann
University of Southern Denmark

Susan Marqusee
University of California, Berkeley

C. Robert Matthews
University of Massachusetts

Stephen L. Mayo
California Institute of Technology

J. Andrew McCammon
University of California, San Diego

C. Nick Pace
Texas A&M University

Mike Parker
St. Vincent's Institute of Medical Research, Victoria

Andreas Plückthun
University of Zurich

Douglas C. Rees
California Institute of Technology

Lynne Regan
Yale University

S. James Remington
University of Oregon, Eugene

Andrew D. Robertson
University of Iowa

George D. Rose
Johns Hopkins University

Luis Serrano
EMBL, Heidelberg

Manfred J. Sippl
University of Salzburg, Austria

Janet L. Smith
Purdue University

Michael F. Summers
University of Maryland, Baltimore County

Tom C. Terwilliger
Los Alamos National Laboratories

Gerhard Wagner
Harvard Medical School

Alex Wlodawer
NCI-Frederick

Editorial Office: *Protein Science*, Department of Biochemistry, Purdue University, Biochemistry Building, West Lafayette, IN 47907-1153, USA. Phone: 765-496-3459; Fax: 765-496-3460; E-mail: proteinscience@purdue.edu.

Publishing and Advertising Offices: Cold Spring Harbor Laboratory Press, 1 Bungtown Road, Cold Spring Harbor, New York 11724-2203. For advertising rates and data, contact the Advertising Manager Marcie Siconolfi: (Phone: 516-422-4010; Fax: 516-422-4092).

Instructions for Authors: See January 2001 issue of *Protein Science* for Detailed Instructions for Authors.

Books for Review: Publications should be sent to Richard N. Perham (Department of Biochemistry, University of Cambridge, Tennis Court Road, Cambridge CB2 1QW, England). Authors and readers are encouraged to draw to the attention of the Book Review Coordinator recent publications that may be of interest to readers of *Protein Science*.

Subscriptions: Published monthly. Institutional: \$967 (U.S.); \$1,023 (Canada and Mexico); \$1,047 (R.O.W.). Personal rate (must be paid by check, credit card, or money order): \$235 (U.S.); \$291 (Canada and Mexico); \$315 (R.O.W.). Contact Kathleen Cicone, Subscription Manager, Fulfillment Department, 500 Sunnyside Boulevard, Woodbury, New York 11797-2924. Phone: Continental U.S. and Canada 1-800-843-4388; all other locations 516-422-4100. Fax: 516-422-4097. Claims for missing issues must be received within 4 months of issue date.

Member Subscriptions: Members of The Protein Society automatically receive *Protein Science* as part of their annual membership dues.

The Protein Society: Membership information and application forms may be obtained from: The Protein Society, 9650 Rockville Pike, Bethesda, MD 20852-3998, USA. Phone: 301-571-0662 or 800-99-AMINO; Fax: 301-571-0666; E-mail: newburgh@protein.faseb.org.

Protein Science is covered by Chemical Abstracts, CASI, Cambridge Scientific Abstracts, Current Contents/Life Sciences, SciSearch, Science Citation Index, Research Alert, Macromolecular Structures, EMBASE/Excerpta Medica, Index Medicus, MEDLINE, and MEDLARS.

All rights reserved. No part of this publication may be reproduced, in any form or by any means, electronic, photocopying or otherwise, without permission in writing from Cold Spring Harbor Laboratory Press, except when permission is obtained through a bona fide copyright clearinghouse, such as the Copyright Clearance Center (USA) or the Copyright Licensing Agency (UK). The CCC Item-Fee Code for this publication is 0961-8368/01 \$12.50 + .10. Specific written permission must be obtained for all other copying. **Photocopying for Classroom Use:** Permission to reproduce parts of this publication for course anthologies may be obtained directly from Cold Spring Harbor Laboratory Press or through duly authorized services such as the Academic Permissions Service of the CCC, 222 Rosewood Drive, Danvers, MA 01923 (Telephone 978-750-8400) and the Copyright Permissions Service of the National Association of College Stores, or through any photoduplication service that properly secures permission from Cold Spring Harbor Laboratory Press or the CCC.

Periodicals postage paid at Hicksville, NY, and additional mailing offices. Forwarding and Return Postage Guaranteed. Address Correction Requested. **Postmaster:** Send address changes in the USA and Canada to: *Protein Science*, Cold Spring Harbor Laboratory Press, Journals Fulfillment Dept., 10 Skyline Drive, Plainview, NY 11803-2510.

Copyright © 2001 The Protein Society

Protein Science

A PUBLICATION OF THE PROTEIN SOCIETY

Volume 10, Number 9

September 2001

CONTENTS

REVIEWS

- A structural basis for processivity 1699
Wendy A. Breyer and Brian W. Matthews
- Sequence-structure analysis of FAD-containing proteins 1712
Orly Dym and David Eisenberg

ARTICLES

- Role of the single cysteine residue, Cys 3, of human and bovine cystatin B (stefin B) in the inhibition of cysteine proteinases 1729
Ewa Pol and Ingemar Björk
- Oligomerization and ligand binding in a homotetrameric hemoglobin: Two high-resolution crystal structures of hemoglobin Bart's (γ_4), a marker for α -thalassemia 1739
Richard D. Kidd, Heather M. Baker, Antony J. Mathews, Thomas Brittain, and Edward N. Baker
- Mechanism of pressure-induced thermostabilization of proteins: Studies of glutamate dehydrogenases from the hyperthermophile *Thermococcus litoralis* 1750
Michael M.C. Sun, Raphaele Caillot, Gary Mak, Frank T. Robb, and Douglas S. Clark
- Phenylalanine fluorescence studies of calcium binding to N-domain fragments of *Paramecium* calmodulin mutants show increased calcium affinity correlates with increased disorder 1758
Wendy S. VanScyoc and Madeline A. Shea
- Rational design of enantioselective enzymes requires considerations of entropy 1769
Jenny Ottosson, Johanna C. Rotticci-Mulder, Didier Rotticci, and Karl Hult
- Role of the single disulphide bond of β_2 -microglobulin in amyloidosis in vitro 1775
David P. Smith and Sheena E. Radford
- Substitution of conserved methionines by leucines in chloroplast small heat shock protein results in loss of redox-response but retained chaperone-like activity 1785
Niklas Gustavsson, Bas P.A. Kokke, Björn Anzeliuss, Wilbert C. Boelens, and Cecilia Sundby

(continued)

Cover Illustration: Ribbon diagram of the carbonmonoxy form of hemoglobin (Hb) Bart's, a non-cooperative homotetramer of Hb γ chains. Hb Bart's spontaneously forms in the blood of sufferers of α -thalassemia. (See Kidd et al., pp. 1739–1749.)

Published by Cold Spring Harbor Laboratory Press



Contents (continued)

Effects of turn residues in directing the formation of the β -sheet and in the stability of the β -sheet	1794
<i>Pei-Yeh Chen, Chih-Kai Lin, Chung-Tien Lee, Howard Jan, and Sunney I. Chan</i>	
The sequence determinants of cadherin molecules	1801
<i>Alexander E. Kister, Michael A. Roytberg, Cyrus Chothia, Jurii M. Vasiliev, and Israel M. Gelfand</i>	
Arginine-140 and isoleucine-141 determine the 17 β -estradiol-binding specificity of the sex-steroid-binding protein (SBP, or SHBG) of human plasma	1811
<i>Philip H. Petra, Elinor T. Adman, William R. Orr, Katherine T. Woodcock, Christine Groff, and Li-Ming Sui</i>	
Influence of Glu-376 $\rightarrow$ Gln mutation on enthalpy and heat capacity changes for the binding of slightly altered ligands to medium chain acyl-CoA dehydrogenase	1822
<i>Karen M. Peterson, K.V. Gopalan, Andreas Nandy, and D.K. Srivastava</i>	
A cofactor-dependent phosphoglycerate mutase homolog from <i>Bacillus stearothermophilus</i> is actually a broad specificity phosphatase	1835
<i>Daniel J. Rigden, Irina Bagyan, Ejvis Lamani, Peter Setlow, and Mark J. Jedrzejewski</i>	
Site-directed mutations of human hemoglobin at residue 35 β : A residue at the intersection of the α 1 β 1, α 1 β 2, and α 1 α 2 interfaces	1847
<i>Jeffrey S. Kavanaugh, Jamie A. Weydert, Paul H. Rogers, Arthur Arnone, Hilda L. Hui, Anita M. Wierzbicka, Laura D. Kwiatkowski, Paul Paily, Robert W. Noble, Stefano Bruno, and Andrea Mozzarelli</i>	
NMR of hydrogen bonding in cold-shock protein A and an analysis of the influence of crystallographic resolution on comparisons of hydrogen bond lengths	1856
<i>Andrei T. Alexandrescu, Doug R. Snyder, and Frits Abildgaard</i>	
Structures of an unliganded neurophysin and its vasopressin complex: Implications for binding and allosteric mechanisms	1869
<i>Chia Kuei Wu, Bing Hu, John P. Rose, Zhi-Jie Liu, Tam L. Nguyen, Changsheng Zheng, Esther Breslow, and Bi-Cheng Wang</i>	
Circularly permuted proteins in the protein structure database	1881
<i>Jongsun Jung and Byungkook Lee</i>	
Chemical denaturation and elevated folding temperatures are required for wild-type activity and stability of recombinant <i>Methanococcus jannaschii</i> 20S proteasome	1887
<i>Rob J. Frankenberg, Tina S. Hsu, Hisao Yakota, Rosalind Kim, and Douglas S. Clark</i>	
Characterization of bothrojaracin interaction with human prothrombin	1897
<i>Robson Q. Monteiro, Paul E. Bock, M. Lucia Bianconi, and Russolina B. Zingali</i>	

FOR THE RECORD

N-terminal extension changes the folding mechanism of the FK506-binding protein	1905
<i>Alla Korepanova, Chanel Douglas, Ilya Leyngold, and Timothy M. Logan</i>	
The PYRIN domain: A member of the death domain-fold superfamily	1911
<i>Wayne J. Fairbrother, Nathaniel C. Gordon, Eric W. Humke, Karen M. O'Rourke, Melissa A. Starovasnik, Jian-Ping Yin, and Vishva M. Dixit</i>	

(continued)

Contents (continued)

NEWS FROM THE PROTEIN SOCIETY	1919
UPCOMING MEETINGS	1920
ERRATUM.....	1922
PRODUCT NEWS	1923
FORTHCOMING PAPERS	vi

REVIEW

Sequence-structure analysis of FAD-containing proteins

ORLY DYM AND DAVID EISENBERG

University of California, Los Angeles-DOE Laboratory of Structural Biology and Molecular Medicine, University of California, Los Angeles, Los Angeles, California 90095-1570, USA

(RECEIVED April 4, 2001; FINAL REVISION June 21, 2001; ACCEPTED June 21, 2001)

Abstract

We have analyzed structure-sequence relationships in 32 families of flavin adenine dinucleotide (FAD)-binding proteins, to prepare for genomic-scale analyses of this family. Four different FAD-family folds were identified, each containing at least two or more protein families. Three of these families, exemplified by glutathione reductase (GR), ferredoxin reductase (FR), and *p*-cresol methylhydroxylase (PCMH) were previously defined, and a family represented by pyruvate oxidase (PO) is newly defined. For each of the families, several conserved sequence motifs have been characterized. Several newly recognized sequence motifs are reported here for the PO, GR, and PCMH families. Each FAD fold can be uniquely identified by the presence of distinctive conserved sequence motifs. We also analyzed cofactor properties, some of which are conserved within a family fold while others display variability. Among the conserved properties is cofactor directionality: in some FAD-structural families, the adenine ring of the FAD points toward the FAD-binding domain, whereas in others the isoalloxazine ring points toward this domain. In contrast, the FAD conformation and orientation are conserved in some families while in others it displays some variability. Nevertheless, there are clear correlations among the FAD-family fold, the shape of the pocket, and the FAD conformation. Our general findings are as follows: (a) no single protein 'pharmacophore' exists for binding FAD; (b) in every FAD-binding family, the pyrophosphate moiety binds to the most strongly conserved sequence motif, suggesting that pyrophosphate binding is a significant component of molecular recognition; and (c) sequence motifs can identify proteins that bind phosphate-containing ligands.

It is generally accepted that the three-dimensional structure of a polypeptide chain is determined by its amino acid sequence. Nevertheless, similar folds can have very different sequences. One of the ultimate goals in sequence analysis is to predict the structure and function of a protein based solely on its sequence. In cases where the protein of interest shares at least 30% amino acid identity with another protein, the two proteins generally exhibit similar three-dimensional

structure (Doolittle 1986). Alternatively, when proteins are known to have similar structure but divergent sequences, consensus sequence motifs can be used to assess the function of unassigned sequences. These consensus motifs usually correspond to residues interacting with cofactors, substrate, or other proteins.

The increasing number of three-dimensional structures of proteins in the Protein Data Bank, complexed with appropriate ligands, provides an important tool for understanding the mechanisms of molecular recognition. In this study, we focussed on flavin adenine dinucleotide (FAD) because it and its related cofactors, nicotinamide adenine dinucleotide (NADH) and adenosine triphosphate (ATP), appear in many biological processes.

Previous comparative structural studies of mononucleotide- and dinucleotide-binding proteins reveal that some exhibit a similar three-dimensional structure with conserved

Reprint requests to: David Eisenberg, UCLA-DOE Laboratory of Structural Biology and Molecular Medicine, UCLA, Box 951570, Los Angeles, CA 90095-1570, USA; e-mail: david@mbi.ucla.edu; fax: 310-206-3914.

Abbreviations: FAD, flavin adenine dinucleotide; FMN, flavin mononucleotide; ATP, adenosine triphosphate; NADH, nicotinamide adenine dinucleotide; NADPH, nicotinamide adenine dinucleotide phosphate; GR, glutathione reductase; FR, ferredoxin reductase; PO, pyruvate oxidase; PCMH, *p*-cresol methylhydroxylase.

Article and publication are at <http://www.proteinscience.org/cgi/doi/10.1101/ps.12801>.

sequence motifs at positions crucial for binding, although the remaining sequence can vary greatly. One of the first folds was identified by Rossmann (Rossmann et al. 1974) who discovered the correlation between the fold of dehydrogenases that bind the cofactor NADH and conserved sequence motifs. The basic structure consists of six parallel β -strands interspersed by α -helices that appear on both sides of the six-stranded β sheet. This symmetrical α/β structure is built from two halves, $\beta_1\alpha_1\beta_2\alpha_2\beta_3$ and $\beta_4\alpha_4\beta_5\alpha_5\beta_6$, with a crossover α -helix (α_3) connecting β_3 and β_4 (Fig. 1). Each of these folds is known as the classical mononucleotide-binding fold or the Rossmann fold.

A variation of the fold observed for dehydrogenases is found in FAD-containing proteins. This fold consists of one $\beta_1\alpha_1\beta_2\alpha_2\beta_3$ Rossmann fold, and a variation of both the second Rossmann fold and the crossover α -helix. This variation includes a three-stranded antiparallel β -sheet connecting β_3 and β_4 , instead of the crossover α -helix observed in dehydrogenases. Moreover the sixth strand, from the second Rossmann fold, is missing whereas the fifth strand is retained but is close to the end of the sequence. Different variations in which structural elements are added or deleted were found in proteins containing other mono- and/or dinucleotides such as flavin mononucleotide (FMN) (Rao and Rossmann 1973), and nicotinamide adenine dinucleotide phosphate (NADPH) (Schulz and Schirmer 1974). Most of these proteins show a series of conserved amino acid residues at positions interacting with the mono/dinucleotide molecule.

We selected a nonredundant set of 32 protein-FAD complex structures from the Protein Data Base (PDB) for structure-sequence analysis with the goal of deepened under-

standing of principles of molecular recognition. On the basis of sequence-structure comparison and the interaction of cofactor atoms with the different protein residues, we identified several conserved motifs for each structural family. Some of these were previously characterized by others (Schulz and Schirmer 1974; Schulz et al. 1982; Wierenga et al. 1983; Schulz 1992; Correll et al. 1993; Lu et al. 1994; Ingelman et al. 1997; Fraaije et al. 1998) and some are newly derived. These conserved sequence motifs are called "most conserved" when they are present in all family members, and "partially conserved" when present in some but not all family members. In addition to the sequence-structure analysis, we investigated a more complete set of variables, including cofactor conformation, characteristics of the protein pocket wherein the cofactor is bound, cofactor directionality, and correlation of cofactor moieties (adenine, pyrophosphate isoalloxazine, etc.) interacting with conserved sequence motifs in the different family folds. Such fundamental discriminators may improve our understanding of protein evolution, in particular for FAD-binding proteins where many tertiary structures, often distantly related, are known. Furthermore, the presence of a variety of conserved sequence motifs in FAD families, specifically those that are pyrophosphate-binding, can be used as a tool for molecular recognition of phosphate analog-binding proteins.

Results

FAD cofactor

The dinucleotide FAD consists of adenosine monophosphate (AMP) linked to flavin mononucleotide (FMN) by a pyrophosphate bond (Fig. 2A). The AMP moiety is composed of the adenine ring bonded to a ribose that is linked to a phosphate group. The FMN moiety, also known as the riboflavin moiety, is composed of the isoalloxazine-flavin, linked to a sugar, ribitol, which is coupled to a phosphate group. The catalytic function of the FAD is concentrated in the isoalloxazine ring, whereas the ribityl phosphate and the AMP moiety mainly stabilize cofactor binding to protein residues. The flavin functions mainly in a redox capacity, being able to take up two electrons from one substrate and release them two at a time to a substrate or coenzyme, or one at a time to an electron acceptor. The surrounding protein controls many of the catalytic properties of the flavin ring, such as the rate of accepting electrons, the pathway of electron flow within the flavin ring, and the flavin's oxidation-reduction potential (Mathews 1991). This control is asserted in part by the cofactor conformation, which adapts to the protein as either an elongated or bent butterfly conformation (Fig. 2A and B, respectively). In the bent conformation, the AMP portion is folded back, placing the adenine and isoalloxazine rings in close proximity, whereas in the

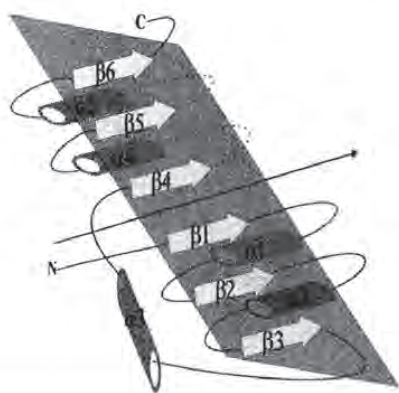


Fig. 1. Topology diagram of dehydrogenases that bind NADH. This symmetrical α/β structure is composed of two halves, $\beta_1\alpha_1\beta_2\alpha_2\beta_3$ and $\beta_4\alpha_4\beta_5\alpha_5\beta_6$, with a crossover α -helix (α_3) connecting strands β_3 and β_4 . Each of these motifs is known as the classical mononucleotide-binding motif or the Rossmann fold. Cylinders represent α -helices and arrows denote β -strands. Dashed lines indicate elements of secondary structure below the plane of the fold. The overall pseudo two-fold rotation symmetry between the two $\beta\alpha\beta$ motifs is shown by an arrow.

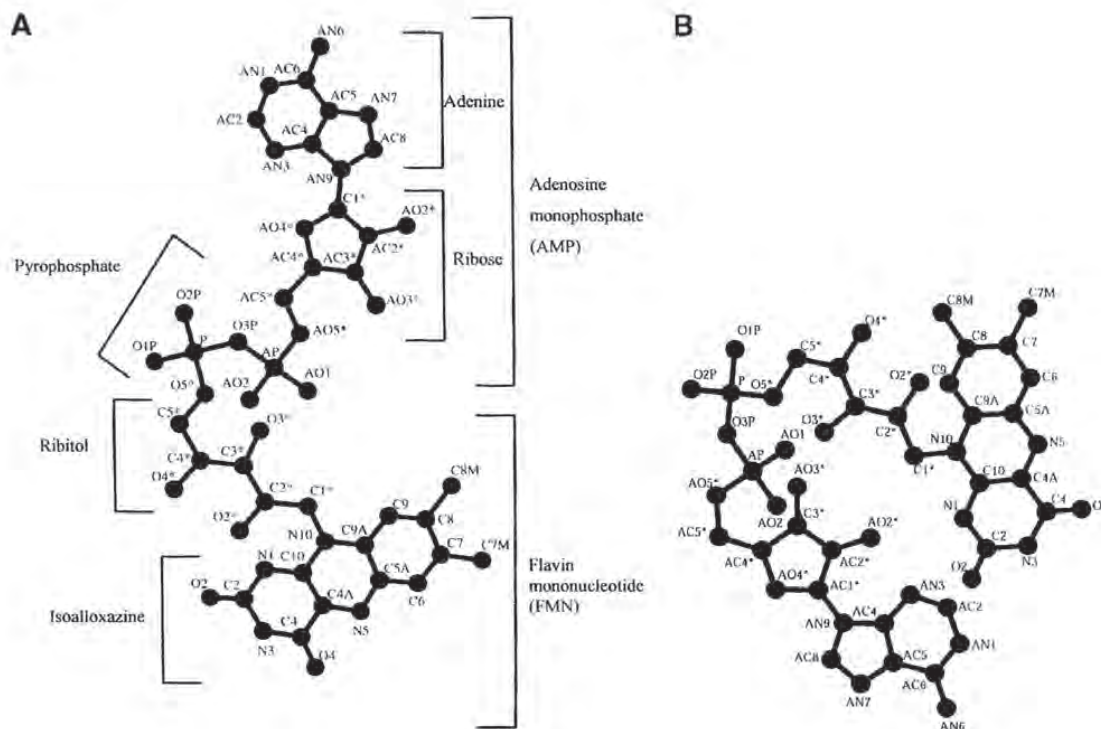


Fig. 2. The FAD cofactor conformations. (A) An elongated conformation where the adenine ring is distal from the isoalloxazine ring. The division between the component parts FMN and AMP composing the FAD is shown. The AMP is composed of an adenine ring connected to a ribose that is connected to a phosphate group. The FMN moiety is composed of the isoalloxazine-flavin ring linked to a ribitol, which is connected to a phosphate group. (B) Bent conformation where the AMP portion is folded back, placing the adenine and isoalloxazine rings in close proximity. Variation in the proximity of the two rings determines the degree of cofactor flexibility. All atoms composing the FAD cofactor are labeled.

elongated conformation the adenine ring is distal from the isoalloxazine ring.

The glutathione reductase (GR) structural family

Structural conservation

One of the most thoroughly studied FAD-containing families is represented by the enzyme glutathione reductase (GR). The protein members of this populous family catalyze diverse reactions (Karplus and Schulz 1987; Chen et al. 1994; Mande et al. 1996; Mizutani et al. 1996; Yeh et al. 1996; Enroth et al. 1998; Eppink et al. 1998; Binda et al. 1999; Bond et al. 1999; Lennon et al. 1999; Leys et al. 1999; Trickey et al. 1999; Wohlfahrt et al. 1999; Yue et al. 1999; Ziegler et al. 1999) (Table 1). All family members adopt the Rossmann fold ($\beta_1\alpha_1\beta_2\alpha_2\beta_3$ in Fig. 3A). The topology of the GR family consists of a central five-stranded parallel β -sheet ($\beta_1, \beta_2, \beta_3, \beta_7$, and β_8) surrounded by α -helices (α_1 and α_2) and an additional crossover connection composed of a three-stranded antiparallel β -sheet (β_{4-6}) (Fig. 3A).

Of the 15 GR family members listed in Table 1, two separate subfamilies were identified using the CE program for comparing a polypeptide chain to each chain in the PDB (Shindyalov and Bourne 1998). One subfamily, GR₁, contains the first seven proteins listed in Table 1; the last eight proteins belong to the second subfamily, GR₂. Each of the two subfamilies is obtained when any of the proteins belonging to it is used as a query in the sequence-structure search. Whereas proteins from the GR₁ subfamily align well through the entire FAD-binding domain, those belonging to the GR₂ subfamily align well only in their N-terminal (~30 residues). Nevertheless, all GR family members share a similar overall three-dimensional structure in their FAD-binding domain as well as at least one conserved sequence motif (Table 1). Terminal additions as well as various insertions within the fold are present in the members of GR₂, however. Specifically, insertions of several secondary structures are observed in the connections between β_2 to α_2 and α_2 to β_3 of the Rossmann fold (Fig. 3A). This is in agreement with the observation that these proteins align well only in their N-terminal residues comprising β_1, α_1 , and β_2 . Notably, proteins from GR₁, with the exception of sulfide de-

Table 1. Cofactors and conserved sequence motifs in the nonredundant FAD-containing protein structures retrieved from the Protein Data Bank and used in our sequence-structure analysis

PDB Code	Family	Protein (Reference)	Cofactors	C, NC ^a	B, E ^b	Sequence motifs
1joa	GR ₁	Glutathione reductase (Karplus and Schulz 1987)	FAD, NADH	NC	E	GxGxxG(x) ₁₇ E, hhhxGxGxxAxEx, T(x) ₅ TxxGD, D(x) ₆ GxxP
1icd		Sulfide dehydrogenase (Chen et al. 1994)	FAD	C	E	GxGxxG(x) ₁₉ E, hhhxPxPxPxEx, S(x) ₆ HxxGD, D(x) ₆ PxxA
1ebd		Dihydrolipoamide dehydrogenase (Mande et al. 1996)	FAD, NADH	NC	E	GxGxxG(x) ₁₇ E, hhhxGxGxxGxE, T(x) ₅ FxxGD, D(x) ₆ GxxP
1bzl		Trypanothione reductase (Bond et al. 1999)	FAD, NADPH	NC	E	GxGxxG(x) ₁₈ D, hhhxGvxxxSxE, T(x) ₅ YxxGD, D(x) ₆ GxxP
3grs	GR ₂	NADH peroxidase (Yue et al. 1999)	FAD, NADPH	NC	E	GxGxxG(x) ₁₉ E, hhhxGxGxxGxE, T(x) ₅ FxxGD, D(x) ₆ GxxP
1cjc		Adrenodoxin reductase (Ziegler et al. 1999)	FAD, NADPH	NC	E	GxGxxG(x) ₁₉ E, hhhxGxGxxAxEx, V(x) ₆ YxxGW, GxGxxG(x) ₂₁ , FxxGD,
1c10		Thioredoxin reductase (Lennon et al. 1999)	FAD, NADPH	NC	E	D(x) ₈ GxxP
1foh		Phenol hydroxylase (Enroth et al. 1998)	FAD, NADPH	NC	E	GxGxxG(x) ₁₆ D, S(x) ₅ FxxGD
1bf3		<i>p</i> -Hydroxybenzoate hydroxylase (Eppink et al. 1998)	FAD, NADPH	NC	E	GxGxxG(x) ₁₇ E, FxxGD
1cf3		Glucose oxidase (Wohlfahrt et al. 1999)	FAD	NC	E	GxGxxG(x) ₁₈ E
1b37		Polyamine oxidase (Binda et al. 1999)	FAD	NC	E	GxGxxG(x) ₁₈ E, T(x) ₅ FxxGD
1d4c		Fumarate reductase (Leys et al. 1999)	FAD	C	E	GxGxxG(x) ₁₇ E
1b4v		Cholesterol oxidase (Yue et al. 1999)	FAD	NC	E	GxAxxG
1aa8		D-Amino acid oxidase (Mizutani et al. 1996)	FAD	NC	E	GxGxxG(x) ₁₉ D
1b3m	FR	Sarcosine oxidase (Trickey et al. 1999)	FAD	C	E	GxGxxG(x) ₁₇ D
1fdr		Flavodoxin reductase (Ingelman et al. 1997)	FAD, NADPH	NC	B	RxYS, GxxSxxL(x) ₅ G(x) ₈ AxG, MxxxGTAlxP
1a8p		Ferredoxin reductase (Sridhar Prasad et al. 1998)	FAD, NADPH	NC	B	RxYS, GxxTxxL(x) ₅ G(x) ₈ PxG, MxxxGTGlxP
1ndh		NADH-Cytochrome b ₅ reductase (Nishida et al. 1995)	FAD, NADH	NC	E	RxYT, GxxSxxL(x) ₅ G(x) ₇ PxG, MxxxGTGlxP
2cnd		Nitrate reductase (Lu et al. 1994)	FAD, NADH	NC	E	RxYT, GxxTxxL(x) ₅ G(x) ₇ PxG, MxxxGSGlxP
1amo		NADPH-P450 reductase (Wang et al. 1997)	FAD, NADPH	NC	E	RxYS, GxxTxxL(x) ₆ G, MxxxGTGlxP
1cqx	PCMH	Flavohemoglobin (Ermler et al. 1995)	FAD, NADH	NC	E	RxYS, GxxSxxL(x) ₆ G(x) ₇ PxG
1qlt		<i>p</i> -Cresol methylhydroxylase (Cunane et al. 2000)	FAD	C	E	P(x) ₆ GxN, G(x) ₇ GY, K(x) ₆ E(x) ₂ YxxVxxG(x) ₈ Y, R, GxxL
1dii		Vanillyl-alcohol oxidase (Fraaije et al. 2000)	FAD	C	E	P(x) ₆ GxN, G(x) ₇ GY, R(x) ₆ E(x) ₂ YxxVxxG(x) ₈ Y, R, GxxL
2mbr		Uridine diphospho-N-acetylenol-pyruvylglucosamine reductase (Benson et al. 1997)	FAD, NADPH	NC	E	P(x) ₆ GxN, G(x) ₈ AY, K(x) ₆ E(x) ₄ YxxVxxG(x) ₈ Y, R, GxxL
1qj2		Carbon monoxide dehydrogenase (Dobbe et al. 1999)	FAD	NC	E	P(x) ₈ GxN, G(x) ₃ AY, R(x) ₅ D(x) ₅ YxxAxxG(x), R
1f0x		D-Lactate dehydrogenase (Dym et al. 2000)	FAD	NC	E	A(x) ₇ AxN, G(x) ₇ GS, R
1pow	PO	Pyruvate Oxidase (Muller et al. 1994)	FAD	NC	E	R(x) ₅ GxG, VGxN, K(x) ₇ IxxDP(x) ₉ D(x) ₃ ADxxK, KxLxxLxxxL(x) ₆ T(x) ₄ GxV
1efv		Electron transfer (Roberts et al. 1996)	FAD	NC	E	K(x) ₅ GxG, K(x) ₇ VGxS, IxxDP(x) ₈ D(x) ₄ ADxxK, KxLxxLxxxL(x) ₆ S(x) ₆ GxV
1ivh		Acyl-CoA dehydrogenase (Tiffany et al. 1997)	FAD	NC	E	
1dnp		DNA Photolyase (Park et al. 1995)	FAD	NC	B	
1b5t		Methylenetetrahydrofolate reductase (Guenther et al. 1999)	FAD, NADPH	NC	E	
1qr2		Quinone oxidoreductase (Foster et al. 1999)	FAD, NADPH	NC	E	

^a FAD bond type: C, covalent; NC, noncovalent.^b Conformation: B, bent; E, elongated.^c SM signifies the four single-membered families.

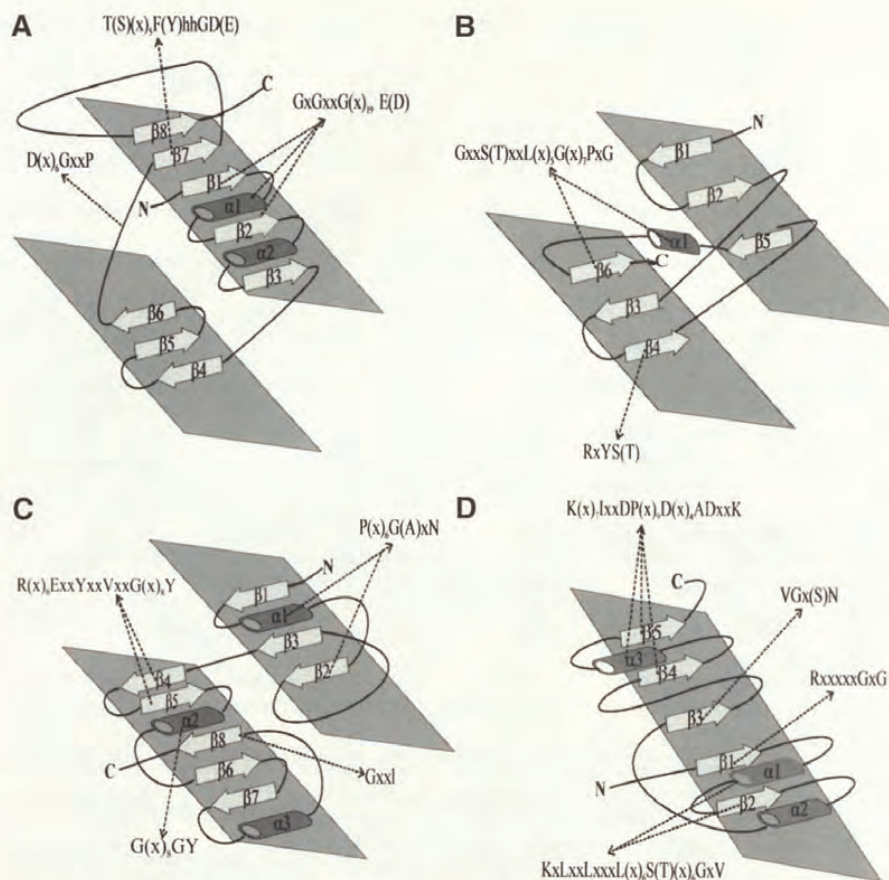


Fig. 3. Topological diagram of FAD-binding domain of the four FAD-family folds. (A) Rossmann fold ($\beta_1\alpha_1\beta_2\alpha_2\beta_3$) adopted by the glutathione reductase (GR) family members. For a full description of this fold, it must be noted that there are two subfamilies, GR₁ and GR₂ (see text), and that there are exceptions to the generalizations described here. For example, D-amino acid oxidase of the GR₂ subfamily is an exception to the rule that the FAD-binding fold in the GR family contains a 3-strand β -meander connecting β_3 and β_4 ; instead, it has a crossover α -helix. (B) Ferredoxin reductase (FR) family fold adopting a cylindrical β -domain organized into two orthogonal sheets, $\beta_1\beta_2\beta_3$ and $\beta_3\beta_4\beta_6$. (C) The *p*-cresol methylhydroxylase (PCMH) family fold consists of two $\alpha + \beta$ subdomains; one is composed of three parallel β -strands (β_{1-3}) and the second contains five antiparallel β -strands (β_{4-8}) surrounded by α -helices. (D) The pyruvate oxidase (PO) family fold consists of five parallel β -strands (β_{1-5}) interspersed by α -helices similar to the double Rossmann fold found in dehydrogenases. Cylinders represent α -helices and arrows denote β -strands. The location, indicated in dashed lines, of the conserved sequence motifs in each of the FAD-family folds is listed in Table 1.

hydrogenase (Van Driessche et al. 1996), comprise NAD(P)H-binding domains (between β_7 and β_8 in Fig. 3A), adopting the Rossmann fold (Rossmann et al. 1974), in addition to the FAD-binding domain. Phenol hydroxylase (Enroth et al. 1998) and *p*-hydroxybenzoate hydroxylase (Eppink et al. 1998) are the only proteins from the second subfamily known to bind NADPH. However, both lack the Rossmann-type NADPH-binding fold.

FAD binding and conformation

The FAD cofactor in the GR family members adopt elongated conformations with the adenine and isoalloxazine moieties distal from each other (Table 1 and Fig. 4A). The

degree of extension of the cofactor can vary considerably. In addition, the cofactor may be either covalently or noncovalently bound to the protein, and there is no apparent correlation of cofactor conformation with any of the conserved sequence motifs (Table 1). In contrast, the direction of the FAD cofactor is conserved among all GR family members: the adenine ring of the cofactor points toward the FAD-binding domain, while the isoalloxazine ring points away from it (Fig. 4A).

Conserved sequence motifs

The GR family members show several conserved amino acid residues at a few crucial positions, and the rest of the

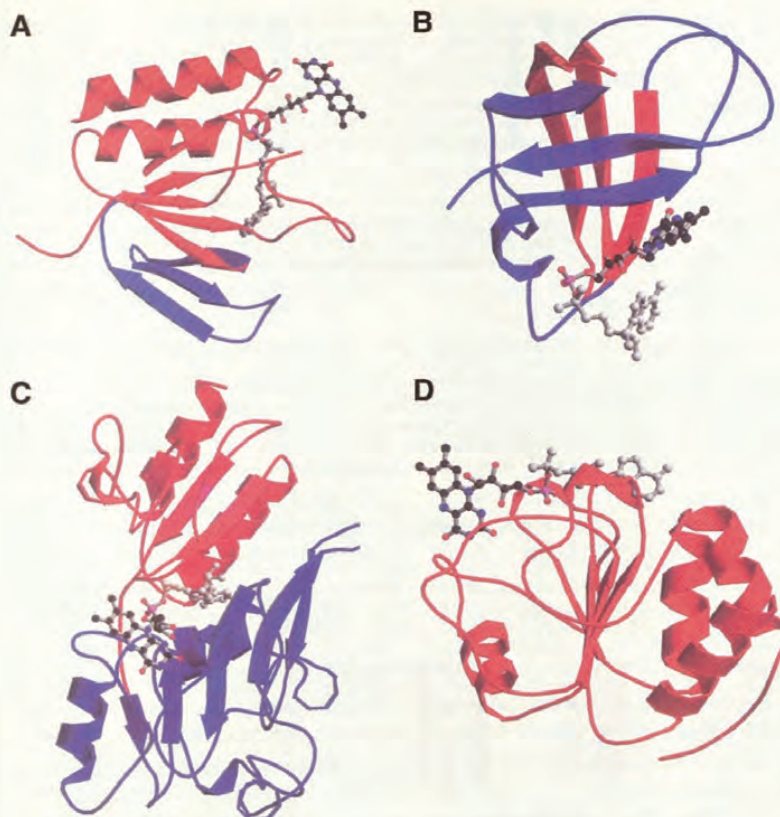


Fig. 4. Ribbon representation of the FAD-binding domain of the four FAD-family folds complexed with FAD. (A) Rossmann fold adopted by the glutathione reductase (GR) family members. The blue shading indicates the crossover connection composed of a three-stranded antiparallel β -sheet; the red shading indicates the central five-stranded parallel β -sheet surrounded by two α -helices. The FAD cofactor adopts an elongated conformation with the adenosine moiety (gray circles) pointing toward the FAD-binding domain. (B) Ferredoxin reductase (FR) family. The two antiparallel three-stranded β -sheets are shown in red and blue. The FAD in its bent conformation is shown with the isoalloxazine ring (black circles) pointing toward the FAD-binding domain. (C) The $\alpha + \beta$ fold adopted by the *p*-cresol methylhydroxylase (PCMH) family members. In red are three parallel β -strands surrounded by α -helices, and in blue are five antiparallel β -strands surrounded by α -helices. The FAD molecule adopts an elongated conformation and is located in between the two subdomains with the adenine ring pointing toward them. (D) Rossmann fold adopted by pyruvate oxidase (PO) family members, shown in red. The cofactor adopts an elongated conformation and lies perpendicular to the β -strands. The stick drawing of the cofactor is depicted with gray circles for atoms in the adenine and sugar rings, with red circles for phosphate and oxygen atoms, and black circles for atoms for the isoalloxazine ring. Figure created by MOLSCRIPT (Kraulis 1991) and RASTER3D (Bacon and Anderson 1988; Merritt and Murphy 1994).

sequence varies. We conducted a sequence-structure alignment of the two GR subfamily members using the program CE. Conserved sequence motifs found in each of the GR family members are listed in Table 1. Specifically, four conserved motifs for FAD binding are found in GR₁ members (depicted in colored boxes in Fig. 5), two of which are present in the GR₂ subfamily as well (Table 1).

The most conserved and well-studied sequence motif is part of the Rossmann fold, xhxhGxGxxGxxxhxxh(x)₈hxhE(D) (Fig. 5, blue box), where x is any residue and h is a hydrophobic residue (Schulz and Schirmer 1974; Schulz et al. 1982; Wierenga et al. 1983; Schulz 1992). This motif is found at the N-terminal part of the sequence, and in fact it

is the only conserved sequence motif present in all GR family members (Table 1). This is in agreement with the observation that while members of GR₁ align well throughout the entire FAD-binding domain, GR₂ members align well mainly at the N-terminus. This consensus is known as the dinucleotide-binding motif (DBM) (Wierenga et al. 1983) or the phosphate-binding sequence signature (Moller and Amons 1985) and is a common motif among FAD- and NAD(P)H-dependent oxidoreductases. The central part of this consensus motif, GxGxxG, is part of the loop connecting the first β -strand and α -helix in the $\beta\alpha\beta\alpha\beta$ Rossmann fold with the N-terminal end of helix α_1 pointing toward the pyrophosphate moiety for charge compensation (Fig. 3A).

1JOA:	1/2	MKVIVLSSHGYYEAVEELNLHPDAEIQWYK-G-DF-----IS----FLSXGMQL
1FCD:A	3/4	RKVVVVGSGTGATAAKYIKLADPSIEVTLEP-N-TD-----Y----YTCYLSNE
1EBD:A	5/12	-ETLVVVGAGPGGYVAARAA-QL-GQKVTIVEK-GN-----LGGVCLNVGCI PSK
1BZL:A	6/8	-DLVVIGAGSGGLEAAWNAATLY-KKRVAVIDV-Q-MVHGPPFFSALGGTCVNVGCVPKK
3GRS:	22/23	-DYLVIIGSGGLASARRAA-EL-GARAAVVES-HK-----LGGTCVNVGCVPKK
1CJC:A	7/8	PQICVVGSGPAGFYTAQHLLKHHSRAHVDIYEK-Q-LV-----PFGL--VRFVAPD
1CL0:A	6/7	SKLLILGSGPAGYTAAYVAAR--ANLQPVLTGME-KG-----GQ----LTTTTEVE
1JOA:	47/48	----YLE-----GKVKDV-NSV-----RYMTGEKM
1FCD:A	48/49	----VIG-----GDRK-L-ESI-----K-HGYDGL
1EBD:A	51/58	----ALISASHRYEQAKHSEEMGIKAENVITIDPAK--VQEWKA-SVVKK--LTGGVEGL
1BZL:A	62/64	----LMVTGAQYMEHLRESAGFGWEFDRTTLRAEWKNLIAVKD-EAVLN--INKSYDEM
3GRS:	68/69	----VMWNTAVHSEPMHDHADYGFPSCEGKFNWRV--IKEKRD-AYVSR--LNAIQNN
1CJC:A	55/56	----H-----PEVKNVINTFTQTA
1CL0:A	51/52	NWPGD-----PNDLTGPLL-----MERMHEHA
1JOA:	67/68	ESRG--VNVFSNTEITAIQ--P---KEHQVTVKD----LVSGEER--VE--NYDKLII
1FCD:A	66/67	RAHG--IQVVH-DSATGID--P---DKKLKTA-----GGA--EF--GYDRCVV
1EBD:A	101/108	LKGNK-VEIVK-GEAYFVD--A-----NTVRVVG--DSAQ--TY--TFKNAIL
1BZL:A	114/116	PRDTEGLEFFL-GWGSLES--K-----NVNVNRESADPAS--AVKE--RL--ETEHILL
3GRS:	118/119	LTKSH-IEIIR-GHAAFTSDPK-----PTIEV-----SGK--KY--TAPHILI
1CJC:A	70/71	RSDR--CAFYGNVEVG-----RDVTVQELQDAYHAVVL
1CL0:A	73/74	TKFE--TEII--FDHINKV--DLQNRPFRLNGD-----NG--EY--TCDALII
1JOA:	110/111	SPGAV-PFELD---IPGK---DL---DNIYLM-RGRQWAIKLL-----OQTVDFEV
1FCD:A	103/104	APGIE-LIYDK---IEGYSEAA---AKLPHAWKAGEQTALIR-----KQLEDMADG
1EBD:A	140/147	ATGSR-PIELP---N--F---KFSNR--ILDS-T--GALNL-----GEV---P-
1BZL:A	159/161	ASGSW-PHPMN---IPGI---EH---CISS-N--EAFYL-----PEP---P-
3GRS:	155/156	ATGGM-PSTPHESQIPGA--SL---GITS-D--GFFQL-----EEL---P-
1CJC:A	101/102	SYGAEDHQALD---IPGE---EL---PGVFSR-R--AFVWYNGLPENRELAP--DLSC-
1CL0:A	111/112	ATGAS-ARYL-----G--LP--SEEAF-----KGR-----
1JOA:	150/151	NNVVVIS---SYI---AAEAFKA-----G-----KKVT
1FCD:A	148/149	GTVVIAPPAAPFRCPGPFYERASQVAYY-----LKAHKPMKVI
1EBD:A	171/178	KSLAVIIG---SYI---ELGTAYANF-----G-----TKVT
1BZL:A	189/191	RRVLTIVG---CFISVFAGIFNAY-----GPKD---GQVT
3GRS:	188/189	GRSVIVGA---GYIAVVMAGILSAL-----G-----SKTS
1CJC:A	146/147	DTAVILQ---SNVLLVARILLTPPDHLEKTDITEAALGALRQSRV-----KTVW
1CL0:A	131/132	--GVSA-----CATCDGFFYRN-----QKVA
1JOA:	236/237	D--LVVVAVGVN--TA--WLKG--TLELHPNGLIKTD-E-----YMR-T-SE--P
1FCD:A	248/249	D--VINLIIPQAGKIAQ--IAG-----LTNDAGWCPVDIK-----TFESSIH--K
1EBD:A	260/267	D--YVLVTVGRRPN--TD--ELGLEQIGIKMTNRGLIEVD-Q-----QCRT-SV--P
1BZL:A	279/281	D--LVMMIAIGRSP--TK--DLQLQAGVMIKNG-GVQVD-E-----YSRT-NV--S
3GRS:	283/284	D--CLLWAIGRVN--TK--DLSLNKLGITDDKGHIIVD-E-----FQNT-NV--K
1CJC:A	323/324	G--LVLSSIGYKSR--PIDPSVPF-----DPKLGVPVNM-----EGR-VVDVP
1CL0:A	235/236	DVAGLFVAIGHSEN--TA---IFE---GQLELENGYIKVQ-SGIHGNTQTSTI---P
1JOA:	275/276	DVFAVGDATLIKYNPADTEVNIALA--TNARKQGRFAVKNL-----E-EPV-KPPFGVQG
1FCD:A	288/289	GIHVIDGASIA---NMPKSG--YSANSQGVAAAVVLLKG-BEP-GTPSYL-N
1EBD:A	302/309	NIFAIGDIVP-----GPALA--HKASYEGKVAAEAI---A-GHP-SAVDYVAI
1BZL:A	320/322	NIYAIGDVTN-----RVMLT--PVAINEAAALVDTV--FGTTP-RKTDHTRV
3GRS:	325/326	GIYAVGDCVG-----KALLT--PVAIAAGRKLHRL--FEYKESKLDYNNI
1CJC:A	361/362	GLYCSEWVKR-----GPTGVI--TTMTDTSFLTQQL--L-Q-----
1CL0:A	280/281	GVFAAGDVMD-----HIYRQAITSAITGCMAALDAE-----R-YL-----
1JOA:	326/327	SSGLAVF-DYKFASTGINEVMAQKLKGET-----KAVTVVEDYLMDFNPDQK-----
1FCD:A	336/337	TCYSILAPAYGISVAA-----
1EBD:A	343/350	PAVVFSD--PECASVGYFEQ-----QAKDEGIDV--IAAKFPF-----AANGRA
1BZL:A	362/364	ASAVFSI--PPIGTCGLIEE-----VASKRYEVV--AVYLSSF-----T-----
3GRS:	368/369	PTVVFHSH--PPIGTVGLTED-----EATHKYGIENVKTYSTSF-----T-----
1CJC:A		-----
1CL0:A		-----
1JOA:	372/373	K-----AWFKLVYDPETTTQILGAQLMSKADLTANINAI SLAIQAKMTIEDLAY
1FCD:A		-----
1EBD:A	383/390	LALNDTD-----GFLKLVRKEDGVIIGAIIGP-NASDMIAELGLAIEAGMTAEDIAL
1BZL:A	397/399	PLMHKVSQSKYKTFVAKIITNHSDDGTVLGVHLLGD-NAPEIIQIGIGICLKLNAKISDFYN
3GRS:	405/406	PMYHAVTKRKTK-CVMKMCANKEEKVVGIIHQGL-GCDEMLQGFVAVKMGATKADFDN
1CJC:A		-----
1CL0:A		-----

Fig. 5. Structure-based amino acid alignment for proteins belonging to the GR₁ family of proteins as obtained with the program CE (Shindyalov and Bourne 1998) (first seven proteins listed in Table 1). The first column gives the PDB accession code, and the second column gives two numbers that each specify the sequence position that follows. The left-hand number corresponds to the sequence and the right-hand to the PDB. The conserved sequence motifs are depicted in boxes: GxGxxGxxxhxxh(x)₆hxD(E) in blue, VVV-V(I)GxGxxGxE in red, T(S)xxxxF(Y)hhGD(E) in pink, and D(x)₆GxxP in green. In yellow are all conserved residues not directly involved in binding FAD. Proteins: 1JOA, NADH peroxidase from *Enterococcus faecalis*; 1FCD, flavocytochrome C sulfide dehydrogenase from *Chromatium vinosum*; 1EBD, dihydrolipoamide dehydrogenase from *Bacillus stearothermophilus*; 1BZL, trypanothione reductase from *Trypanosoma cruzi*; 3GRS, glutathione reductase from *Homo sapiens*; 1CJC, adrenodoxin reductase from *Bos taurus*; 1CL0, thioredoxin reductase from *Escherichia coli*.

The importance of the Gly residues in the conserved central GxGxxG is well understood (Wierenga et al. 1986): the first strictly conserved Gly allows for a tight turn of the main chain, which is important for positioning the second Gly. The second Gly, because of its missing side chain, permits close contact of the main chain to the pyrophosphate of FAD, specifically oxygen atoms O_{P1} or O_{P2} . The third Gly allows close packing of the helix with the β -sheet. The hydrophobic residues provide hydrophobic interactions of the α -helix with the β -sheet. The conserved negatively charged terminal residue, Glu or Asp, hydrogen bonds to the ribose 2' hydroxyl of the adenosine moiety. A variation of this glycine-rich sequence motif is hhhxGxGxxGxE (Fig. 5, red box), in which the central GxGxxG is common (Wierenga et al. 1986). It is part of the $\beta\alpha\beta$ Rossmann NAD(P)H-binding motif and is present only in GR₁ subfamily members containing the NAD(P)H-binding domain in addition to the FAD-binding domain (first seven proteins in Table 1). Sulfide dehydrogenase belongs to the GR₁ subfamily; however, it lacks the NAD(P)H-binding domain, consistent with the observation that glycines are replaced by prolines in this conserved motif (Fig. 5, red box). Two enzymes of GR₂, *p*-hydroxybenzoate (Eppink et al. 1998) and phenol hydroxylase (Enroth et al. 1998), are exceptional: they bind NADPH (Table 1) but lack a Rossmann-type NADPH-binding domain and therefore lack the conserved hhhxGxGxxGxE sequence motif typical of the NAD(P)H-binding domain (Wierenga et al. 1983).

Another highly conserved FAD-binding sequence motif was first identified from studies of rubredoxin reductase (Eggink et al. 1990), and is an 11 amino acid segment, T(S)xxxxxF(Y)hhGD(E) (Fig. 5, pink box). This motif is present in all members of GR₁ and in some of GR₂ (Table 1). The hydrophobic residues belong to the seventh β -strand of the FAD-binding domain, found near the C-terminus of the protein (Fig. 3A). The initial Thr residue participates in the formation of the 'greek key' motif found just before the seventh strand, and the invariant Gly and the terminal Asp residues are part of the loop connecting the β -strand to the α -helix, and hydrogen bonded to the O_{P1} or O_{P2} atoms of the pyrophosphate group.

There are some partially conserved sequence motifs in the GR family. Vallon (2000) has reported short GG and ATG motifs, present in only a few of the protein members. We found another partially conserved sequence motif, D(x)₆GxxP (Fig. 5, green box). This motif is situated at the interface between the NAD(P)H- and FAD-binding domains and one of the x residues, located between the Gly and Pro, often an Arg residue, making a polar contact with the isoalloxazine ring. This newly derived sequence motif is absent in GR family members lacking NAD(P)H-binding domain (GR₂ proteins in Table 1).

The ferredoxin reductase (FR) structural family

Structural conservation

Another well-studied FAD-family fold, represented by the enzyme ferredoxin reductase (FR) (Karplus et al. 1991), consists of six proteins listed in Table 1 (Lu et al. 1994; Ermler et al. 1995; Nishida et al. 1995; Ingelman et al. 1997; Wang et al. 1997; Sridhar Prasad et al. 1998). All family members were identified using the program CE (Shindyalov and Bourne 1998). In addition to the FAD-binding domain, all of the FR family members have the NAD(P)H-binding domain (Table 1), adopting the Rossmann fold found in the dehydrogenases (Fig. 1; Rossmann et al. 1974). However, the number of strands in the NAD(P)H-binding domain differs among the various family members. The FAD-binding fold characteristic of the FR family is a cylindrical β -domain with a flattened six-stranded antiparallel β -barrel organized into two orthogonal sheets ($\beta_1\beta_2\beta_5$ and $\beta_4\beta_3\beta_6$) separated by one α -helix, and is distinguished from the $\beta\alpha\beta\alpha\beta$ Rossmann fold (Fig. 3B). The cylinder is open between strands β_4 and β_5 . This opening of the cylinder makes space for the isoalloxazine and ribityl moieties of the FAD, to which hydrogen bonds are formed from the open edges of the strands (Fig. 4B). One end of the cylinder is covered by the only helix of the domain, which is essential for the binding of the pyrophosphate groups of the FAD. The structural differences in the FAD-binding domain are manifested mainly as loops of different length and extra extending structural elements, which may be important for interactions with their redox partners. The structural core of all FR family members is highly conserved.

FAD binding and conformation

The binding pocket for the FAD cofactor is located between the FAD- and NAD(P)H-binding domains, whose relative orientations vary among the different family members. As a result, the FAD cofactor can adopt either an elongated or bent conformation (Table 1). In contrast to this variability of the general cofactor conformation, the conformation of the FMN part of the FAD molecule is nearly identical in all family members. In particular, the isoalloxazine ring is always inserted into the protein at the FAD- and NAD(P)H-domain boundaries, whereas the adenine and ribose moieties are mostly exposed (Fig. 4B). As a result, the binding of the isoalloxazine, ribityl, and pyrophosphate moieties are well preserved in the FR family, whereas the adenosine displays large conformational differences and a small number of contacts with protein residues. This observation implies that the adenine and ribose moieties are more flexible, contributing to the variability in cofactor flexibility observed in the FR family. Furthermore, the loop between β_5 and the α -helix involved in the adenine binding differs among the various family members. Its length varies, and in

some family members it is altogether missing. This variation in loop length possibly contributes to the varied cofactor conformations.

Conserved sequence motifs

Despite their significant structural similarities, the FR sequences vary greatly, showing conserved residues at only a few crucial positions (Correll et al. 1993; Lu et al. 1994; Ingelman et al. 1997) (Fig. 6 and Table 1). One of the most conserved sequence motifs is RxYS(T) (Fig. 6, red box), which is part of β_4 (Fig. 3B), where the invariant positively

charged Arg residue forms hydrogen bonds to the negative pyrophosphate oxygen atom (O_{P1} or O_{P2}) in all family members (Ingelman et al. 1997). Another side chain involved in cofactor binding is Tyr, which makes extensive van der Waals contacts with the flavin and hydrogen bonds to the ribityl 4' hydroxyl (O_4^*). An additional highly conserved sequence motif is GxxS(T)xxL(x)₅G(x)₇PxG (Fig. 6, green box), which is part of $\alpha_1\beta_6$ and is known as the phosphate-binding motif (Fig. 3B) (Sridhar Prasad et al. 1998). In all family members, the Ser or Thr residues are part of the loop connecting β_5 to α_1 , and are within hydro-

1FDR: _	2/3	ADWVTGKVTQVQNW--T---DALFSLTVHAP---V--L--PFTAGQFTKLGLGLEIDGE-
1A8P: _	3/4	-NLNVERVLSVHHW--N---DTLFSFKTRN---PS-L--RFENGQFVMIGLEVDGR-
1NDH: _	11/12	DIKYPLRLIDKEVV--N---HDTRRFRFALPSFEHI--L--GLPVGQHIYLSARIDGN-
1CNF: _	11/12	-GRIHCLVAKKEL--S---RDVRLFRFSLSPDQV--L--GLPTGKHIFVCATIEGK-
1B2R: A	21/21	PFIGKVISNEPLVKEGG---IGIVQHIKFDL---TG-GNLKYIEGQSIGIIPPGVDKN
1AMO: A	216/280	KNPFLAAVTANRKL--NQGTERHLMHLELDIS---DSKI--RYESGDHVAVYPANDSAL
1FDR: _		-----
1A8P: _		-----
1NDH: _		-----
1CNF: _		-----
1B2R: A	72/72	GKP-----
1AMO: A	268/332	VNQIGEILGADLDVIMSLNLDDEESNKKHPFPCPTTYRTALTYLDTNPPRTNVLVELA
1FDR: _		-----
1A8P: _		-----
1NDH: _		-----
1CNF: _		-----
1B2R: A		-----
1AMO: A	328/392	QYASEPSEQEHLHKMASSSGEGKELYLSWVVEARRHILAILQDYPSLRPPIDHLCCELLPR
1FDR: _	47/48	RVQRAKSYVNSP--DNP--D---LEFYLVTV-----P-----D-----
1A8P: _	48/49	PLMRAYSIASPN--YEE--H---LEFFSIKV-----Q-----N-----
1NDH: _	60/61	LVIRPYTPVSSD--DDKGFV---DLVIKVVFKDTHPKFP-A-----G-----
1CNF: _	59/60	LCMRAYTPTSMV--DEIGH-----FDLLVKVYFKNEHPKFPN-----G-----
1B2R: A	75/75	EKLRLYSIASTR--HGD--DVDDKTISLCVRQL-----E-----YKHPESGE
1AMO: A	388/452	LQARYYSIASSSKVHPN--S---VHICAVAV-----EYEAKSGRVNK-----
1FDR: _	73/74	---GKLSPRLAALKPGD---EVQVVSEAAQFFVLD-----EVPHCETL
1A8P: _	74/75	---GPLTSRLQHLKEGD---ELMVSRRKPTGLVTS-----DLLPGKHL
1NDH: _	96/97	---GKMSQYLESMKIGD---TIEFR--GENGLLVYQGGKFAIRPDKKSSPVIKTVKSV
1CNF: _	95/96	---GLMTQYLDLSLPVGS---YIDVK--GPLGHVEYTGGRGSFVINGKQ---RNA--RRL
1B2R: A	113/113	TVYGVCSYLT--HIEPC---SEVKITGPEVKEMLL-----PDDPEANV
1AMO: A	425/489	---GVATSWLRAKEPAGENGGRALVPMFVRKSGFRLP-----FKSTTPV
1FDR: _	110/111	WMLATGTAIGFYLILRL--G-----KD-L---DRFKNLVLVHAARYA-ADLS
1A8P: _	111/112	YMLSTCTGLAPFMSLIQD--P-----EV-Y---ERFEKVVLHGVRQV-NELA
1NDH: _	147/148	GMIAGGTGITPMLQVIRAIMK-----DP-D---DHT-VCHLLFANQTE-KDIL
1CNF: _	140/141	AMICGSGGITPMYQIIQAVLR-----DQPE---DHT-EMHLVYANRTE-DDIL
1B2R: A	152/152	IMLATGTGIAPMRTYLWR--MFKDAERAANPEYQ-F---K--GFSWLVPVGPPT-PNIL
1AMO: A	466/530	IMVGPGTGIAFMGFIQE--R-----AW-LREQGKEVGETLLYYGCRSDEEDYL
1FDR: _	151/152	YLPLMQELEK-----RYEGKLRIQTVVSRET---AAGSL---TGRIPALIESGELE
1A8P: _	152/153	YQQFITEHLPQSEYFGEAVKEKLIYYPTVTRES---FH--N---QGRLTDLMRSGKLF
1NDH: _	189/190	LRPELEELRN-----EHSARFKLWYTVDRAP---AWDYS---QGFVN---EEMIRD
1CNF: _	183/184	LRDELDRWAA-----EYDRLKVWYVIDQVKRPEGWKYS---VGFTV---EAVLRE
1B2R: A	202/202	YKEELEETIQQ-----KYPDNRFLTYSISREQ---K---NPQGGRMVYQDRV--AEHA
1AMO: A	512/576	YREELARFHK-----DGA-LTQLNVAFSREQ---A---H---KVYVQHLL--KRDR

Fig. 6. Sequence alignment of all FAD-containing proteins resembling the ferredoxin reductase structure based on structure-sequence alignment as obtained using the CE program (Shindyalov and Bourne 1998). Conserved sequence motifs are depicted in boxes: RxYS(T) in red, GxxS(T)xxL(x)₅G(x)₇PxG in green, and MxxxGT(S)G(A)IxP in blue. In yellow are conserved residues not interacting with the cofactor. The data bank accession code and position numbers according to the sequence and according to the PDB are given. Proteins: 1FDR, flavodoxin reductase from *Escherichia coli*; 1A8P, ferredoxin oxidoreductase from *Azotobacter vinelandii*; 1NDH, cytochrome b₅ reductase from *Sus scrofa*; 2CND, nitrate reductase from *Zea mays*; 1AMO, NADPH-cytochrome P450 reductase from *Rattus norvegicus*; 1CQX, flavohemoprotein from *Alcaligenes eutrophus*.

gen bonding distance to one of the pyrophosphate oxygen atoms (O_{P1} or O_{P2}). The absence of side chain at the first Gly allows a close approach of the main chain to the pyrophosphate moiety. The remaining conserved residues cluster in the helix and β_6 of the FAD-binding domain.

Another conserved sequence motif, MxxxGT(S)G(A)IxP (Fig. 6, blue box), is part of the NAD(P)H-binding domain and comprises the $\beta\alpha\beta$ structure motif. The conserved Gly residue is located near the N-terminus of the sequence involved in pyrophosphate binding of the NAD(P)H cofactor, and is known as the phosphate-binding loop (Karplus et al. 1991). In addition, the Thr residue interacts with the isoalloxazine ring of the FAD cofactor in selected family members, and in the other family members it is in the vicinity of the cofactor. The remaining residues cluster in the interface between the NAD(P)H- and FAD-binding domains and around the NAD(P)H binding site. There is similarity between the GxGxxG motif observed in dehydrogenases and the central part of the phosphate-binding motif, GT(S)-G(A)IxP, present in the NAD(P)H-binding domain of FR members. Both of these motifs are rich in Gly, but the specific residues interacting with the NAD(P)H cofactor are different in the two types of domains (Ingelman et al. 1997).

The p-cresol methylhydroxylase (PCMH) structural family

Structural conservation

The third structural family of FAD-binding proteins is exemplified by *p*-cresol methylhydroxylase (PCMH); all family members are listed in Table 1 (Benson et al. 1997; Dobbek et al. 1999; Cunane et al. 2000; Dym et al. 2000; Fraaije et al. 2000). These enzymes catalyze diverse redox reactions. The FAD-binding domain consists of two $\alpha + \beta$ subdomains: one is composed of three parallel β -strands (β_{1-3}) surrounded by α -helices, and is packed against the second subdomain containing five antiparallel β -strands (β_{4-8}) surrounded by α -helices (Fig. 4C). A schematic diagram of the topological arrangement of the secondary structure of a PCMH family representative is shown in Figure 3C.

FAD binding and conformation

The binding pocket for the FAD cofactor is essentially identical for all family members. The two subdomains accommodate the FAD cofactor between them (Fig. 4C). They envelope most of the FAD cofactor and interact with it by hydrogen bonds and van der Waals contacts. In fact, the extended opening between the two subdomains in PCMH family members allows the FAD to penetrate in between them and adopt an elongated conformation (Table 1). In all family members, the adenine ring points towards the FAD-binding domain (Fig. 4C). Although the conformation of

FAD is conserved, it may be either covalently or noncovalently bound to the protein (Table 1).

Conserved sequence motifs

Although the sequence conservation of the PCMH family along the entire alignment is very small, members share some conserved residues in the N-terminal and C-terminal regions comprising the FAD-binding domain. Sequence-structure alignment is shown in Figure 7, and conserved sequence motifs are listed in Table 1. One of the most conserved and well studied residues is an Arg residue situated at the C terminus which is within hydrogen bonding distance of both the O_2 atom of the isoalloxazine ring and O_3^* of the ribitol, to provide charge compensation (Fig. 7, pink box). An additional most conserved sequence motif is P(x)₆G(A)xN (Fig. 7, blue box) in which the middle x residue is within hydrogen bonding distance of O_{P1} or O_{P2} and forms a coil segment connecting β_2 to α_1 (Fraaije et al. 1998) (Fig. 3C). This loop binds to the adenine ring and compensates for the negative charge of the FAD molecule through the formation of hydrogen bonds between backbone nitrogen atoms of Gly and some other residues belonging to the sequence motif. Another well-studied conserved sequence is G(x)₈GY (Fig. 6, green box) in which the x residues are part of α_2 and are within hydrogen bonding distance of the pyrophosphate and the isoalloxazine moieties (Fraaije et al. 1998) (Fig. 3c). Similar to the FR and PO family members, the negatively charged pyrophosphate group of the FAD is compensated by a hydrogen bond to a positive residue (Arg if it is present) or by the backbone to an amide group (in this sequence motif, a Gly residue). In contrast to some other flavoproteins (e.g., Mande et al. 1996; Mizutani et al. 1996; Sridhar Prasad et al. 1998; Yue et al. 1999) however, there is no helix dipole oriented toward the pyrophosphate moiety to help compensate for the negative charge.

Based on our sequence-structure analysis, we identified two partially conserved sequence motifs. The first is GxxL, part of β_8 , which interacts with the adenine ring (Fig. 7, orange box). The second motif we found is R(x)₆ExxYxxVxxG(x)₈Y (Fig. 7, red box), part of β_4 and β_5 with the Glu, Tyr, and Gly residues conserved. The central part of this motif, VxxG, is located near the adenine ring and is in close proximity to the GxxL and P(x)₆G(A)xN sequence motifs.

The pyruvate oxidase (PO) structural family

Structural conservation

Pyruvate oxidase (PO) was previously considered a single-member family (Muller et al. 1994). We found that electron transfer flavoprotein (Roberts et al. 1996) has a similar fold and shares several conserved sequence motifs with PO (Table 1). In both proteins, the FAD-binding domain fold is similar to the Rossmann fold of the dehydro-

1QLT:A	10/11	LTLPPKLSLSDFNFIQDIIRIVGSENVEV----	ISSKDQIVDGSYMKPTHTDHPHHVMD
1DII:A	7/8	AVLPKGVTVQGEFNKAVQKFRALLGDDNVLVESDQL-----	VPYNKIMMPVE
2MBR: _	4/7	-----	KPWNTFGI----
1QLT:A	66/67	QDYFLASAIIVAPRNADVQSIIVGLANKFSFPLWPISIGRNSGYGGAAPRVSGSVVLDMGK	
1DII:A	53/54	NAAHAPSAAVTATTVEQVQGVVKICNEHKIPITWTISTGRNFGYGSAAFPVQRQVILDLK-	
2MBR: _	12/15	--DHNAQHIVCAEDEQQLLNAWQYATAEGQFVLLILGEGSN----	VLFLIEDYRGTVIIN--
1QLT:A	126/127	NMNRVLEVNVE--GAYCVTEPGVTYHDLHNYLEANNLRDKLWLDVPLDG--GGSVLENAVE	
1DII:A	112/113	KMNKIIKIDPE--MCYALVEPGVTFGQMYDYIQENNL--PVMLSFSAPSIAIGPVGNTMD	
2MBR: _	64/67	-RIKIGIEIHDPEDAWVLLHVGAGENWHRLVKITLQE--GMPGLENLALI--PGCVSSPIQ	
1QLT:A	183/184	RGVGYTPYGDHWMHSGMEVVLANGELLRTGMGALPDPKRPETMGLKPEDQPWSKIAHLF	
1DII:A	168/169	RGVGYTPYGEHFMMQCGMEVVLANGDVYRTGMGGVPG--SNTWQIF	
2MBR: _	119/122	NIGAYG--VELQRVCAYVDSVELATGKQVRL-----	T
1QLT:A	243/244	PYGFPGPYID----GLFSQ--SNMGIVTKIGIWMNPNGGYQSYLITLPKDGDLKQAVDI	
1DII:A	212/213	KWGYGPTLD----GMFTQ--ANYGICTKMGFWLMPKPPVFKPFVIFEDADIVEIVDA	
2MBR: _	149/152	AK----ECRFGYRDSIFKHIEYQDRFAIVAGLRILPKKEWQP-----	
1QLT:A	296/297	IRPLRLGMALQNVPTIRHILLDAAVL-GDKRSYSSRTEPLSDEELDKIAQLNLGRWNFY	
1DII:A	265/266	LRPLRMSNTIPNSVVIASLTWEAGSAHLTRAQYTTEPGHTPDSPVQKQKDTGMGAWNLY	
2MBR: _	185/188	-----VLTY-----	GD
1QLT:A	355/356	GA-LYGPEPIRRVLWETIKDAFSAIPGVKFFYPEDTPENSVLRVRD----	KTMQGIPTYD
1DII:A	325/326	AA-LYGTQEQVDVNWKIVTDVFKKLKGRIVTQEEAGDTQPFKYRA----	QLMSGVPNLQ
2MBR: _	191/194	LTRLDPSTT-----	VTPQQVFNAVCHMRT
1QLT:A	410/411	ELKWI--DWLPNGAA--LFFSPIAKVSGEDAMMQ-----	YAVTKKRCQEAGLDFIGTFTV
1DII:A	380/381	EFGLY--NWRGGGGS--MWFAPVSEARGSECKKQ-----	AAMAKRVLHKYGLDYVAEFIV
2MBR: _	214/217	TKLPDPKV----NGNAGSFFKNPVVSAETAKALLSQFPTA-----	PNY
1QLT:A	461/462	GMR---EMHHIV-----	CIVFNKKD---LIQKRKVQWLMRT
1DII:A	431/432	APR---DMHHVI-----	DVLYDRTN---PEETKRADACFNE
2MBR: _	253/256	PQADGSVKLAGWLDQCQLKGMQIGGAHVHRQQALVLINEDNAKSED--VVQLAHVVRQ	
1QLT:A	491/492	LID---DCAANGWGEYRTHLA-----	FMDQIMETYNWNSSFLRFNEVLKNAVDPNGII
1DII:A	461/462	LLD---EFEKEGYAVYRVNTR-----	FQDRVAQSYG---PVKRKLEHAIKRAVDNNIL
2MBR: _	311/314	KVGEKFNWV---LEPEVRFVIGASGEVSAVET-----	
1QLT:A	542/543	APGKSGVWP	
1DII:A	509/510	APGRSGIDL	
2MBR: _		-----	

Fig. 7. The sequence alignment of FAD-containing proteins resembles the *p*-cresol methylhydroxylase structure. Conserved sequence motifs are depicted in boxes: R(x)₆ExxYxxVxxG(x)₈Y in red, P(x)₆G(A)xN in blue, GxxL in orange, R in pink, and G(x)₈GY in green. The data bank accession code and position numbers according to the sequence and according to the PDB are given. Proteins: 1DII, *p*-cresol methylhydroxylase from *Pseudomonas putida*; 1QLT, vanillyl-alcohol oxidase from *Penicillium simplicissimum*; 2MBR, uridine diphospho-N-acetylenolpyruvylglucosamine reductase from *Escherichia coli*.

genases that binds NADH (Rossmann et al. 1974) (Fig. 3D and Fig. 1, respectively). The basic structure consists of five instead of six parallel β -strands (β_{1-5}) interspersed by α -helices that lie on both sides of the five-stranded β -sheet.

FAD binding and conformation motifs

The FAD cofactor adopts an elongated conformation and lies roughly perpendicular to the core β -strands (Fig. 4D, Table 1). In contrast to NADH in dehydrogenases, the FAD cofactor in the PO structural family is bound in the opposite direction with the adenosine moiety at the second $\beta\alpha\beta\alpha\beta$ unit rather than at the first such unit. In addition, the conserved motif GxGxxG found in both the FAD-binding domain in the GR family and in NADH-binding protein in dehydrogenases is absent in the PO family. A characteristic of the FAD-binding site in the PO family members

involves internal hydrogen bonding in the FAD molecule between the ribityl 4'-hydroxyl group and N₁ of the isoalloxazine ring (Roberts et al. 1999).

Conserved sequence

Although the determination of the consensus sequence motif must await the identification of the structures of similar proteins, possible conserved motifs, based on the sequence-structure alignment, are proposed in Figure 8 and Table 1. The more conserved sequence motif KxLxxLxxxL(x)₆S(T)(x)₆GxV (Fig. 7, red box), is located in α_1 and β_2 , which are part of the $\beta\alpha\beta\alpha\beta$ Rossmann fold (Fig. 3D). The Thr or Ser are within hydrogen bonding distance of the O_{P1} or O_{P2} atoms of the pyrophosphate moiety. An additional conserved sequence motif is RxxxxGxG (Fig. 8, blue box), in which the backbone ni-

```

1POW:A 206/215 RPLIYYGICARK--AGAEIEQSKTKIPLMSYPAK--GIVADRYPAYLGSANRVAQKP
1EFV:A 198/217 KVVVSGGRGLKSGENFRLIYDADQHAAVGAHAAVDAGFVFN----DMQVGQT-----

1POW:A 262/271 ANEALAQADVVLFGNNYFF--AEVSKAFNTRYFLOIDIIPAKLGKRHKTIIVAVLADAQ
1EFV:A 249/268 GK--IVAPELYIAVGLSGAIQHL---AGMDSKTIVAENKDEAPTIFQV-ADYGVIVADLF

1POW:A 320/329 KTLAAILAQVS
1EFV:A 303/322 KVVPEMTEILK

```

Fig. 8. Structure-sequence alignment for the two members of the PO family fold as derived from the program CE (Shindyalov and Bourne 1998). Potential conserved sequence motifs are shown in boxes: KxLxxLxxxL(x)₆S(T)(x)₆GxV in red, RxxxxGxG in blue, VGxS(N) in green, and K(X)₇IXXDP(x)₉D(X)₄AD(X)₂K in pink. The data bank accession code and position numbers according to the sequence (starting from one) and according to the PDB are given. Proteins: IPOW, pyruvate oxidase from *Lactobacillus plantarum*; IEFV, electron transfer flavoprotein from *Homo sapiens*.

trogen atom from the last x residue, Arg in the case of electron transfer flavoprotein and Ile in pyruvate oxidase, interacts with the pyrophosphate moiety for charge compensation. These two conserved sequence motifs are potentially phosphate-binding motifs in the PO protein family. Another conserved motif is VGxS(N) (Fig. 8, green box), which is part of the β_3 of the second Rossmann fold, where Ser in electron transfer flavoprotein and Asn in pyruvate oxidase interact with the pyrophosphate group (Fig. 3D). It is noteworthy that although the residue hydrogen bonded to O_{P1} or O_{P2} atoms is different in the two proteins, they both align well in the sequence-structure alignment (Fig. 8). Moreover, residues in the electron transfer flavoprotein family are highly conserved (Roberts et al. 1999). These include Gly 223 and Gly 225, part of RxxxxGxG; Ser 249, part of KxLxxLxxxL(x)₆S(T)(x)₆GxV; and Ser 282, part of VGxS(N). An additional conserved motif, K(x)₇IXXDP(x)₉D(x)₄AD(x)₂K (Fig. 8, pink box), is part of $\alpha_3\beta_4$ of the second Rossmann fold, in which the first Asp residue interacts with the adenine ring (Fig. 3D).

Single-member FAD folds

There are four FAD-containing proteins (the last four proteins listed in Table 1) with different folds, all distinct from the above four family folds. Our amino acid sequence analysis revealed that none of them contains the conserved FAD-binding sequence motifs identified in the FAD families.

The FAD-binding domain fold of Acyl coenzyme A dehydrogenase (MCAD) is composed of three domains. Domains 1 and 3 are mostly α -helical and are closely associated. Domain 2 consists of a four-stranded parallel β -sheet sandwich between three-stranded antiparallel β -sheets (Tiffany et al. 1997). The FAD in the enzyme has an elongated conformation.

The structure of methylenetetrahydrofolate reductase is a $\beta_8\alpha_8$ -barrel, which resembles the structure of flavin mononucleotide (FMN)-binding proteins rather than FAD-binding proteins. However, the FAD cofactor binds to a different part of the FAD-binding domain compared to the barrels that bind FMN (Guenther et al. 1999).

In DNA photolyase, the FAD-binding domain is composed of 14 α -helices grouped into two clusters with the FAD-binding site in between (Park et al. 1995). Cluster I comprises α_7 to α_{11} and contains all residues that interact with the phosphate oxygen of the FAD. Cluster II consists of α_{12} to α_{20} , and residues within this bundle have polar interactions with the riboflavin part of the FAD. The catalytic cofactor FAD is bound to photolyase in a U-shaped conformation with the isalloxazine and adenine rings in close proximity.

The topology of the FAD-binding domain in quinone oxidoreductase is composed of a twisted central parallel β -sheet surrounded on both sides by connecting helices (Foster et al. 1999). Although at first glance it seems that this fold is reminiscent of the dehydrogenase fold or the PO family fold, the exact topology differs. The most prominent difference is the absence of the conserved FAD-binding sequence motifs identified in dehydrogenases and in the PO family (Smith et al. 1977).

Identifying phosphate-binding sites

In attempts to detect phosphate analog-binding sites, we used conserved phosphate-binding sequence motifs identified in the FAD-family folds as a query in searching the Protein Data Bank. This search was carried on using the option FINDPATTERNS in the GCG server (Williams and Gibbs 1990). Specifically, there are nine frequently occurring conserved phosphate-binding sequence motifs: GxGxxG(x)₁₇₋₁₉E(D) and T(S)(x)₅F(Y)hhGD(E) from the GR protein family, RxxS(T) and GxxS(T)xxL(x)₅G found in the FR family, P(x)₆G(A)xN and R(x)₆ExxYxxV from the PCMH family, and K(R)xLxxLxxxL, R(x)₅GxG, and K(R)(x)₇IXXDP(x)₉D from the PO family.

Forty nonFAD-containing protein structures were identified from the above query sequence motifs, each containing a phosphate-containing ligand. Approximately half of these protein structures contain a ligand closely related to the FAD cofactor (e.g., NAD(P)H, ATP, and FMN), and most sequences contain the dinucleotide-binding motif GxGxxG(x)₁₇₋₁₉E of the GR family, known to bind these

cofactors (Schulz 1992). Interestingly, two of these structures were solved in the presence of NAD(P)H and contain one of our newly derived phosphate-binding sequence motifs from the new PO family, adopting a fold similar to the Rossmann fold of dehydrogenases that bind NAD(P)H (Table 2). The structures of the remaining half (~20) of the phosphate-containing proteins were solved in the presence of ligands distantly related to FAD cofactor. Nevertheless, in only half of them is the phosphate-containing ligand within hydrogen bonding or van der Waals distance of the phosphate-binding sequence motif (Table 2). In all examples, the pyrophosphate moiety of the cofactor forms hydrogen bonds to residues from the phosphate-binding motifs. In addition, in phospholipase A2 and basic fibroblast growth factor, the phosphate-binding motif interacts with more than one phosphate-containing ligand (Table 2).

Discussion

FAD-family folds

Protein motif identification has become an essential part of sequence analysis in general and genome research in particular. FAD-containing proteins offer a useful case study, as the three-dimensional structures of 32 different flavoproteins are known to atomic resolution and their folding pattern appears much more similar than would be suggested by comparison of their sequences. Our comparative sequence-structure study of these proteins revealed four different FAD-folds as well as various conserved sequence motifs distinguished from one fold to another. Three of the family

folds, GR, FR, and PCMH, were previously defined, while PO is newly derived. Among all of the conserved sequence motifs listed in Table 1, D(x)₆GxxP in the GR family, R(x)₆ExxYxxVxxG(x)₈Y and GxxL in the PCMH family, and all four in the PO family are newly derived. In addition, there are four proteins that exhibit FAD-binding domains with folds different from the ones displayed by the four FAD families. Conserved sequence motifs identified in the FAD families are absent in these proteins.

The FAD-binding domain folds adopted by the FAD families vary. The largest family is represented by GR enzymes, and all family members adopt the $\beta\alpha\beta\alpha\beta$ fold known as the Rossmann fold. A related fold is found in the newly derived PO family members, adopting a fold similar to the double ($\beta\alpha\beta\alpha\beta$ and $\beta\alpha\beta\alpha\beta$) Rossmann fold found in dehydrogenases. Nevertheless, sequence-structure homology revealed that their sequences diverge greatly and in fact share no conserved sequence motifs. This could potentially be explained by the finding that in contrast to NADH in dehydrogenases, the FAD cofactor in the PO structural family is bound in the opposite direction with the adenosine moiety at the second $\beta\alpha\beta\alpha\beta$ unit rather than at the first. Although NADH and FAD share moieties, they are not identical and would therefore be expected to interact with different parts of the polypeptide chain. Therefore, despite the topological and functional similarities of FAD-binding domains found in PO family members to NAD(P)H-binding domains found in dehydrogenases, a divergent evolutionary relationship between these structures cannot be assumed. The FAD-binding fold adopted by the FR and PCMH family members is composed of a cylindrical β -domain and two $\alpha + \beta$ subdomains respectively. Both folds are distinct from the Rossmann fold.

Table 2. Results of a search of the Protein Data Bank for phosphate containing ligands binding to proteins, with known three-dimensional structures, containing one of the pyrophosphate-binding consensus motifs identified in the FAD-family folds of Table 1

Sequence Motif	FAD fold	PDB Code	Protein name	Phosphate-containing ligands
GxGxxG(x) ₁₇₋₁₉ E	GR	1agr	Guanine nucleotide-binding protein	guanosine-5'-diphosphate
	GR	1vip	Phospholipase A2	SO ₄
	GR	1pbo	Phospholipase A2	1-o-octyl-2-heptylphosphonyl-sn-glycerole-3-3-phosphoethanolamin
	GR	1gia	Gi α 1	5'-guanosine-diphosphate monothiophosphate
	GR	1csn	Casein Kinase-1	ATP, SO ₄
	GR	1pgn	6-phosphogluconate dehydrogenase	pyrophosphate 2
	GR	2cae	Catalase	S-dioxynehyonine
RxxxxxGxG	PO	1kvr	Udp-galactose 4-epimerase	uridine-5'-diphosphate, NADH
K(x) ₇ LxxD(x) ₁₀ D	PO	1bfb	Basic Fibroblast Growth Factor	N,O6-disulfoglucosamine, 1,4-dideoxy-o2-sulfo-glucuronic acid, 1,4-dideoxy-5-dehydro-o2-sulfo-glucuronic acid, SO ₄
	PO	1gts	Glutamyl-tRNA synthetase	AMP, ATP
	PO	1ad3	Aldehyde dehydrogenase	NADH
KxLxxLxxxL	PO	1tpu	Triosephosphate isomerase	Phosphoglycolohydroxamic acid
	PO	1cwn	Aldehyde reductase	NADPH
GxSxxL(x) ₃ G	FD	1ajr	Aspartate transaminase	2-lysine(3-hydroxy-2-2-methyl-5 phosphonooxymethyl-pyridin-4 ylmethane)

Atomic interactions of cofactor with proteins

For each FAD family we analyzed the correlation between the fold and various cofactor properties. Although some of the FAD properties are conserved among proteins belonging to the same FAD-family fold, others are not. One of the least conserved properties is the bond type (covalent versus noncovalent). When there is covalent interaction, FAD is bound to protein residues through atoms from the isoalloxazine ring. This is in agreement with the observation that the isoalloxazine ring interacts with residues located in the poorly conserved regions along the polypeptide sequence.

Another less conserved cofactor property is the number of protein residues-cofactor atoms interactions. Whereas all proteins studied have at least one contact with the pyrophosphate moiety (O_{P1} or O_{P2} atoms), the number of contacts varies from one protein to another and there is no strong preference for a particular number of contacts in the different families. This observation is reinforced by the variety of conserved sequence motifs present in the different family folds. Moreover, some members of a FAD-fold family contain all conserved sequence motifs whereas others contain only some.

A partially conserved property is the geometry of the FAD, which is controlled by the surrounding protein as either an elongated or bent-butterfly conformation. The cofactor conformation in most of the FAD-family folds is elongated with slight flexibility that varies from one protein to another. In the FR family, the cofactor adopts either an elongated or bent conformation (Table 1). This variation can be explained by the observation that although the general fold of the individual FAD- and NAD(P)H-binding domains in the FR family is invariant, their relative orientation differs among the family members. As a result the FAD, situated at the junction between the two domains, can adopt either an elongated or bent conformation depending on the proximity of the two domains. In contrast, in the PCMH family, the FAD-binding domain is composed of two subdomains distant from each other, which accommodate the FAD in between them. In fact the FAD holds the two subdomains together and therefore can adopt only an elongated conformation. Similarly in the PO family, the FAD molecule is situated on top of the core β -strands and adopts an elongated conformation for maximal interaction with the FAD-binding domain. Therefore, it seems that the fold of the FAD-binding domain and the cofactor conformation are not independent, because the fold determines the position and shape of the pocket on which the cofactor conformation depends. This variability in the cofactor conformation and shape of cofactor pocket in the FAD-binding domain of the different FAD-family folds implies that there is no single protein 'pharmacophore' for binding FAD.

One of the most conserved properties of the FAD is the direction in which it approaches the FAD-binding domain. In all FR family members, the isoalloxazine ring points toward the FAD-binding domain, whereas in the rest of the FAD-family folds it is the adenosine moiety that does so.

Cofactor atoms interacting with conserved sequence motifs

Another characteristic of the cofactor is evident from our analysis of the correlation between FAD atoms interacting with the conserved sequence motifs identified for each of the families. These conserved sequence motifs are distinctive in the four FAD families. Nevertheless, residues from the most conserved sequence motifs in a specific family fold make hydrophobic and/or hydrophilic contacts with cofactor atoms. In contrast, residues from partially conserved sequence motifs are distant ($>4.0\text{\AA}$) from cofactor atoms and therefore probably form at best weak interactions with the cofactor. Residues involved in binding oxygen atoms (O_{P1} and O_{P2}) of the pyrophosphate moiety are more highly conserved relative to the alignment as a whole, and in fact are part of the most conserved sequence motifs. In contrast, residues involved in binding the adenine, ribose, ribitol, and isoalloxazine moieties are part of less conserved regions. Because the pyrophosphate moiety is negatively charged, it is expected to bind to regions in the protein with positive charges. This can be achieved by interacting with positively charged amino acids, peptide groups, or α -helix dipoles. Notably, when there is no helix to stabilize the negative charge of the pyrophosphate group, Arg is the most common residue and is capable of forming multiple hydrogen bonds with the pyrophosphate moiety. When, however, there is an α -helix dipole, as observed in most of the GR family members, Gly is the most common amino acid, and its main chain is within hydrogen bonding distance of the pyrophosphate group.

In contrast to the conserved pyrophosphate binding, the residues involved in binding the isoalloxazine and adenosine moieties are usually part of the partially conserved sequence motifs. This is consistent with the fact that the catalytic function of FAD is restricted to the isoalloxazine ring, and with the variety of family members involved in diverse redox reactions. Consequently, diverse residues from the different proteins are involved in catalytic function and contact the isoalloxazine ring. Similarly, the function of the adenosine moiety is mainly to stabilize cofactor binding through interaction with the protein. This stability can be achieved by interaction with different residues in the different proteins belonging to the same FAD-family fold.

The variety of redox functions may also explain the variability in cofactor conformation observed in the FAD families. As mentioned above, the flexibility is achieved by movement of the adenine ring toward or away from the isoalloxazine ring. As a result, the two rings can interact

with different parts of the protein, usually in less conserved regions. We therefore propose that the pyrophosphate moiety anchors the FAD-binding domain by binding to the most conserved sequence motif, with less specific binding of the isoalloxazine and adenine rings to less conserved sequence motifs. This is consistent with the observation that some of the most conserved sequence motifs described above are known to bind ATP, FMN, and GTP as well. They all share a common pyrophosphate group. This observation further implies that it is possible to predict the pyrophosphate-binding site by identifying the most conserved sequence motif in a multiple alignment of other prosthetic groups containing pyrophosphate moieties such as AMP, FMN, and GTP. It is conceivable to assume that inspection of the residues interacting with the O_{P1} or O_{P2} atoms can be used as a tool for identifying the most conserved sequence motif in the FAD-containing proteins adopting a unique fold, as for the four single-member (SM) protein families of Table 1.

These conserved phosphate-binding sequence motifs have a predictive use: they can be applied to searches for phosphate analog-containing proteins. In fact, we identified a dozen nonFAD-containing proteins that bind phosphate-containing ligands as cofactors (Table 2). Each contains one of the phosphate-binding motifs assigned for the FAD families. These motifs are within hydrogen bond distance of the phosphate moiety of the phosphate-containing ligand. Interestingly, three of the phosphate analog ligands were actually sulphate ions. This implies that conserved sulfate- and sulphate analog-binding sequence motifs could be considered potential binding sites for phosphate ions.

Conclusion

The FAD-dependent enzymes illustrate how similar cofactors are utilized by nature in a wide variety of protein families. The availability of the three-dimensional structures of a large collection of FAD-containing proteins provides an attractive opportunity for sequence-function-structure analysis. Four different FAD-family folds, each with distinctive conserved sequence motifs were identified. Furthermore, the FAD pocket shape can be distinguished from one fold to another as well, implying that there is no unique 'pharmacophore' for binding FAD. In contrast to the diversity of the FAD fold and its sequence motifs, the pyrophosphate moiety makes hydrogen bonds with residues from the most conserved sequence motifs in each fold family. Therefore, we propose that the pyrophosphate moiety is crucial for molecular recognition, whereas the isoalloxazine ring is involved in catalytic function, and the adenine ring stabilizes cofactor binding, both rings interacting with protein residues in partially conserved sequence motifs. As a result, the isoalloxazine and adenine rings interact with different residues in the different proteins belonging to the same FAD-family fold. This is consistent with the variability in

bond type (covalent versus noncovalent) and cofactor conformation (elongated versus bent) observed in the FAD-family folds. Nonetheless, in each FAD-family fold there exists a clear correlation between the fold, the shape and position of the pocket, and the cofactor conformation. This is in agreement with the finding that the cofactor exhibits directionality, which is highly conserved within a family fold.

An unexpected outcome of our analysis is that the conserved pyrophosphate-binding sequence motifs that were identified in FAD-dependent proteins may be applied to discover phosphate-binding motifs in sequences of non-FAD-containing proteins. Where the structures of these proteins are known, the identified motifs are found to bind to phosphate- or sulfate-containing ligands. Building on this result, the sequence motifs found to bind to pyrophosphate in four single-membered FAD protein families (last four entries in Table 1) may serve to identify new members of these families.

Materials and methods

Sequences and three-dimensional structures of all flavoproteins were retrieved from the Protein Data Bank (www.rcsb.org/pdb). A total of 150 flavoprotein X-ray structures solved in the presence of FAD were available from the October 2000 release, and consist of 32 nonredundant proteins. Table 1 lists the proteins studied, data bank accession codes, and references to one of the structural papers on each protein. Homology searching for identifying three-dimensional similarities in protein structure was performed using the program Combinatorial Extension (CE) (<http://cl.sdsc.edu/ce.html>). The program aligns two or more polypeptide chains using characteristics of their local geometry as defined by vectors between $C\alpha$ positions (Shindyalov and Bourne 1998). Each of the 32 proteins was used as a query in the sequence-structure search, resulting in the identification of four FAD-family folds. Based on this sequence-structure alignment, conserved sequence motifs unique for each of the FAD families were identified (Table 1). Next, protein residues forming van der Waals interactions and hydrogen bond contacts with the cofactor were analyzed using the program LigPlot (Wallace et al. 1995). We then correlated these protein residues with residues from the most conserved sequence motif. Finally, each of the conserved sequence motifs was used as query in a search of the Protein Data Bank for nonFAD-containing proteins solved in the presence of phosphate-containing ligands. This search was aided by the option FINDPATTERNS on the Genetic Computer Group (GCG) server (Williams and Gibbs 1990).

Acknowledgments

We thank Dr. I. Xenarios for discussion and NIH and DOE for support.

References

- Bacon, D.J. and Anderson, W.F. 1988. A fast algorithm for rendering space-filling molecule pictures. *J. Mol. Graph.* 6: 219-220.
- Benson, T.E., Walsh, C.T., and Hogle, J.M. 1997. X-ray crystal structures of the

- S229A mutant and wild-type MurB in the presence of the substrate enol-pyruvyl-UDP-N-acetylglucosamine at 1.8-Å resolution. *Biochemistry* **36**: 806–811.
- Binda, C., Coda, A., Angelini, R., Federico, R., Ascenzi, P., and Mattevi, A. 1999. A 30-angstrom-long U-shaped catalytic tunnel in the crystal structure of polyamine oxidase. *Structure Fold. Des.* **7**: 265–276.
- Bond, C.S., Zhang, Y., Berriman, M., Cunningham, M.L., Fairlamb, A.H., and Hunter, W.N. 1999. Crystal structure of *Trypanosoma cruzi* trypanothione reductase in complex with trypanothione, and the structure-based discovery of new natural product inhibitors. *Structure Fold. Des.* **7**: 81–89.
- Chen, Z.W., Koh, M., Van Driessche, G., Van Beeumen, J.J., Bartsch, R.G., Meyer, T.E., Cusanovich, M.A., and Mathews, F.S. 1994. The structure of flavocytochrome c sulfide dehydrogenase from a purple phototrophic bacterium. *Science* **266**: 430–432.
- Correll, C.C., Ludwig, M.L., Bruns, C.M., and Karplus, P.A. 1993. Structural prototypes for an extended family of flavoprotein reductases: Comparison of phthalate dioxygenase reductase with ferredoxin reductase and ferredoxin. *Protein Sci.* **2**: 2112–2133.
- Cunane, L.M., Chen, Z.W., Shamala, N., Mathews, F.S., Cronin, C.N., and McIntire, W.S. 2000. Structures of the flavocytochrome p-cresol methylhydroxylase and its enzyme-substrate complex: Gated substrate entry and proton relays support the proposed catalytic mechanism. *J. Mol. Biol.* **295**: 357–374.
- Dobbek, H., Gremer, L., Meyer, O., and Huber, R. 1999. Crystal structure and mechanism of CO dehydrogenase, a molybdo iron-sulfur flavoprotein containing S-sclanylcysteine. *Proc. Natl. Acad. Sci.* **96**: 8884–8889.
- Doolittle, R.F. 1986. *Of URFs and ORFs: A primer on how to analyse derived amino acid sequences*, University Science Books, Mill Valley, CA.
- Dym, O., Pratt, E.A., Ho, C., and Eisenberg, D. 2000. The crystal structure of D-lactate dehydrogenase, a peripheral membrane respiratory enzyme. *Proc. Natl. Acad. Sci.* **97**: 9413–9418.
- Eggink, G., Engel, H., Vriend, G., Terpstra, P., and Witholt, B. 1990. Rubredoxin reductase of *Pseudomonas oleovorans*. Structural relationship to other flavoprotein oxidoreductases based on one NAD and two FAD fingerprints. *J. Mol. Biol.* **212**: 135–142.
- Enroth, C., Neujahr, H., Schneider, G., and Lindqvist, Y. 1998. The crystal structure of phenol hydroxylase in complex with FAD and phenol provides evidence for a concerted conformational change in the enzyme and its cofactor during catalysis. *Structure* **6**: 605–617.
- Eppink, M.H., Schreuder, H.A., and van Berkel, W.J. 1998. Lys42 and Ser42 variants of p-hydroxybenzoate hydroxylase from *Pseudomonas fluorescens* reveal that Arg42 is essential for NADPH binding. *Eur. J. Biochem.* **253**: 194–201.
- Ermier, U., Siddiqui, R.A., Cramm, R., and Friedrich, B. 1995. Crystal structure of the flavohemoglobin from *Alcaligenes eutrophus* at 1.75 Å resolution. *EMBO J.* **14**: 6067–6077.
- Foster, C.E., Bianchet, M.A., Talalay, P., Zhao, Q., and Amzel, L.M. 1999. Crystal structure of human quinone reductase type 2, a metalloflavoprotein. *Biochemistry* **38**: 9881–9886.
- Fraaije, M.W., Van Berkel, W.J., Benen, J.A., Visser, J., and Mattevi, A. 1998. A novel oxidoreductase family sharing a conserved FAD-binding domain [published erratum appears in *Trends Biochem. Sci.* 1998 Jul;23(7):271]. *Trends Biochem. Sci.* **23**: 206–207.
- Fraaije, M.W., van Den Heuvel, R.H., van Berkel, W.J., and Mattevi, A. 2000. Structural analysis of flavinylation in vanillyl-alcohol oxidase. *J. Biol. Chem.* **275**: 38654–38658.
- Guenther, B.D., Sheppard, C.A., Tran, P., Rozen, R., Mathews, R.G., and Ludwig, M.L. 1999. The structure and properties of methylenetetrahydrofolate reductase from *Escherichia coli* suggest how folate ameliorates human hyperhomocysteinemia. *Nat. Struct. Biol.* **6**: 359–365.
- Ingelman, M., Bianchi, V., and Eklund, H. 1997. The three-dimensional structure of flavodoxin reductase from *Escherichia coli* at 1.7 Å resolution. *J. Mol. Biol.* **268**: 147–157.
- Karplus, P.A., Daniels, M.J., and Herriott, J.R. 1991. Atomic structure of ferredoxin-NADP+ reductase: Prototype for a structurally novel flavoenzyme family. *Science* **251**: 60–66.
- Karplus, P.A. and Schulz, G.E. 1987. Refined structure of glutathione reductase at 1.54 Å resolution. *J. Mol. Biol.* **195**: 701–729.
- Kraulis, P.J. 1991. MOLSCRIPT: A program to produce both detailed and schematic plots of protein structures. *J. Appl. Cryst.* **24**: 946–950.
- Lennon, B.W., Williams, C.H., Jr., and Ludwig, M.L. 1999. Crystal structure of reduced thioredoxin reductase from *Escherichia coli*: Structural flexibility in the isoalloxazine ring of the flavin adenine dinucleotide cofactor. *Protein Sci.* **8**: 2366–2379.
- Leys, D., Tsapin, A.S., Nealson, K.H., Meyer, T.E., Cusanovich, M.A., and Van Beeumen, J.J. 1999. Structure and mechanism of the flavocytochrome c fumarate reductase of *Shewanella putrefaciens* MR-1. *Nat. Struct. Biol.* **6**: 1113–1117.
- Lu, G., Campbell, W.H., Schneider, G., and Lindqvist, Y. 1994. Crystal structure of the FAD-containing fragment of corn nitrate reductase at 2.5 Å resolution: Relationship to other flavoprotein reductases. *Structure* **2**: 809–821.
- Mande, S.S., Sarfaty, S., Allen, M.D., Perham, R.N., and Hol, W.G. 1996. Protein-protein interactions in the pyruvate dehydrogenase multienzyme complex: Dihydrolipoamide dehydrogenase complexed with the binding domain of dihydrolipoamide acetyltransferase. *Structure* **4**: 277–286.
- Mathews, F.S. 1991. New flavoenzymes. *Curr. Opin. Struct. Biol.* **1**: 954–967.
- Merritt, E.A. and Murphy, M.E.P. 1994. Raster 3D Version 2.0 — A program for photorealistic molecular graphics. *Acta. Crystallogr. D. Biol. Crystallogr.* **50**: 869–873.
- Mizutani, H., Miyahara, I., Hirotsu, K., Nishina, Y., Shiga, K., Setoyama, C., and Miura, R. 1996. Three-dimensional structure of porcine kidney D-amino acid oxidase at 3.0 Å resolution. *J. Biochem. (Tokyo)* **120**: 14–17.
- Moller, W. and Amons, R. 1985. Phosphate-binding sequences in nucleotide-binding proteins. *FEBS Lett.* **186**: 1–7.
- Muller, Y.A., Schumacher, G., Rudolph, R., and Schulz, G.E. 1994. The refined structures of a stabilized mutant and of wild-type pyruvate oxidase from *Lactobacillus plantarum*. *J. Mol. Biol.* **237**: 315–335.
- Nishida, H., Inaka, K., Yamanaka, M., Kaida, S., Kobayashi, K., and Miki, K. 1995. Crystal structure of NADH-cytochrome b5 reductase from pig liver at 2.4 Å resolution. *Biochemistry* **34**: 2763–2767.
- Park, H.W., Kim, S.T., Sancar, A., and Deisenhofer, J. 1995. Crystal structure of DNA photolyase from *Escherichia coli* [see comments]. *Science* **268**: 1866–1872.
- Rao, S.T., and Rossmann, M.G. 1973. Comparison of super-secondary structures in proteins. *J. Mol. Biol.* **76**: 241–256.
- Roberts, D.L., Fremman, F.E., and Kim, J.J. 1996. Three-dimensional structure of human electron transfer flavoprotein to 2.1-Å resolution. *Proc. Natl. Acad. Sci.* **93**: 14355–14360.
- Roberts, D.L., Salazar, D., Fulmer, J.P., Fremman, F.E., and Kim, J.J. 1999. Crystal structure of *Paracoccus denitrificans* electron transfer flavoprotein: Structural and electrostatic analysis of a conserved flavin binding domain. *Biochemistry* **38**: 1977–1989.
- Rossmann, M.G., Moras, D., and Olsen, K.W. 1974. Chemical and biological evolution of nucleotide-binding protein. *Nature* **250**: 194–199.
- Schulz, G.E. 1992. Binding of nucleotides by proteins. *Curr. Opin. Struct. Biol.* **2**: 61–67.
- Schulz, G.E. and Schirmer, R.H. 1974. Topological comparison of adenylyl kinase with other proteins. *Nature* **250**: 142–144.
- Schulz, G.E., Schirmer, R.H., and Pai, E.F. 1982. FAD-binding site of glutathione reductase. *J. Mol. Biol.* **160**: 287–308.
- Shindyalov, I.N. and Bourne, P.E. 1998. Protein structure alignment by incremental combinatorial extension (CE) of the optimal path. *Protein Eng.* **11**: 739–747.
- Smith, W.W., Burnett, R.M., Darling, G.D., and Ludwig, M.L. 1977. Structure of the semiquinone form of flavodoxin from *Clostridium MP*. Extension of 1.8 Å resolution and some comparisons with the oxidized state. *J. Mol. Biol.* **117**: 195–225.
- Sridhar Prasad, G., Kresge, N., Muhlberg, A.B., Shaw, A., Jung, Y.S., Burgess, B.K., and Stout, C.D. 1998. The crystal structure of NADPH: Ferredoxin reductase from *Azotobacter vinelandii*. *Protein Sci.* **7**: 2541–2549.
- Tiffany, K.A., Roberts, D.L., Wang, M., Paschke, R., Mohsen, A.W., Vockley, J., and Kim, J.J. 1997. Structure of human isovaleryl-CoA dehydrogenase at 2.6 Å resolution: Structural basis for substrate specificity. *Biochemistry* **36**: 8455–8464.
- Trickey, P., Wagner, M.A., Jorns, M.S., and Mathews, F.S. 1999. Monomeric sarcosine oxidase: Structure of a covalently flavinylated amine oxidizing enzyme. *Structure Fold. Des.* **7**: 331–345.
- Vallon, O. 2000. New sequence motifs in flavoproteins: Evidence for common ancestry and tools to predict structure. *Proteins* **38**: 95–114.
- Van Driessche, G., Koh, M., Chen, Z.W., Mathews, F.S., Meyer, T.E., Bartsch, R.G., Cusanovich, M.A., and Van Beeumen, J.J. 1996. Covalent structure of the flavoprotein subunit of the flavocytochrome c: Sulfide dehydrogenase from the purple phototrophic bacterium *Chromatium vinosum*. *Protein Sci.* **5**: 2753–2764.
- Wallace, A.C., Laskowski, R.A., and Thornton, J.M. 1995. LIGPLOT: A program to generate schematic diagrams of protein-ligand interactions. *Protein Eng.* **8**: 127–134.
- Wang, M., Roberts, D.L., Paschke, R., Shea, T.M., Masters, B.S., and Kim, J.J. 1997. Three-dimensional structure of NADPH-cytochrome P450 reductase: Prototype for FMN- and FAD-containing enzymes. *Proc. Natl. Acad. Sci.* **94**: 8411–8416.

- Wierenga, R.K., Drenth, J., and Schulz, G.E. 1983. Comparison of the three-dimensional protein and nucleotide structure of the FAD-binding domain of p-hydroxybenzoate hydroxylase with the FAD- as well as NADPH-binding domains of glutathione reductase. *J. Mol. Biol.* **167**: 725-739.
- Wierenga, R.K., Terpstra, P., and Hol, W.G. 1986. Prediction of the occurrence of the ADP-binding beta alpha beta-fold in proteins, using an amino acid sequence fingerprint. *J. Mol. Biol.* **187**: 101-107.
- Williams, G.W., and Gibbs, G.P. 1990. Automatic updating of the EMBL database via EMBNet. *Comput. Appl. Biosci.* **6**: 122-123.
- Wohlfahrt, G., Witt, S., Hendle, J., Schomburg, D., Kalisz, H.M., and Hecht, H.J. 1999. 1.8 and 1.9 Å resolution structures of the *Penicillium amagasakiense* and *Aspergillus niger* glucose oxidases as a basis for modelling substrate complexes. *Acta Crystallogr. D. Biol. Crystallogr.* **55**: 969-977.
- Yeh, J.I., Claiborne, A., and Hol, W.G. 1996. Structure of the native cysteine-sulfenic acid redox center of enterococcal NADH peroxidase refined at 2.8 Å resolution. *Biochemistry* **35**: 9951-9957.
- Yue, Q.K., Kass, I.J., Sampson, N.S., and Vrielink, A. 1999. Crystal structure determination of cholesterol oxidase from *Streptomyces* and structural characterization of key active site mutants. *Biochemistry* **38**: 4277-4286.
- Ziegler, G.A., Vönnhein, C., Hanukoglu, I., and Schulz, G.E. 1999. The structure of adrenodoxin reductase of mitochondrial P450 systems: Electron transfer for steroid biosynthesis. *J. Mol. Biol.* **289**: 981-990.