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3D-JURY model of
SARS methyltransferase

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Cover: 3D-Jury model of the C-terminal RNA methyltransferase module of the large ORF1ab protein of the coronavirus isolated from patients suffering from SARS (Severe Acute Respiratory Syndrome). This kind of structure assignment is currently not feasible using standard sequence comparison methods.

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S, Supplementary Material available at NAR Online

EDITORIAL

Ten years ago *Nucleic Acids Research* published a special issue devoted to factual databases in the biological sciences. The Database Issue, which appears as the first issue of each year, has now become a regular feature and provides an essential resource for scientists interested in locating databases that encapsulate specific biological knowledge in a computer-accessible form. More recently, many web servers have appeared that perform useful computations on DNA, RNA and protein sequences and structures. Between them they provide an unparalleled array of useful computational services. This new special Web Server Issue is devoted to descriptions of these internet resources and aims to provide both a single source where authors of these web servers can highlight their offerings and where readers can find out what is available. It is the natural companion to the Database Issue and one that will be repeated in 2004.

In the current issue there are reports of 131 web servers that run the gamut from simple pattern prediction to three-dimensional protein structure prediction. The servers described have all been subjected to rigorous peer review, are available free-of-charge and provide invaluable resources to the scientific community. The scientists and programmers who have provided these resources deserve our immense thanks. They illustrate the very best of the scientific spirit that transcends national boundaries and promotes the cooperation and sharing of resources.

Authors interested in submitting articles for the 2004 Web Server Issue should contact Dr R. J. Roberts (roberts@neb.com) before sending in their articles, preferably before December 31, 2003. The deadline for submission of articles will be February 15, 2004. Prospective authors for papers in either the Database Issue or the Web Server Issue should consult the NAR web site (www.nar.oupjournals.org) for further details and submission instructions. Note that the 2004 Database Issue will be edited by Dr Alex Bateman, who should be contacted at nardatabase@mrc-lmb.cam.ac.uk, no later than July 1 by all authors considering new submissions to that issue.

SIFT: predicting amino acid changes that affect protein function

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ABSTRACT

Single nucleotide polymorphism (SNP) studies and random mutagenesis projects identify amino acid substitutions in protein-coding regions. Each substitution has the potential to affect protein function. SIFT (Sorting Intolerant From Tolerant) is a program that predicts whether an amino acid substitution affects protein function so that users can prioritize substitutions for further study. We have shown that SIFT can distinguish between functionally neutral and deleterious amino acid changes in mutagenesis studies and on human polymorphisms. SIFT is available at <http://blocks.fhcrc.org/sift/SIFT.html>.

INTRODUCTION

Single nucleotide polymorphisms (SNPs) are used as markers in linkage and association studies to detect which regions in the human genome may be involved in disease. SNPs in coding and regulatory regions may be implicated in disease themselves. Non-synonymous SNPs that lead to an amino acid change in the protein product are of major interest, because amino acid substitutions currently account for approximately half of the known gene lesions responsible for human inherited disease (1). SIFT (Sorting Intolerant From Tolerant) uses sequence homology to predict whether an amino acid substitution will affect protein function and hence, potentially alter phenotype (2,3).

SIFT has been applied to human variant databases and was able to distinguish mutations involved in disease from neutral polymorphisms (3). Assuming that disease-causing amino acid substitutions are damaging to protein function, we applied SIFT to a database of missense substitutions associated with or involved in disease (4). SIFT predicted 69% to be damaging. When SIFT was applied to the non-synonymous SNPs in dbSNP (5), a database of putative SNPs, 25% of the variants were predicted to be deleterious. This was similar to SIFT's 20% false positive error which suggested that most non-synonymous SNPs are functionally neutral. Furthermore, a subset of the variants from dbSNP predicted to affect function were involved in disease which confirmed SIFT sensitivity.

The SIFT algorithm relies solely on sequence for prediction, yet performs similarly to tools that use structure (3,6-8). An

advantage of not requiring structure is that a larger number of substitutions can be predicted on. Of the non-synonymous SNPs identified by the SNP Consortium, 74% were sufficiently similar to homologs in protein sequence databases for SIFT prediction. The number of substitutions that SIFT can predict on is expected to increase as more genomes are sequenced and more protein sequences become available.

SIFT PREDICTION METHOD

SIFT presumes that important amino acids will be conserved in the protein family, and so changes at well-conserved positions tend to be predicted as deleterious. For example, if a position in an alignment of a protein family only contains the amino acid isoleucine, it is presumed that substitution to any other amino acid is selected against and that isoleucine is necessary for protein function. Therefore, a change to any other amino acid will be predicted to be deleterious to protein function. If a position in an alignment contains the hydrophobic amino acids isoleucine, valine and leucine, then SIFT assumes, in effect, that this position can only contain amino acids with hydrophobic character. At this position, changes to other hydrophobic amino acids are usually predicted to be tolerated but changes to other residues (such as charged or polar) will be predicted to affect protein function.

To predict whether an amino acid substitution in a protein will affect protein function, SIFT considers the position at which the change occurred and the type of amino acid change. Given a protein sequence, SIFT chooses related proteins and obtains an alignment of these proteins with the query. Based on the amino acids appearing at each position in the alignment, SIFT calculates the probability that an amino acid at a position is tolerated conditional on the most frequent amino acid being tolerated. If this normalized value is less than a cutoff, the substitution is predicted to be deleterious (2). The SIFT algorithm and software have been described previously (2,3).

SIFT WEBSITE

Input

Users can obtain predictions for amino acid changes of interest at <http://www.blocks.fhcrc.org/sift/SIFT.html>. From this page,

Substitution at pos 1426 from S to P is predicted to AFFECT PROTEIN FUNCTION with a score of 0.02.
Median sequence conservation: 2.90
Sequences represented at this position:26

Substitution at pos 1432 from N to K is predicted to be TOLERATED with a score of 0.08.
Median sequence conservation: 2.90
Sequences represented at this position:26

Substitution at pos 1445 from D to N is predicted to AFFECT PROTEIN FUNCTION with a score of 0.01.
Median sequence conservation: 3.66
Sequences represented at this position:21

WARNING: This substitution may have been predicted to affect function just because the sequences used were not diverse enough. There is LOW CONFIDENCE in this prediction.

Figure 1. An example of SIFT prediction on amino acid changes in a protein. Substitutions with score less than 0.05 are predicted to affect protein function. In the last prediction, the median conservation of the sequences does not meet the threshold so a warning is issued.

there are links to three submission pages which allow users different levels of involvement in order to control the quality of their predictions.

For minimal involvement, users can simply submit their protein sequences and amino acid substitutions. In its fully automated mode, SIFT will search for protein sequences homologous to the query protein and based on these sequences, calculate probabilities for each possible amino acid change. Users can select from among SWISS-PROT, SWISS-PROT/TrEMBL, or NCBI's non-redundant protein databases for SIFT to search (4,9).

Although SIFT can choose sequences automatically, better prediction results may be obtained when all of the sequences that are provided are orthologous to the query protein. This is because inclusion of paralogous sequences confounds prediction at residues conserved only among the orthologues. If a user already has sequences that are thought to be functionally similar to the protein of interest, these sequences can be directly submitted and SIFT's step for choosing sequences skipped. Given the query protein and homologous sequences, SIFT obtains the alignment.

If regions are misaligned, SIFT will not recognize conserved positions and therefore miss potentially damaging substitutions. For best prediction quality, a third mode of operation allows users to submit their own alignments.

Output

Predictions are given for all 20 possible amino acid changes at each position in the protein. The alignment is also returned so that users can examine the sequences used for prediction and modify them for resubmission. This option is also useful for removing uncertain, erroneous and misaligned sequences from alignment output generated by SIFT in its automatic mode.

For amino acid substitutions submitted by the user, a more detailed synopsis is provided (Fig. 1). The score is the normalized probability that the amino acid change is tolerated. SIFT predicts substitutions with scores less than 0.05 as deleterious. Some SIFT users have found that substitutions with scores less than 0.1 provide better sensitivity for detecting deleterious SNPs (Cornelia Ulrich, personal communication

and 10). The quantitative score allows users to prioritize their amino acid changes by ranking them from the lowest scores to the highest.

Confidence in a substitution predicted to be deleterious depends on the diversity of the sequences in the alignment. If the sequences used for prediction are closely related, then many positions will appear conserved and SIFT will predict most substitutions to affect protein function. This leads to a high false positive error where functionally neutral substitutions are predicted to be deleterious.

To alert the user to these situations, SIFT calculates the median conservation value which measures the diversity of the sequences in the alignment. Conservation, as measured by information content (11), is calculated for each position in the alignment and the median of these values is obtained. Conservation ranges from $\log_2 20$ ($= 4.32$), when a position is completely conserved and only one amino acid is observed, to zero, when all 20 amino acids are observed at a position. By default, SIFT builds alignments with a median conservation value of 3.0. Predictions based on sequence alignments with higher median conservation values are less diverse and will have a higher false positive error (Fig. 2).

Even if there are few homologous sequences available, SIFT performs better than simply predicting non-conservative amino acid substitutions as deleterious, where non-conservative changes are defined as having negative scores in an amino acid substitution scoring matrix. We have shown that with only one sequence homologous to the test protein, SIFT can predict at twice as many neutral substitutions correctly compared to a substitution scoring matrix (2). Even with few homologous sequences, there will be positions that differ between the test sequences and the other sequences. Depending on the amino acids appearing at these positions, SIFT may predict these positions to be unimportant for protein function. This additional information can eliminate functionally neutral substitutions and increase selectivity to deleterious substitutions.

In summary, a large number of substitutions can be obtained from mutagenesis projects, SNP datasets, and changes between closely related organisms. When it is not feasible to conduct experiments on all substitutions, SIFT and other similar prediction tools (13) may be useful in prioritizing which

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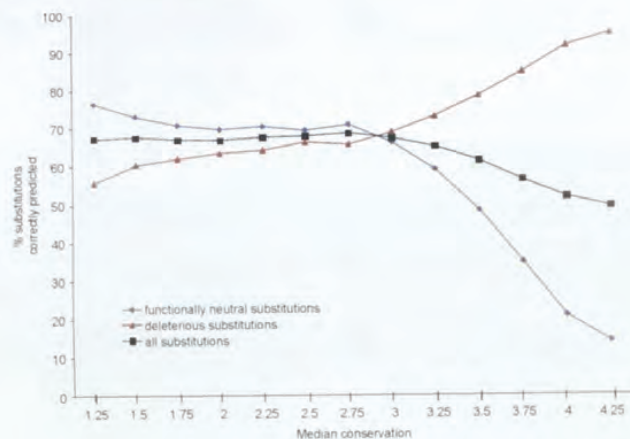


Figure 2. Prediction depends on the diversity of the sequences used in the alignment. Percentage of substitutions correctly predicted is based on (over 4000) substitutions that were assayed throughout the LacI protein of *Escherichia coli* (2,12). When the sequences in the alignment used for prediction are closely related (high median conservation) then many positions appear conserved and important for function. In this situation, prediction accuracy on deleterious substitutions is high but many functionally neutral substitutions are erroneously predicted to be deleterious. To obtain an alignment with a predicted median conservation, the LacI protein sequence of *E. coli* was submitted to the SIFT website and the median conservation setting adjusted. Because the homologous sequences available are distantly related to *E. coli* LacI, alignments with higher median conservation values could not be obtained. In order to obtain alignments with median conservation values more than 3.25, closely related sequences were simulated by starting with an alignment of identical *E. coli* LacI sequences. A position and a sequence were randomly selected from the LacI alignment with median conservation 2.75. The amino acid corresponding to this location was substituted in the starting alignment. Amino acids continued to be randomly selected and substituted until the desired median conservation was met. The simulated alignment was then evaluated for its performance as previously described (2) and the plotted value is the average performance of 100 simulated alignments.

changes affect protein function and may contribute to phenotypic differences.

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ActionMap: a web-based software that automates loci assignments to framework maps

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ABSTRACT

Genetic linkage map computation may be a repetitive and time consuming task, especially when numerous loci are assigned to a framework map. We thus developed ActionMap, a web-based software that automates genetic mapping on a fixed framework map without adding the new markers to the map. Using this tool, hundreds of loci may be automatically assigned to the framework in a single process. ActionMap was initially developed to map numerous ESTs with a small plant mapping population and is limited to inbred lines and backcrosses. ActionMap is highly configurable and consists of Perl and PHP scripts that automate command steps for the MapMaker program. A set of web forms were designed for data import and mapping settings. Results of automatic mapping can be displayed as tables or drawings of maps and may be exported. The user may create personal access-restricted projects to store raw data, settings and mapping results. All data may be edited, updated or deleted. ActionMap may be used either online or downloaded for free (<http://moulon.inra.fr/~bioinfo/>).

INTRODUCTION

Identification of genes involved in plant traits is a first step to understanding biological processes underlying these traits. Transcriptome and proteome studies, functional annotation and ultimately co-localization of a gene with a QTL must be taken into account to track candidate genes down. As only two complete plant genome sequences are available, large efforts are still required to genetically map genes in numerous other plant species. ESTs databases are a major source for new sequences to map. The throughput of genetic mapping does not allow systematic mapping of all these ESTs, but hundreds of cDNAs can still be mapped within a year by a single laboratory. For such projects, data storage and mapping computation need to be optimized. Automation of genetic linkage map computation may increase the throughput as well as the reliability of large-scale mapping projects.

Genetic linkage map computation may be achieved by using different softwares such as MapMaker (1), JoinMap (2) or GIMEL (3). MapMaker is a command line software widely-used in plant genetics. It is well suited for automation as command lines may be read directly from text files that have been written by the user or by other software. However, building a genetic map *de novo* requires too much human expertise to be easy to automate. Expert systems developed for automated construction of human genetic linkage maps remain to be improved (4), and cannot be applied to the genetic structure of usual plant mapping populations. Nevertheless, if loci are assigned to a fixed framework map, MapMaker command steps required to assign new markers to the framework are similar for each marker. We thus developed ActionMap: a software that automates the assignment of markers to a fixed framework map without adding the new markers to the map. This choice was driven by the requirements of projects involving high numbers of loci (thousands) mapped on a small number of segregating plants (less than one hundred). In such projects, three point orders between closely linked loci are not statistically significant, so it is better not to include all the markers to the map.

ActionMap performs mapping step by step by generating command line files for MapMaker. Outputs from MapMaker are captured and analyzed in order to generate the next step command line file and so on, until mapping is completed. For quality control purposes, it is critical to store all the intermediate data. Therefore, a database was integrated into our software to save the automatic mapping settings, the intermediate data and the results as well as the raw segregation data. This database can then be considered as a mapping data management system. As pointed out by Li *et al.* (5), such a database must manage the fact that segregation data may change over time (repeated experiments) and are not 100% accurate. Therefore our software provides functions for easy importation, deletion and also edition or update of the data. Reliability of the data is also an important feature to address. For instance, consistency and formats of data as well as their compatibility with MapMaker are checked when data files are transferred to ActionMap.

IMPLEMENTATION

ActionMap is packaged as a web server, which may be used online or downloaded for free (<http://moulon.inra.fr/~bioinfo/>).

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