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(54) **RECOMBINANT FACTOR C AND METHOD FOR PRODUCING THE SAME, AND METHOD FOR MEASURING ENDOTOXIN**

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Related U.S. Application Data

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(30) **Foreign Application Priority Data**

Dec. 10, 2012 (JP) 2012-269840

(51) **Int. Cl.**

C12N 9/64 (2006.01)
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C12N 5/10 (2006.01)
A61K 38/48 (2006.01)
G01N 33/579 (2006.01)
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CPC **C12N 9/6408** (2013.01); **C12Y 304/21069** (2013.01); **C12Y 304/21084** (2013.01); **G01N 33/56911** (2013.01); **G01N 33/579** (2013.01); **G01N 2333/96411** (2013.01)

(58) **Field of Classification Search**

None
See application file for complete search history.

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(74) *Attorney, Agent, or Firm* — Sughrue Mion, PLLC

(57) **ABSTRACT**

To provide a method for producing a horseshoe crab recombinant Factor C. The horseshoe crab recombinant Factor C is produced through expression thereof by use of mammalian cells such as CHO DG44 and HEK293 as host cells.

13 Claims, 10 Drawing Sheets

Specification includes a Sequence Listing.

CRL Exhibit 1060
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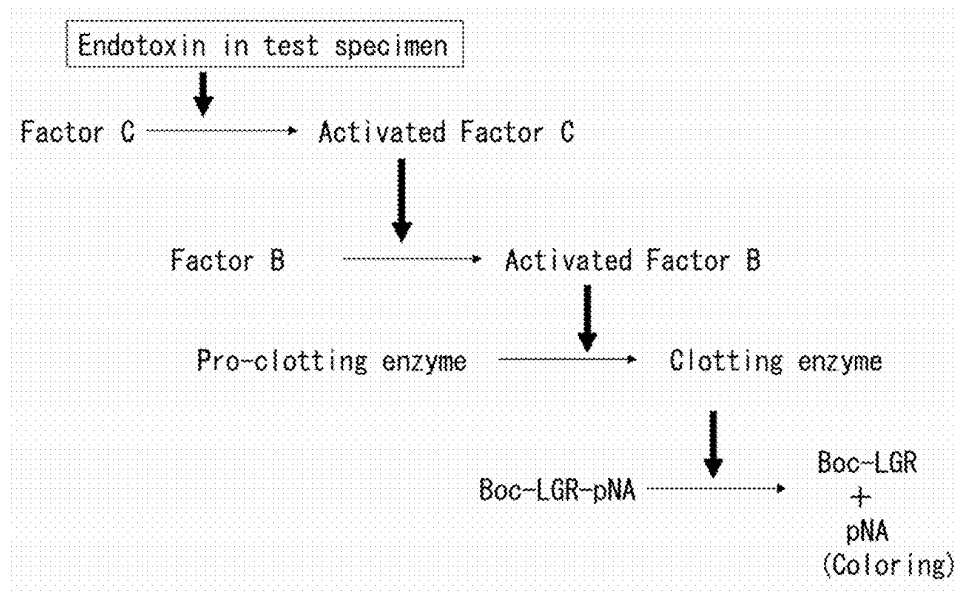
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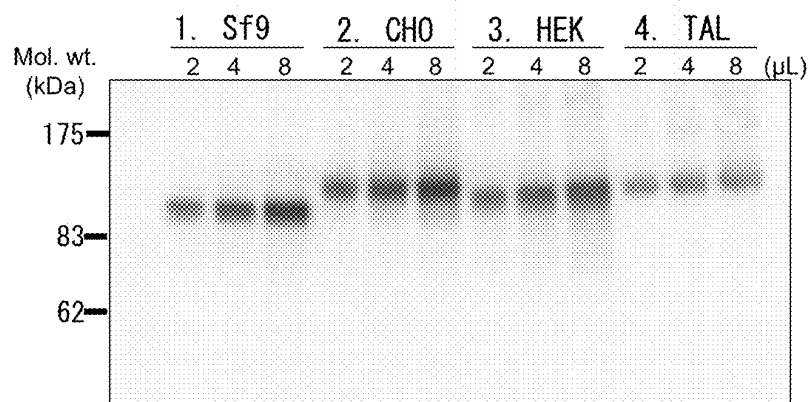
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[Fig. 1]



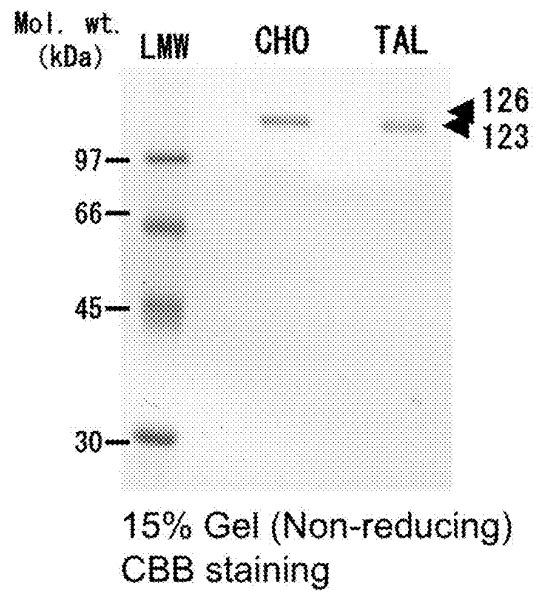
[Fig. 2]



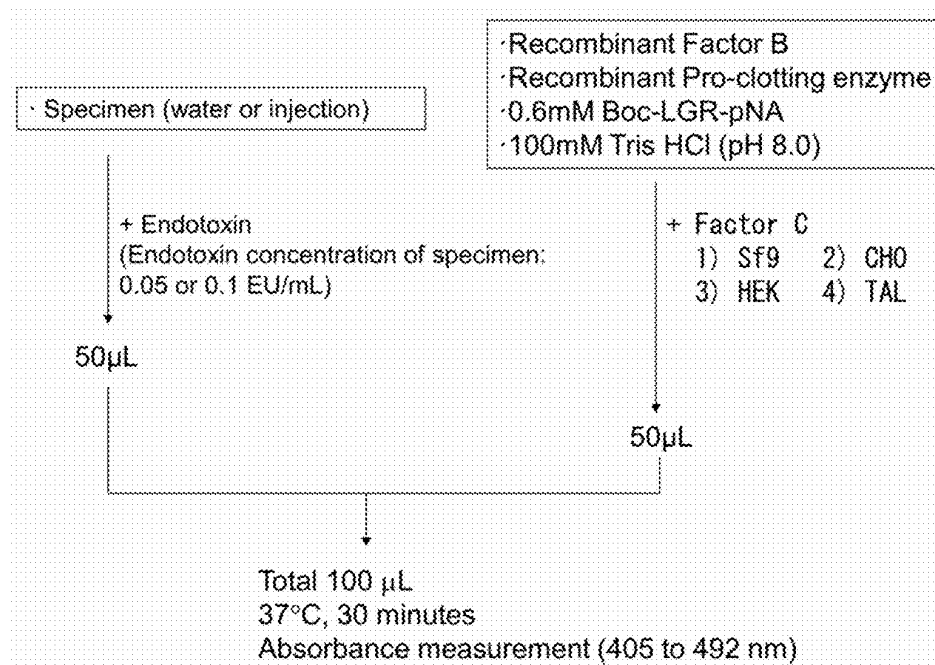
Primary antibody: Anti-factor C monoclonal antibody (2C12)

Electrophoresis conditions: non-reducing

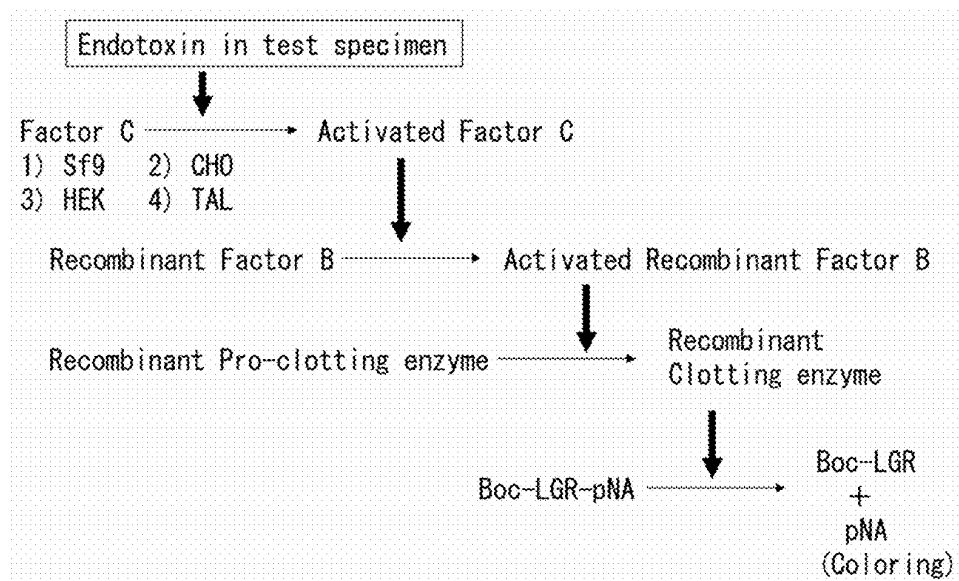
[Fig. 3]



[Fig. 4]



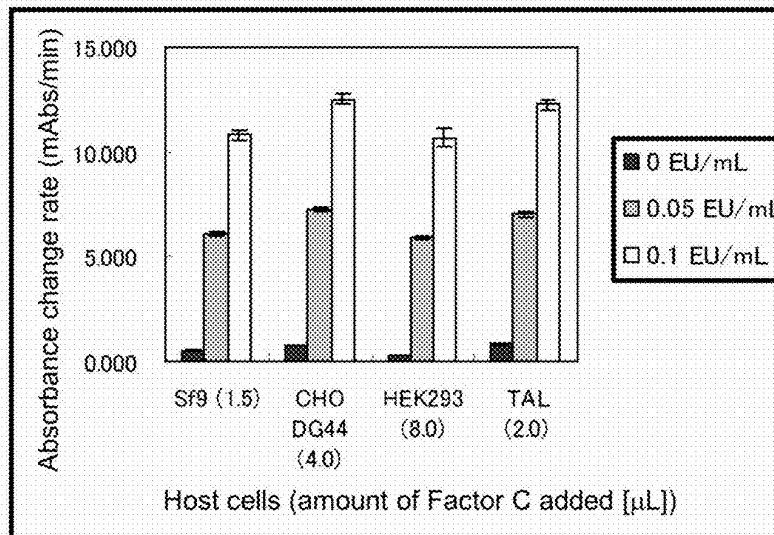
[Fig. 5]



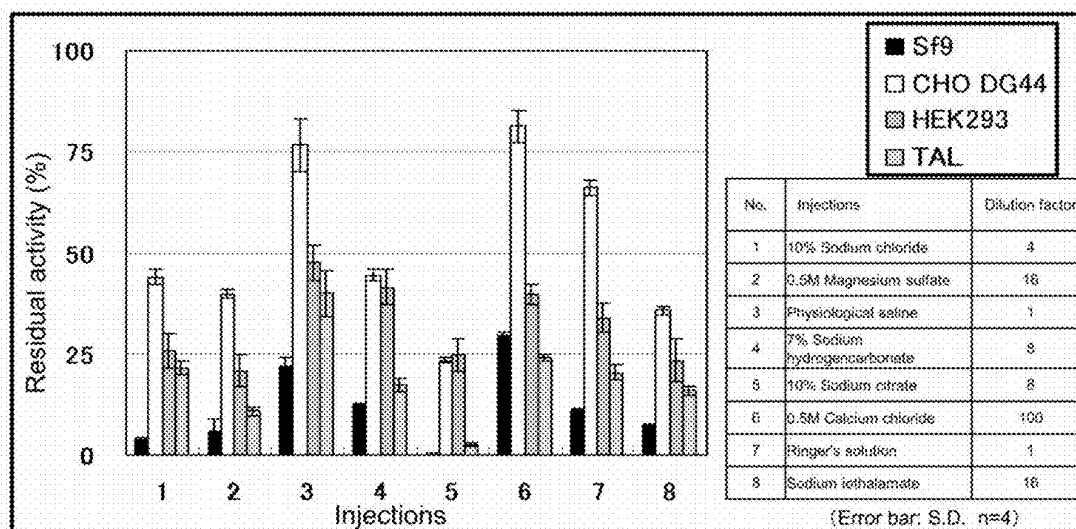
[Fig. 6]

Host cells for Factor C	Amount of Factor C added (μ L)	Endotoxin concentration (EU/mL)	Absorbance change rate (av. $\pm$ S.D.) (mAbs/min)
Sf9	1.5	0	0.520 $\pm$ 0.015
		0.05	6.095 $\pm$ 0.108
		0.1	10.795 $\pm$ 0.246
CHO DG44	4	0	0.770 $\pm$ 0.021
		0.05	7.260 $\pm$ 0.087
		0.1	12.510 $\pm$ 0.255
HEK293	8	0	0.300 $\pm$ 0.017
		0.05	5.885 $\pm$ 0.125
		0.1	10.685 $\pm$ 0.417
TAL	2	0	0.840 $\pm$ 0.055
		0.05	7.030 $\pm$ 0.140
		0.1	12.270 $\pm$ 0.255

n=3



[Fig. 7]

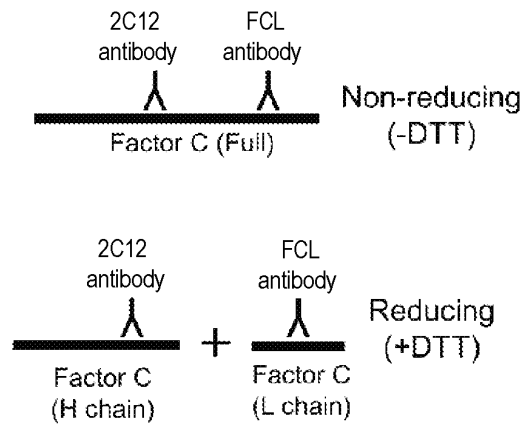


TtFc	MVLASFLVSGLVGLGILAQQMRPVQSRGVLDGLCDETRFECKCGDPGYVFNVPMKQCTYFY	
834Cr ID4	MVLASFLVSGLVGLLGAQKMRPVQSKGVLDGLCDETRFECKCGDPGYVFNI PVKQCTYFY *****[****]:*****[*****]*****	60
TtFc	RWRPYCKPCDDLEAKDICPKYKRCQECKAGLDSVCVTPPNKYGTWCSGECQCKNGGICDQ	
834Cr ID4	RWRPYCKPCDDLEAKDICPKYKRCQECKAGLDSVCVTPPNKYGTWCSGECQCKNGGICDQ *****	120
TtFc	RTGACTCRDRYEGAHCEILKGCPLLPSDSQVQEVNRNPPDNPTIDYSCSPGFKLKGMARI	
834Cr ID4	RTGACACRDRYEGVHCEILKGCPLLPSDSQVQEVNRNPPDNPTIDYSCSPGFKLKGMARI *****[*****]:*****[*****]	180
TtFc	SCLPNGQWSSFPFKCI RECAKVSSPEHGKVNAPSGNMIEGATLRFCDSPYYLIQGETLT	
834Cr ID4	SCLPNGQWSNFPPKCI RECAMVSSPEHGKVNALSGDMIEGATLRFCDSPYYLIQGETLT *****[*****] ***** *	240
TtFc	CQGNQGWSGQIPQCKKL VFCPLDPVNHAHQVKIGVEQKYGGFPGTEVTYTCSGNYFL	
834Cr ID4	CQGNQGWNGQIPQCKNL VFCPLDPVNHAHKVKIGVEQKYGGFPGTEVTYTCSGNYFL *****[*****]:*****[*****]	300
TtFc	MGFN TLKCNPDGSWSGSQPSCVKVADREVD CDSKA VD FLDDVGEPVRI HCPAGCSL TAGT	
834Cr ID4	MGFDTLKCNPDGSWSGSQPSCVKVADREVD CDSKA VD FLDDVGEPVRI HCPAGCSL TAGT ***[*****] *****	360
TtFc	VWGTAIYHELSSVCRAAI HAGKL PNSGGAVHVNNGPYSDFLGSDLNGIKSEELKSLARS	
834Cr ID4	VWGTAIYHELSSVCRAAI HAGKL PNSGGAVHVNNGPYSDFLGSDLNGIKSEELKSLARS *****	420
TtFc	FRFDYVSSSTAGRSGCPDGWF EVECNCVYVTSQR AWE RA QGVCTNMAARLA VL D KDL IP	
834Cr ID4	FRFDYVRSSTAGKSGCPDGWF EVDENCVYVTSQR AWE RA QGVCTNMAARLA VL D KDV IP ***** [****][*****] *****[*****]	480
TtFc	SSLTETLRGKGLTTTWIGLHRLDAEKPFVWELMDRSNVVLNDNLTFWASGEPGNETNCVY	
834Cr ID4	NSLTETLRGKGLTTTWIGLHRLDAEKPFVWELMDRSNVVLNDNLTFWASGEPGNETNCVY [*****] ***** <u>*****</u> <u>*****</u>	540
TtFc	LDIRDQLQPVWKTKSCFQPS SFACMMDLSDRNKAKCDDPGLEN GHAT LH GQS ID GFY AG	
834Cr ID4	MDIQDQLQSVWKTKSCFQPS SFACMMDLSDRNKAKCDDPGS LENGHAT LH GQS ID GFY AG [*]*[****] *****[*****] *****	600
TtFc	SSI RYSCEVLHYLSGTETVTCTTNGTWSAPKPRCIKVITCQNPPVP SYGSVEIKPPSRTN	
834Cr ID4	SSI RYSCEVLHYLSGTETVTCTTNGTWSAPKPRCIKVITCQNPPVP SYGSVEIKPPSRTN *****	660

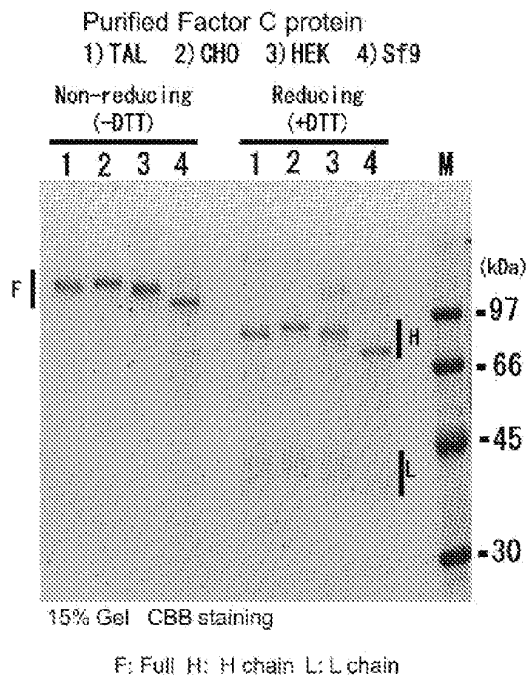
[Fig. 9]

TtFC 834CrID4	SISRVGSPFLRLPRLPLPLARAAPPPKPRSSQPSTVDLASKVKLPEGHYRVGSRAIYTC SISRVGSPFLRLPRLPLPLARAAPPPKPRSSQPSTVDLASKVKLPEGHYRVGSRAIYTC *****	720
TtFC 834CrID4	ESRYEYELLSQGRRCDNNGNSGRPASCIPVCGRSDSPRSPFIWNGNSTEIGQWPWQAGI ESRYEYELLSQGRRCDNNGNSGRPASCIPVCGRSDSPRSPFIWNGNSTEIGQWPWQAGI *****	780
TtFC 834CrID4	SRWLADHNMFLLQCGGSLLNEKWIVTAAHCVTYSATAEIIDPSQFKIYLGKYYRDDSRRD SRWLADHNMFLLQCGGSLLNEKWIVTAAHCVTYSATAEIIDPNQFKMYLGKYYRDDSRRD *****	840
TtFC 834CrID4	DYVQVREALEIHVNPNYDPGNLNFIDIALIQLKTPVTLTTRVQPICLPTDITTREHLKEGT DYVQVREALEIHVNPNYDPGNLNFIDIALIQLKTPVTLTTRVQPICLPTDITTREHLKEGT *****	900
TtFC 834CrID4	LAVVTGWGLNENNTYSEMIQQAFLPVVAASTCEEYKADLPLTVTENMFCAGYKKGRYD LAVVTGWGLNENNTYSEMIQQAFLPVVAASTCEEYKADLPLTVTENMFCAGYKKGRYD *****	960
TtFC 834CrID4	ACSGDSGGPLVFADDSRTERRWVLEGI VSWGSPSGCGKANQYGGFTKVNFLSWIRQFI ACSGDSGGPLVFADDSRTERRWVLEGI VSWGSPSGCGKANQYGGFTKVNFLSWIRQFI *****	1019

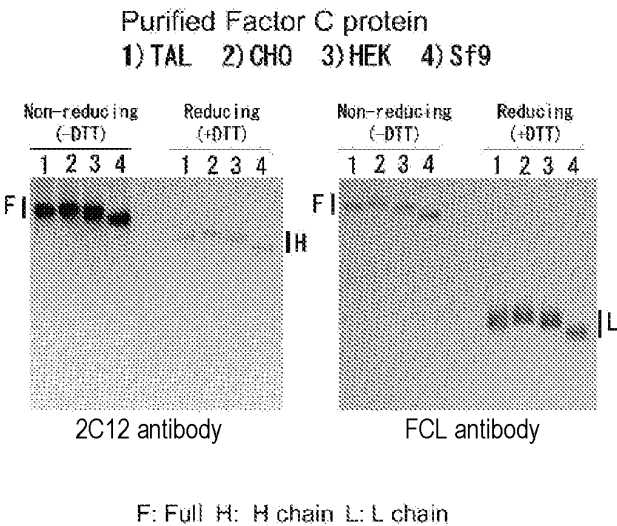
[Fig. 10]



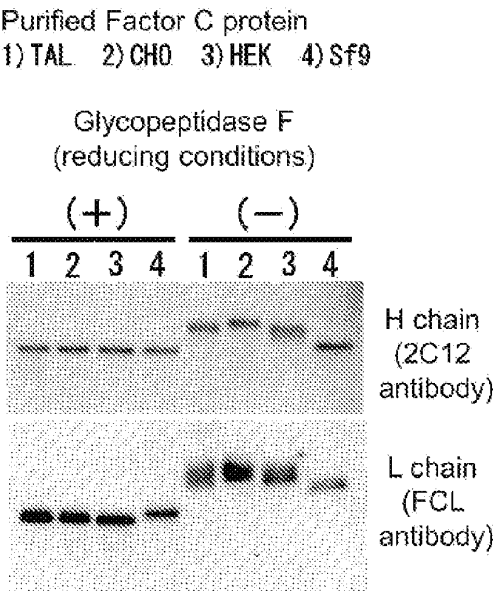
[Fig. 11]



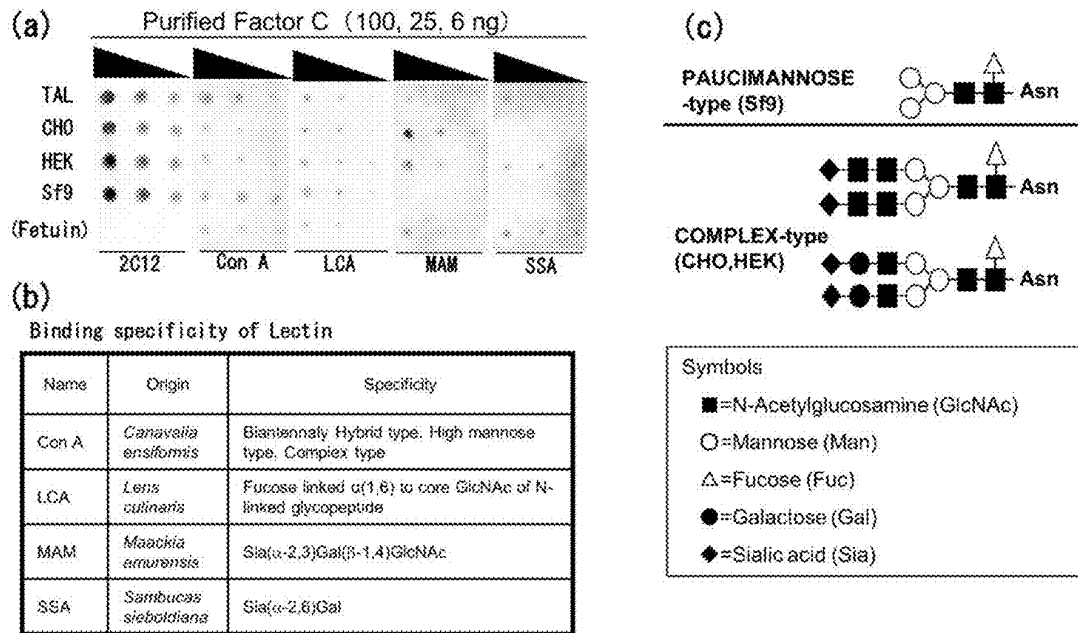
[Fig. 12]



[Fig. 13]



[Fig. 14]



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RECOMBINANT FACTOR C AND METHOD FOR PRODUCING THE SAME, AND METHOD FOR MEASURING ENDOTOXIN

CROSS REFERENCE TO RELATED APPLICATIONS

This is a Continuation of U.S. patent application Ser. No. 14/650,767 filed Jun. 9, 2015 now U.S. Pat. No. 10,144,923, which is a continuation of National Stage of International Application No. PCT/JP2013/083082 filed Dec. 10, 2013, claiming priority based on Japanese Patent Application No. 2012-269840 filed Dec. 10, 2012, the contents of all of which are incorporated herein by reference in their entirety.

TECHNICAL FIELD

The present invention relates to a novel recombinant Factor C, to a method for producing the recombinant Factor C, and to a method for measuring endotoxin.

BACKGROUND ART

Endotoxin is a lipopolysaccharide which is present in the outer membrane of Gram-negative bacteria and is known as a strong pyrogen. It is also known that even a very small amount of endotoxin causes various pathological conditions via bacterial infection. Such conditions include not only fever but also release of an inflammatory cytokine concomitant with activation of macrophages, induction of endotoxin shock, and the like. Therefore, detection of endotoxin is essential in pharmaceutical products such as an injection, water, medical devices, and the like. From another aspect, endotoxin is a conceivable main cause for a shock involved in an infection with a Gram-negative bacterium. Thus, the presence of infection and a therapeutic effect can be determined through a blood endotoxin analysis.

Meanwhile, it has been known that the American horseshoe crab (*Limulus polyphemus*) undergoes blood clotting when it is infected with a Gram-negative bacterium. This phenomenon has been conventionally employed for the detection of endotoxin.

Specifically, there is known a method for measuring endotoxin by use of a hemocyte extract of a horseshoe crab (i.e. an amebocyte lysate of a horseshoe crab, hereinafter also referred to simply as a lysate) (see, for example, Non-Patent Document 1). This method is called a "limulus test," which employs a cascade reaction of a variety of proteins present in the lysate, which reaction occurs via contact between endotoxin and the lysate. FIG. 1 shows the scheme of the cascade reaction.

When endotoxin comes into contact with the lysate, Factor C, which is a serine protease zymogen present in the lysate, is activated to thereby form activated Factor C. The thus-formed activated Factor C activates Factor B present in the lysate, to thereby form activated Factor B. The thus-formed activated Factor B activates Pro-clotting enzyme present in the lysate, to thereby form a corresponding Clotting enzyme.

The Clotting enzyme hydrolyzes a specific site of a coagulogen molecule present in the lysate, thereby coagulin gel is formed, and thus the lysate is coagulated. Thus, endotoxin can be measured through measuring the lysate coagulation reaction.

Alternatively, endotoxin may also be measured through coloring reaction between the Clotting enzyme and a synthetic substrate. For example, the Clotting enzyme acts on

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t-butoxycarbonyl-leucyl-glycyl-arginyl-pNA (Boc-Leu-Gly-Arg-pNA), which is a synthetic substrate, to hydrolyze the amino bonds thereof, and thereby pNA is released. Thus, when the synthetic substrate has been added to the reaction system, endotoxin can be measured through measuring the absorbance (at 405 nm) of the coloring substance (pNA).

Furthermore, it is known that a cascade reaction system can be reconstituted by use of Factor C, Factor B, and Pro-clotting enzyme, which are purified from a lysate of the Japanese horseshoe crab (Non-Patent Document 2).

However, for using such a lysate, or Factor C, Factor B, and Pro-clotting enzyme purified from the lysate, horseshoe crabs must be caught and blood must be recovered therefrom. Hence, from the viewpoint of protection of biological resources, difficulty is encountered in supply of such ingredients in an inexhaustible manner. Under such circumstances, there is demand for a technique that can produce these ingredients by genetic engineering, to thereby reconstitute a cascade reaction system.

For example, there is known a case where Factor C, Factor B, and Pro-clotting enzyme were expressed in insect cells as host cells, to thereby reconstitute a cascade reaction system (Patent Documents 1 and 2). However, it has been reported that since the reconstituted system contains sodium chloride, magnesium sulfate, or calcium chloride in the reaction system, the cascade reaction is suppressed (Patent Document 1).

Alternatively, there is known a case of using mammalian cells as host cells, wherein Factor C derived from the Singaporean horseshoe crab (*Carcinoscorpius rotundicauda*) was expressed in COS-1, which is a cell line derived from African green monkey kidney cells, as host cells. However, it has been reported that when COS-1 was used as a host, the expressed Factor C was insoluble (Non-Patent Documents 3 and 4). That is, there has never been known a case where functional Factor C is produced using mammalian cells as host cells.

PRIOR ART DOCUMENTS

Patent Documents

Patent Document 1: WO 2008/004674, pamphlet
Patent Document 2: WO 2012/118226, pamphlet

Non-Patent Documents

Non-Patent Document 1: Iwanaga S., Curr. Opin. Immunol. February; 5(1): 74-82 (1993)
Non-Patent Document 2: Nakamura T., et al., J. Biochem. March; 99(3): 847-57 (1986)
Non-Patent Document 3: Roopashree S. Dwarakanath, et al., Biotechnology letters 19(4): 357-361 (1997)
Non-Patent Document 4: Jing Wang, Bow Ho and Jeak L. Ding, Biotechnology letters 23: 71-76 (2001)

SUMMARY OF THE INVENTION

Problems to be Solved by the Invention

An object of the present invention is to provide a novel recombinant horseshoe crab Factor C, and a method for producing the recombinant Factor C.

Meanwhile, injections to be administered in vivo generally contain a variety of salts (ions). In production of such injections, detection of endotoxin is obligated by the pharmacopoeia of the relevant country. Under such circum-

stances, in an aspect, another object of the present invention is to provide a reconstituted cascade reaction system which is not susceptible to reaction inhibition even in the presence of a salt (ion).

Means for Solving the Problems

The present inventors have found that a recombinant horseshoe crab Factor C can be produced by use of human cells or Chinese hamster cells as host cells, and that endotoxin can be measured by use of the thus-produced recombinant Factor C while reaction inhibition in the presence of a salt (ion) is mitigated. The present invention has been accomplished on the basis of these findings.

Accordingly, the present invention encompasses the following modes.

[1]

A horseshoe crab Factor C having activity of Factor C.

[1.1.1]

The above-described Factor C, which contains sialic acid.

[1.1.2]

The above-described Factor C, which contains (α -2,3) linked terminal sialic acid.

[1.1.3]

The above-described Factor C, which contains (α -2,3) linked terminal sialic acid in a greater amount, as compared with a native Factor C.

[1.1.4]

The above-described Factor C, which contains (α -2,3) linked terminal sialic acid in a greater amount, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell.

[1.1.5]

The above-described Factor C, which exhibits a higher reactivity in lectin blotting by use of *Maackia amurensis* agglutinin, as compared with a native Factor C.

[1.1.6]

The above-described Factor C, which exhibits a higher reactivity in lectin blotting by use of *Maackia amurensis* agglutinin, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell.

[1.2.1]

The above-described Factor C, which exhibits a residual activity of 10% or higher in the presence of 21 mM sodium citrate.

[1.2.2]

The above-described Factor C, which exhibits a residual activity of 20% or higher in the presence of 21 mM sodium citrate.

[1.2.3]

The above-described Factor C, which exhibits a residual activity of 25% or higher in the presence of 52 mM sodium hydrogencarbonate.

[1.2.4]

The above-described Factor C, which exhibits a residual activity of 35% or higher in the presence of 52 mM sodium hydrogencarbonate.

[1.2.5]

The above-described Factor C, which exhibits a residual activity of 25% or higher in the presence of 214 mM sodium chloride.

[1.2.6]

The above-described Factor C, which exhibits a residual activity of 35% or higher in the presence of 214 mM sodium chloride.

[1.2.7]

The above-described Factor C, which exhibits a residual activity of 15% or higher in the presence of 16 mM magnesium sulfate.

5 [1.2.8]

The above-described Factor C, which exhibits a residual activity of 25% or higher in the presence of 16 mM magnesium sulfate.

[1.2.9]

10 The above-described Factor C, which exhibits a residual activity of 35% or higher in the presence of 2.5 mM calcium chloride.

[1.2.10]

15 The above-described Factor C, which exhibits a residual activity of 45% or higher in the presence of 2.5 mM calcium chloride.

[1.2.11]

20 The above-described Factor C, which exhibits a higher residual activity in the presence of 21 mM sodium citrate, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[1.2.12]

25 The above-described Factor C, which exhibits a higher residual activity in the presence of 52 mM sodium hydrogencarbonate, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[1.2.13]

30 The above-described Factor C, which exhibits a higher residual activity in the presence of 214 mM sodium chloride, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[1.2.14]

35 The above-described Factor C, which exhibits a higher residual activity in the presence of 16 mM magnesium sulfate, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[1.2.15]

40 The above-described Factor C, which exhibits a higher residual activity in the presence of 2.5 mM calcium chloride, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[1.3.1]

45 The Factor C, which is a protein shown in the following (A), (B), (C), or (D):

(A) a protein comprising the amino acid sequence shown in SEQ ID NO: 2;

50 (B) a protein comprising the amino acid sequence shown in SEQ ID NO:2 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Factor C;

(C) a protein comprising the amino acid sequence shown in SEQ ID NO: 4;

55 (D) a protein comprising the amino acid sequence shown in SEQ ID NO:4 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Factor C.

[1.3.2]

60 The Factor C, which is a protein shown in the following (A) or (B):

(A) a protein comprising the amino acid sequence shown in SEQ ID NO: 2;

65 (B) a protein comprising the amino acid sequence shown in SEQ ID NO:2 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Factor C.

[1.4]

The above-described Factor C, which is a recombinant protein.

[1.5.1]

The above-described Factor C, which has a molecular weight of 115 kDa to 140 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.2]

The above-described Factor C, which has a molecular weight of 115 kDa to 130 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.3]

The above-described Factor C, which has a molecular weight of 120 kDa to 130 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.4]

The above-described Factor C, which has a molecular weight of 120 kDa to 128 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.5]

The above-described Factor C, which has a molecular weight of 127 kDa $\pm$ 5 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.6]

The above-described Factor C, which has a molecular weight of 128 kDa $\pm$ 2 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.7]

The above-described Factor C, which has a molecular weight of 127 kDa $\pm$ 2 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.8]

The above-described Factor C, which has a molecular weight of 126 kDa $\pm$ 2 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.5.9]

The above-described Factor C, which has a molecular weight of 127 kDa $\pm$ 1 kDa, as measured through SDS-PAGE under a non-reducing condition.

[1.6]

The above-described Factor C, which is water-soluble.

[1.7]

The above-described Factor C, which is a culture supernatant containing Factor C.

[1.8]

The above-described Factor C, which has the following characteristics (1) to (3):

(1) having activity of Factor C;

(2) having a molecular weight of 115 kDa to 130 kDa, as measured through SDS-PAGE under a non-reducing condition; and

(3) exhibiting a residual activity of 10% or higher in the presence of 21 mM sodium citrate.

[2]

A method for producing a horseshoe crab Factor C, the method comprising expressing the horseshoe crab Factor C in a mammalian cell as a host cell.

[2.1.1]

The above-described method, wherein the mammalian cell is a mammalian cell other than COS-1.

[2.1.2]

The above-described method, wherein the mammalian cell is a cell of a mammal selected from the group consisting of a primate and a rodent.

[2.1.3]

The above-described method, wherein the mammalian cell is a primate cell.

[2.1.4]

The above-described method, wherein the mammalian cell is a rodent cell.

[2.1.5]

The above-described method, wherein the mammalian cell is a cell of a mammal selected from the group consisting of a primate and a rodent, other than monkey.

[2.1.6]

The above-described method, wherein the mammalian cell is a Chinese hamster cell or a human cell.

[2.1.7]

The above-described method, wherein the mammalian cell is a Chinese hamster cell.

[2.1.8]

The above-described method, wherein the mammalian cell is a human cell.

[2.1.9]

The above-described method, wherein the mammalian cell is CHO or HEK.

[2.1.10]

The above-described method, wherein the mammalian cell is CHO.

[2.1.11]

The above-described method, wherein the mammalian cell is HEK.

[2.1.12]

The above-described method, wherein the mammalian cell is CHO DG44 or HEK293.

[2.1.13]

The above-described method, wherein the mammalian cell is CHO DG44.

[2.1.14]

The above-described method, wherein the mammalian cell is HEK293.

[2.2.1]

The above-described method, wherein the Factor C is a protein shown in the following (A), (B), (C), or (D):

(A) a protein comprising the amino acid sequence shown in SEQ ID NO: 2;

(B) a protein comprising the amino acid sequence shown in SEQ ID NO:2 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Factor C;

(C) a protein comprising the amino acid sequence shown in SEQ ID NO: 4;

(D) a protein comprising the amino acid sequence shown in SEQ ID NO:4 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Factor C.

[2.2.2]

The above-described method, wherein the Factor C is a protein shown in the following (A) or (B):

(A) a protein comprising the amino acid sequence shown in SEQ ID NO: 2;

(B) a protein comprising the amino acid sequence shown in SEQ ID NO:2 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Factor C.

[3]

A horseshoe crab Factor C producible by the above-described method.

[3.1]

The above-described Factor C, which has activity of Factor C.

[3.2.1]

The above-described Factor C, which contains sialic acid.

[3.2.2]

The above-described Factor C, which contains (α -2,3) linked terminal sialic acid.

[3.2.3]

The above-described Factor C, which contains (α -2,3) linked terminal sialic acid in a greater amount, as compared with a native Factor C.

[3.2.4]

The above-described Factor C, which contains (α -2,3) linked terminal sialic acid in a greater amount, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell.

[3.2.5]

The above-described Factor C, which exhibits a higher reactivity in lectin blotting by use of *Maackia amurensis* agglutinin, as compared with a native Factor C.

[3.2.6]

The above-described Factor C, which exhibits a higher reactivity in lectin blotting by use of *Maackia amurensis* agglutinin, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell.

[3.3.1]

The above-described Factor C, which exhibits a residual activity of 10% or higher in the presence of 21 mM sodium citrate.

[3.3.2]

The above-described Factor C, which exhibits a residual activity of 20% or higher in the presence of 21 mM sodium citrate.

[3.3.3]

The above-described Factor C, which exhibits a residual activity of 25% or higher in the presence of 52 mM sodium hydrogencarbonate.

[3.3.4]

The above-described Factor C, which exhibits a residual activity of 35% or higher in the presence of 52 mM sodium hydrogencarbonate.

[3.3.5]

The above-described Factor C, which exhibits a residual activity of 25% or higher in the presence of 214 mM sodium chloride.

[3.3.6]

The above-described Factor C, which exhibits a residual activity of 35% or higher in the presence of 214 mM sodium chloride.

[3.3.7]

The above-described Factor C, which exhibits a residual activity of 15% or higher in the presence of 16 mM magnesium sulfate.

[3.3.8]

The above-described Factor C, which exhibits a residual activity of 25% or higher in the presence of 16 mM magnesium sulfate.

[3.3.9]

The above-described Factor C, which exhibits a residual activity of 35% or higher in the presence of 2.5 mM calcium chloride.

[3.3.10]

The above-described Factor C, which exhibits a residual activity of 45% or higher in the presence of 2.5 mM calcium chloride.

[3.3.11]

The above-described Factor C, which exhibits a higher residual activity in the presence of 21 mM sodium citrate, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[3.3.12]

The above-described Factor C, which exhibits a higher residual activity in the presence of 52 mM sodium hydrogencarbonate, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[3.3.13]

The above-described Factor C, which exhibits a higher residual activity in the presence of 214 mM sodium chloride, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[3.3.14]

The above-described Factor C, which exhibits a higher residual activity in the presence of 16 mM magnesium sulfate, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[3.3.15]

The above-described Factor C, which exhibits a higher residual activity in the presence of 2.5 mM calcium chloride, as compared with a horseshoe crab Factor C expressed in Sf9 as a host cell and/or a native Factor C.

[3.4]

The above-described Factor C, which is a recombinant protein.

[3.5.1]

The above-described Factor C, which has a molecular weight of 115 kDa to 140 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.2]

The above-described Factor C, which has a molecular weight of 115 kDa to 130 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.3]

The above-described Factor C, which has a molecular weight of 120 kDa to 130 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.4]

The above-described Factor C, which has a molecular weight of 120 kDa to 128 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.5]

The above-described Factor C, which has a molecular weight of 127 kDa $\pm$ 5 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.6]

The above-described Factor C, which has a molecular weight of 128 kDa $\pm$ 2 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.7]

The above-described Factor C, which has a molecular weight of 127 kDa $\pm$ 2 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.8]

The above-described Factor C, which has a molecular weight of 126 kDa $\pm$ 2 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.5.9]

The above-described Factor C, which has a molecular weight of 127 kDa $\pm$ 1 kDa, as measured through SDS-PAGE under a non-reducing condition.

[3.6]

The above-described Factor C, which is water-soluble.

[3.7]

The above-described Factor C, which is a culture supernatant containing Factor C.

[4]

An endotoxin assay agent containing the above-described Factor C.

[4.1]

The above-described assay agent, which further contains a horseshoe crab Factor B and a horseshoe crab Pro-clotting enzyme.

[4.2]

The above-described assay agent, wherein the Factor B is a protein shown in the following (E) or (F) and the Pro-clotting enzyme is a protein shown in the following (G) or (H):

(E) a protein comprising the amino acid sequence shown in SEQ ID NO: 6;

(F) a protein comprising the amino acid sequence shown in SEQ ID NO:6 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Factor B;

(G) a protein comprising the amino acid sequence shown in SEQ ID NO: 8;

(H) a protein comprising the amino acid sequence shown in SEQ ID NO:8 but which includes substitution, deletion, insertion, or addition of one or several amino acid residues, and having activity of Pro-clotting enzyme.

[5]

An endotoxin assay kit comprising the above-described assay agent.

[6]

A method for measuring endotoxin in a test specimen, the method comprising a step of mixing the above-described assay agent and the test specimen, and a step of measuring the progress of a cascade reaction.

[6.1.1]

The above-described method, wherein the test specimen contains an ion.

[6.1.2]

The above-described method, wherein the test specimen contains a cation.

[6.1.3]

The above-described method, wherein the test specimen contains a metal ion.

[6.1.4]

The above-described method, wherein the test specimen contains an alkali metal ion or an alkaline earth metal ion.

[6.1.5]

The above-described method, wherein the test specimen contains an alkali metal ion.

[6.1.6]

The above-described method, wherein the test specimen contains an alkaline earth metal ion.

[6.2.1]

The above-described method, wherein the test specimen contains one or more kinds of ions selected from the group consisting of sodium ion, potassium ion, calcium ion, and magnesium ion.

[6.2.2]

The above-described method, wherein the test specimen contains sodium ion.

[6.2.3]

The above-described method, wherein the test specimen contains potassium ion.

[6.2.4]

The above-described method, wherein the test specimen contains calcium ion.

[6.2.5]

The above-described method, wherein the test specimen contains magnesium ion.

[6.3.1]

The above-described method, wherein the contained amount of the ion in the test specimen is such an amount that

the concentration of the cation derived from the test specimen becomes 1 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

5 [6.3.2]

The above-described method, wherein the contained amount of the ion in the test specimen is such an amount that the concentration of the cation derived from the test specimen becomes 2 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.3.3]

The above-described method, wherein the contained amount of the ion in the test specimen is such an amount that the concentration of the cation derived from the test specimen becomes 5 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

15 [6.3.4]

The above-described method, wherein the contained amount of the ion in the test specimen is such an amount that the concentration of the cation derived from the test specimen becomes 10 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

25 [6.3.5]

The above-described method, wherein the contained amount of the ion in the test specimen is such an amount that the concentration of the cation derived from the test specimen becomes 20 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

30 [6.3.6]

The above-described method, wherein the contained amount of the ion in the test specimen is such an amount that the concentration of the cation derived from the test specimen becomes 50 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

40 [6.3.7]

The above-described method, wherein the contained amount of the ion in the test specimen is such an amount that the concentration of the cation derived from the test specimen becomes 100 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.4]

The above-described method, wherein the test specimen is an injection.

50 [6.5.1]

The above-described method, wherein the test specimen contains one or more kinds of salts selected from the group consisting of sodium chloride, magnesium sulfate, sodium hydrogencarbonate, sodium citrate, calcium chloride, potassium chloride, sodium iothalamate, calcium disodium edetate, and dihydrogen sodium phosphate.

[6.5.2]

The above-described method, wherein the test specimen contains sodium chloride.

60 [6.5.3]

The above-described method, wherein the test specimen contains magnesium sulfate.

65 [6.5.4]

The above-described method, wherein the test specimen contains sodium hydrogencarbonate.

[6.5.5]

The above-described method, wherein the test specimen contains sodium citrate.

[6.5.6]

The above-described method, wherein the test specimen contains calcium chloride.

[6.5.7]

The above-described method, wherein the test specimen contains potassium chloride.

[6.5.8]

The above-described method, wherein the test specimen contains sodium iohalamate.

[6.5.9]

The above-described method, wherein the test specimen contains calcium disodium edetate.

[6.5.10]

The above-described method, wherein the test specimen contains dihydrogen sodium phosphate.

[6.6.1]

The above-described method, wherein the contained amount of the salt in the test specimen is such an amount that the concentration of the salt derived from the test specimen becomes 1 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.6.2]

The above-described method, wherein the contained amount of the salt in the test specimen is such an amount that the concentration of the salt derived from the test specimen becomes 2 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.6.3]

The above-described method, wherein the contained amount of the salt in the test specimen is such an amount that the concentration of the salt derived from the test specimen becomes 5 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.6.4]

The above-described method, wherein the contained amount of the salt in the test specimen is such an amount that the concentration of the salt derived from the test specimen becomes 10 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.6.5]

The above-described method, wherein the contained amount of the salt in the test specimen is such an amount that the concentration of the salt derived from the test specimen becomes 20 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.6.6]

The above-described method, wherein the contained amount of the salt in the test specimen is such an amount that the concentration of the salt derived from the test specimen becomes 50 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.6.7]

The above-described method, wherein the contained amount of the salt in the test specimen is such an amount that the concentration of the salt derived from the test specimen becomes 100 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the assay agent.

[6.7]

The above-described method, which further comprises a step of adding, to the reaction system, a substrate for detecting the progress of the cascade reaction.

[6.8]

The above-described method, which further comprises a step of calculating the amount of endotoxin in the test specimen.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 A chart showing a cascade reaction system of the limulus test.

FIG. 2 A photograph showing the results of western blotting of four types of Factor C (Sf9, CHO, HEK, and TAL) derived from Japanese horseshoe crab (*Tachypleus tridentatus*), under non-reducing conditions.

FIG. 3 A photograph (SDS-PAGE) showing the molecular weight of each of a native Factor C (TAL) derived from Japanese horseshoe crab (*Tachypleus tridentatus*) and a recombinant Factor C (CHO).

FIG. 4 A scheme of measuring the activity of Factor C.

FIG. 5 A chart showing a reconstituted cascade reaction system.

FIG. 6 A table and a graph showing the activity of the four types of Factor C (Sf9, CHO, HEK, and TAL) derived from Japanese horseshoe crab (*Tachypleus tridentatus*).

FIG. 7 A table and a graph showing the activity inhibition by injections of the four types of Factor C (Sf9, CHO, HEK, and TAL) derived from Japanese horseshoe crab (*Tachypleus tridentatus*).

FIG. 8 Alignment of Factor C of *Tachypleus tridentatus* (TiFC; SEQ ID NO: 2) and Factor C of *Carcinoscorpius rotundicauda* (834CrID4; SEQ ID NO: 4) (continued to FIG. 9). Consensus sequences which can undergo N-type sugar chain (N-linked sugar chain) modification are underlined.

FIG. 9 Alignment of Factor C of *Tachypleus tridentatus* (TiFC; SEQ ID NO: 2) and Factor C of *Carcinoscorpius rotundicauda* (834CrID4; SEQ ID NO: 4) (continued from FIG. 8). Consensus sequences which can undergo N-type sugar chain (N-linked sugar chain) modification are underlined.

FIG. 10 A schematic representation of reaction sites between Factor C and specific antibodies under reducing and non-reducing conditions.

FIG. 11 A photograph (SDS-PAGE) showing the molecular weight of the four types of purified Factor C derived from Japanese horseshoe crab (*Tachypleus tridentatus*).

FIG. 12 A photograph (western blotting) showing the molecular weight of the four types of purified Factor C derived from Japanese horseshoe crab (*Tachypleus tridentatus*).

FIG. 13 A photograph (western blotting) showing the results of removal of N-type sugar chain, by glycopeptidase F, from the four types of purified Factor C derived from Japanese horseshoe crab (*Tachypleus tridentatus*).

FIG. 14 (a) A photograph (lectin blotting) showing the results of detection of N-type sugar chain of the four types of purified Factor C derived from Japanese horseshoe crab (*Tachypleus tridentatus*); (b) a table showing the binding specificity of lectin; and (c) a schematic representation of N-type sugar chain structures of a recombinant Factor C, assumed by the experimental results and reports by documents.

MODES FOR CARRYING OUT THE INVENTION

In the present invention, a series of reactions wherein endotoxin activates Factor C to generate active-type Factor C; the active-type Factor C activates Factor B to generate active-type Factor B; and the active-type Factor B activates Pro-clotting enzyme to generate a corresponding Clotting enzyme; may be referred to as “cascade reaction”. In the present invention, Factor C, Factor B, and Pro-clotting enzyme may be collectively referred to as “factor”. In the present invention, a cascade reaction system constituted by combining the factors may be referred to as “reconstituted system”. In such a reconstituted system, an effect of a contaminant contained in an amebocyte lysate of a horse-

(1) Factor C of the Present Invention

The Factor C of the present invention is a Factor C derived from a horseshoe crab. The Factor C of the present invention is a recombinant protein. The term “recombinant protein” refers to a protein obtained through introduction of a gene encoding the protein into a host cell and heterologous expression of the gene. The Factor C of the present invention has activity of Factor C.

Examples of the horseshoe crab include *Tachypleus tridentatus* (Japanese horseshoe crab), *Limulus polyphemus* (American horseshoe crab), *Carcinoscorpius rotundicauda* (Southeast Asian (Singaporean) horseshoe crab), and *Tachypleus gigas* (Southeast Asian horseshoe crab). Among them, *Tachypleus tridentatus* (Japanese horseshoe crab) and *Carcinoscorpius rotundicauda* (Southeast Asian (Singaporean) horseshoe crab) are preferred as a Factor C source. Furthermore, among them, *Tachypleus tridentatus* (Japanese horseshoe crab) is more preferred as a Factor C source. The amino acid sequence of the Factor C derived from *Tachypleus tridentatus* is shown in SEQ ID NO: 2. The nucleotide sequence of a gene encoding the Factor C derived from *Tachypleus tridentatus* is shown in SEQ ID NO: 1. The amino acid sequence of the Factor C derived from *Carcinoscorpius rotundicauda* is shown in SEQ ID NO: 4. The nucleotide sequence of a gene encoding the Factor C derived from *Carcinoscorpius rotundicauda* is shown in SEQ ID NO: 3.

The Factor C of the present invention may undergo N-type sugar chain (N-linked sugar chain) modification. The term “N-type sugar chain” refers to a sugar chain which is linked to an asparagine residue of a protein. In the Factor C derived from *Tachypleus tridentatus* and the Factor C derived from *Carcinoscorpius rotundicauda*, the amino acid residues which can undergo N-type sugar chain modification are conserved. FIGS. 8 and 9 show alignments and consensus sequences (Asn-Xaa-Ser/Thr, wherein Xaa represents any amino acid residue) which can undergo N-type sugar chain modification of both types of Factor C. Thus, in particular, the Factor C derived from *Carcinoscorpius rotundicauda* is thought to be a suitable Factor C as with the Factor C derived from *Tachypleus tridentatus*.

The Factor C of the present invention may contain a sialic acid. Examples of the sialic acid include N acetylneuraminic acid (Neu5Ac) and N-glycolylneuraminic acid (Neu5Gc). The expression “containing a sialic acid” refers to the state that a sialic acid is linked to the Factor C of the present invention. The sialic acid may be linked to Factor C directly or indirectly. In one case, a sugar chain containing a sialic acid may be linked to Factor C, whereby the sialic acid may be indirectly linked to Factor C. The sialic acid may be, for

example, an (α -2,3)-linked sialic acid, more specifically, an (α -2,3)-linked terminal sialic acid. The term “(α -2,3)-linked sialic acid” refers to a sialic acid linked to a sugar via (α -2,3) glycoside linkage. The term “(α -2,3)-linked terminal sialic acid” refers to a sialic acid which is linked to a sugar via (α -2,3) glycoside linkage and which is located at a terminus of the sugar chain. An example of the sugar chain is an N-type sugar chain (N-linked sugar chain). The sialic acid may be added to Factor C through, for example, expression of Factor C in a host cell which can add a sialic-acid-containing sugar chain to a protein through modification after translation. Examples of such a host cell include mammalian cells. Sialic acid may be detected through, for example, sugar chain analysis. Sugar chain analysis may be carried out through a known technique. For example, sugar chain analysis can be carried out by using lectin or a sugar-chain-recognizing antibody. Specifically, for example, an (α -2,3)-linked terminal sialic acid can be specifically detected through lectin blotting using *Maackia amurensis* agglutinin (MAM).

The Factor C of the present invention may contain sialic acid in a greater amount, as compared with a control Factor C. The expression “containing sialic acid in a greater amount, as compared with a control Factor C” refers to the state that the amount of sialic acid linked to the Factor C of the present invention is greater than the amount of sialic acid linked to the control Factor C, in terms of the amount by mole of sialic acid per molecule of Factor C. For example, the amount of sialic acid linked to the Factor C of the present invention may be 1.1 times or greater, 1.2 times or greater, 1.3 times or greater, 1.5 times or greater, 2.0 times or greater, 2.5 times or greater, or 3.0 times or greater than the amount of sialic acid linked to the control Factor C, in terms of the amount by mole of sialic acid per molecule of Factor C.

Examples of the control Factor C include a recombinant Factor C expressed in insect cell Sf9, and a native Factor C. The expression “Factor C expressed in Sf9 as a host cell” refers to Factor C obtainable through expression of a gene encoding Factor C consisting of the same amino acid sequence as that of the Factor C of the present invention, in Sf9 as a host cell. The term “native Factor C” refers to Factor C isolated and purified from a horseshoe crab lysate. The native Factor C may be Factor C isolated and purified from a lysate of a horseshoe crab from which the Factor C of the present invention is derived. That is, specifically, for example, when the Factor C of the present invention is a recombinant Factor C of *Tachypleus tridentatus* or a variant thereof, the native Factor C may be Factor C isolated and purified from a lysate of *Tachypleus tridentatus*.

Whether or not “the Factor C of the present invention contains sialic acid in a greater amount, as compared with a control Factor C” may be determined through comparison between the sialic acid amount of the Factor C of the present invention and that of the control Factor C. The amount of sialic acid may be determined through, for example, sugar chain analysis. Sugar chain analysis may be carried out through a known technique. For example, sugar chain analysis can be carried out by using lectin or a sugar-chain-recognizing antibody. Specifically, for example, an (α -2,3)-linked terminal sialic acid can be specifically quantitated through lectin blotting using *Maackia amurensis* agglutinin (MAM). It can be confirmed that the higher the reactivity in lectin blotting using MAM, the greater the amount of (α -2,3)-linked terminal sialic acid, and that is, the greater amount of (α -2,3)-linked terminal sialic acid the Factor C contains. In other words, the Factor C of the present invention may exhibit higher reactivity, as compared with the

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control Factor C, in lectin blotting using MAM. The expression “exhibit higher reactivity, as compared with the control Factor C, in lectin blotting using MAM” refers to the state that, when the equiamount of the Factor C of the present invention and the control Factor C are subjected to lectin blotting using MAM, the detection intensity (such as the extent of coloring) of the spot attributed to the Factor C of the present invention is higher than that of the spot attributed to the control Factor C.

The Factor C of the present invention may have no His-tag added to the C-terminus. Also, the Factor C of the present invention may have no V5-tag added to the C-terminus. Also, the Factor C of the present invention may have no peptide added to the C-terminus. Also, the Factor C of the present invention may have no peptide added to the N-terminus. Also, the Factor C of the present invention may have no peptide added to any of the N-terminus and the C-terminus.

So long as the Factor C of the present invention has activity of Factor C, the Factor C may be a variant of any of the aforementioned horseshoe crab Factor C proteins (e.g., a protein having the amino acid sequence shown in SEQ ID NO: 2 or 4). Such a variant includes, for example, a homologue and an artificially modified product of the aforementioned Factor C. Notably, the amino acid sequence of the variant is not necessarily found in an actual horseshoe crab. That is, the Factor C “of a horseshoe crab or derived from a horseshoe crab” includes a variant of Factor C found in a horseshoe crab and having an amino acid sequence which is not found in any horseshoe crab.

The activity of Factor C refers to such an activity that the Factor C is activated in the presence of endotoxin to convert Factor B into activated Factor B. Whether “the Factor C of the present invention has activity of Factor C” can be confirmed, for example, by combining the Factor C of the present invention with a suitable Factor B and a suitable Pro-clotting enzyme in the presence of endotoxin, and detecting the progress of the cascade reaction. Specifically, the protein of SEQ ID NO: 6 and the protein of SEQ ID NO: 8 may be used as a suitable Factor B and a suitable Pro-clotting enzyme, respectively. The progress of the cascade reaction may be determined by use of the below-mentioned substrate for detection.

So long as the Factor C of the present invention has activity of Factor C, the Factor C of the present invention may be a protein having the amino acid sequence of the above-mentioned Factor C such as the amino acid sequence shown in SEQ ID NO: 2 or 4, but which includes substitution, deletion, insertion, or addition of one or several amino acid residues. Although the meaning of the term “one or several” varies depending on the positions of the amino acid residues in the three-dimensional structure of the protein and the types of the amino acid residues, the term specifically refers to preferably 1 to 20, more preferably 1 to 10, still more preferably 1 to 5, particularly preferably 1 to 3. The substitution, deletion, insertion, or addition of one or several amino acid residues is a conservative variation where the function of the protein is appropriately maintained. A typical conservative variation is conservative substitution. Conservative substitution is a variation among Phe, Trp, and Tyr, in the case where the substitution occurs at an aromatic amino acid residue; a variation among Leu, Ile, and Val, in the case where the substitution occurs at a hydrophobic amino acid residue; a variation between Gln and Asn, in the case where the substitution occurs at a polar amino acid residue; a variation among Lys, Arg, and His, in the case where the substitution occurs at a basic amino acid residue; a variation

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between Asp and Glu, in the case where the substitution occurs at an acidic amino acid residue; or a variation between Ser and Thr, in the case where the substitution occurs at an amino acid residue having a hydroxyl group. Specific examples of the substitution which can be recognized as conservative substitution include a substitution of Ala by Ser or Thr; a substitution of Arg by Gln, His, or Lys; a substitution of Asn by Glu, Gln, Lys, His, or Asp; a substitution of Asp by Asn, Glu, or Gln; a substitution of Cys by Ser or Ala; a substitution of Gln by Asn, Glu, Lys, His, Asp, or Arg; a substitution of Glu by Gly, Asn, Gln, Lys, or Asp; a substitution of Gly by Pro; a substitution of His by Asn, Lys, Gln, Arg, or Tyr; a substitution of Ile by Leu, Met, Val, or Phe; a substitution of Leu by Ile, Met, Val, or Phe; a substitution of Lys by Asn, Glu, Gln, His, or Arg; a substitution of Met by Ile, Leu, Val, or Phe; a substitution of Phe by Trp, Tyr, Met, Ile, or Leu; a substitution of Ser by Thr or Ala; a substitution of Thr by Ser or Ala; a substitution of Trp by Phe or Tyr; a substitution of Tyr by His, Phe, or Trp; and a substitution of Val by Met, Ile, or Leu. The aforementioned substitution, deletion, insertion, addition, or the like of an amino acid residue(s) also encompasses naturally occurring variations such as those based on individual differences between horseshoe crabs and the strain and species of the horseshoe crabs, from which the gene is derived.

The Factor C of the present invention may also be a protein which has a homology or identity of 80% or higher, preferably 90% or higher, more preferably 95% or higher, still more preferably 97% or higher, particularly preferably 99% or higher with respect to the entire amino acid sequence of Factor C (e.g., the entire amino acid sequence shown in SEQ ID NO: 2 or 4) and which has activity of Factor C.

The gene encoding the Factor C of the present invention is not particularly limited, so long as the gene codes for the aforementioned Factor C of the present invention. The gene encoding the Factor C of the present invention may be a DNA encoding a protein having activity of Factor C, wherein the DNA fragment is hybridized with a probe prepared from a known gene sequence, e.g., with the complementary sequence to the entirety or a part of the nucleotide sequence shown SEQ ID NO: 1 or 3, under stringent conditions. As used herein, the term “stringent conditions” generally refer to the conditions under which a specific hybrid is formed but a non-specific hybrid is not formed. An example of such conditions is conditions under which DNAs having a high homology, e.g., a homology of 80% or higher, preferably 90% or higher, more preferably 95% or higher, still more preferably 97% or higher, particularly preferably 99% or higher, are hybridized with each other, and DNAs having a homology lower than the above level are not hybridized with each other. Another example of the conditions is conditions generally employed in washing of southern hybridization; i.e., washing once, preferably 2 to 3 times, at temperature and salt concentration corresponding to 60° C., 1×SSC, 0.1% SDS, preferably 60° C., 0.1×SSC, 0.1% SDS, more preferably 68° C., 0.1×SSC, 0.1% SDS.

In the gene encoding the Factor C of the present invention, any codon may be substituted by its equivalent codon. For example, the codon combination of the gene encoding the Factor C of the present invention may be modified such that the codon combination is optimized for expression in a mammalian cell. The optimization may be performed through employment of, for example, a conventional contract service. Notably, the gene encoding the Factor C of the

present invention may be a variant of a DNA in which the codon combination is optimized for expression in a mammalian cell.

The above descriptions about the variants of genes and proteins may also be applied mutatis mutandis to Factor B, Pro-clotting enzyme, and gene encoding the same.

The Factor C of the present invention may have a molecular weight of 115 kDa to 140 kDa, 115 kDa to 130 kDa, 120 kDa to 130 kDa, or 120 kDa to 128 kDa, as measured through SDS-PAGE under a non-reducing condition. Also, the Factor C of the present invention may have a molecular weight of 127 kDa $\pm$ 5 kDa, 128 kDa $\pm$ 2 kDa, 127 kDa $\pm$ 2 kDa, 126 kDa $\pm$ 2 kDa, or 127 kDa $\pm$ 1 kDa, as measured through SDS-PAGE under a non-reducing condition. Also, the Factor C of the present invention may have a molecular weight of 128 kDa or 126 kDa, as measured through SDS-PAGE under a non-reducing condition.

The Factor C of the present invention may have low susceptibility to inhibition of activity by an ion. The expression "low susceptibility to inhibition of activity by an ion" refers to the state that the Factor C of the present invention exhibits a high residual activity in the presence of an ion, as compared with a control Factor C. Examples of the control Factor C include a recombinant Factor C expressed in insect cell Sf9 and a native Factor C. The meaning of the expression "exhibiting a high residual activity" is not particularly limited, so long as the residual activity of the Factor C of the present invention is higher than that of the control Factor C. The expression "exhibiting a high residual activity" may refer to, for example, the state that the residual activity of the Factor C of the present invention is 1.1 times or higher, 1.2 times or higher, 1.3 times or higher, 1.5 times or higher, 2.0 times or higher, 2.5 times or higher, or 3.0 times or higher than the residual activity of the control Factor C. Specific examples of "low susceptibility to inhibition of activity by an ion" include such a property that the Factor C of the present invention exhibits a higher residual activity in the presence of 21 mM sodium citrate, as compared with the control Factor C; such a property that the Factor C of the present invention exhibits a higher residual activity in the presence of 52 mM sodium hydrogencarbonate, as compared with the control Factor C; such a property that the Factor C of the present invention exhibits a higher residual activity in the presence of 214 mM sodium chloride, as compared with the control Factor C; such a property that the Factor C of the present invention exhibits a higher residual activity in the presence of 16 mM magnesium sulfate, as compared with the control Factor C; and such a property that the Factor C of the present invention exhibits a higher residual activity in the presence of 2.5 mM calcium chloride, as compared with the control Factor C. Specific examples of "low susceptibility to inhibition of activity by an ion" further include such a property that the Factor C of the present invention exhibits a residual activity of 10% or higher or 20% or higher in the presence of 21 mM sodium citrate, as compared with the control Factor C; such a property that the Factor C of the present invention exhibits a residual activity of 25% or higher or 35% or higher in the presence of 52 mM sodium hydrogencarbonate, as compared with the control Factor C; such a property that the Factor C of the present invention exhibits a residual activity of 25% or higher or 35% or higher in the presence of 214 mM sodium chloride, as compared with the control Factor C; such a property that the Factor C of the present invention exhibits a residual activity of 15% or higher or 25% or higher in the presence of 16 mM magnesium sulfate, as compared with the control Factor C; and such a property that the Factor C of the present

invention exhibits a residual activity of 35% or higher or 45% or higher in the presence of 2.5 mM calcium chloride, as compared with the control Factor C. The Factor C of the present invention may have any one of the above properties singly or two or more of the above properties in combination. The "residual activity" is defined as a relative reactivity (%) observed for a measurement specimen (endotoxin concentration: 0.05 EU/mL) prepared by adding endotoxin to an ion-containing test specimen, with respect to the reactivity observed for another measurement specimen (endotoxin concentration: 0.05 EU/mL) prepared by adding endotoxin to injection water as 100%, in the assay system shown in FIG. 4.

The Factor C of the present invention may be water-soluble. As used herein, the term "water-soluble" refers to the state that, when the Factor C of the present invention is expressed in an appropriate host cell, the thus-expressed Factor C is detected in a soluble fraction. The expression "detected in a soluble fraction" may refer to the state that 20% or more, 50% or more, 80% or more, 90% or more, or 95% or more of the expressed Factor C is detected in a soluble fraction.

(2) Method for Producing the Factor C of the Present Invention

The Factor C of the present invention may be produced through, for example, expression in a mammalian cell as a host. That is, the present invention provides a method for producing a horseshoe crab Factor C, the method comprising expressing the horseshoe crab Factor C in a mammalian cell as a host cell. This method may also be referred to as "the production method of the present invention." The Factor C producible through production method of the present invention is an embodiment of the Factor C of the present invention.

The mammalian cell is not particularly limited, so long as the cell can functionally express a horseshoe crab Factor C. The term "functional expression" refers to the state that the expressed Factor C exhibits activity of Factor C. Specifically, the mammalian cell may be a cell other than COS-1 cell. The mammalian cell is preferably a cell of a mammal selected from the group consisting of a rodent and a primate. Examples of the rodent include, but are not particularly limited to, Chinese hamster, hamster, mouse, rat, and guinea pig. Of these, Chinese hamster is preferred. Examples of the Chinese hamster cell include Chinese hamster ovary (CHO) cell line. Examples of CHO include CHO DG44 and CHO K1. Examples of the primate include, but are not particularly limited to, human, monkey, and chimpanzee. The primate may be a primate other than monkey. Of these, human is preferred. Examples of the human cell include human embryo kidney (HEK) cell line. Examples of HEK include HEK293.

When the mammalian cell retains a gene encoding Factor C, the cell can express Factor C. The gene encoding Factor C may be retained in the mammalian cell, such that it can be expressed under the control of a promoter which functions in the cell. In the mammalian cell, the gene encoding Factor C, for example, may be present on a vector such as a plasmid which undergoes autonomous replication outside a chromosome, or may be introduced to a chromosome. The technique of introducing the gene encoding Factor C into a mammalian cell is not particularly limited. Examples of the vector which undergoes autonomous replication outside a chromosome include pCA7 (Makoto Takeda, et al., J. Virol. 79(22): 14346-54 (2005)). Alternatively, the gene encoding Factor C may be introduced to a chromosome of a mammalian cell by

use of pCI-Neo Vector (product of Promega). The mammalian cell may retain one copy of, or two or more copies of the gene encoding Factor C.

The culture conditions under which a mammalian cell is cultured are not particularly limited, so long as the mammalian cell can proliferate. Conditions generally employed in culturing mammalian cells may be employed, after appropriate modification has been performed, if needed. A culture medium which is generally employed in culturing mammalian cells may be employed as the culture medium in the invention. Specifically, an RPMI 1640 medium (product of Sigma) or a DMEM medium (product of Sigma) may be used. Culturing can be performed, for example, as static culture at 36° C. to 38° C. under feeding of 5% CO₂.

Whether or not Factor C has been functionally expressed can be confirmed by measuring activity of Factor C. Expression of Factor C may also be confirmed by measuring the amount of mRNA formed via transcription of a gene encoding Factor C, or detecting Factor C through western blotting by use of an antibody.

The expressed Factor C can be recovered as a solution containing Factor C, and can be used as a component of the endotoxin assay agent of the present invention. The solution containing Factor C may be, for example, a culture broth, a culture supernatant, a disrupted cell extract, a mixture thereof, or the like. Factor C may be used after being purified to a desired extent, or may be used without any purification. In the present invention, even when a cell culture supernatant containing expressed Factor C is used as it is without any purification, satisfactory endotoxin assay performance can be attained. Purification of Factor C may be performed through, for example, a known protein purification technique. Examples of such a technique include ammonium sulfate precipitation, gel filtration chromatography, ion exchange chromatography, hydrophobic interaction chromatography, hydroxyapatite chromatography, and so forth. In the case where a tag such as His-tag has been added to Factor C, the Factor C may be purified through affinity chromatography based on affinity to the tag. A solution containing Factor C may be used after being filtered by means of a filter. For example, a 0.22-μm filter can be used as the filter.

(3) Endotoxin Assay Agent of the Present Invention

The Factor C of the present invention may be used in endotoxin assay. That is, the present invention provides an endotoxin assay agent containing the Factor C of the present invention. This endotoxin assay agent may also be referred to as an “endotoxin assay agent of the present invention.” The endotoxin assay agent of the present invention may further contain a factor or factors other than Factor C, depending on the mode of endotoxin assay.

For example, the Factor C of the present invention may be used in combination with Factor B and Pro-clotting enzyme, for endotoxin assay. In other words, an embodiment of the endotoxin assay agent of the present invention is an endotoxin assay agent containing the Factor C of the present invention, Factor B, and Pro-clotting enzyme. In this case, endotoxin can be measured by detecting Clotting enzyme, which is the final product of the cascade reaction. The Factor B and the Pro-clotting enzyme contained in the embodiment of the endotoxin assay agent of the present invention may also be referred to as the “Factor B of the present invention” and the “Pro-clotting enzyme of the present invention”, respectively.

Alternatively, endotoxin can be measured by detecting an intermediate stage of the cascade reaction. In this case, it is sufficient that the endotoxin assay agent of the present invention contains a factor or factors involved in from the

start of the cascade reaction to the relevant intermediate stage. Specifically, for example, when endotoxin is measured by detecting the activated Factor C, which is an intermediate of the cascade reaction, it is sufficient that the endotoxin assay agent of the present invention contains Factor C.

The Factor B of the present invention is a Factor B derived from a horseshoe crab. The Pro-clotting enzyme of the present invention is a Pro-clotting enzyme derived from a horseshoe crab. Examples of the horseshoe crab include *Tachypleus tridentatus* (Japanese horseshoe crab), *Limulus polyphemus* (American horseshoe crab), *Carcinoscorpius rotundicauda* (Southeast Asian (Singaporean) horseshoe crab), and *Tachypleus gigas* (Southeast Asian horseshoe crab). Among them, *Tachypleus tridentatus* (Japanese horseshoe crab) is preferred as a source of Factor B and Pro-clotting enzyme.

The amino acid sequence of the Factor B of *Tachypleus tridentatus* and that of the Pro-clotting enzyme of *Tachypleus tridentatus* are shown in SEQ ID NOs: 6 and 8, respectively. The nucleotide sequence of a gene encoding the Factor B of *Tachypleus tridentatus* and that of a gene encoding the Pro-clotting enzyme of *Tachypleus tridentatus* are shown in SEQ ID NOs: 5 and 7, respectively.

The Factor B of the present invention may be a variant of any of the aforementioned horseshoe crab Factor B proteins (e.g., a protein having the amino acid sequence shown in SEQ ID NO: 6), so long as the variant has activity of Factor B. Such a variant includes, for example, a homologue and an artificially modified product of the aforementioned Factor B. The gene encoding the Factor B of the present invention is not particularly limited, so long as the gene codes for the Factor B of the present invention. The above descriptions about the variants of Factor C and the gene encoding Factor C may also be applied mutatis mutandis to the variants of Factor B and the gene encoding Factor B.

The activity of Factor B refers to such an activity that the Factor B is activated in the presence of activated Factor C to convert Pro-clotting enzyme into the activated form thereof, i.e. the corresponding Clotting enzyme. Whether “the Factor B of the present invention has activity of Factor B” can be confirmed, for example, by combining the Factor B of the present invention with a suitable Factor C and a suitable Pro-clotting enzyme in the presence of endotoxin, and detecting the progress of the cascade reaction. Specifically, the protein of SEQ ID NO: 2 and the protein of SEQ ID NO: 8 may be used as a suitable Factor C and a suitable Pro-clotting enzyme, respectively. The progress of the cascade reaction may be determined by use of the below-mentioned substrate for detection.

The Pro-clotting enzyme of the present invention may be a variant of any of the aforementioned horseshoe crab Pro-clotting enzymes (e.g., a protein having the amino acid sequence shown in SEQ ID NO: 8), so long as the variant has activity of Pro-clotting enzyme. Such a variant includes, for example, a homologue and an artificially modified product of the aforementioned Pro-clotting enzyme. The gene encoding the Pro-clotting enzyme of the present invention is not particularly limited, so long as the gene codes for the Pro-clotting enzyme of the present invention. The above descriptions about the variants of Factor C and the gene encoding Factor C may also be applied mutatis mutandis to the variants of Pro-clotting enzyme and the gene encoding Pro-clotting enzyme.

The activity of Pro-clotting enzyme is such an activity that the Pro-clotting enzyme is converted into the activated form thereof, i.e. the corresponding Clotting enzyme, in the

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presence of activated Factor B, to react with the below-mentioned substrate for detection. The activity of reacting with the detection substrate refers to, for example, an activity of reacting with a coagulogen to induce coagulation, or an activity of reacting with the below-described substrate represented by X-Y-Z (wherein X represents a protection group, Y represents a peptide, and Z represents a dye linked to Y via amide bond), to thereby release dye Z (e.g., an activity of reacting with Boc-Leu-Gly-Arg-pNA, to thereby release pNA). Whether "the Pro-clotting enzyme of the present invention has the activity of Pro-clotting enzyme" can be confirmed, for example, by combining the Pro-clotting enzyme of the present invention with a suitable Factor C and a suitable Factor B in the presence of endotoxin, and detecting the progress of the cascade reaction. Specifically, the protein of SEQ ID NO: 2 and the protein of SEQ ID NO: 6 may be used as a suitable Factor C and a suitable Factor B, respectively. The progress of the cascade reaction may be measured by use of the below-mentioned substrate for detection.

So long as the Factor B of the present invention and the Pro-clotting enzyme of the present invention has activity of Factor B and activity of Pro-clotting enzyme, respectively, the Factor B of the present invention and/or the Pro-clotting enzyme of the present invention may have any peptide or the like added thereto. Examples of such a peptide include tag sequences such as His-tag and V5-tag. Similar to Factor C of the present invention, the Factor B of the present invention and/or the Pro-clotting enzyme of the present invention may have no His-tag added to the C-terminus; may have no V5-tag added to the C-terminus; may have no peptide added to the C-terminus; may have no peptide added to the N-terminus; or may have no peptide added to any of the N-terminus and the C-terminus.

In the gene encoding the Factor B of the present invention and/or the gene encoding the Pro-clotting enzyme of the present invention, any codon may be substituted by its equivalent codon. For example, the codon combination of the gene encoding the Factor B of the present invention and/or the gene encoding the Pro-clotting enzyme of the present invention may be modified such that the codon combination is optimized for expression in a host cell. An example of DNA which code for Factor B of SEQ ID NO: 6 and whose codon combination is optimized for expression in an insect cell is a DNA of SEQ ID NO: 9. Notably, the gene encoding the Factor B of the present invention and/or the gene encoding the Pro-clotting enzyme of the present invention may be a variant of a DNA in which the codon combination is optimized for expression in a host cell.

The Factor B of the present invention and the Pro-clotting enzyme of the present invention may each be a native protein or a recombinant protein.

A native Factor B and a native Pro-clotting enzyme may each be obtained from the hemocyte extract of the aforementioned various horseshoe crabs. These factors may be used after being purified to a desired extent. Purification may be performed through, for example, a known technique (Nakamura T. et al., J. Biochem. 1986 March; 99(3): 847-57).

The recombinant Factor B and the recombinant Pro-clotting enzyme may each be obtained through expression in a host cell. The host cell is not particularly limited, so long as a factor of interest can be expressed. For example, a host cell generally employed for heterologous protein expression may be used. Examples of the host cell include insect cells, animal cells, plant cells, yeast cells, and bacterial cells. Among them, an eukaryotic cell is preferred from the

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viewpoint of modification after translation. Specifically, an insect cell and an animal cell are more preferred.

Examples of the animal cells include mammalian cells. In the case where an animal cell such as a mammalian cell is used, the above descriptions about production of the Factor C of the present invention may also be applied mutatis mutandis to production of the Factor B and the Pro-clotting enzyme.

Examples of the insect cells include Sf9, Sf21, SF+, and High-Five. Of these, Sf9 is preferred as the insect cell.

The technique for expressing a factor of interest in an insect cell as a host is not particularly limited, so long as the factor can be expressed. For example, techniques generally employed for heterologous protein expression may be suitably used. For example, a factor of interest can be expressed by infecting an insect cell with a virus into which a gene encoding the factor has been incorporated (the technique being called a "virus method"). Also, for example, a factor of interest can be expressed by incorporating into an insect cell a vector into which a gene encoding the factor has been incorporated, to thereby incorporate the gene into a chromosome of the host cell (the technique being called a "stably expressing cell line method").

<Virus Method>

The virus employed in the virus method is not particularly limited, so long as an insect cell can be infected with the virus, to thereby express a factor of interest. A virus generally employed for protein expression in insect cells may be suitably used. Examples of such viruses include Baculovirus. Baculovirus is preferably Nucleopolyhedrovirus (NPV). Examples of NPV include *Autographa californica* NPV (AcNPV) and *Bombyx mori* NPV (BmNPV). NPV is preferably AcNPV.

Incorporation of a nucleic acid into a virus may be performed through a conventional method. For example, incorporation of a nucleic acid into a virus may be performed through homologous recombination by use of a transfer vector. Examples of the transfer vector include pPSC8 (product of Protein Sciences), pFastBac (product of Life technologies corporation), and pVL1393 (product of Pharmingen). Among them, pPSC8 is a preferable transfer vector.

An insect cell can be infected, through a conventional technique, with the virus into which a gene encoding a factor of interest has been incorporated, to thereby obtain the insect cell which retain the virus and express the factor.

<Stably Expressing Cell Line Method>

Through incorporation of a gene encoding a factor of interest into a chromosome of an insect cell, a stably expressing cell line which stably expresses the factor can be established. The method of establishing the stably expressing cell line is not particularly limited, and a conventional technique may be employed. For example, a stably expressing cell line can be established by use of pIZ-V5 (product of Life Technologies Corporation), according to a user's instruction attached thereto.

The conditions under which an insect cell is cultured are not particularly limited, so long as the insect cell can proliferate. Conditions generally employed in culturing insect cells may be employed, after appropriate modification has been performed, if needed. A culture medium which is generally employed in culturing insect cells may be employed as the culture medium in the invention. Examples of such a culture medium include a commercially available serum-free medium for culturing insect cells. Specifically, an Sf900III serum-free medium (product of Life Technolo-

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gies Corporation) or the like may be suitably used. Culturing can be performed, for example, as shaking culture at 27° C. to 28° C.

Whether or not a factor of interest has been expressed can be confirmed by measuring activity of the factor. Expression of a factor of interest may also be confirmed by measuring the amount of mRNA formed via transcription of a gene encoding the factor, or detecting the factor through western blotting by use of an antibody.

The expressed factor can be recovered as a solution containing the factor, and can be used as a component of the endotoxin assay agent of the present invention. The solution containing the factor may be, for example, a culture broth, a culture supernatant, a disrupted cell extract, a mixture thereof, or the like. The factor may be used after being purified to a desired extent, or may be used without any purification. In the present invention, even when a cell culture supernatant containing the expressed factor is used as it is without any purification, satisfactory endotoxin assay performance can be attained. Purification of the factor may be performed through, for example, a known protein purification technique. Examples of such a technique include ammonium sulfate precipitation, gel filtration chromatography, ion exchange chromatography, hydrophobic interaction chromatography, hydroxyapatite chromatography, and so forth. In the case where a tag such as His-tag has been added to the factor, the factor may be purified through affinity chromatography based on affinity to the tag. A solution containing the factor may be used after being filtered by means of a filter. For example, a 0.22-μm filter can be used as the filter.

In the case each factor is produced through the virus method, the virus is preferably removed. The method of removing virus is not particularly limited, and a conventional technique may be employed. For example, the virus may be removed by means of a hollow fiber filtration membrane having a pore size of 500 kDa.

In the present invention, the Factor C of the present invention, the Factor B of the present invention, and the Pro-clotting enzyme of the present invention may be separately expressed in individually established expression cells.

The endotoxin assay agent of the present invention may contain, for example, only the Factor C of the present invention. Alternatively, the agent may contain, for example, only the Factor C of the present invention, the Factor B of the present invention, and the Pro-clotting enzyme of the present invention.

The endotoxin assay agent of the present invention may further contain a substrate for detecting the progress of the cascade reaction. As used herein, such a substrate may be referred to as a detection substrate.

Examples of the detection substrate include coagulogen. Coagulogen is a detection substrate with respect to Clotting enzyme, which is the final product of the cascade reaction. When coagulogen comes into contact with Clotting enzyme, a coagulated product, coagulin, is formed. The progress of the coagulation reaction can be determined by measuring the turbidity of the reaction mixture. Coagulogen can be recovered from a hematocyte extract of a horseshoe crab (i.e., lysate). Since the nucleotide sequence of the gene encoding coagulogen has been determined (Miyata et al, Protein Nucleic acid Enzyme, extra issue, No. 29; p. 30-43; 1986), coagulogen can also be produced genetically according to a conventional technique.

The detection substrate may be a synthetic substrate. The synthetic substrate is not particularly limited, so long as it has a property suitable for detection of the cascade reaction.

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Examples of the “property suitable for detection of the cascade reaction” include a property for detecting the presence of Clotting enzyme and a property for detecting an intermediate stage of the cascade reaction. Examples of the “property for detecting the presence of Clotting enzyme” include a property of coloring by catalytic reaction with the Clotting enzyme and a property of generating fluorescence by catalytic reaction with the Clotting enzyme. Examples of the “property for detecting an intermediate stage of the cascade reaction” include a property of coloring by catalytic reaction of a cascade reaction intermediate such as activated Factor C and a property of generating fluorescence by catalytic reaction of a cascade reaction intermediate such as activated Factor C. Examples of the synthetic substrate include a substrate represented by formula X-Y-Z (wherein X represents a protection group, Y represents a peptide, and Z represents a dye linked to Y via amide bond). When endotoxin is present in the reaction system, an amide linkage Y-Z is cleaved by the catalytic reaction of Clotting enzyme or an intermediate, which is yielded by the cascade reaction, to release the dye Z, whereby coloring occurs or fluorescence is generated. The protective group X is not particularly limited, and a known protective group for peptide may be suitably used. Examples of such a protective group include a t-butoxycarbonyl group and a benzoyl group. The dye Z is not particularly limited, and it may be a dye detectable under visible light or may be a fluorescent dye. Examples of the dye Z include p-nitroaniline (pNA), 7-methoxycoumarin-4-acetic acid (MCA), 2,4-dinitroaniline (DNP), and Dansyl dyes. Examples of the peptide Y include Leu-Gly-Arg (LGR), Ile-Glu-Gly-Arg (IEGR) (SEQ ID NO: 10), Val-Pro-Arg (VPR), and Asp-Pro-Arg (DPR). A synthetic substrate which reacts with the Clotting enzyme or the intermediate may be appropriately selected as the synthetic substrate in accordance with the mode of endotoxin assay. For example, from the viewpoint of substrate specificity, a substrate comprising LGR as peptide Y can be suitably used for detecting the Clotting enzyme, and a substrate comprising VPR or DPR as peptide Y can be suitably used for detecting activated Factor C. The released dye Z may be determined through a technique in accordance with the property of the target dye.

The endotoxin assay agent of the present invention may further contain an additional component other than the relevant factors and a detection substrate, so long as the assay agent can be used for endotoxin assay. The additional component is not particularly limited, and it may be selected in consideration of storage stability, handling easiness, stability of the factors and detection substrate, and so forth.

The endotoxin assay agent of the present invention may be formulated into any preparation such as solid, liquid, or gel. In the formulation process, additives such as an excipient, a binder, a disintegrant, a lubricant, a flavor/corrigent, a diluent, and a solvent, which are generally used in formulation, may be used. The endotoxin assay agent of the present invention as is may be used for endotoxin assay. Alternatively, the assay agent may be used for endotoxin assay after being diluted with or dispersed or dissolved in water, physiological saline, buffer, etc. Needless to say, the endotoxin assay agent of the present invention also encompasses such diluted, dispersed, dissolved, or similarly processed form of the agent.

In the endotoxin assay agent of the present invention, the factors and other components may be present as a mixture or present separately. For example, the factors may be mixed at any proportions in formulation, or individual factors may be formulated separately.

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The concentrations of the factors and other components contained in the endotoxin assay agent of the present invention are not particularly limited. However, each concentration is preferably adjusted so as to fall within the below-mentioned suitable concentration range for assaying endotoxin. The concentration of each factor in the endotoxin assay agent of the present invention (as a solution prepared before contact with a test specimen) may be, for example, 5 to 200 $\mu\text{g/mL}$, and is preferably 20 to 100 $\mu\text{g/mL}$, more preferably 40 to 80 $\mu\text{g/mL}$, particularly preferably about 60 $\mu\text{g/mL}$. Also, the concentration of each factor in the endotoxin assay agent of the present invention may be adjusted in accordance with the method of producing the factor. When Factor C is produced with a rodent cell as a host, the concentration of Factor C in the endotoxin assay agent of the present invention (as a solution prepared before contact with a test specimen) can also be, for example, 5 to 200 $\mu\text{g/mL}$, 10 to 50 $\mu\text{g/mL}$, or 15 to 40 $\mu\text{g/mL}$. When Factor C is produced with a primate cell as a host, the concentration of Factor C in the endotoxin assay agent of the present invention (as a solution prepared before contact with a test specimen) can also be, for example, 5 to 200 $\mu\text{g/mL}$, 50 to 150 $\mu\text{g/mL}$, or 80 to 120 $\mu\text{g/mL}$. The above-exemplified concentrations may be calculated, for example, on the assumption that all the protein species present in the filtrate of a culture supernatant which has been obtained through expression of a factor of interest via the above-exemplified technique consists of the factor of interest.

The endotoxin assay agent of the present invention may be provided as an endotoxin assay kit. The endotoxin assay kit is not particularly limited, so long as the kit comprises the endotoxin assay agent of the present invention. The endotoxin assay kit may further comprise, for example, one or more elements selected from an endotoxin standard, a reaction container (e.g., a tube or a microplate), an instruction, and the like.

(4) Endotoxin Assay Method of the Present Invention

When a test specimen contains endotoxin, the cascade reaction proceeds by mixing the endotoxin assay agent of the present invention with the test specimen. Through measuring the progress of the cascade reaction, endotoxin contained in the test specimen can be measured. That is, the present invention provides a method for measuring endotoxin present in a test specimen, the method comprising a step of mixing the endotoxin assay agent of the present invention and the test specimen, and a step of measuring the progress of a cascade reaction. The method is also referred to as the "endotoxin assay method of the present invention."

In an embodiment of the endotoxin assay method of the present invention (hereinafter may be referred to as the "first embodiment"), all the factors are mixed with the test specimen. In the first embodiment, the factors contained in the endotoxin assay agent of the present invention may be contained in the reaction system from the start of the step of mixing the endotoxin assay agent of the present invention with the test specimen, or may be successively added to the reaction system.

For example, the step of mixing the endotoxin assay agent of the present invention with the test specimen may comprise the following steps (A) to (C):

(A) a step of adding the Factor C of the present invention to the reaction system;

(B) a step of adding the Factor B of the present invention to the reaction system; and

(C) a step of adding the Pro-clotting enzyme to the reaction system.

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The steps (A) to (C) may be performed separately, partially simultaneously, or totally simultaneously. The steps (A) to (C) may be performed in any order. For example, the step (B) may be performed after the step (A), and the step (C) may be performed after the step (B).

In the first embodiment, the progress of the cascade reaction can be measured by adding a detection substrate to the reaction system and measuring the response (coloring, coagulation, etc.) of the substrate. The detection substrate may be contained in the reaction system from the start of the step of mixing the endotoxin assay agent of the present invention with the test specimen, or may be added to the reaction system during the progress of the step or after completion of the step. Needless to say, the first embodiment also encompasses an endotoxin assay method employing the endotoxin assay agent of the present invention which contains a detection substrate in advance.

In the endotoxin assay method of the present invention, so long as the cascade reaction proceeds when the test specimen contains endotoxin, the Factor B and the Pro-clotting enzyme of the present invention per se are not need to be in contact with the test specimen. That is, another embodiment of the endotoxin assay method of the present invention (hereinafter may be referred to as a "second embodiment") is a method for measuring endotoxin present in a test specimen, the method comprising the following steps (A) to (D):

(A) a step of mixing the Factor C of the present invention with the test specimen;

(B) a step of mixing the Factor B of the present invention with the Factor C after mixing of step (A);

(C) a step of mixing the Pro-clotting enzyme of the present invention with the Factor B after mixing of step (B); and

(D) a step of measuring the progress of a cascade reaction.

In the second embodiment, the steps (A) to (D) may be performed separately, partially simultaneously, or totally simultaneously. For example, step A may be initiated, and then the Factor B and the Pro-clotting enzyme may be added to the reaction system during the progress of the step A or after completion of step A. Also, step B may be initiated, and then the Pro-clotting enzyme may be added to the reaction system during the progress of the step B or after completion of step B. Alternatively, all three factors may be incorporated into the reaction system from the start of step A. Still alternatively, the Factor C after the contact of step A may be recovered and used in step B, or the Factor B after the contact of step B may be recovered and used in step C.

In the second embodiment, the progress of the cascade may be measured by adding a detection substrate to the reaction system and measuring the response (coloring, coagulation, etc.) of the substrate. The detection substrate may be contained in the reaction system from the start of step A, or may be added to the reaction system during the progress of any step or after completion of any step.

Alternatively, endotoxin can be measured by detecting an intermediate stage of the cascade reaction. Specifically, endotoxin can be measured by detecting activated Factor C, which is an intermediate of the cascade reaction. That is, still another embodiment of the endotoxin assay method of the present invention (hereinafter may be referred to as a "third embodiment") may be a method for measuring endotoxin present in a test specimen, the method comprising a step of mixing the Factor C of the present invention with the test specimen and a step of detecting activated Factor C. In the third embodiment, the activated Factor C can be detected by adding a detection substrate to the reaction system and

measuring the response (coloring, etc.) of the substrate. The detection substrate may be contained in the reaction system from the start of the step of mixing the Factor C of the present invention with a test specimen, or may be added to the reaction system during the progress of the step or after completion of the step.

The endotoxin assay method of the present invention, so long as the cascade reaction proceeds when the test specimen contains endotoxin, may further comprise any other arbitrary step. For example, the endotoxin assay method of the present invention may comprise a step of adding a detection substrate to the reaction system, or a step of mixing Clotting enzyme or an intermediate, which was yielded by the cascade reaction, with the detection substrate. Also, for example, the endotoxin assay method of the present invention may comprise a step of calculating the endotoxin level of the test specimen based on the reaction of the detection substrate.

In the endotoxin assay method of the present invention, reaction is preferably carried out in an aqueous medium such as water or a buffer.

In the assay method of the present invention, the concentration of each factor in the reaction mixture is not particularly limited, so long as the cascade reaction proceeds when the test specimen contains endotoxin. The concentration of each factor may be appropriately tuned in accordance with properties of the factor and so forth. For example, the concentration of each factor, as a final concentration, may be, for example, 2.5 to 100 $\mu\text{g/mL}$, and is preferably 10 to 50 $\mu\text{g/mL}$, more preferably 20 to 40 $\mu\text{g/mL}$, particularly preferably about 30 $\mu\text{g/mL}$. Also, the concentration of each factor in the reaction mixture may be adjusted in accordance with the method of producing the factor. When Factor C is produced with a rodent cell as a host, the concentration of Factor C in the reaction mixture, as a final concentration, may also be, for example, 2.5 to 100 $\mu\text{g/mL}$, 5 to 25 $\mu\text{g/mL}$, or 7.5 to 20 $\mu\text{g/mL}$. When Factor C is produced with a primate cell as a host, the concentration of Factor C in the reaction mixture, as a final concentration, may also be, for example, 2.5 to 100 $\mu\text{g/mL}$, 25 to 75 $\mu\text{g/mL}$, or 40 to 60 $\mu\text{g/mL}$. The above-exemplified concentrations may be calculated, for example, on the assumption that all the protein species present in the filtrate of a culture supernatant which has been obtained through expression of a factor of interest via the above-exemplified technique consists of the factor of interest.

In the assay method of the present invention, the concentration of the detection substrate in the reaction mixture is not particularly limited, so long as the cascade reaction proceeds when the test specimen contains endotoxin. The concentration of the detection substrate may be appropriately tuned in accordance with properties of the detection substrate and so forth. In the case where the detection substrate is a synthetic substrate, the concentration of the detection substrate is, for example, generally 0.001 mM to 100 mM, preferably 0.01 mM to 10 mM, as a final concentration.

In any embodiment, so long as the cascade reaction proceeds when the test specimen contains endotoxin, the reaction system may further contain any other arbitrary component, in addition to the factors, the detection substrate, and the test specimen.

The pH of the reaction mixture is not particularly limited, so long as the cascade reaction proceeds when the test specimen contains endotoxin. The pH of the reaction mixture may be appropriately tuned in accordance with prop-

erties of the factors. The pH of the reaction mixture is, for example, generally 5 to 10, preferably 7 to 8.5.

The reaction temperature is not particularly limited, so long as the cascade reaction proceeds when the test specimen contains endotoxin. The reaction temperature may be appropriately tuned in accordance with properties of the factors. The reaction temperature is, for example, generally 10° C. to 80° C., preferably 20° C. to 50° C. For example, the reaction temperature may be 37° C.

The reaction time is not particularly limited. The reaction time may be appropriately tuned in accordance with various conditions such as properties of the factors and reaction temperature. The reaction time is, for example, generally 5 minutes to 2 hours, preferably 15 minutes to 90 minutes. The reaction time may be, for example, 30 minutes to 40 minutes.

In any embodiment, the test specimen, the factors, and other components may be further added singly or in any combination to the reaction system during the course of reaction. These components may be added once or a plurality of times, or in a continuous manner. The reaction conditions may be constant from the start to the end of the reaction, or may be varied in the course of the reaction.

The progress of the cascade reaction attributable to the presence of endotoxin can be measured by measuring the response (coloring, coagulation, etc.) of the detection substrate, whereby endotoxin in the test specimen can be measured. The response (coloring, coagulation, etc.) of the detection substrate may be measured through a technique in accordance with the property of the detection substrate employed.

In the case of quantitatively measuring endotoxin, correlation data between endotoxin level and the extent of response (coloring, coagulation, etc.) of the detection substrate are obtained by use of an endotoxin standard sample with known concentration, and then the amount of endotoxin present in the specimen can be quantitatively determined on the basis of the obtained correlation data. The correlation data is, for example, a calibration curve. The quantitation may be carried out through a kinetic method or an end-point method.

The test specimen which can be subjected to endotoxin assay is not particularly limited. Examples of the specimen include water for medical use, pharmaceutical products, infusion solutions, blood products, medical apparatuses, medical instruments, cosmetics, foods and beverages, environmental specimens (e.g., air, river water, and soil), biological specimens (e.g., blood, body fluid, and tissue), native proteins, recombinant proteins, nucleic acids, and saccharides. The test specimen may be, for example, an injection. An injection refers to an agent that can be administered to the biological body via injection. Such an injection is not necessarily fluid during commercial distribution, and may be a lyophilized product. The test specimen itself, or an extract or wash liquid thereof may be mixed, dispersed, or dissolved in the reaction system, to thereby be subjected to endotoxin assay.

As described above, the Factor C of the present invention may have low susceptibility to inhibition of activity by an ion. Thus, in an embodiment of the present invention, endotoxin present in a test specimen containing an ion can be suitably measured.

In other words, the test specimen of the invention may be a test specimen containing an ion. The expression "containing an ion" refers to the state that the test specimen contains a substance which is present in the form of an ion during endotoxin assay. That is, "ion" referred to in the present

invention may be species which are present inherently in an ionized form in the test specimen, or may be species which are present inherently in a salt form in the test specimen but are ionized during endotoxin assay. The type of ions is not particularly limited. An example of the ion is a cation, and an example of the cation is a metal ion. Examples of the metal ion include an alkali metal ion and an alkaline earth metal ion. Specific examples of the alkali metal ion and the alkaline earth metal ion include sodium ion, potassium ion, calcium ion, and magnesium ion. The contained amount of the ion in the test specimen may be such an amount that, for example, the concentration of the cation derived from the test specimen becomes 1 mM or higher, 2 mM or higher, 5 mM or higher, 10 mM or higher, 20 mM or higher, 50 mM or higher, or 100 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the endotoxin assay agent. Specific examples of the salt include sodium chloride, magnesium sulfate, sodium hydrogencarbonate, sodium citrate, calcium chloride, potassium chloride, sodium iohalamate, calcium disodium edetate, and dihydrogen sodium phosphate. The contained amount of the salt in the test specimen may be such an amount that, for example, the concentration of the salt derived from the test specimen becomes 1 mM or higher, 2 mM or higher, 5 mM or higher, 10 mM or higher, 20 mM or higher, 50 mM or higher, or 100 mM or higher, as a final concentration in the reaction system after mixing the test specimen with the endotoxin assay agent. The test specimen may contain one kind of ion (or salt) or two or more kinds of ions (or salts).

Examples

The present invention will next be described more particularly by way of examples. However, these are merely examples of the present invention, and the scope of the present invention is not limited to these.

1. Preparation of Factor C Proteins

(1) Expression of Factor C in Insect Cells (Sf9)

In this section, Factor C was prepared by use of insect cell Sf9 as a host in the following manner.

A DNA encoding Factor C of Japanese horseshoe crab, *Tachypleus tridentatus*, was inserted between the EcoRV recognition site and the MluI recognition site of pIZ-V5 (product of Life Technologies Corporation), which is a vector for expression in insect cells, to thereby construct a Factor C expression plasmid. The nucleotide sequence of the DNA is shown in SEQ ID NO: 1, and the amino acid sequence of Factor C encoded by the DNA is shown in SEQ ID NO: 2 (the same applies throughout the Example).

The Factor C expression plasmid was introduced into cultured Sf9 cells through transfection by use of Celfectin reagent (product of Life Technologies Corporation). The Sf9 cells were statically cultured at 28° C. in an Sf900 III serum-free medium (product of Life Technologies Corporation) containing 300 to 600 µg/mL Zeocin (product of Life Technologies Corporation), whereby Sf9 strains into which the genome the expression plasmid had been incorporated were selected.

Candidates for Sf9 in which the Factor C gene had been stably expressed were isolated through the cloning cylinder technique, and a plurality of Sf9 cell lines was cloned. The thus-obtained clones were individually cultured, to thereby obtain culture supernatants. Secretion of the recombinant Factor C protein in each culture supernatant was confirmed through western blotting by use of an anti-Factor C antibody (2C12 antibody; Yoshiki Miura, et al., J. Biochem. 112: 476-481 (1992)). Also, activity of Factor C (property of

being activated by endotoxin) in the culture supernatant was confirmed through activity measurement by use of a synthetic substrate Boc-LGR-pNA. Notably, the procedure of activity measurement is described in "4. Activity measurement" below (the same applies throughout the Example). Based on the results of western blotting and activity measurement, an Sf9 cell line highly expressing Factor C was selected.

The Sf9 cell line highly expressing Factor C was suspension-cultured (28° C.) in an Sf900 III medium containing 1× penicillin/streptomycin (product of Life Technologies Corporation) and 50 µg/mL Zeocin to a later phase of the logarithmic growth phase (6 to 8×10⁶ cells/mL). Then, the culture was centrifuged (3,000×g, 30 minutes, 4° C.), to thereby recover a culture supernatant. The supernatant was filtered through a 0.22-µm filter, and the thus-recovered fraction (i.e. filtrate) was employed as recombinant Factor C (Sf9).

(2) Expression of Factor C in Mammalian Cells (CHO DG44)

In this section, Factor C was prepared by use of CHO DG44, which is a Chinese hamster ovary cell line, as a host through the following procedure.

A DNA encoding Factor C of Japanese horseshoe crab, *Tachypleus tridentatus*, was inserted between the EcoRI recognition site and the XbaI recognition site of pCI-neo (product of Promega), which is a vector for expression in mammalian cells, to thereby construct a Factor C expression plasmid. A kozak sequence (GCCACC) was added just before the start codon of Factor C.

The Factor C expression plasmid and a dhfr expression plasmid were incorporated into cultured CHO DG44 cells through simultaneous transfection using a lipofection reagent (Lipofectamine LTX) (product of Life Technologies Corporation). The CHO DG44 cells were statically cultured at 37° C. under feeding of 5% CO₂ in an RPMI 1640 medium (5% dialyzed serum, containing 1× penicillin/streptomycin) (product of Sigma) added with 1 mg/mL Geneticin (product of Life Technologies Corporation), whereby CHO DG44 strains into which the genome the expression plasmids had been incorporated were selected.

Fifteen days after the simultaneous transfection, MTX (methotrexate) (product of Wako Pure Chemical Industries, Ltd.) was added to the culture medium. The MTX concentration was elevated stepwise from 50 nM to 5,000 nM over three months, and CHO DG44 cell lines highly expressing Factor C were selected as polyclones. Effective amplification of the incorporated Factor C gene by virtue of MTX was confirmed by detecting Factor C in a culture supernatant through western blotting using a 2C12 antibody, and quantitating the transcriptional amount of mRNA of Factor C through real-time PCR.

The group of polyclonal CHO DG44 cell lines highly expressing Factor C was diluted by the medium to obtain a cell concentration of 0.5 cells/well, and the dilute was inoculated to a 96-well plate. From about 10 to 20 days after inoculation, the formed cell colonies were successively subjected to expansion culture, whereby CHO DG44 cell lines highly expressing Factor C were cloned. The transcriptional amount of mRNA of Factor C was confirmed through real-time PCR. The amount of Factor C protein secreted to the medium was confirmed through western blotting using a 2C12 antibody. The Factor C activity (property of being activated by endotoxin) in the culture supernatant was confirmed through activity measurement using a synthetic

substrate Boc-LGR-pNA. Based on these results, a monoclonal CHO DG44 cell line highly expressing Factor C was selected.

The CHO DG44 cell line highly expressing Factor C (monoclonal) was conditioned in a serum-free complete synthetic medium containing 5 μ M MTX and thus floated. The cell line was grown at 37° C. under feeding of 5% CO₂ to a later phase of the logarithmic growth phase. The thus-obtained floating culture broth was centrifuged (3,000×g, 30 minutes, 4° C.), to thereby recover a culture supernatant. The supernatant was filtered through a 0.22- μ m filter, and the thus-recovered fraction (i.e. filtrate) was employed as recombinant Factor C (CHO).

(3) Expression of Factor C in Mammalian Cells (HEK293)
In this section, Factor C was prepared by use of HEK293, which is a human embryo kidney cell line, as a host through the following procedure.

A DNA encoding Factor C of a Japanese horseshoe crab, *Tachypleus tridentatus*, was inserted between the EcoRI recognition site and the XhoI recognition site of pCA7 (Makoto Takeda, et al., J. Virol. 79 (22): 14346-54 (2005)), which is a vector for expression in mammalian cells, to thereby construct a Factor C expression plasmid. A kozak sequence (GCCACC) was added just before the start codon of Factor C.

The Factor C expression plasmid was incorporated into cultured HEK293 cells through transfection using polyethylenimine (product of Sigma) (Hashiguti et al., Expression of Recombinant Protein Using Cultured Human Cells—Standard Protocol by 293-type cells-, PSSJ Archives, 1, e017 (2008); available on the world wide web at pssj.jp/archives/Protocol/Expression/293_01/293_01_01.html). HEK293 cells were statically cultured in a DMEM medium (containing 10% fetal bovine serum and 2 mM L-glutamine) (product of Sigma) at 37° C. under feeding of 5% CO₂ without replacement of the medium. Four days after transfection, the culture broth was recovered, and was centrifuged (3,300×g, 30 minutes, 4° C.), to thereby recover a culture supernatant. The supernatant was filtered through a 0.22- μ m filter, and the thus-recovered fraction (i.e. filtrate) was employed as recombinant Factor C (HEK).

(4) Preparation of Native Factor C (Derived from Amebocyte of *Tachypleus tridentatus*)

A fraction containing Factor B and Factor C was recovered from an amebocyte extract of Japanese horseshoe crab, *Tachypleus tridentatus*, through column chromatography by use of dextran sulfate resin (Takanori Nakamura, et al., Eur. J. Biochem. 154: 511-521 (1986)). The fraction (5 mL) was subjected to gel filtration by means of a column (ϕ 2.2 cm×97 cm) filled with Sepharose CL6B gel (product of GE Healthcare). Gel filtration was performed under almost the same conditions as employed in a known technique (Jeak Ling Ding and Bow Ho, U.S. Pat. No. 5,712,144; Jeak L. Ding, et al., Biochem. Biophys. Acta 1202: 149-156). The flow rate was adjusted to 15 mL/h, and 50 mM Tris buffer (pH 8.0) containing 154 mM NaCl, 1 mM EDTA, and 5% DMSO was employed as a solvent.

Gel filtration eluted fractions (3.7 mL/tube, 150 tubes) were subjected to SDS-PAGE followed by CBB staining and western blotting using a 2C12 antibody, and activity measurement using a synthetic substrate Boc-LGR-pNA, whereby fractions containing Factor C protein were identified. These fractions containing the purified Factor C protein were employed as native Factor C (TAL).

2. Preparation of Recombinant Factor B and Recombinant Pro-Clotting Enzyme

A DNA encoding Factor B of Japanese horseshoe crab, *Tachypleus tridentatus*, was inserted between the EcoRV recognition site and the MluI recognition site of pIZ-V5 (product of Life Technologies Corporation), which is a vector for expression in insect cells, to thereby construct a Factor B expression plasmid. The nucleotide sequence of the DNA is shown in SEQ ID NO: 9, and the amino acid sequence of Factor B encoded by the DNA is shown in SEQ ID NO: 6. Notably, the nucleotide sequence shown in SEQ ID NO: 9 is a nucleotide sequence optimized for expression in insect cells.

A DNA encoding Pro-clotting enzyme of Japanese horseshoe crab, *Tachypleus tridentatus*, was inserted between the EcoRV recognition site and the MluI recognition site of pIZ-V5 (product of Life Technologies Corporation), which is a vector for expression in insect cells, to thereby construct a Pro-clotting enzyme expression plasmid. The nucleotide sequence of the DNA is shown in SEQ ID NO: 7, and the amino acid sequence of Pro-clotting enzyme encoded by the DNA is shown in SEQ ID NO: 8.

Through the same method as employed in the case of Factor C, the expression plasmids were each incorporated into Sf9 cells, high-expression clones were selected, and culture supernatants thereof were prepared. The culture supernatants were individually filtered through a 0.22- μ m filter, and the obtained fractions (i.e. filtrates) were employed as a recombinant Factor B and a recombinant Pro-clotting enzyme. Recovery of the factors was confirmed through western blotting employing an antibody specific to each factor and activity measurement.

3. Comparison of Molecular Weights of Factor C Proteins (1)

The molecular weights of the above-prepared four types of Factor C proteins were compared with one another through SDS-PAGE and western blotting.

Firstly, each Factor C in an amount shown in FIG. 2 was subjected to SDS-PAGE under a non-reducing condition employing a 5 to 20% gradient gel. In FIG. 2, Sf9, CHO, and HEK denote the Factor C expressed in Sf9 cells, CHO DG44 cells, and HEK293 cells, respectively, and TAL refers to native Factor C (hereinafter the same applies). Notably, Factor C is cleaved into an H chain and an L chain under reducing conditions. In the present Example, SDS-PAGE was carried out under a non-reducing condition in order to compare the molecular weights of Factor C in an intact state.

After SDS-PAGE, proteins were transferred to a PVDF membrane (product of Bio RAD) by means of a semi-dry blotter. The PVDF membrane was recovered and blocked with 5% skim milk, and then sequentially treated with a primary antibody (2C12 antibody) and a secondary antibody (HRP-labeled anti-mouse goat antibody) (product of Dako). Bands attributed to Factor C were made to emit light with a detection reagent, SuperSignal West Dura (product of Thermo Scientific), and the light emission was recorded by a CCD camera.

The four types of Factor C were found to have different molecular weights, and the molecular weight was the greatest in the case of CHO, followed by TAL, HEK, and Sf9 in the descending order (FIG. 2).

Regarding Singaporean horseshoe crab, *Carcinoscorpius rotundicauda*, it has been reported that all of the native Factor C protein purified from an amebocyte extract, and recombinant Factor C proteins produced in insect cells Sf9 and S2, and in African green monkey kidney cell line COS-1 as a host cell have the same molecular weight of 132 kDa

(Jeak Ling Ding and Bow Ho, U.S. Pat. No. 5,712,144, and Jeak L. Ding, et al., Biochem. Biophys. Acta 1202: 149-156 (1993) (for native Factor C protein); Jing Wang, et al., J. Biol. Chem. 277(39): 36363-72 (2002) (for Sf9 and S2); and Roopashree S. Dwarakanath, et al., Biotechnology letters 19(4): 357-361 (1997) (for COS-1)).

Thus, among the above-prepared four types of Factor C, the molecular weight of each of CHO and TAL was correctly measured. Firstly, recombinant Factor C (CHO) was purified in a manner similar to that employed in the case of native Factor C (TAL). Then, electrophoresis (15% gel, LMW Marker kit (product of GE Healthcare)) was performed under the same conditions as employed in a known technique (Jeak Ling Ding and Bow Ho, U.S. Pat. No. 5,712, 144; and Jeak L. Ding, et al., Biochem. Biophys. Acta 1202: 149-156), and the molecular weight of each Factor C was then determined through Ferguson plotting. As a result, recombinant Factor C (CHO) was found to have a molecular weight of 126 kDa, and native Factor C (TAL) was found to have a molecular weight of 123 kDa (FIG. 3). Thus, native Factor C (TAL) and recombinant Factor C (CHO) derived from Japanese horseshoe crab, *Tachypleus tridentatus*, were found to have a molecular weight smaller than that of the Factor C derived from Singaporean horseshoe crab, *Carcinoscropsius rotundicauda* (132 kDa).

The difference in molecular weight among the above-prepared four types of Factor C is conceived to be attributed to the difference in post-translation modification (sugar chain modification) depending on the type of host cells employed. For example, it has been known that sialic acid is linked to the N-type sugar chain after translation in mammalian cells, whereby a recombinant protein produced in mammalian cells has dimensions larger than those of a recombinant protein produced in insect cells (Robert L. Harrison and Donald L. Jarvis, Methods in Molecular Biology 338: 341-356 (2007)). This supports the results of the present Example (FIG. 2).

4. Activity Measurement

Each of the above-prepared four types of Factor C was combined with other factors, and activity measurement was carried out through the following procedure. FIG. 4 shows the scheme of activity measurement.

As shown in FIG. 4, common materials other than Factor C (a recombinant Factor B, a recombinant Pro-clotting enzyme, a synthetic substrate Boc-LGR-pNA, and Tris buffer) were mixed together, to thereby prepare a common liquid mixture. To the common liquid mixture, Factor C in an amount specified in FIG. 6 was added, and the total volume was adjusted to 50 μ L. The thus-obtained mixture was mixed with 50 μ L of a specimen (water containing United States Pharmacopoeia standard endotoxin (USP-RSE) (product of Seikagaku corporation)), and the resultant mixture was heated at 37° C. for 30 minutes. In this assay system, Factor C is activated by endotoxin, to thereby form activated Factor C; Factor B is activated by the activated Factor C, to thereby form activated Factor B; Pro-clotting enzyme was activated by the activated Factor B, to thereby form a corresponding Clotting enzyme; and the Clotting enzyme cleaves a synthetic substrate Boc-LGR-pNA, whereby p-nitroaniline is released (FIG. 5). The change in absorbance (A 405 nm) attributed to formation of p-nitroaniline was measured in a time-dependent manner, and the absorbance change rate per unit time (mAbs/min) was calculated and employed as Factor C activity. The Measurement was conducted thrice for each sample.

In the above assay system, the final concentration of each factor was as follows: recombinant Factor B 30 μ g/mL,

recombinant Pro-clotting enzyme 30 μ g/mL, recombinant Factor C (Sf9) 11.3 μ g/mL, recombinant Factor C (CHO) 12 μ g/mL, recombinant Factor C (HEK) 51.5 μ g/mL, and Factor C (TAL) 0.25 μ g/mL. The above-exemplified concentrations (except for the case of TAL) were calculated, on the assumption that all the protein species present in the sample (i.e. filtrate) prepared through expression of a factor of interest consisted of the factor of interest. The Factor C (TAL) concentration was measured after purification of Factor C (TAL).

As a result, reconstituted systems each exhibiting almost equivalent activity were successfully constituted for the four types of Factor C (FIG. 6). In each reconstituted system, the activity was increased with an increase in endotoxin concentration (0.05 and 0.1 EU/mL) of a specimen.

The recombinant Factor C derived from *Carcinoscropsius rotundicauda* and produced in mammalian cell COS-1 (132 kDa) is reported to be recovered in an insoluble fraction (Roopashree S. Dwarakanath, et al., Biotechnology letters 19(4): 357-361 (1997); and Jing Wang, Bow Ho and Jeak L. Ding, Biotechnology letters 23: 71-76 (2001)). Also, the recombinant Factor C derived from *Carcinoscropsius rotundicauda* and produced in insect cell S2 (132 kDa) is reported to exhibit no protease activity in response to endotoxin (Jing Wang, et al., J. Biol. Chem. 277(39): 36363-72 (2002)). Lack of activity of the latter Factor C (produced in S2) is thought to be attributed to the difference in sugar chain structure from the native Factor C derived from *Carcinoscropsius rotundicauda* and the recombinant Factor C produced in insect cell Sf9, due to difference in reactivity to lectin.

Meanwhile, in the present Example, it was found that the recombinant Factor C derived from *Tachypleus tridentatus* (Sf9, CHO, and HEK) exhibited an endotoxin-dependent activity, even though the molecular weight thereof differs from that of native Factor C (TAL); i.e., even though the sugar chain structure thereof differs from that of native Factor C (TAL).

5. Inhibition of Activity by Injection

Endotoxin is a strong immune reaction-inducing substance. Thus, when an injection contaminated with endotoxin is administered to the body, a grave side effect occurs. Therefore, in production of injections, detection of endotoxin by use of a limulus amoebocyte lysate reagent is obligated by the pharmacopoeia of the relevant country, to thereby confirm no endotoxin contamination.

Thus, the above-prepared four types of Factor C were compared in terms of reactivity to endotoxin in various injections.

As a control specimen, injection water containing endotoxin (endotoxin concentration: 0.05 EU/mL) was used. Measurement specimens used were as follows: 4-fold dilute of 10 w/v % sodium chloride injection (10% sodium chloride injection, product of Otsuka Pharmaceutical Co., Ltd.); 16-fold dilute of 0.5 M magnesium sulfate injection (Mg sulfate corrective injection 1 mEq/mL, product of Otsuka Pharmaceutical Co. Ltd.); undiluted solution of physiological saline (Otsuka isotonic sodium chloride solution, product of Otsuka Pharmaceutical Co., Ltd.); 8-fold dilute of 7 w/v % sodium hydrogencarbonate injection (Meylon injection 7%, product of Otsuka Pharmaceutical Co., Ltd.); 8-fold dilute of 10 w/v % sodium citrate injection (sodium citrate injection for transfusion, product of Fuso Pharmaceutical Industries, Ltd.); 100-fold dilute of 0.5 M calcium chloride injection (Ca chloride corrective injection 1 mEq/mL, product of Otsuka Pharmaceutical Co., Ltd.); undiluted solution of Ringer's solution (Ringer's solution "Fuso", product of

Fuso Pharmaceutical Industries, Ltd.); and 16-fold dilute of sodium iothalamate injection (Conray 400 injection, product of Daiichi Sankyo Co., Ltd.), each spiked with endotoxin (endotoxin concentration: 0.05 EU/mL). When the measurement specimen was 8-fold dilute of 10 w/v % sodium citrate injection, the final sodium citrate concentration was about 21 mM. When the measurement specimen was 8-fold dilute of 7 w/v % sodium hydrogencarbonate injection, the final sodium hydrogencarbonate concentration was about 52 mM. When the measurement specimen was 4-fold dilute of 10 w/v % sodium chloride injection, the final sodium chloride concentration was about 214 mM. When the measurement specimen was undiluted solution of physiological saline, the final sodium chloride concentration was about 77 mM. When the measurement specimen was 16-fold dilute of 0.5 M magnesium sulfate injection, the final magnesium sulfate concentration was about 16 mM. When the measurement specimen was 100-fold dilute of 0.5 M calcium chloride injection, the final calcium chloride concentration was about 2.5 mM. Activity of Factor C was measured through the procedure shown in FIG. 4. A residual activity was defined as a relative reactivity (%) for endotoxin in an injection specimen, with respect to the reactivity for endotoxin in the corresponding control specimen as 100%. The measurement was carried out four times for each injection specimen.

FIG. 7 shows the results. When a typical injection was employed as a measurement specimen, reaction with endotoxin was inhibited, as compared with the case in which injection water was used. The degree of inhibition varied in accordance with the type of injection and the type of host for expressing Factor C. Recombinant Factor C (CHO) and recombinant Factor C (HEK) showed a tendency to have low susceptibility to reaction inhibition to all the tested injections, as compared with native Factor C (TAL). Particularly, recombinant Factor C (CHO) exhibited the highest residual activity in many cases. In contrast, recombinant Factor C (Sf9) was most susceptible to reaction inhibition in many injections (7 injections except sample No. 6).

The test results indicate that Factor C expressed in mammalian cells can have low susceptibility to activity inhibition by ions. Such a property is particularly advantageous for detection of endotoxin contamination in production of injections.

6. Preparation of L Chain-Specific Factor C Protein Antibody

A Factor C protein is known to be present as a single chain (Full chain) under non-reducing conditions, and to be cleaved into two chains (H chain and L chain) under reducing conditions. The anti-Factor C monoclonal antibody (2C12 antibody) is reported to recognize the Full chain and the H chain (Yoshiki Miura, et al., J. Biochem. 112: 476-481 (1992)). Thus, a peptide antigen present in the L chain was produced. A rabbit was immunized with the peptide antigen through a conventional method, whereby a polyclonal antibody which can recognize the L chain and the Full chain (FCL antibody) was produced (FIG. 10).

7. Purification of Recombinant Factor C Proteins

According to the method employed in "(4) Preparation of native Factor C (derived from *Tachypleus tridentatus*)" of "1. Preparation of Factor C proteins", three types of recombinant Factor C proteins were purified from culture supernatants of mammalian cells (CHO DG44, and HEK293) and insect cells (Sf9). Briefly, each supernatant (100 to 300 mL) was subjected to column chromatography by use of dextran sulfate resin, and elution fractions containing the corresponding recombinant Factor C were isolated through gel filtration. Purification of the recombinant Factor C proteins

was confirmed through SDS-PAGE and CBB staining, western blotting employing the 2C12 antibody, and activity measurement.

8. Comparison of Molecular Weights of Factor C Proteins (2)

The molecular weights of the above-purified four types of Factor C proteins were compared again with one another through SDS-PAGE and western blotting.

The four types of purified Factor C proteins, derived from a native horseshoe crab (TAL), a culture supernatant of CHO DG44 (CHO), a culture supernatant of HEK293 (HEK), and a culture supernatant of Sf9 (Sf9), were applied to a single gel (15% gel) for SDS-PAGE, and subjected to CBB staining (FIG. 11). Each of the four types of purified Factor C proteins presented as a single chain (Full chain) under non-reducing conditions, and was cleaved into two chains (H chain and L chain) under reducing conditions. After transfer to the PVDF membrane, western blotting was performed by use of the 2C12 antibody and the FCL antibody (FIG. 12). In relation to antibody recognition sites (FIG. 10), the 2C12 antibody specifically recognized the Full chain and the H chain, and the FCL antibody specifically recognized the Full chain and the L chain. The CBB-stained bands shown in FIG. 11 were identified to bands derived from Factor C. Regarding molecular weight of Factor C, CHO had the largest molecular weight; TAL and HEK had almost the same molecular weight; and Sf9 has the smallest molecular weight, in all cases of the Full chain under non-reducing conditions and of the H-chain and L chain under reducing conditions.

In order to more precisely compare the molecular weights of the four types of purified Factor C, electrophoresis (15% gel, LMW marker (product of GE Healthcare)) was performed under the same conditions as employed in a known technique (Jeak Ling Ding and Bow Ho, U.S. Pat. No. 5,712,144; and Jeak L. Ding, et al., Biochem. Biophys. Acta 1202: 149-156), followed by CBB staining, and the molecular weights were then determined through Ferguson plotting.

Table 1 shows the results. Under non-reducing conditions, TAL had a molecular weight of 124 kDa, which is almost equivalent to 123 kDa reported by the reference. The molecular weight of CHO was higher, 128 kDa, and that of HEK was 124 kDa, which is equivalent to that of TAL. The molecular weight of Sf9 was 109 kDa, which is smaller than that of TAL. As a result, when recombinant Factor C was expressed in insect cells (Sf9), the molecular weight thereof was found to be smaller than that of native Factor C (TAL), whereas when recombinant Factor C was expressed in mammalian cells (CHO, HEK), the molecular weight thereof was found to be equivalent to or larger than that of native Factor C (TAL). This tendency was also confirmed in the case of the H chain or the L chain under reducing conditions.

The molecular weight of native Factor C derived from *Carcinoscorpius rotundicauda* (Singaporean horseshoe crab) and that of recombinant Factor C thereof obtained via gene expression in insect cells have been reported to be the same value, 132 kDa, under non-reducing conditions. The four types of purified Factor C derived from *Tachypleus tridentatus* (Japanese horseshoe crab) and obtained in the present Example were found to have a molecular weight smaller than 132 kDa (Table 1).

TABLE 1

Species of		Mol. wt. (kDa)			Reference
horseshoe crab	Source	Full	H	L	
<i>Tachypleus tridentatus</i>	Natural lysate	124	85	42	Present Example
<i>Tachypleus tridentatus</i>	CHO DG44	128	88	43	Present Example
<i>Tachypleus tridentatus</i>	HEK293	124	85	42	Present Example
<i>Tachypleus tridentatus</i>	Sf9	109	75	39	Present Example
<i>Tachypleus tridentatus</i>	Natural lysate	123	80	43	Takanori Nakamura, et al., Eur. J. Biochem. 154: 511-521 (1986)
<i>Carcinoscorpius rotundicauda</i>	Natural lysate	132	80	52	Jeak L. Ding, et al., Biochem. Biophys. Acta 1202: 149-156 (1993)
<i>Carcinoscorpius rotundicauda</i>	Sf9 (+baculo virus)	132	No data	No data	Jing Wang, et al., J. Biol. Chem. 277 (39): 36363-72 (2002)
<i>Carcinoscorpius rotundicauda</i>	S2	132	No data	No data	Jing Wang, et al., J. Biol. Chem. 277 (39): 36363-72 (2002)

9. Confirmation of N-Type Sugar Chain Modification

The genes employed in the present Example for expressing the recombinant Factor C were sequences all derived from *Tachypleus tridentatus*, and the expression plasmids were designed so that no tag was added to any terminus of each sequence. Therefore, the translated amino acid sequences of the four types of Factor C obtained in the present Example are thought to be the same one. In addition, the partial sequences of the N-terminal amino acid residues of the recombinant Factor C (HEK, Sf9) were determined. These amino acid sequences are identical to the sequence of the native Factor C of *Tachypleus tridentatus*, reported by the reference (data not shown).

Generally, the molecular weight of protein is thought to vary considerably by modification after translation. In addition, it has also been reported that the structure of an N-type sugar chain added to protein through modification after translation differs between the case of insect cells and the case of mammalian cells. Thus, in order to confirm whether the above-observed difference in molecular weight among the purified Factor C proteins is attributed to the difference in N-type sugar chain structure, the Factor C proteins each were treated with glycopeptidase F. This enzyme is known to cleave and remove an N-type sugar chain added to protein. Therefore, when the purified Factor C proteins have the same molecular weight after treatment with the enzyme, N-type sugar chain is conceived to induce the difference in molecular weight among the Factor C proteins.

According to an attached instruction, each purified Factor C protein (1 µg) was treated with 1 mU glycopeptidase F (product of Takara) under modification conditions in the presence of SDS and a reducing agent. The thus-treated samples were separated through SDS-PAGE, and then analyzed through western blotting. FIG. 13 shows the results. In all cases of the four types of Factor C, the molecular weight of H chain decreased by the glycopeptidase treatment and became the same value. By contrast, although the molecular weight of L chain decreased after the glycopeptidase treatment in all cases of the four types of Factor C, the molecular weight became the same value only in the cases of CHO, HEK, and TAL. Specifically, among four types of L chains, the L chain of the Factor C derived from Sf9 was found to have a molecular weight larger than that of the other three

cases, after the glycopeptidase treatment. Therefore, in the L chain of the Factor C derived from Sf9, a certain modification other than N-type sugar chain modification was conceived to occur, after translation.

Thus, the aforementioned analysis indicates that the difference in mode of N-type sugar chain after translation plays a main role in providing the difference in molecular weight of Factor C.

10. Analysis of N-Type Sugar Chain of Factor C

In N-type sugar chain modification of a protein in mammalian cells, sugar molecules are stepwise added to an asparagine residue of the protein, and eventually, a sugar chain containing terminal sialic acid (i.e., complex-type) is formed. In contrast, in insect cells, sialic acid is not generally added, and a sugar chain having a mannose terminus (i.e., paucimannose-type) is formed (Robert L. Harrison and Donald L. Jarvis, Methods in Molecular Biology 338: 341-356 (2007)). In order to confirm the difference in N-type sugar chain structure among the four types of purified Factor C proteins, lectin blotting was performed by use of various lectins such as terminal sialic acid-specific lectins (e.g., *Maackia amurensis* agglutinin (MAM) and *Sambucus sieboldiana* agglutinin (SSA)).

Dilution series (100, 25, and 6 ng) samples of each of the four types of purified Factor C and bovine fetuin protein as a positive control were dot-blotted onto a nitrocellulose membrane, and the membrane was blocked with 3% BSA. Then, the membrane was sequentially reacted with biotin-labeled lectin (product of J-Oil Mills, Inc.) and streptavidin-labeled HRP (product of Thermo Scientific). Light emission attributed to decomposition of the HRP substrate (product of Thermo Scientific) was detected. Instead of lectin, a 2C12 anti-Factor C antibody was used as a control, to thereby confirm that the same amounts of four types of Factor C were blotted.

FIG. 14(a) shows the results of lectin blotting, and FIG. 14(b) shows the binding specificity of lectins employed. *Lens culinaris* agglutinin (LCA) (fucose-specific lectin) and *Canavalia ensiformis* agglutinin (Con A) (bi-antenna-type, high-mannose-type, and complex-type sugar chain-specific lectin) were bonded to all the samples; i.e., four types of Factor C and fetuin. In contrast, MAM ((α-2,3)-linked terminal sialic acid-specific lectin) was strongly bonded to

CHO and less strongly bonded to HEK. TAL and fetuin have almost the same weak reactivity, but did not react with Sf9. SSA ((α -2,6)-linked terminal sialic acid-specific lectin) reacted with HEK, TAL, and fetuin, but did not react with CHO and Sf9.

The above analysis indicates that the N-type sugar chain of recombinant Factor C expressed in mammalian cells (CHO and HEK) contains (α -2,3)-linked terminal sialic acid, and that the N-type sugar chain of recombinant Factor C expressed in insect cells (Sf9) contains no (α -2,3)-linked terminal sialic acid. FIG. 14(c) shows schematic structures of the N-type sugar chain of recombinant Factor C, estimated by the results of the above experiments and reports by the reference. The results of the above experiments support that the difference in mode of N-type sugar chain modification after translation is mainly attributed to the difference in molecular weight of Factor C.

INDUSTRIAL APPLICABILITY

According to the present invention, a recombinant horseshoe crab Factor C can be efficiently produced. In an embodiment of the present invention, endotoxin present in a

test specimen containing an ion can be measured, while inhibition of reaction is reduced.

SEQUENCE LISTING

- 5 SEQ ID NO: 1: DNA sequence of the Factor C gene of Japanese horseshoe crab
 SEQ ID NO: 2: Amino acid sequence of the Factor C of Japanese horseshoe crab
 SEQ ID NO: 3: DNA sequence of the Factor C gene of Southeast Asian (Singaporean) horseshoe crab
 10 SEQ ID NO: 4: Amino acid sequence of the Factor C of Southeast Asian (Singaporean) horseshoe crab
 SEQ ID NO: 5: DNA sequence of the Factor B gene of Japanese horseshoe crab
 15 SEQ ID NO: 6: Amino acid sequence of the Factor B of Japanese horseshoe crab
 SEQ ID NO: 7: DNA sequence of the Pro-clotting enzyme gene of Japanese horseshoe crab
 SEQ ID NO: 8: Amino acid sequence of the Pro-clotting enzyme of Japanese horseshoe crab
 20 SEQ ID NO: 9: DNA sequence of a Factor B gene of Japanese horseshoe crab, the gene having a codon optimized for expression in insect cells
 SEQ ID NO: 10: Peptide sequence

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 10

<210> SEQ ID NO 1

<211> LENGTH: 3060

<212> TYPE: DNA

<213> ORGANISM: Tachypleus tridentatus

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (1) .. (3060)

<400> SEQUENCE: 1

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1 5 10 15	
gcc caa caa atg cgt cca gtt cag tcc aga gga gta gat ctg ggc ttg	96
Ala Gln Gln Met Arg Pro Val Gln Ser Arg Gly Val Asp Leu Gly Leu	
20 25 30	
tgt gat gaa acg agg ttc gag tgt aag tgt gga gat cca ggc tat gtg	144
Cys Asp Glu Thr Arg Phe Glu Cys Lys Cys Gly Asp Pro Gly Tyr Val	
35 40 45	
ttc aac gtc cct atg aaa caa tgc acg tac ttc tat cga tgg agg cct	192
Phe Asn Val Pro Met Lys Gln Cys Thr Tyr Phe Tyr Arg Trp Arg Pro	
50 55 60	
tat tgt aaa cca tgt gat gac ctg gag gct aag gac att tgt cca aag	240
Tyr Cys Lys Pro Cys Asp Asp Leu Glu Ala Lys Asp Ile Cys Pro Lys	
65 70 75 80	
tac aaa cga tgt caa gag tgt aag gct ggt ctt gat agt tgt gtt act	288
Tyr Lys Arg Cys Gln Glu Cys Lys Ala Gly Leu Asp Ser Cys Val Thr	
85 90 95	
tgt cca cct aac aaa tat ggt act tgg tgt agc ggt gaa tgt caa tgt	336
Cys Pro Pro Asn Lys Tyr Gly Thr Trp Cys Ser Gly Glu Cys Gln Cys	
100 105 110	
aag aat gga ggt atc tgt gac cag agg aca gga gct tgt acc tgt cgt	384
Lys Asn Gly Gly Ile Cys Asp Gln Arg Thr Gly Ala Cys Thr Cys Arg	
115 120 125	
gac aga tat gaa gga gcg cac tgt gaa att ctc aaa ggt tgt cct ctt	432
Asp Arg Tyr Glu Gly Ala His Cys Glu Ile Leu Lys Gly Cys Pro Leu	
130 135 140	

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ctt cca tcg gat tct caa gtt cag gaa gtc aga aac cca cca gat aat Leu Pro Ser Asp Ser Gln Val Gln Glu Val Arg Asn Pro Pro Asp Asn 145 150 155 160	480
ccc caa act att gac tac agc tgt tca cca ggg ttc aag ctt aaa ggc Pro Gln Thr Ile Asp Tyr Ser Cys Ser Pro Gly Phe Lys Leu Lys Gly 165 170 175	528
gtg gca cga att agc tgt ctc cca aat gga cag tgg agt agc ttt cca Val Ala Arg Ile Ser Cys Leu Pro Asn Gly Gln Trp Ser Ser Phe Pro 180 185 190	576
ccc aaa tgt att cga gaa tgt gcc aag gtt tca tct cca gaa cac ggg Pro Lys Cys Ile Arg Glu Cys Ala Lys Val Ser Ser Pro Glu His Gly 195 200 205	624
aaa gtg aat gct cct agt ggc aat atg ata gaa ggg gct act tta cgg Lys Val Asn Ala Pro Ser Gly Asn Met Ile Glu Gly Ala Thr Leu Arg 210 215 220	672
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gaa gtg acc tat acg tgt tcg ggt aac tac ttc ttg atg ggt ttt aac Glu Val Thr Tyr Thr Cys Ser Gly Asn Tyr Phe Leu Met Gly Phe Asn 290 295 300	912
acc tta aaa tgt aac cct gat ggg tcc tgg tca gga tca cag cca tcc Thr Leu Lys Cys Asn Pro Asp Gly Ser Trp Ser Gly Ser Gln Pro Ser 305 310 315 320	960
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gac ttc ttg gat gat gtt ggt gaa cct gtc agg atc cac tgt cct gct Asp Phe Leu Asp Asp Val Gly Glu Pro Val Arg Ile His Cys Pro Ala 340 345 350	1056
ggc tgt tct ttg aca gct ggt act gtg tgg ggt aca gcc ata tac cac Gly Cys Ser Leu Thr Ala Gly Thr Val Trp Gly Thr Ala Ile Tyr His 355 360 365	1104
gaa ctt tcc tca gtg tgt cgt gca gcc atc cat gct ggc aag ctt cca Glu Leu Ser Ser Val Cys Arg Ala Ala Ile His Ala Gly Lys Leu Pro 370 375 380	1152
aac tct gga ggg gcg gtg cat gta gtg aac aat ggc ccc tac tcg gac Asn Ser Gly Gly Ala Val His Val Val Asn Asn Gly Pro Tyr Ser Asp 385 390 395 400	1200
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ctt gcc cgc agt ttt cga ttt gat tat gtc agt tca tcc aca gca ggt Leu Ala Arg Ser Phe Arg Phe Asp Tyr Val Ser Ser Ser Thr Ala Gly 420 425 430	1296
aga tca gga tgt cct gat gga tgg ttt gag gta gaa gag aac tgt gtg Arg Ser Gly Cys Pro Asp Gly Trp Phe Glu Val Glu Glu Asn Cys Val 435 440 445	1344
tac gtt aca tca aaa cag aga gcc tgg gaa aga gct caa ggt gtg tgt Tyr Val Thr Ser Lys Gln Arg Ala Trp Glu Arg Ala Gln Gly Val Cys	1392

-continued

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Thr Asn Met Ala Ala Arg Leu Ala Val Leu Asp Lys Asp Leu Ile Pro			
465	470	475	480
agt tcc ttg act gag act cta cga ggg aaa ggg tta aca acc aca tgg			1488
Ser Ser Leu Thr Glu Thr Leu Arg Gly Lys Gly Leu Thr Thr Trp			
	485	490	495
ata gga ttg cac aga cta gat gct gag aag ccc ttt gtt tgg gag cta			1536
Ile Gly Leu His Arg Leu Asp Ala Glu Lys Pro Phe Val Trp Glu Leu			
	500	505	510
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Met Asp Arg Ser Asn Val Val Leu Asn Asp Asn Leu Thr Phe Trp Ala			
	515	520	525
tct ggc gaa cct gga aat gaa act aac tgt gta tat ctg gac atc cga			1632
Ser Gly Glu Pro Gly Asn Glu Thr Asn Cys Val Tyr Leu Asp Ile Arg			
	530	535	540
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Asp Gln Leu Gln Pro Val Trp Lys Thr Lys Ser Cys Phe Gln Pro Ser			
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Ser Phe Ala Cys Met Met Asp Leu Ser Asp Arg Asn Lys Ala Lys Cys			
	565	570	575
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Ser Ile Asp Gly Phe Tyr Ala Gly Ser Ser Ile Arg Tyr Ser Cys Glu			
	595	600	605
gtt ctc cac tac ctc agt gga act gag acc gta act tgt aca aca aat			1872
Val Leu His Tyr Leu Ser Gly Thr Glu Thr Val Thr Cys Thr Thr Asn			
	610	615	620
ggc aca tgg agt gct cct aaa cct cga tgt atc aaa gtc atc acc tgc			1920
Gly Thr Trp Ser Ala Pro Lys Pro Arg Cys Ile Lys Val Ile Thr Cys			
	625	630	635
caa aac cct cct gta cca tca tat ggt tct gtg gaa atc aaa ccc cca			1968
Gln Asn Pro Pro Val Pro Ser Tyr Gly Ser Val Glu Ile Lys Pro Pro			
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agt cgg aca aac tcg atc agt cgt gtt ggg tca cct ttc ttg agg ttg			2016
Ser Arg Thr Asn Ser Ile Ser Arg Val Gly Ser Pro Phe Leu Arg Leu			
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cca cgg tta ccc ctc cca tta gcc aga gca gcc aaa cct cct cca aaa			2064
Pro Arg Leu Pro Leu Pro Leu Ala Arg Ala Ala Lys Pro Pro Pro Lys			
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Pro Arg Ser Ser Gln Pro Ser Thr Val Asp Leu Ala Ser Lys Val Lys			
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cta cct gaa ggt cat tac cgg gta ggg tct cga gcc att tac acg tgc			2160
Leu Pro Glu Gly His Tyr Arg Val Gly Ser Arg Ala Ile Tyr Thr Cys			
	705	710	715
gag tcg aga tac tac gaa cta ctt gga tct caa ggc aga aga tgt gac			2208
Glu Ser Arg Tyr Tyr Glu Leu Leu Gly Ser Gln Gly Arg Arg Cys Asp			
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Ser Asn Gly Asn Trp Ser Gly Arg Pro Ala Ser Cys Ile Pro Val Cys			
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Gly Arg Ser Asp Ser Pro Arg Ser Pro Phe Ile Trp Asn Gly Asn Ser			
	755	760	765
aca gaa ata ggt cag tgg cgg tgg cag gca gga atc tct cga tgg ctt			2352

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Ala	Asp	His	Asn	Met	Trp	Phe	Leu	Gln	Cys	Gly	Gly	Ser	Leu	Leu	Asn	
785					790					795					800	
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Glu	Lys	Trp	Ile	Val	Thr	Ala	Ala	His	Cys	Val	Thr	Tyr	Ser	Ala	Thr	
			805						810					815		
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Ala	Glu	Ile	Ile	Asp	Pro	Ser	Gln	Phe	Lys	Ile	Tyr	Leu	Gly	Lys	Tyr	
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Tyr	Arg	Asp	Asp	Ser	Arg	Asp	Asp	Asp	Tyr	Val	Gln	Val	Arg	Glu	Ala	
		835					840					845				
ctc	gag	atc	cac	gta	aat	cct	aac	tac	gac	ccc	ggc	aat	ctc	aac	ttt	2592
Leu	Glu	Ile	His	Val	Asn	Pro	Asn	Tyr	Asp	Pro	Gly	Asn	Leu	Asn	Phe	
		850				855					860					
gac	ata	gcc	cta	att	caa	ctg	aaa	act	cct	gtt	act	ttg	aca	aca	cga	2640
Asp	Ile	Ala	Leu	Ile	Gln	Leu	Lys	Thr	Pro	Val	Thr	Leu	Thr	Thr	Arg	
865					870					875					880	
gtc	caa	cca	atc	tgt	ctg	cct	act	gac	atc	aca	aca	aga	gaa	cac	ttg	2688
Val	Gln	Pro	Ile	Cys	Leu	Pro	Thr	Asp	Ile	Thr	Thr	Arg	Glu	His	Leu	
			885						890					895		
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Lys	Glu	Gly	Thr	Leu	Ala	Val	Val	Thr	Gly	Trp	Gly	Leu	Asn	Glu	Asn	
			900					905					910			
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Asn	Thr	Tyr	Ser	Glu	Met	Ile	Gln	Gln	Ala	Val	Leu	Pro	Val	Val	Ala	
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Ala	Ser	Thr	Cys	Glu	Glu	Gly	Tyr	Lys	Glu	Ala	Asp	Leu	Pro	Leu	Thr	
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			945		950				955						960	
gcc	tgc	agt	ggg	gac	agt	gga	gga	cca	tta	gtg	ttt	gct	gat	gat	tcc	2928
Ala	Cys	Ser	Gly	Asp	Ser	Gly	Gly	Pro	Leu	Val	Phe	Ala	Asp	Asp	Ser	
			965					970						975		
cgt	acc	gaa	agg	cgg	tgg	gtc	ttg	gaa	ggg	att	gtc	agc	tgg	ggc	agt	2976
Arg	Thr	Glu	Arg	Arg	Trp	Val	Leu	Glu	Gly	Ile	Val	Ser	Trp	Gly	Ser	
			980					985					990			
ccc	agt	gga	tgt	ggc	aag	gct	aac	cag	tat	ggg	ggc	ttc	act	aaa	gtt	3024
Pro	Ser	Gly	Cys	Gly	Lys	Ala	Asn	Gln	Tyr	Gly	Gly	Phe	Thr	Lys	Val	
		995					1000					1005				
aac	gtt	ttt	cta	tca	tgg	att	agg	cag	ttc	att	tga					3060
Asn	Val	Phe	Leu	Ser	Trp	Ile	Arg	Gln	Phe	Ile						
		1010				1015										

<210> SEQ ID NO 2

<211> LENGTH: 1019

<212> TYPE: PRT

<213> ORGANISM: Tachyplesus tridentatus

<400> SEQUENCE: 2

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1				5					10					15	

Ala	Gln	Gln	Met	Arg	Pro	Val	Gln	Ser	Arg	Gly	Val	Asp	Leu	Gly	Leu
			20					25					30		

Cys	Asp	Glu	Thr	Arg	Phe	Glu	Cys	Lys	Cys	Gly	Asp	Pro	Gly	Tyr	Val
		35					40					45			

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Phe	Asn	Val	Pro	Met	Lys	Gln	Cys	Thr	Tyr	Phe	Tyr	Arg	Trp	Arg	Pro
50						55						60			
Tyr	Cys	Lys	Pro	Cys	Asp	Asp	Leu	Glu	Ala	Lys	Asp	Ile	Cys	Pro	Lys
65					70					75					80
Tyr	Lys	Arg	Cys	Gln	Glu	Cys	Lys	Ala	Gly	Leu	Asp	Ser	Cys	Val	Thr
				85					90					95	
Cys	Pro	Pro	Asn	Lys	Tyr	Gly	Thr	Trp	Cys	Ser	Gly	Glu	Cys	Gln	Cys
			100					105					110		
Lys	Asn	Gly	Gly	Ile	Cys	Asp	Gln	Arg	Thr	Gly	Ala	Cys	Thr	Cys	Arg
		115					120					125			
Asp	Arg	Tyr	Glu	Gly	Ala	His	Cys	Glu	Ile	Leu	Lys	Gly	Cys	Pro	Leu
		130				135						140			
Leu	Pro	Ser	Asp	Ser	Gln	Val	Gln	Glu	Val	Arg	Asn	Pro	Pro	Asp	Asn
145					150					155					160
Pro	Gln	Thr	Ile	Asp	Tyr	Ser	Cys	Ser	Pro	Gly	Phe	Lys	Leu	Lys	Gly
				165					170					175	
Val	Ala	Arg	Ile	Ser	Cys	Leu	Pro	Asn	Gly	Gln	Trp	Ser	Ser	Phe	Pro
				180				185						190	
Pro	Lys	Cys	Ile	Arg	Glu	Cys	Ala	Lys	Val	Ser	Ser	Pro	Glu	His	Gly
		195					200					205			
Lys	Val	Asn	Ala	Pro	Ser	Gly	Asn	Met	Ile	Glu	Gly	Ala	Thr	Leu	Arg
		210				215						220			
Phe	Ser	Cys	Asp	Ser	Pro	Tyr	Tyr	Leu	Ile	Gly	Gln	Glu	Thr	Leu	Thr
225					230					235					240
Cys	Gln	Gly	Asn	Gly	Gln	Trp	Ser	Gly	Gln	Ile	Pro	Gln	Cys	Lys	Lys
				245					250					255	
Leu	Val	Phe	Cys	Pro	Asp	Leu	Asp	Pro	Val	Asn	His	Ala	Glu	His	Gln
			260					265					270		
Val	Lys	Ile	Gly	Val	Glu	Gln	Lys	Tyr	Gly	Gln	Phe	Pro	Gln	Gly	Thr
			275				280						285		
Glu	Val	Thr	Tyr	Thr	Cys	Ser	Gly	Asn	Tyr	Phe	Leu	Met	Gly	Phe	Asn
		290				295					300				
Thr	Leu	Lys	Cys	Asn	Pro	Asp	Gly	Ser	Trp	Ser	Gly	Ser	Gln	Pro	Ser
305					310					315					320
Cys	Val	Lys	Val	Ala	Asp	Arg	Glu	Val	Asp	Cys	Asp	Ser	Lys	Ala	Val
				325					330					335	
Asp	Phe	Leu	Asp	Asp	Val	Gly	Glu	Pro	Val	Arg	Ile	His	Cys	Pro	Ala
			340					345					350		
Gly	Cys	Ser	Leu	Thr	Ala	Gly	Thr	Val	Trp	Gly	Thr	Ala	Ile	Tyr	His
		355				360						365			
Glu	Leu	Ser	Ser	Val	Cys	Arg	Ala	Ala	Ile	His	Ala	Gly	Lys	Leu	Pro
		370				375					380				
Asn	Ser	Gly	Gly	Ala	Val	His	Val	Val	Asn	Asn	Gly	Pro	Tyr	Ser	Asp
385					390					395					400
Phe	Leu	Gly	Ser	Asp	Leu	Asn	Gly	Ile	Lys	Ser	Glu	Glu	Leu	Lys	Ser
				405					410					415	
Leu	Ala	Arg	Ser	Phe	Arg	Phe	Asp	Tyr	Val	Ser	Ser	Ser	Thr	Ala	Gly
				420				425						430	
Arg	Ser	Gly	Cys	Pro	Asp	Gly	Trp	Phe	Glu	Val	Glu	Glu	Asn	Cys	Val
			435				440					445			
Tyr	Val	Thr	Ser	Lys	Gln	Arg	Ala	Trp	Glu	Arg	Ala	Gln	Gly	Val	Cys
					455						460				

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Thr	Asn	Met	Ala	Ala	Arg	Leu	Ala	Val	Leu	Asp	Lys	Asp	Leu	Ile	Pro	465	470	475	480
Ser	Ser	Leu	Thr	Glu	Thr	Leu	Arg	Gly	Lys	Gly	Leu	Thr	Thr	Thr	Trp	485	490	495	
Ile	Gly	Leu	His	Arg	Leu	Asp	Ala	Glu	Lys	Pro	Phe	Val	Trp	Glu	Leu	500	505	510	
Met	Asp	Arg	Ser	Asn	Val	Val	Leu	Asn	Asp	Asn	Leu	Thr	Phe	Trp	Ala	515	520	525	
Ser	Gly	Glu	Pro	Gly	Asn	Glu	Thr	Asn	Cys	Val	Tyr	Leu	Asp	Ile	Arg	530	535	540	
Asp	Gln	Leu	Gln	Pro	Val	Trp	Lys	Thr	Lys	Ser	Cys	Phe	Gln	Pro	Ser	545	550	555	560
Ser	Phe	Ala	Cys	Met	Met	Asp	Leu	Ser	Asp	Arg	Asn	Lys	Ala	Lys	Cys	565	570	575	
Asp	Asp	Pro	Gly	Pro	Leu	Glu	Asn	Gly	His	Ala	Thr	Leu	His	Gly	Gln	580	585	590	
Ser	Ile	Asp	Gly	Phe	Tyr	Ala	Gly	Ser	Ser	Ile	Arg	Tyr	Ser	Cys	Glu	595	600	605	
Val	Leu	His	Tyr	Leu	Ser	Gly	Thr	Glu	Thr	Val	Thr	Cys	Thr	Thr	Asn	610	615	620	
Gly	Thr	Trp	Ser	Ala	Pro	Lys	Pro	Arg	Cys	Ile	Lys	Val	Ile	Thr	Cys	625	630	635	640
Gln	Asn	Pro	Pro	Val	Pro	Ser	Tyr	Gly	Ser	Val	Glu	Ile	Lys	Pro	Pro	645	650	655	
Ser	Arg	Thr	Asn	Ser	Ile	Ser	Arg	Val	Gly	Ser	Pro	Phe	Leu	Arg	Leu	660	665	670	
Pro	Arg	Leu	Pro	Leu	Pro	Leu	Ala	Arg	Ala	Ala	Lys	Pro	Pro	Pro	Lys	675	680	685	
Pro	Arg	Ser	Ser	Gln	Pro	Ser	Thr	Val	Asp	Leu	Ala	Ser	Lys	Val	Lys	690	695	700	
Leu	Pro	Glu	Gly	His	Tyr	Arg	Val	Gly	Ser	Arg	Ala	Ile	Tyr	Thr	Cys	705	710	715	720
Glu	Ser	Arg	Tyr	Tyr	Glu	Leu	Leu	Gly	Ser	Gln	Gly	Arg	Arg	Cys	Asp	725	730	735	
Ser	Asn	Gly	Asn	Trp	Ser	Gly	Arg	Pro	Ala	Ser	Cys	Ile	Pro	Val	Cys	740	745	750	
Gly	Arg	Ser	Asp	Ser	Pro	Arg	Ser	Pro	Phe	Ile	Trp	Asn	Gly	Asn	Ser	755	760	765	
Thr	Glu	Ile	Gly	Gln	Trp	Pro	Trp	Gln	Ala	Gly	Ile	Ser	Arg	Trp	Leu	770	775	780	
Ala	Asp	His	Asn	Met	Trp	Phe	Leu	Gln	Cys	Gly	Gly	Ser	Leu	Leu	Asn	785	790	795	800
Glu	Lys	Trp	Ile	Val	Thr	Ala	Ala	His	Cys	Val	Thr	Tyr	Ser	Ala	Thr	805	810	815	
Ala	Glu	Ile	Ile	Asp	Pro	Ser	Gln	Phe	Lys	Ile	Tyr	Leu	Gly	Lys	Tyr	820	825	830	
Tyr	Arg	Asp	Asp	Ser	Arg	Asp	Asp	Asp	Tyr	Val	Gln	Val	Arg	Glu	Ala	835	840	845	
Leu	Glu	Ile	His	Val	Asn	Pro	Asn	Tyr	Asp	Pro	Gly	Asn	Leu	Asn	Phe	850	855	860	
Asp	Ile	Ala	Leu	Ile	Gln	Leu	Lys	Thr	Pro	Val	Thr	Leu	Thr	Thr	Arg	865	870	875	880
Val	Gln	Pro	Ile	Cys	Leu	Pro	Thr	Asp	Ile	Thr	Thr	Arg	Glu	His	Leu				

[illegible]

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165	170	175	
atg gca cga att agc tgt ctc cca aat gga cag tgg agt aac ttt cca Met Ala Arg Ile Ser Cys Leu Pro Asn Gly Gln Trp Ser Asn Phe Pro 180 185 190			576
ccc aaa tgt att cga gaa tgt gcc atg gtt tca tct cca gaa cat ggg Pro Lys Cys Ile Arg Glu Cys Ala Met Val Ser Ser Pro Glu His Gly 195 200 205			624
aaa gtg aat gct ctt agt ggt gat atg ata gaa ggg gct act tta cgg Lys Val Asn Ala Leu Ser Gly Asp Met Ile Glu Gly Ala Thr Leu Arg 210 215 220			672
ttc tca tgt gat agt ccc tac tac ttg att ggt caa gaa aca tta acc Phe Ser Cys Asp Ser Pro Tyr Tyr Leu Ile Gly Gln Glu Thr Leu Thr 225 230 235 240			720
tgt cag ggt aat ggt cag tgg aat gga cag ata cca caa tgt aag aac Cys Gln Gly Asn Gly Gln Trp Asn Gly Gln Ile Pro Gln Cys Lys Asn 245 250 255			768
ttg gtc ttc tgt cct gac ctg gat cct gta aac cat gct gaa cac aag Leu Val Phe Cys Pro Asp Leu Asp Pro Val Asn His Ala Glu His Lys 260 265 270			816
gtt aaa att ggt gtg gaa caa aaa tat ggt cag ttt cct caa ggc act Val Lys Ile Gly Val Glu Gln Lys Tyr Gly Gln Phe Pro Gln Gly Thr 275 280 285			864
gaa gtg acc tat acg tgt tcg ggt aac tac ttc ttg atg ggt ttt gac Glu Val Thr Tyr Thr Cys Ser Gly Asn Tyr Phe Leu Met Gly Phe Asp 290 295 300			912
acc tta aaa tgt aac cct gat ggg tct tgg tca gga tca cag cca tcc Thr Leu Lys Cys Asn Pro Asp Gly Ser Trp Ser Gly Ser Gln Pro Ser 305 310 315 320			960
tgt gtt aaa gtg gca gac aga gag gtc gac tgt gac agt aaa gct gta Cys Val Lys Val Ala Asp Arg Glu Val Asp Cys Asp Ser Lys Ala Val 325 330 335			1008
gac ttc ttg gat gat gtt ggt gaa cct gtc agg atc cac tgt cct gct Asp Phe Leu Asp Asp Val Gly Glu Pro Val Arg Ile His Cys Pro Ala 340 345 350			1056
ggc tgt tct ttg aca gct ggt act gtg tgg ggt aca gcc ata tac cat Gly Cys Ser Leu Thr Ala Gly Thr Val Trp Gly Thr Ala Ile Tyr His 355 360 365			1104
gaa ctt tcc tca gtg tgt cgt gca gcc atc cat gct ggc aag ctt cca Glu Leu Ser Ser Val Cys Arg Ala Ala Ile His Ala Gly Lys Leu Pro 370 375 380			1152
aac tct gga gga gcg gtg cat gtt gtg aac aat ggc ccc tac tcg gac Asn Ser Gly Gly Ala Val His Val Val Asn Asn Gly Pro Tyr Ser Asp 385 390 395 400			1200
ttt ctg ggt agt gac ctg aat ggg ata aaa tcg gaa gag ttg aag tct Phe Leu Gly Ser Asp Leu Asn Gly Ile Lys Ser Glu Glu Leu Lys Ser 405 410 415			1248
ctt gcc cgg agt ttc cga ttc gat tat gtc cgt tcc tcc aca gca ggt Leu Ala Arg Ser Phe Arg Phe Asp Tyr Val Arg Ser Ser Thr Ala Gly 420 425 430			1296
aaa tca gga tgt cct gat gga tgg ttt gag gta gac gag aac tgt gtg Lys Ser Gly Cys Pro Asp Gly Trp Phe Glu Val Asp Glu Asn Cys Val 435 440 445			1344
tac gtt aca tca aaa cag aga gcc tgg gaa aga gct caa ggt gtg tgt Tyr Val Thr Ser Lys Gln Arg Ala Trp Glu Arg Ala Gln Gly Val Cys 450 455 460			1392
acc aat atg gct gct cgt ctt gct gtg ctg gac aaa gat gta att cca Thr Asn Met Ala Ala Arg Leu Ala Val Leu Asp Lys Asp Val Ile Pro 465 470 475 480			1440
aat tcg ttg act gag act cta cga ggg aaa ggg tta aca acc acg tgg			1488

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Asn	Ser	Leu	Thr	Glu	Thr	Leu	Arg	Gly	Lys	Gly	Leu	Thr	Thr	Thr	Trp	
				485					490						495	
ata	gga	ttg	cac	aga	cta	gat	gct	gag	aag	ccc	ttt	att	tg	gag	tta	1536
Ile	Gly	Leu	His	Arg	Leu	Asp	Ala	Glu	Lys	Pro	Phe	Ile	Trp	Glu	Leu	
			500					505					510			
atg	gat	cgt	agt	aat	gtg	gtt	ctg	aat	gat	aac	cta	aca	ttc	tg	gcc	1584
Met	Asp	Arg	Ser	Asn	Val	Val	Leu	Asn	Asp	Asn	Leu	Thr	Phe	Trp	Ala	
		515					520					525				
tct	ggc	gaa	cct	gga	aat	gaa	act	aac	tgt	gta	tat	atg	gac	atc	caa	1632
Ser	Gly	Glu	Pro	Gly	Asn	Glu	Thr	Asn	Cys	Val	Tyr	Met	Asp	Ile	Gln	
	530				535					540						
gat	cag	ttg	cag	tct	gtg	tg	aaa	acc	aag	tca	tgt	ttt	cag	ccc	tca	1680
Asp	Gln	Leu	Gln	Ser	Val	Trp	Lys	Thr	Lys	Ser	Cys	Phe	Gln	Pro	Ser	
545				550					555						560	
agt	ttt	gct	tg	atg	atg	gat	ctg	tca	gac	aga	aat	aaa	gcc	aaa	tg	1728
Ser	Phe	Ala	Cys	Met	Met	Asp	Leu	Ser	Asp	Arg	Asn	Lys	Ala	Lys	Cys	
			565						570				575			
gat	gat	cct	gga	tca	ctg	gaa	aat	gga	cac	gcc	aca	ctt	cat	gga	caa	1776
Asp	Asp	Pro	Gly	Ser	Leu	Glu	Asn	Gly	His	Ala	Thr	Leu	His	Gly	Gln	
			580				585					590				
agt	att	gat	ggg	ttc	tat	gct	gg	tct	tct	ata	agg	tac	ag	tg	gag	1824
Ser	Ile	Asp	Gly	Phe	Tyr	Ala	Gly	Ser	Ser	Ile	Arg	Tyr	Ser	Cys	Glu	
	595					600						605				
gtt	ctc	cac	tac	ctc	agt	gga	act	gaa	acc	gta	act	tgt	aca	aca	aat	1872
Val	Leu	His	Tyr	Leu	Ser	Gly	Thr	Glu	Thr	Val	Thr	Cys	Thr	Thr	Asn	
	610					615					620					
ggc	aca	tg	agt	gct	cct	aaa	cct	cga	tgt	atc	aaa	gtc	atc	acc	tg	1920
Gly	Thr	Trp	Ser	Ala	Pro	Lys	Pro	Arg	Cys	Ile	Lys	Val	Ile	Thr	Cys	
625				630					635						640	
caa	aac	ccc	cct	gta	cca	tca	tat	gg	tct	gtg	gaa	atc	aaa	ccc	cca	1968
Gln	Asn	Pro	Pro	Val	Pro	Ser	Tyr	Gly	Ser	Val	Glu	Ile	Lys	Pro	Pro	
			645					650					655			
agt	cgg	aca	aac	tcg	ata	agt	cgt	gtt	ggg	tca	cct	ttc	ttg	agg	ttg	2016
Ser	Arg	Thr	Asn	Ser	Ile	Ser	Arg	Val	Gly	Ser	Pro	Phe	Leu	Arg	Leu	
			660				665					670				
cca	cgg	tta	ccc	ctc	cca	tta	gct	aga	gca	gcc	aaa	cct	cct	cca	aaa	2064
Pro	Arg	Leu	Pro	Leu	Pro	Leu	Ala	Arg	Ala	Ala	Lys	Pro	Pro	Pro	Lys	
		675				680						685				
cct	aga	tcc	tca	caa	ccc	tct	act	gtg	gac	ttg	gct	tct	aaa	gtt	aaa	2112
Pro	Arg	Ser	Ser	Gln	Pro	Ser	Thr	Val	Asp	Leu	Ala	Ser	Lys	Val	Lys	
	690					695					700					
cta	cct	gaa	gg	cat	tac	cgg	gta	ggg	tct	cga	gcc	atc	tac	acg	tg	2160
Leu	Pro	Glu	Gly	His	Tyr	Arg	Val	Gly	Ser	Arg	Ala	Ile	Tyr	Thr	Cys	
	705			710					715					720		
gag	tcg	aga	tac	tac	gaa	cta	ctt	gga	tct	caa	ggc	aga	aga	tg	gac	2208
Glu	Ser	Arg	Tyr	Tyr	Glu	Leu	Leu	Gly	Ser	Gln	Gly	Arg	Arg	Cys	Asp	
			725						730					735		
tct	aat	gga	aac	tg	agt	gg	cgg	cca	gcg	agc	tgt	att	cca	gtt	tg	2256
Ser	Asn	Gly	Asn	Trp	Ser	Gly	Arg	Pro	Ala	Ser	Cys	Ile	Pro	Val	Cys	
			740				745						750			
gga	cgg	tca	gac	tct	cct	cgt	tct	cct	ttt	atc	tg	aat	ggg	aat	tct	2304
Gly	Arg	Ser	Asp	Ser	Pro	Arg	Ser	Pro	Phe	Ile	Trp	Asn	Gly	Asn	Ser	
		755				760						765				
aca	gaa	ata	gg	cag	tg	cgg	tg	cag	gca	gga	atc	tct	aga	tg	ctt	2352
Thr	Glu	Ile	Gly	Gln	Trp	Pro	Trp	Gln	Ala	Gly	Ile	Ser	Arg	Trp	Leu	
		770				775					780					
gca	gac	cac	aat	atg	tg	ttt	ctc	cag	tgt	gga	gga	tct	cta	ttg	aat	2400
Ala	Asp	His	Asn	Met	Trp	Phe	Leu	Gln	Cys	Gly	Gly	Ser	Leu	Leu	Asn	
	785				790					795					800	

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gag aaa tgg atc gtc act gct gcc cac tgt gtc acc tac tct gct act Glu Lys Trp Ile Val Thr Ala Ala His Cys Val Thr Tyr Ser Ala Thr 805 810 815	2448
gct gag att att gac ccc aat cag ttt aaa atg tat ctg ggc aag tac Ala Glu Ile Ile Asp Pro Asn Gln Phe Lys Met Tyr Leu Gly Lys Tyr 820 825 830	2496
tac cgt gat gac agt aga gac gat gac tat gta caa gta aga gag gct Tyr Arg Asp Asp Ser Arg Asp Asp Asp Tyr Val Gln Val Arg Glu Ala 835 840 845	2544
ctt gag atc cac gtg aat cct aac tac gac ccc ggc aat ctc aac ttt Leu Glu Ile His Val Asn Pro Asn Tyr Asp Pro Gly Asn Leu Asn Phe 850 855 860	2592
gac ata gcc cta att caa ctg aaa act cct gtt act ttg aca aca cga Asp Ile Ala Leu Ile Gln Leu Lys Thr Pro Val Thr Leu Thr Thr Arg 865 870 875 880	2640
gtc caa cca atc tgt ctg cct act gac atc aca aca aga gaa cac ttg Val Gln Pro Ile Cys Leu Pro Thr Asp Ile Thr Thr Arg Glu His Leu 885 890 895	2688
aag gag gga aca tta gca gtg gtg aca ggt tgg ggt ttg aat gaa aac Lys Glu Gly Thr Leu Ala Val Val Thr Gly Trp Gly Leu Asn Glu Asn 900 905 910	2736
aac acc tat tca gag acg att caa caa gct gtg cta cct gtt gtt gca Asn Thr Tyr Ser Glu Thr Ile Gln Gln Ala Val Leu Pro Val Val Ala 915 920 925	2784
gcc agc acc tgt gaa gag ggg tac aag gaa gca gac tta cca ctg aca Ala Ser Thr Cys Glu Glu Gly Tyr Lys Glu Ala Asp Leu Pro Leu Thr 930 935 940	2832
gta aca gag aac atg ttc tgt gca ggt tac aag aag gga cgt tat gat Val Thr Glu Asn Met Phe Cys Ala Gly Tyr Lys Lys Gly Arg Tyr Asp 945 950 955 960	2880
gcc tgc agt ggg gac agt gga gga cct tta gtg ttt gct gat gat tcc Ala Cys Ser Gly Asp Ser Gly Gly Pro Leu Val Phe Ala Asp Asp Ser 965 970 975	2928
cgt acc gaa agg cgg tgg gtc ttg gaa ggg att gtc agc tgg ggc agt Arg Thr Glu Arg Arg Trp Val Leu Glu Gly Ile Val Ser Trp Gly Ser 980 985 990	2976
ccc agt gga tgt ggc aag gcg aac cag tac ggg ggc ttc act aaa gtt Pro Ser Gly Cys Gly Lys Ala Asn Gln Tyr Gly Gly Phe Thr Lys Val 995 1000 1005	3024
aac gtt ttc ctg tca tgg att agg cag ttc att tga Asn Val Phe Leu Ser Trp Ile Arg Gln Phe Ile 1010 1015	3060

<210> SEQ ID NO 4

<211> LENGTH: 1019

<212> TYPE: PRT

<213> ORGANISM: Carcinoscorpius rotundicauda

<400> SEQUENCE: 4

Met Val Leu Ala Ser Phe Leu Val Ser Gly Leu Val Leu Gly Leu Leu 1 5 10 15
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Cys Asp Glu Thr Arg Phe Glu Cys Lys Cys Gly Asp Pro Gly Tyr Val 35 40 45
Phe Asn Ile Pro Val Lys Gln Cys Thr Tyr Phe Tyr Arg Trp Arg Pro 50 55 60
Tyr Cys Lys Pro Cys Asp Leu Glu Ala Lys Asp Ile Cys Pro Lys 65 70 75 80

Tyr	Lys	Arg	Cys	Gln	Glu	Cys	Lys	Ala	Gly	Leu	Asp	Ser	Cys	Val	Thr	
				85					90					95		
Cys	Pro	Pro	Asn	Lys	Tyr	Gly	Thr	Trp	Cys	Ser	Gly	Glu	Cys	Gln	Cys	
			100					105					110			
Lys	Asn	Gly	Gly	Ile	Cys	Asp	Gln	Arg	Thr	Gly	Ala	Cys	Ala	Cys	Arg	
		115					120					125				
Asp	Arg	Tyr	Glu	Gly	Val	His	Cys	Glu	Ile	Leu	Lys	Gly	Cys	Pro	Leu	
		130				135					140					
Leu	Pro	Ser	Asp	Ser	Gln	Val	Gln	Glu	Val	Arg	Asn	Pro	Pro	Asp	Asn	
					150					155					160	
Pro	Gln	Thr	Ile	Asp	Tyr	Ser	Cys	Ser	Pro	Gly	Phe	Lys	Leu	Lys	Gly	
			165						170					175		
Met	Ala	Arg	Ile	Ser	Cys	Leu	Pro	Asn	Gly	Gln	Trp	Ser	Asn	Phe	Pro	
			180					185					190			
Pro	Lys	Cys	Ile	Arg	Glu	Cys	Ala	Met	Val	Ser	Ser	Pro	Glu	His	Gly	
		195					200					205				
Lys	Val	Asn	Ala	Leu	Ser	Gly	Asp	Met	Ile	Glu	Gly	Ala	Thr	Leu	Arg	
						215					220					
Phe	Ser	Cys	Asp	Ser	Pro	Tyr	Tyr	Leu	Ile	Gly	Gln	Glu	Thr	Leu	Thr	
					230					235					240	
Cys	Gln	Gly	Asn	Gly	Gln	Trp	Asn	Gly	Gln	Ile	Pro	Gln	Cys	Lys	Asn	
			245					250						255		
Leu	Val	Phe	Cys	Pro	Asp	Leu	Asp	Pro	Val	Asn	His	Ala	Glu	His	Lys	
			260					265					270			
Val	Lys	Ile	Gly	Val	Glu	Gln	Lys	Tyr	Gly	Gln	Phe	Pro	Gln	Gly	Thr	
		275					280					285				
Glu	Val	Thr	Tyr	Thr	Cys	Ser	Gly	Asn	Tyr	Phe	Leu	Met	Gly	Phe	Asp	
		290				295					300					
Thr	Leu	Lys	Cys	Asn	Pro	Asp	Gly	Ser	Trp	Ser	Gly	Ser	Gln	Pro	Ser	
					310					315					320	
Cys	Val	Lys	Val	Ala	Asp	Arg	Glu	Val	Asp	Cys	Asp	Ser	Lys	Ala	Val	
			325						330					335		
Asp	Phe	Leu	Asp	Asp	Val	Gly	Glu	Pro	Val	Arg	Ile	His	Cys	Pro	Ala	
			340					345					350			
Gly	Cys	Ser	Leu	Thr	Ala	Gly	Thr	Val	Trp	Gly	Thr	Ala	Ile	Tyr	His	
		355					360					365				
Glu	Leu	Ser	Ser	Val	Cys	Arg	Ala	Ala	Ile	His	Ala	Gly	Lys	Leu	Pro	
						375					380					
Asn	Ser	Gly	Gly	Ala	Val	His	Val	Val	Asn	Asn	Gly	Pro	Tyr	Ser	Asp	
					390					395					400	
Phe	Leu	Gly	Ser	Asp	Leu	Asn	Gly	Ile	Lys	Ser	Glu	Glu	Leu	Lys	Ser	
			405					410						415		
Leu	Ala	Arg	Ser	Phe	Arg	Phe	Asp	Tyr	Val	Arg	Ser					

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500						505						510					
Met	Asp	Arg	Ser	Asn	Val	Val	Leu	Asn	Asp	Asn	Leu	Thr	Phe	Trp	Ala		
		515					520					525					
Ser	Gly	Glu	Pro	Gly	Asn	Glu	Thr	Asn	Cys	Val	Tyr	Met	Asp	Ile	Gln		
	530					535					540						
Asp	Gln	Leu	Gln	Ser	Val	Trp	Lys	Thr	Lys	Ser	Cys	Phe	Gln	Pro	Ser		
545					550					555					560		
Ser	Phe	Ala	Cys	Met	Met	Asp	Leu	Ser	Asp	Arg	Asn	Lys	Ala	Lys	Cys		
				565					570					575			
Asp	Asp	Pro	Gly	Ser	Leu	Glu	Asn	Gly	His	Ala	Thr	Leu	His	Gly	Gln		
			580					585					590				
Ser	Ile	Asp	Gly	Phe	Tyr	Ala	Gly	Ser	Ser	Ile	Arg	Tyr	Ser	Cys	Glu		
	595						600					605					
Val	Leu	His	Tyr	Leu	Ser	Gly	Thr	Glu	Thr	Val	Thr	Cys	Thr	Thr	Asn		
	610					615					620						
Gly	Thr	Trp	Ser	Ala	Pro	Lys	Pro	Arg	Cys	Ile	Lys	Val	Ile	Thr	Cys		
625					630					635					640		
Gln	Asn	Pro	Pro	Val	Pro	Ser	Tyr	Gly	Ser	Val	Glu	Ile	Lys	Pro	Pro		
				645					650					655			
Ser	Arg	Thr	Asn	Ser	Ile	Ser	Arg	Val	Gly	Ser	Pro	Phe	Leu	Arg	Leu		
			660					665					670				
Pro	Arg	Leu	Pro	Leu	Pro	Leu	Ala	Arg	Ala	Ala	Lys	Pro	Pro	Pro	Lys		
		675					680					685					
Pro	Arg	Ser	Ser	Gln	Pro	Ser	Thr	Val	Asp	Leu	Ala	Ser	Lys	Val	Lys		
	690					695					700						
Leu	Pro	Glu	Gly	His	Tyr	Arg	Val	Gly	Ser	Arg	Ala	Ile	Tyr	Thr	Cys		
705					710					715					720		
Glu	Ser	Arg	Tyr	Tyr	Glu	Leu	Leu	Gly	Ser	Gln	Gly	Arg	Arg	Cys	Asp		
				725					730					735			
Ser	Asn	Gly	Asn	Trp	Ser	Gly	Arg	Pro	Ala	Ser	Cys	Ile	Pro	Val	Cys		
			740					745					750				
Gly	Arg	Ser	Asp	Ser	Pro	Arg	Ser	Pro	Phe	Ile	Trp	Asn	Gly	Asn	Ser		
	755						760					765					
Thr	Glu	Ile	Gly	Gln	Trp	Pro	Trp	Gln	Ala	Gly	Ile	Ser	Arg	Trp	Leu		
	770					775						780					
Ala	Asp	His	Asn	Met	Trp	Phe	Leu	Gln	Cys	Gly	Gly	Ser	Leu	Leu	Asn		
785					790					795					800		
Glu	Lys	Trp	Ile	Val	Thr	Ala	Ala	His	Cys	Val	Thr	Tyr	Ser	Ala	Thr		
			805							810				815			
Ala	Glu	Ile	Ile	Asp	Pro	Asn	Gln	Phe	Lys	Met	Tyr	Leu	Gly	Lys	Tyr		
			820					825					830				
Tyr	Arg	Asp	Asp	Ser	Arg	Asp	Asp	Asp	Tyr	Val	Gln	Val	Arg	Glu	Ala		
		835					840					845					
Leu	Glu	Ile	His	Val	Asn	Pro	Asn	Tyr	Asp	Pro	Gly	Asn	Leu	Asn	Phe		
	850					855					860						
Asp	Ile	Ala	Leu	Ile	Gln	Leu	Lys	Thr	Pro	Val	Thr	Leu	Thr	Thr	Arg		
865					870					875					880		
Val	Gln	Pro	Ile	Cys	Leu	Pro	Thr	Asp	Ile	Thr	Thr	Arg	Glu	His	Leu		
			885						890					895			
Lys	Glu	Gly	Thr	Leu	Ala	Val	Val	Thr	Gly	Trp	Gly	Leu	Asn	Glu	Asn		
			900					905					910				
Asn	Thr	Tyr	Ser	Glu	Thr	Ile	Gln	Gln	Ala	Val	Leu	Pro	Val	Val	Ala		
		915					920					925					

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Ala Ser Thr Cys Glu Glu Gly Tyr Lys Glu Ala Asp Leu Pro Leu Thr
930 935 940

Val Thr Glu Asn Met Phe Cys Ala Gly Tyr Lys Lys Gly Arg Tyr Asp
945 950 955 960

Ala Cys Ser Gly Asp Ser Gly Gly Pro Leu Val Phe Ala Asp Asp Ser
965 970 975

Arg Thr Glu Arg Arg Trp Val Leu Glu Gly Ile Val Ser Trp Gly Ser
980 985 990

Pro Ser Gly Cys Gly Lys Ala Asn Gln Tyr Gly Gly Phe Thr Lys Val
995 1000 1005

Asn Val Phe Leu Ser Trp Ile Arg Gln Phe Ile
1010 1015

<210> SEQ ID NO 5
<211> LENGTH: 1203
<212> TYPE: DNA
<213> ORGANISM: Tachypleus tridentatus
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1) .. (1203)

<400> SEQUENCE: 5

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Met Thr Trp Ile Cys Val Ile Thr Leu Phe Ala Leu Ala Ser Ala Thr
1 5 10 15

ttg ggt aac aaa gtt agt aga gtg ggg gtc ctc ttc ccc aag aca cgg 96
Leu Gly Asn Lys Val Ser Arg Val Gly Val Leu Phe Pro Lys Thr Arg
20 25 30

aac gac aat gag tgt aca gca aga ggg gga ttg aaa gga tcc tgc aaa 144
Asn Asp Asn Glu Cys Thr Ala Arg Gly Gly Leu Lys Gly Ser Cys Lys
35 40 45

tcc ctc ata gac tgt cct agt gtc ttg gct acg ttg aag gac agt ttt 192
Ser Leu Ile Asp Cys Pro Ser Val Leu Ala Thr Leu Lys Asp Ser Phe
50 55 60

cct gtc gtt tgc tct tgg aat ggt cga ttt cag cct att gtc tgc tgt 240
Pro Val Val Cys Ser Trp Asn Gly Arg Phe Gln Pro Ile Val Cys Cys
65 70 75 80

cct gat gca ata gca cca cca cct gta acc aca aca gct gta act gta 288
Pro Asp Ala Ile Ala Pro Pro Pro Val Thr Thr Ala Val Thr Val
85 90 95

ata tct aca aaa gaa cca aag ctt cca aga tta cat ata tca ggt tgt 336
Ile Ser Thr Lys Glu Pro Lys Leu Pro Arg Leu His Ile Ser Gly Cys
100 105 110

gga aaa aga aaa gtc aaa ata gat att aca act gtt gga cgc tct gga 384
Gly Lys Arg Lys Val Lys Ile Asp Ile Thr Thr Val Gly Arg Ser Gly
115 120 125

tca cca ata ctt cct ccg ata tct act cct caa aat tca aca ggt ggg 432
Ser Pro Ile Leu Pro Pro Ile Ser Thr Pro Gln Asn Ser Thr Gly Gly
130 135 140

aga gga att att gct gga ggc gta gaa gcc aaa att ggc gcg tgg cct 480
Arg Gly Ile Ile Ala Gly Gly Val Glu Ala Lys Ile Gly Ala Trp Pro
145 150 155 160

tgg atg gca gct gtt ttt gtg aaa aac ttt ggc att ggc aga ttc cac 528
Trp Met Ala Ala Val Phe Val Lys Asn Phe Gly Ile Gly Arg Phe His
165 170 175

tgt gct ggt agc ata atc agt aac aag tac att ttg tca gct gcc cac 576
Cys Ala Gly Ser Ile Ile Ser Asn Lys Tyr Ile Leu Ser Ala Ala His
180 185 190

gcc ttc ctt atc gga ggt cga aag ttg acc cca act cgc tta gct gtc 624

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<210> SEQ ID NO 6
<211> LENGTH: 400
<212> TYPE: PRT
<213> ORGANISM: Tachyplesus tridentatus

<400> SEQUENCE: 6
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Met	Thr	Trp	Ile	Cys	Val	Ile	Thr	Leu	Phe	Ala	Leu	Ala	Ser	Ala	Thr
1				5					10					15	
Leu	Gly	Asn	Lys	Val	Ser	Arg	Val	Gly	Val	Leu	Phe	Pro	Lys	Thr	Arg
			20					25					30		
Asn	Asp	Asn	Glu	Cys	Thr	Ala	Arg	Gly	Gly	Leu	Lys	Gly	Ser	Cys	Lys
		35					40					45			
Ser	Leu	Ile	Asp	Cys	Pro	Ser	Val	Leu	Ala	Thr		Leu	Lys	Asp	Ser
	50					55						60			Phe
Pro	Val	Val	Cys	Ser	Trp	Asn	Gly	Arg	Phe	Gln	Pro	Ile	Val	Cys	Cys
65					70					75				80	
Pro	Asp	Ala	Ile	Ala	Pro	Pro	Pro	Val	Thr	Thr	Thr	Ala	Val	Thr	Val
			85						90					95	

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Ile Ser Thr Lys Glu Pro Lys Leu Pro Arg Leu His Ile Ser Gly Cys
      100                      105                      110

Gly Lys Arg Lys Val Lys Ile Asp Ile Thr Thr Val Gly Arg Ser Gly
      115                      120                      125

Ser Pro Ile Leu Pro Pro Ile Ser Thr Pro Gln Asn Ser Thr Gly Gly
      130                      135                      140

Arg Gly Ile Ile Ala Gly Gly Val Glu Ala Lys Ile Gly Ala Trp Pro
      145                      150                      155                      160

Trp Met Ala Ala Val Phe Val Lys Asn Phe Gly Ile Gly Arg Phe His
      165                      170                      175

Cys Ala Gly Ser Ile Ile Ser Asn Lys Tyr Ile Leu Ser Ala Ala His
      180                      185                      190

Ala Phe Leu Ile Gly Gly Arg Lys Leu Thr Pro Thr Arg Leu Ala Val
      195                      200                      205

Arg Val Gly Gly His Tyr Ile Lys Arg Gly Gln Glu Tyr Pro Val Lys
      210                      215                      220

Asp Val Ile Ile His Pro His Tyr Val Glu Lys Glu Asn Tyr Asn Asp
      225                      230                      235                      240

Ile Ala Ile Ile Glu Leu Lys Glu Glu Leu Asn Phe Thr Asp Leu Val
      245                      250                      255

Asn Pro Ile Cys Leu Pro Asp Pro Glu Thr Val Thr Asp Pro Leu Lys
      260                      265                      270

Asp Arg Ile Val Thr Ala Ala Gly Trp Gly Asp Leu Asp Phe Ser Gly
      275                      280                      285

Pro Arg Ser Gln Val Leu Arg Glu Val Ser Ile Pro Val Val Pro Val
      290                      295                      300

Asp Lys Cys Asp Gln Ala Tyr Glu Lys Leu Asn Thr Pro Ser Leu Lys
      305                      310                      315                      320

Asn Gly Ile Thr Asn Asn Phe Leu Cys Ala Gly Leu Glu Gly Gly
      325                      330                      335

Lys Asp Ala Cys Gln Gly Asp Ser Gly Gly Pro Leu Met Leu Val Asn
      340                      345                      350

Asn Thr Arg Trp Ile Val Val Gly Val Val Ser Phe Gly His Lys Cys
      355                      360                      365

Ala Glu Glu Gly Tyr Pro Gly Val Tyr Ser Arg Val Ala Ser Tyr Leu
      370                      375                      380

Asp Trp Ile Ala Lys Val Thr Asn Ser Leu Asp His Ala Val Thr Asn
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<210> SEQ ID NO 7
<211> LENGTH: 1128
<212> TYPE: DNA
<213> ORGANISM: Tachypleus tridentatus
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)..(1128)

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<400> SEQUENCE: 7

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1          5          10          15

tct gtg gtt aga tgc agt act ctc agc aga cag cgt aga cag ttt gtt      96
Ser Val Val Arg Cys Ser Thr Leu Ser Arg Gln Arg Arg Gln Phe Val
20         25         30

ttc cct gac gag gaa gaa ctt tgc tca aac cga ttt act gaa gaa gga      144
Phe Pro Asp Glu Glu Glu Leu Cys Ser Asn Arg Phe Thr Glu Glu Gly
35         40         45

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aca tgc aaa aat gtc ttg gat tgt aga ata ctt tta caa aaa aat gat Thr Cys Lys Asn Val Leu Asp Cys Arg Ile Leu Leu Gln Lys Asn Asp 50 55 60	192
tat aat tta ctc aaa gaa tca ata tgc ggc ttt gaa ggc ata aca ccc Tyr Asn Leu Leu Lys Glu Ser Ile Cys Gly Phe Glu Gly Ile Thr Pro 65 70 75 80	240
aaa gtt tgt tgt ccg aaa tca agc cat gta att tca agt aca cag gca Lys Val Cys Cys Pro Lys Ser Ser His Val Ile Ser Ser Thr Gln Ala 85 90 95	288
cct cca gaa acc act acg act gaa cgc cca cca aaa cag ata cca ccc Pro Pro Glu Thr Thr Thr Thr Glu Arg Pro Pro Lys Gln Ile Pro Pro 100 105 110	336
aat ctt cat gaa gtg tgt gga att cac aat act aca act acc agg att Asn Leu His Glu Val Cys Gly Ile His Asn Thr Thr Thr Arg Ile 115 120 125	384
att gga ggt cgg gaa gca cct att gga gcc tgg ccg tgg atg act gct Ile Gly Gly Arg Glu Ala Pro Ile Gly Ala Trp Pro Trp Met Thr Ala 130 135 140	432
gtc tac ata aaa caa gga gga atc aga agt gtt cag tgt ggt ggc gca Val Tyr Ile Lys Gln Gly Gly Ile Arg Ser Val Gln Cys Gly Gly Ala 145 150 155 160	480
ctt gtc act aac agg cac gtg att aca gct tcg cac tgt gtt gta aac Leu Val Thr Asn Arg His Val Ile Thr Ala Ser His Cys Val Val Asn 165 170 175	528
agt gca gga aca gat gtg atg cca gct gat gta ttc tcg gtt cgt ctg Ser Ala Gly Thr Asp Val Met Pro Ala Asp Val Phe Ser Val Arg Leu 180 185 190	576
ggg gaa cac aat tta tac agt acc gat gac gat tcg aat cca ata gat Gly Glu His Asn Leu Tyr Ser Thr Asp Asp Asp Ser Asn Pro Ile Asp 195 200 205	624
ttt gca gtt acg tcg gtg aaa cat cac gaa cac ttt gta ctc gcg acg Phe Ala Val Thr Ser Val Lys His His Glu His Phe Val Leu Ala Thr 210 215 220	672
tat ttg aat gac atc gca att cta acg tta aat gac aca gtt acg ttt Tyr Leu Asn Asp Ile Ala Ile Leu Thr Leu Asn Asp Thr Val Thr Phe 225 230 235 240	720
aca gac aga att cga ccc att tgt cta cct tat cgt aag ttg aga tac Thr Asp Arg Ile Arg Pro Ile Cys Leu Pro Tyr Arg Lys Leu Arg Tyr 245 250 255	768
gat gat cta gca atg aga aaa ccg ttt atc act gga tgg gga aca aca Asp Asp Leu Ala Met Arg Lys Pro Phe Ile Thr Gly Trp Gly Thr Thr 260 265 270	816
gca ttt aac ggc cca tct agt gca gtg ttg aga gaa gta cag tta cca Ala Phe Asn Gly Pro Ser Ser Ala Val Leu Arg Glu Val Gln Leu Pro 275 280 285	864
ata tgg gaa cac gag gcc tgt aga cag gcc tac gag aag gat tta aat Ile Trp Glu His Glu Ala Cys Arg Gln Ala Tyr Glu Lys Asp Leu Asn 290 295 300	912
att aca aac gtg tat atg tgt gct ggc ttt gca gat ggc ggg aag gat Ile Thr Asn Val Tyr Met Cys Ala Gly Phe Ala Asp Gly Gly Lys Asp 305 310 315 320	960
gct tgc cag ggt gat tct gga ggt cca atg atg ttg cct gtt aaa acc Ala Cys Gln Gly Asp Ser Gly Gly Pro Met Met Leu Pro Val Lys Thr 325 330 335	1008
gga gag ttt tat ctc att gga att gtg tct ttc gga aag aaa tgc gca Gly Glu Phe Tyr Leu Ile Gly Ile Val Ser Phe Gly Lys Lys Cys Ala 340 345 350	1056
ttg cct gga ttt cct ggg gtt tac aca aaa gtg aca gag ttt tta gat Leu Pro Gly Phe Pro Gly Val Tyr Thr Lys Val Thr Glu Phe Leu Asp	1104

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355	360	365	
tgg att gca gaa cat atg gtg tag			1128
Trp Ile Ala Glu His Met Val			
370	375		

<210> SEQ ID NO 8
 <211> LENGTH: 375
 <212> TYPE: PRT
 <213> ORGANISM: Tachypleus tridentatus
 <400> SEQUENCE: 8

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Ser Val Val Arg Cys Ser Thr Leu Ser Arg Gln Arg Arg Gln Phe Val	
20 25 30	
Phe Pro Asp Glu Glu Glu Leu Cys Ser Asn Arg Phe Thr Glu Glu Gly	
35 40 45	
Thr Cys Lys Asn Val Leu Asp Cys Arg Ile Leu Leu Gln Lys Asn Asp	
50 55 60	
Tyr Asn Leu Leu Lys Glu Ser Ile Cys Gly Phe Glu Gly Ile Thr Pro	
65 70 75 80	
Lys Val Cys Cys Pro Lys Ser Ser His Val Ile Ser Ser Thr Gln Ala	
85 90 95	
Pro Pro Glu Thr Thr Thr Thr Glu Arg Pro Pro Lys Gln Ile Pro Pro	
100 105 110	
Asn Leu His Glu Val Cys Gly Ile His Asn Thr Thr Thr Thr Arg Ile	
115 120 125	
Ile Gly Gly Arg Glu Ala Pro Ile Gly Ala Trp Pro Trp Met Thr Ala	
130 135 140	
Val Tyr Ile Lys Gln Gly Gly Ile Arg Ser Val Gln Cys Gly Gly Ala	
145 150 155 160	
Leu Val Thr Asn Arg His Val Ile Thr Ala Ser His Cys Val Val Asn	
165 170 175	
Ser Ala Gly Thr Asp Val Met Pro Ala Asp Val Phe Ser Val Arg Leu	
180 185 190	
Gly Glu His Asn Leu Tyr Ser Thr Asp Asp Asp Ser Asn Pro Ile Asp	
195 200 205	
Phe Ala Val Thr Ser Val Lys His His Glu His Phe Val Leu Ala Thr	
210 215 220	
Tyr Leu Asn Asp Ile Ala Ile Leu Thr Leu Asn Asp Thr Val Thr Phe	
225 230 235 240	
Thr Asp Arg Ile Arg Pro Ile Cys Leu Pro Tyr Arg Lys Leu Arg Tyr	
245 250 255	
Asp Asp Leu Ala Met Arg Lys Pro Phe Ile Thr Gly Trp Gly Thr Thr	
260 265 270	
Ala Phe Asn Gly Pro Ser Ser Ala Val Leu Arg Glu Val Gln Leu Pro	
275 280 285	
Ile Trp Glu His Glu Ala Cys Arg Gln Ala Tyr Glu Lys Asp Leu Asn	
290 295 300	
Ile Thr Asn Val Tyr Met Cys Ala Gly Phe Ala Asp Gly Gly Lys Asp	
305 310 315 320	
Ala Cys Gln Gly Asp Ser Gly Gly Pro Met Met Leu Pro Val Lys Thr	
325 330 335	
Gly Glu Phe Tyr Leu Ile Gly Ile Val Ser Phe Gly Lys Lys Cys Ala	
340 345 350	

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Leu Pro Gly Phe Pro Gly Val Tyr Thr Lys Val Thr Glu Phe Leu Asp
355 360 365

Trp Ile Ala Glu His Met Val
370 375

<210> SEQ ID NO 9
<211> LENGTH: 1203
<212> TYPE: DNA
<213> ORGANISM: Tachypleus tridentatus

<400> SEQUENCE: 9

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gtgtcccggtg tgggtgtcct gttccccaag acccgtaacg acaacgagtg caccgctcgt 120
gggtggtctga agggctcctg caagtccctg atcgactgcc cctccgtgct ggctaccctg 180
aaggactcct tccccgtcgt gtgctcctgg aacgggtcgtt tccagcccat cgtgtgctgc 240
cccgcgcta tcgtccccc cctgtgacc accaccgctg tgaccgtgat ctccaccaag 300
gagcccaagc tgccccgtct gcacatctcc ggttgctggca agcgcaaggt caagatcgac 360
atcaccacgg tgggcccgttc cggttccccc atcctgcccc ccatctccac cccccagaac 420
tccactgggtg gtctgtggtat catcgctggc ggtgtcgagg ctaagatcgg tgettggccc 480
tggatggctg ctgtgttcgt gaagaacttc ggtatcggtc gttccactg cgtggttcc 540
atcatctcca acaagtacat cctgtccgct gctcacgctt tctcatcgg tggtcgcaag 600
ctgaccccc cccgtctggc tgtgcgtgtg ggtgggtcact acatcaagcg tggccaggag 660
taccocgtca aggacgtgat catccacccc cactacgtgg agaaggagaa ctacaacgac 720
atcgccatca tcgagctgaa ggaggagctg aacttcacgg acctgggtcaa ccccatctgc 780
ctgcccgaac ccgagactgt gaccgaccc ctgaaggacc gtatcgtgac cgctgctggc 840
tggggcgacc tggacttctc cggtecccggt tccaggtgc tgcgtgaggt gtccatcccc 900
gtggtgcccc tggacaagtg cgaccaggct tacgagaagc tgaacacccc ctccctgaag 960
aacggtatta ccaacaactt cctctcgccc ggactcgagg aggggtggaa ggacgcttgc 1020
cagggcgact ccggtggtcc cctgatgctg gtcaacaaca cccgttggtat cgtcgtgggt 1080
gtcgtgtcct tcggtcacaa gtgcgctgag gagggttacc ccggcgtcta ctcccggtg 1140
gttctctacc tggactggat cgctaaggtc accaactccc tggaccacgc tgtcaccaac 1200
taa 1203

<210> SEQ ID NO 10
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic peptide

<400> SEQUENCE: 10

Ile Glu Gly Arg
1

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The invention claimed is:

1. A method for imparting a salt ion resistance for a horseshoe crab Factor C, the method comprising:

recombinantly expressing and secreting the horseshoe crab Factor C into a culture broth or a culture supernatant using a mammalian cell as a host cell, and recovering the recombinant horseshoe crab Factor C secreted from the mammalian cell,

wherein the horseshoe crab Factor C recombinantly expressed and secreted using the mammalian cell contains (α -2,3)-linked terminal sialic acid in a greater amount, as compared with a corresponding horseshoe crab Factor C expressed using a Sf9 insect cell as a host cell,

wherein the mammalian cell is a Chinese hamster ovary (CHO) cell or a human embryonic kidney (HEK) cell, and

wherein the horseshoe crab Factor C recombinantly expressed and secreted into a culture broth or a culture supernatant using the mammalian cell is a protein selected from the following (A), (B), (C), or (D):

(A) a protein comprising the amino acid sequence of Factor C of *Tachypleus tridentatus*, wherein the protein has no peptide added to the N-terminus of Factor C of *Tachypleus tridentatus*, and wherein the protein has activity of Factor C;

(B) a protein comprising the amino acid sequence of Factor C of *Limulus polyphemus*, wherein the protein has no peptide added to the N-terminus of Factor C of *Limulus polyphemus*, and wherein the protein has activity of Factor C;

(C) a protein comprising the amino acid sequence of Factor C of *Carcinoscorpius rotundicauda*, wherein the protein has no peptide added to the N-terminus of Factor C of *Carcinoscorpius rotundicauda*, and wherein the protein has activity of Factor C;

(D) a protein comprising the amino acid sequence of Factor C of *Tachypleus gigas*, wherein the protein has no peptide added to the N-terminus of Factor C of *Tachypleus gigas*, and wherein the protein has activity of Factor C.

2. The method according to claim 1, wherein the salt ion is selected from the group consisting of sodium ion, potassium ion, calcium ion, and magnesium ion.

3. The method according to claim 1, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in the presence of 21 mM sodium citrate as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

4. The method according to claim 1, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in the presence of 52 mM sodium hydrogencarbonate as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

5. The method according to claim 1, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in the presence of 214 mM sodium chloride as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

6. The method according to claim 1, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in

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the presence of 16 mM magnesium sulfate as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

7. The method according to claim 1, wherein the protein comprises the amino acid sequence of Factor C of *Tachypleus tridentatus*, *Limulus polyphemus*, or *Carcinoscorpius rotundicauda*.

8. A method for producing an endotoxin assay agent comprising a horseshoe crab Factor C, the method comprising:

recombinantly expressing and secreting the horseshoe crab Factor C into a culture broth or a culture supernatant using a mammalian cell as a host cell,

recovering the recombinant horseshoe crab Factor C secreted from the mammalian cell, and

producing an endotoxin assay agent comprising the recombinant horseshoe crab Factor C,

wherein the horseshoe crab Factor C recombinantly expressed and secreted using the mammalian cell contains (α -2,3)-linked terminal sialic acid in a greater amount, as compared with a corresponding horseshoe crab Factor C expressed using a Sf9 insect cell as a host cell,

wherein the mammalian cell is a Chinese hamster ovary (CHO) cell or a human embryonic kidney (HEK) cell, and

wherein the horseshoe crab Factor C recombinantly expressed and secreted into a culture broth or a culture supernatant using the mammalian cell is a protein selected from the following (A), (B), (C), or (D):

(A) a protein comprising the amino acid sequence of Factor C of *Tachypleus tridentatus*, wherein the protein has no peptide added to the N-terminus of Factor C of *Tachypleus tridentatus*, and wherein the protein has activity of Factor C;

(B) a protein comprising the amino acid sequence of Factor C of *Limulus polyphemus*, wherein the protein has no peptide added to the N-terminus of Factor C of *Limulus polyphemus*, and wherein the protein has activity of Factor C;

(C) a protein comprising the amino acid sequence of Factor C of *Carcinoscorpius rotundicauda*, wherein the protein has no peptide added to the N-terminus of Factor C of *Carcinoscorpius rotundicauda*, and wherein the protein has activity of Factor C;

(D) a protein comprising the amino acid sequence of Factor C of *Tachypleus gigas*, wherein the protein has no peptide added to the N-terminus of Factor C of *Tachypleus gigas*, and wherein the protein has activity of Factor C.

9. The method according to claim 8, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in the presence of 21 mM sodium citrate as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

10. The method according to claim 8, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in the presence of 52 mM sodium hydrogencarbonate as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

11. The method according to claim 8, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in

the presence of 214 mM sodium chloride as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

12. The method according to claim 8, wherein the horseshoe crab Factor C recombinantly expressed and secreted using a mammalian cell exhibits a higher residual activity in the presence of 16 mM magnesium sulfate as compared with a corresponding horseshoe crab Factor C expressed in a Sf9 insect cell as a host cell.

13. The method according to claim 8, wherein the protein comprises the amino acid sequence of Factor C of *Tachyplesus tridentatus*, *Limulus polyphemus*, or *Carcinoscorpius rotundicauda*.

* * * * *

UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 10,982,202 B2
APPLICATION NO. : 15/983725
DATED : April 20, 2021
INVENTOR(S) : Hikaru Mizumura, Toshio Oda and Shun-ichiro Kawabata

Page 1 of 1

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

On the Title Page

(56) References Cited

Page 2, Under OTHER PUBLICATIONS, left column, Line 7, delete “Retrived” and insert --Retrieved-- therefor.

Page 2, Under OTHER PUBLICATIONS, left column, Line 8, delete “19,” and insert --29,-- therefor.

Page 2, Under OTHER PUBLICATIONS, left column, Line 28, delete “iin” and insert --in-- therefor.

In the Specification

Column 1, Line 9, please delete “Jun. 9, 2015” and insert --Jun. 9, 2015,-- therefor.

Column 13, Line 60, please delete “N acetylneuraminic” and insert --N-acetylneuraminic-- therefor.

Column 29, Line 51, please delete “Celfectin” and insert --Cellfectin-- therefor.

In the Claims

Claim 7, Column 76, Line 6, please delete “*Limuluspolyphemus*,” and insert --*Limulus polyphemus*,-- therefor.

Claim 13, Column 77, Line 12, please delete “*Limuluspolyphemus*,” and insert --*Limulus polyphemus*, - therefor.

Signed and Sealed this
Twenty-second Day of June, 2021



Drew Hirshfeld
Performing the Functions and Duties of the
Under Secretary of Commerce for Intellectual Property and
Director of the United States Patent and Trademark Office